

HIGHLIGHTS OF PRESCRIBING INFORMATION

These highlights do not include all the information needed to use HPC, Cord Blood safely and effectively. See full prescribing information for HPC, Cord Blood.

HPC, Cord Blood

Injectable Suspension for Intravenous Use

Initial U.S. Approval: XXXX

WARNING: FATAL INFUSION REACTIONS, GRAFT VERSUS HOST DISEASE, ENGRAFTMENT SYNDROME, AND GRAFT FAILURE

See full prescribing information for complete boxed warning.

- **Fatal infusion reactions: Monitor patients during infusion and discontinue for severe reactions. Use is contraindicated in patients with known allergy to dimethyl sulfoxide (DMSO), Dextran 40, hydroxyethylstarch, or human serum albumin (4, 5.1, 5.2).**
- **Graft-vs.-host disease (GVHD): GVHD may be fatal. Administration of immunosuppressive therapy may decrease the risk of GVHD (5.3).**
- **Engraftment syndrome: Engraftment syndrome may be fatal. Treat engraftment syndrome promptly with corticosteroids (5.4).**
- **Graft failure: Graft failure may be fatal. Monitor patients for laboratory evidence of hematopoietic recovery (5.5).**

INDICATIONS AND USAGE

HPC, Cord Blood is an allogeneic cord blood hematopoietic progenitor cell therapy indicated for use in unrelated donor hematopoietic progenitor cell transplantation procedures in conjunction with an appropriate preparative regimen for hematopoietic and immunologic reconstitution in patients with disorders affecting the hematopoietic system that are inherited, acquired, or result from myeloablative treatment (1).

The risk benefit assessment for an individual patient depends on the patient characteristics, including disease, stage, risk factors, and specific manifestations of the disease, on characteristics of the graft, and on other available treatments or types of hematopoietic progenitor cells (1).

DOSAGE AND ADMINISTRATION

- For intravenous use only
- Do not irradiate

- Unit selection and administration of HPC, Cord Blood should be done under the direction of a physician experienced in hematopoietic progenitor cell transplantation (2).
- The recommended minimum dose is 2.5×10^7 nucleated cells/kg at cryopreservation (2.1).
- Do not administer HPC, Cord Blood through the same tubing with other products except for normal saline (2.3).

DOSAGE FORMS AND STRENGTHS

Each unit contains a minimum of 5×10^8 total nucleated cells with at least 1.25×10^6 viable CD34+ cells at the time of cryopreservation. The exact pre-cryopreservation nucleated cell content of each unit is provided on the accompanying records (3).

CONTRAINDICATIONS

Known sensitivity to dimethyl sulfoxide (DMSO), Dextran 40, hydroxyethylstarch, or plasma proteins (4).

WARNINGS AND PRECAUTIONS

- Hypersensitivity Reactions (5.1)
- Infusion Reactions (5.2)
- Graft-versus-Host Disease (5.3)
- Engraftment Syndrome (5.4)
- Graft Failure (5.5)
- Malignancies of Donor Origin (5.6)
- Transmission of Serious Infections (5.7)
- Transmission of Rare Genetic Diseases (5.8)

ADVERSE REACTIONS

Mortality, from all causes, at 100 days post-transplant was 25% (5, 6.1).

The most common infusion-related adverse reactions ($\geq 5\%$) are hypertension, vomiting, nausea, bradycardia, and fever (6.1).

To report SUSPECTED ADVERSE REACTIONS, contact the quality department for LifeSouth Community Blood Centers at 1-888-795-2707 and FDA at 1-800-FDA-1088 or www.fda.gov/medwatch.

USE IN SPECIFIC POPULATIONS

Pregnancy: No animal or human data. Use only if clearly needed (8.1).

See 17 for PATIENT COUNSELING INFORMATION

Revised: XX/XXXX

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2 **FULL PRESCRIBING INFORMATION**
3

4 **WARNING: FATAL INFUSION REACTIONS, GRAFT VERSUS HOST DISEASE,**
5 **ENGRAFTMENT SYNDROME AND GRAFT FAILURE**
6

7 **Fatal infusion reactions:** HPC, Cord Blood administration can result in serious, including
8 fatal, infusion reactions. Monitor patients and discontinue HPC, Cord Blood infusion for
9 severe reactions. Use is contraindicated in patients with known allergy to dimethyl
10 sulfoxide (DMSO), Dextran 40, hydroxyethylstarch, or human serum albumin. [See
11 *Contraindications (4) and Warnings and Precautions (5.1, 5.2)*].
12

13 **Graft-vs.-host disease (GVHD):** GVHD is expected after administration of HPC, Cord
14 Blood, and may be fatal. Administration of immunosuppressive therapy may decrease the
15 risk of GVHD. [See *Warnings and Precautions (5.3)*].
16

17 **Engraftment syndrome:** Engraftment syndrome may progress to multiorgan failure and
18 death. Treat engraftment syndrome promptly with corticosteroids. [See *Warnings and*
19 *Precautions (5.4)*].
20

21 **Graft failure:** Graft failure may be fatal. Monitor patients for laboratory evidence of
22 hematopoietic recovery. Prior to choosing a specific unit of HPC, Cord Blood, consider
23 testing for HLA antibodies to identify patients who are alloimmunized. [See *Warnings and*
24 *Precautions (5.5)*].
25

26 **1 INDICATIONS AND USAGE**
27

28 HPC (Hematopoietic Progenitor Cell), Cord Blood is an allogeneic cord blood hematopoietic
29 progenitor cell therapy indicated for use in unrelated donor hematopoietic progenitor stem cell
30 transplantation procedures in conjunction with an appropriate preparative regimen for
31 hematopoietic and immunologic reconstitution in patients with disorders affecting the
32 hematopoietic system that are inherited, acquired, or result from myeloablative treatment.
33

34 The risk benefit assessment for an individual patient depends on the patient characteristics,
35 including disease, stage, risk factors, and specific manifestations of the disease, on characteristics
36 of the graft, and on other available treatments or types of hematopoietic progenitor cells.
37

38 **2 DOSAGE AND ADMINISTRATION**
39

- 40 • For intravenous use only.
 - 41 • Do not irradiate.
- 42

43 Unit selection and administration of HPC, Cord Blood should be done under the direction of a
44 physician experienced in hematopoietic progenitor cell transplantation.
45

46 **2.1 Dosing**
47

48 The recommended minimum dose is 2.5×10^7 nucleated cells/kg at cryopreservation. Multiple
49 units may be required in order to achieve the appropriate dose.
50

51 Matching for at least 4 of 6 HLA-A antigens, HLA-B antigens, and HLA-DRB1 alleles is
52 recommended. The HLA typing and nucleated cell content for each individual unit of HPC,
53 Cord Blood are documented in the accompanying records.

54 55 **2.2 Preparation for Infusion**

56
57 HPC, Cord Blood should be prepared by a trained healthcare professional.

- 58
- 59 • Do not irradiate HPC, Cord Blood.
- 60 • See the appended detailed instructions for preparation of HPC, Cord Blood for infusion.
- 61 • Once prepared for infusion, HPC, Cord Blood may be stored at room temperature (19-25°C)
62 or 4°C for up to 2 hours when DMSO is removed in a washing procedure [*see Instructions*
63 *for Preparation for Infusion*]. No data are available for the stability of HPC, Cord Blood if
64 DMSO is not removed.
- 65 • The recommended limit on DMSO administration is 1 gram per kg body weight per day [*see*
66 *Warnings and Precautions (5.2) and Overdosage (10)*].

67 68 **2.3 Administration**

69
70 HPC, Cord Blood should be administered under the supervision of a qualified healthcare
71 professional experienced in hematopoietic progenitor cell transplantation.

- 72
- 73 1. Confirm the identity of the patient for the specified unit of HPC, Cord Blood prior to
74 administration.
- 75 2. Confirm that emergency medications are available for use in the immediate area.
- 76 3. Ensure the patient is hydrated adequately.
- 77 4. Premedicate the patient 30 to 60 minutes before the administration of HPC, Cord
78 Blood. Premedication can include any or all of the following: antipyretics, histamine
79 antagonists, and corticosteroids.
- 80 5. Inspect the product for any abnormalities such as unusual particulates and for breaches
81 of container integrity prior to administration. Prior to infusion, discuss all such product
82 irregularities with the laboratory issuing the product for infusion.
- 83 6. Administer HPC, Cord Blood by intravenous infusion. Do not administer in the same
84 tubing concurrently with products other than 0.9% Sodium Chloride, Injection (USP).
85 HPC, Cord Blood may be filtered through a 170 to 260 micron filter designed to
86 remove clots. Do NOT use a filter designed to remove leukocytes.
- 87 7. For adults, begin infusion of HPC, Cord Blood at 100 milliliters per hour and increase
88 the rate as tolerated. For children, begin infusion of HPC, Cord Blood at 1 milliliter per
89 kg per hour and increase as tolerated. Reduce the infusion rate if the fluid load is not
90 tolerated. Discontinue the infusion in the event of an allergic reaction or if the patient
91 develops a moderate to severe infusion reaction [*see Warnings and Precautions (5.2)*
92 *and Adverse Reactions (6)*].
- 93 8. Monitor the patient for adverse reactions during, and for at least six hours after,
94 administration. Because HPC, Cord Blood contains lysed red cells that may cause renal
95 failure, careful monitoring of urine output is also recommended.

96
97 **NOTE:** If product is being prepared for a multi-unit infusion, infuse units independently.
98 Should a reaction occur, appropriately manage the reaction before the second unit is thawed for
99 infusion.

100

101 **3 DOSAGE FORMS AND STRENGTHS**

102

103 Each unit of HPC, Cord Blood contains a minimum of 5×10^8 total nucleated cells with a
104 minimum of 1.25×10^6 viable CD34+ cells, suspended in 10% dimethyl sulfoxide (DMSO) and
105 1% Dextran 40, at the time of cryopreservation.

106

107 The exact pre-cryopreservation nucleated cell content is provided in the accompanying records.

108

109 **4 CONTRAINDICATIONS**

110

111 HPC, Cord Blood is contraindicated in patients with known hypersensitivity to dimethyl
112 sulfoxide (DMSO), Dextran 40, hydroxyethylstarch, or plasma proteins [*see Description (11)*
113 *and Dosage and Administration (2.2)*].

114

115 **5 WARNINGS AND PRECAUTIONS**

116

117 **5.1 Hypersensitivity Reactions**

118

119 Allergic reactions may occur with infusion of HPC, Cord Blood. Reactions include
120 bronchospasm, wheezing, angioedema, pruritus and hives [*see Adverse Reactions (6)*]. Serious
121 hypersensitivity reactions, including anaphylaxis, also have been reported. These reactions may
122 be due to dimethyl sulfoxide (DMSO), Dextran 40, hydroxyethylstarch, or a plasma component
123 of HPC, Cord Blood.

124

125 **5.2 Infusion Reactions**

126

127 Infusion reactions are expected to occur and include nausea, vomiting, fever, rigors or chills,
128 flushing, dyspnea, hypoxemia, chest tightness, hypertension, tachycardia, bradycardia,
129 dysgeusia, hematuria, and mild headache. Premedication with antipyretics, histamine
130 antagonists, and corticosteroids may reduce the incidence and intensity of infusion reactions.

131

132 Severe reactions, including respiratory distress, severe bronchospasm, severe bradycardia with
133 heart block or other arrhythmias, cardiac arrest, hypotension, hemolysis, elevated liver enzymes,
134 renal compromise, encephalopathy, loss of consciousness, and seizure also may occur. Many of
135 these reactions are related to the amount of DMSO administered. Minimizing the amount of
136 DMSO administered may reduce the risk of such reactions, although idiosyncratic responses may
137 occur even at DMSO doses thought to be tolerated. The actual amount of DMSO depends on the
138 method of preparation of the product for infusion. Limiting the amount of DMSO infused to no
139 more than 1 gram per kilogram per day is recommended [*see Overdosage (10)*].

140

141 Infusion reactions may begin within minutes of the start of infusion of HPC, Cord Blood,
142 although symptoms may continue to intensify and not peak for several hours after completion of
143 the infusion. Monitor the patient closely during this period. When a reaction occurs, discontinue
144 the infusion and institute supportive care as needed.

145

146 If infusing more than one unit of HPC, Cord Blood on the same day, do not administer
147 subsequent units until all signs and symptoms of infusion reactions from the prior unit have
148 resolved.

149

150 **5.3 Graft-versus-Host Disease**

151
152 Acute and chronic graft-versus-host disease (GVHD) may occur in patients who have received
153 HPC, Cord Blood. Classic acute GVHD is manifested as fever, rash, elevated bilirubin and liver
154 enzymes, and diarrhea. Patients transplanted with HPC, Cord Blood also should receive
155 immunosuppressive drugs to decrease the risk of GVHD. [See *Adverse Reactions (6.1).*]

157 **5.4 Engraftment Syndrome**

158
159 Engraftment syndrome is manifested as unexplained fever and rash in the peri-engraftment
160 period. Patients with engraftment syndrome also may have unexplained weight gain,
161 hypoxemia, and pulmonary infiltrates in the absence of fluid overload or cardiac disease. If
162 untreated, engraftment syndrome may progress to multiorgan failure and death. Once
163 engraftment syndrome is recognized, begin treatment with corticosteroids to ameliorate the
164 symptoms. [See *Adverse Reactions (6.1).*]

166 **5.5 Graft Failure**

167
168 Primary graft failure, which may be fatal, is defined as failure to achieve an absolute neutrophil
169 count greater than 500 per microliter blood by Day 42 after transplantation. Immunologic
170 rejection is the primary cause of graft failure. Patients should be monitored for laboratory
171 evidence of hematopoietic recovery. Consider testing for HLA antibodies in order to identify
172 patients who are alloimmunized prior to transplantation and to assist with choosing a unit with a
173 suitable HLA type for the individual patient. [See *Adverse Reactions (6.1).*]

175 **5.6 Malignancies of Donor Origin**

176
177 Patients who have undergone HPC, Cord Blood transplantation may develop post-transplant
178 lymphoproliferative disorder (PTLD), manifested as a lymphoma-like disease favoring non-
179 nodal sites. PTLD is usually fatal if not treated.

180
181 The incidence of PTLD appears to be higher in patients who have received antithymocyte
182 globulin. The etiology is thought to be donor lymphoid cells transformed by Epstein-Barr virus
183 (EBV). Serial monitoring of blood for EBV DNA may be warranted in high-risk groups.

184
185 Leukemia of donor origin also has been reported in HPC, Cord Blood recipients. The natural
186 history is presumed to be the same as that for *de novo* leukemia.

188 **5.7 Transmission of Serious Infections**

189
190 Transmission of infectious disease may occur because HPC, Cord Blood is derived from human
191 blood. Disease may be caused by known or unknown infectious agents. Donors are screened for
192 increased risk of infection with human immunodeficiency virus (HIV), human T-cell
193 lymphotropic virus (HTLV), hepatitis B virus (HBV), hepatitis C virus (HCV), *T. pallidum*,
194 *T. cruzi*, West Nile Virus (WNV), transmissible spongiform encephalopathy (TSE) agents, and
195 vaccinia. Donors are also screened for clinical evidence of sepsis, and communicable disease
196 risks associated with xenotransplantation. Maternal blood samples are tested for HIV types 1
197 and 2, HTLV types I and II, HBV, HCV, *T. pallidum*, WNV, and *T. cruzi*. HPC, Cord Blood is
198 tested for sterility. These measures do not totally eliminate the risk of transmitting these or other
199 transmissible infectious diseases and disease agents. Report the occurrence of a suspected

200 transmitted infection to the quality department of LifeSouth Community Blood Centers at 1-888-
201 795-2707.

202
203 Testing is also performed for evidence of donor infection due to cytomegalovirus (CMV). Test
204 results may be found in the accompanying records.

205 206 **5.8 Transmission of Rare Genetic Diseases**

207
208 HPC, Cord Blood may transmit rare genetic diseases involving the hematopoietic system for
209 which donor screening and/or testing has not been performed [*see Adverse Reactions (6.1)*].
210 Cord blood donors have been screened by family history to exclude inherited disorders of the
211 blood and marrow. HPC, Cord Blood has been tested to exclude donors with sickle cell anemia,
212 and anemias due to abnormalities in hemoglobins C, D, and E. Because of the age of the donor
213 at the time HPC, Cord Blood collection takes place, the ability to exclude rare genetic diseases is
214 severely limited.

215 216 **6 ADVERSE REACTIONS**

217
218 Day-100 mortality from all causes was 25%.

219
220 The most common infusion-related adverse reactions ($\geq 5\%$) are hypertension, vomiting, nausea,
221 bradycardia, and fever.

222 223 **6.1 Clinical Trials Experience**

224
225 Because clinical trials are conducted under widely varying conditions, adverse reaction rates
226 observed in the clinical trials of a drug cannot be directly compared to rates in the clinical trials
227 of another drug and may not reflect the rates observed in practice.

228
229 The safety assessment of HPC, Cord Blood is based primarily on review of the data submitted to
230 the FDA dockets from various sources, the dataset for the COBLT Study, and published
231 literature.

232 233 *Infusion Reactions*

234
235 The data described in Table 1 reflect exposure to 442 infusions of HPC, Cord Blood (from
236 multiple cord blood banks) in patients treated using a total nucleated cell dose $\geq 2.5 \times 10^7/\text{kg}$ on
237 a single-arm prospective trial or expanded access use (COBLT Study). The population was 60%
238 male and the median age was 5 years (range 0.05-68 years), and included patients treated for
239 hematologic malignancies, inherited metabolic disorders, primary immunodeficiencies, and bone
240 marrow failure. Preparative regimens and graft-vs.-host disease prophylaxis were not
241 standardized. The most common infusion reactions were hypertension, vomiting, nausea, and
242 sinus bradycardia. Hypertension and any grades 3-4 infusion-related reactions occurred more
243 frequently in patients receiving HPC, Cord Blood in volumes > 150 milliliters and in pediatric
244 patients. The rate of serious adverse cardiopulmonary reactions was 0.8%.

245

Table 1: Incidence of Infusion-Related Adverse Reactions
Occurring in $\geq 1\%$ of Infusions (COBLT Study)

	Any grade	Grade 3-4
Any reaction	65.4%	27.6%
Hypertension	48.0%	21.3%
Vomiting	14.5%	0.2%
Nausea	12.7%	5.7%
Sinus bradycardia	10.4%	0
Fever	5.2%	0.2%
Sinus tachycardia	4.5%	0.2%
Allergy	3.4%	0.2%
Hypotension	2.5%	0
Hemoglobinuria	2.1%	0
Hypoxia	2.0%	2.0%

246

247 No information on the types and rates of infusion reactions were reported with LifeSouth HPC,
248 Cord Blood.

249

250 *Other Adverse Reactions*

251

252 For other adverse reactions, the raw clinical data from the dockets were pooled for 1299 (120
253 adult and 1179 pediatric) patients transplanted with HPC, Cord Blood (from multiple cord blood
254 banks) with total nucleated cell dose $\geq 2.5 \times 10^7/\text{kg}$. Of these, 66% (n=862) underwent
255 transplantation as treatment for hematologic malignancy. The preparative regimens and graft-
256 vs.-host disease prophylaxis varied. The median total nucleated cell dose was $6.4 \times 10^7/\text{kg}$
257 (range, $2.5\text{-}73.8 \times 10^7/\text{kg}$). For these patients, Day-100 mortality from all causes was 25%.
258 Primary graft failure occurred in 16%; 42% developed grades 2-4 acute graft-vs.-host disease;
259 and 19% developed grades 3-4 acute graft-vs.-host disease.

260

261 Data from published literature and from observational registries, institutional databases, and cord
262 blood bank reviews reported to the dockets for HPC, Cord Blood (from multiple cord blood
263 banks) revealed nine cases of donor cell leukemia, one case of transmission of infection, and one
264 report of transplantation from a donor with an inheritable genetic disorder. The data are not
265 sufficient to support reliable estimates of the incidences of these events.

266

267 In the COBLT Study, 15% of the patients developed engraftment syndrome.

268

269 **8 USE IN SPECIFIC POPULATIONS**

270

271 **8.1 Pregnancy**

272

273 Pregnancy Category C. Animal reproduction studies have not been conducted with HPC, Cord
274 Blood. It is also not known whether HPC, Cord Blood can cause fetal harm when administered
275 to a pregnant woman or can affect reproduction capacity. There are no adequate and well-
276 controlled studies in pregnant women. HPC, Cord Blood should be used during pregnancy only
277 if the potential benefit justifies the potential risk to the fetus.

278

279 **8.4 Pediatric Use**

280

281 HPC, Cord Blood has been used in pediatric patients with disorders affecting the hematopoietic
282 system that are inherited, acquired, or resulted from myeloablative treatment [*see Dosage and*
283 *Administration (2), Adverse Reactions (6), and Clinical Studies (14)*].

284

285 **8.5 Geriatric Use**

286

287 Clinical studies of HPC, Cord Blood (from multiple cord blood banks) did not include sufficient
288 numbers of subjects aged 65 years and over to determine whether they respond differently than
289 younger subjects. In general, administration of HPC, Cord Blood to patients over age 65 years
290 should be cautious, reflecting their greater frequency of decreased hepatic, renal, or cardiac
291 function, and of concomitant disease or other drug therapy.

292

293 **8.6 Renal Disease**

294

295 HPC, Cord Blood contains Dextran 40 which is eliminated by the kidneys. The safety of HPC,
296 Cord Blood has not been established in patients with renal insufficiency or renal failure.

297

298 **10 OVERDOSAGE**

299

300 **10.1 Human Overdosage Experience**

301

302 There has been no experience with overdosage of HPC, Cord Blood in human clinical trials.
303 Single doses of LifeSouth HPC, Cord Blood up to 71×10^7 TNC/kg have been administered.
304 HPC, Cord Blood prepared for infusion may contain dimethyl sulfoxide (DMSO). The
305 maximum tolerated dose of DMSO has not been established, but it is customary not to exceed a
306 DMSO dose of 1 gm/kg/day when given intravenously. Several cases of altered mental status
307 and coma have been reported with higher doses of DMSO.

308

309 **10.2 Management of Overdose**

310

311 For DMSO overdosage, general supportive care is indicated. The role of other interventions to
312 treat DMSO overdosage has not been established.

313

314 **11 DESCRIPTION**

315

316 HPC, Cord Blood consists of hematopoietic progenitor cells, monocytes, lymphocytes, and
317 granulocytes from human cord blood for intravenous infusion. Blood recovered from umbilical
318 cord and placenta is volume reduced and partially depleted of red blood cells and plasma.

319

320 The active ingredient is hematopoietic progenitor cells which express the cell surface marker
321 CD34. The potency of cord blood is determined by measuring the numbers of total nucleated
322 cells (TNC) and CD34+ cells, and cell viability. Each unit of HPC, Cord Blood contains a
323 minimum of 5×10^8 total nucleated cells with at least 1.25×10^6 viable CD34+ cells at the time
324 of cryopreservation. The cellular composition of HPC, Cord Blood depends on the composition
325 of cells in the blood recovered from the umbilical cord and placenta of the donor. The actual
326 nucleated cell count, the CD34+ cell count, the ABO group, and the HLA typing are listed in the
327 accompanying records sent with each individual unit.

328

329 HPC, Cord Blood has the following inactive ingredients: dimethyl sulfoxide (DMSO), Dextran
330 40, and hydroxyethylstarch. When prepared for infusion according to instructions, the infusate
331 contains the following inactive ingredients: Dextran 40, human serum albumin, residual DMSO,
332 and residual hydroxyethylstarch.

333

334 **12 CLINICAL PHARMACOLOGY**

335

336 **12.1 Mechanism of Action**

337

338 Hematopoietic stem/progenitor cells from HPC, Cord Blood migrate to the bone marrow where
339 they divide and mature. The mature cells are released into the bloodstream, where some
340 circulate and others migrate to tissue sites, partially or fully restoring blood counts and function,
341 including immune function, of blood-borne cells of marrow origin. [See *Clinical Studies (14)*.]
342

343

344 In patients with enzymatic abnormalities due to certain severe types of storage disorders, mature
345 leukocytes resulting from HPC, Cord Blood transplantation may synthesize enzymes that may be
346 able to circulate and improve cellular functions of some native tissues. However, the precise
347 mechanism of action is unknown.

348

348 **14 CLINICAL STUDIES**

349

350 The effectiveness of HPC, Cord Blood, as defined by hematopoietic reconstitution, was
351 demonstrated in one single-arm prospective study (COBLT Study), and in retrospective reviews
352 of data from an observational database for LifeSouth HPC, Cord Blood and data in the dockets
353 and public information. Of the 1299 patients in the dockets and public data, 66% (n=862)
354 underwent transplantation as treatment for hematologic malignancy. Results for patients who
355 received a total nucleated cell dose $\geq 2.5 \times 10^7$ /kg are shown in Table 2. Neutrophil recovery is
356 defined as the time from transplantation to an absolute neutrophil count more than 500 per
357 microliter. Platelet recovery is the time to a platelet count more than 20,000 per microliter.
358 Erythrocyte recovery is the time to a reticulocyte count greater than 30,000 per microliter. The
359 total nucleated cell dose and degree of HLA match were inversely associated with the time to
360 neutrophil recovery in the docket data.

361

Table 2: Hematopoietic Recovery for Patients Transplanted with HPC, Cord Blood, Total Nucleated Cell (TNC) Dose $\geq 2.5 \times 10^7/\text{kg}$

Data Source	COBLT Study*	Docket* and Public Data*	LifeSouth HPC, Cord Blood
Design	Single-arm prospective	Retrospective	Retrospective
Number of patients	324	1299	22
Median age (years) (range)	4.6 (0.07 – 52.2)	7.0 (<1 – 65.7)	8 (0.6 – 61.8)
Gender	59% male 41% female	57% male 43% female	59% male 41% female
Median TNC Dose ($\times 10^7/\text{kg}$) (range)	6.7 (2.6 – 38.8)	6.4 (2.5 – 73.8)	5.1 (2.8 – 70.6)
Neutrophil Recovery at Day 42 (95% CI)	76% (71% – 81%)	77% (75% – 79%)	91% (71% - 98%)
Platelet Recovery at Day 100 of 20,000/microliter (95% CI)	57% (51% – 63%)	-	95% (79% - 99%)
Platelet Recovery at Day 100 of 50,000/microliter (95% CI)	46% (39% – 51%)	45% (42% – 48%)	95% (79% - 99%)
Erythrocyte Recovery at Day 100 (95% CI)	65% (58% – 71%)	-	-
Median time to Neutrophil Recovery	27 days	25 days	22 days
Median time to Platelet Recovery of 20,000/microliter	90 days	-	44 days
Median time to Platelet Recovery of 50,000/microliter	113 days	122 days	70 days
Median time to Erythrocyte Recovery	64 days	-	-

* HPC, Cord Blood (from multiple cord blood banks)

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16 HOW SUPPLIED/STORAGE AND HANDLING

HPC, Cord Blood is supplied as a cryopreserved cell suspension in a sealed bag containing a minimum of 5×10^8 total nucleated cells with a minimum of 1.25×10^6 viable CD34+ cells in a volume of 25 milliliters (ISBT 128 Product Code S1393, ISBT 128 Facility Identifier Number W2434). The exact pre-cryopreservation nucleated cell content is provided in the accompanying records.

Store HPC, Cord Blood at or below -150°C until ready for thawing and preparation.

17 PATIENT COUNSELING INFORMATION

Discuss the following with patients receiving HPC, Cord Blood:

- Report immediately any signs and symptoms of acute infusion reactions, such as fever, chills, fatigue, breathing problems, dizziness, nausea, vomiting, headache, or muscle aches.

- 381 • Report immediately any signs or symptoms suggestive of graft-vs.-host disease, including
382 rash, diarrhea, or yellowing of the eyes.

383 **INSTRUCTIONS FOR PREPARATION FOR INFUSION**

384

385 The HPC, Cord Blood unit is stored continuously inside a steel canister in liquid nitrogen at
386 temperatures $\leq -150^{\circ}$ C. For shipment, the canister is placed inside a container specifically
387 designed to keep the temperature at or below -150° C (dry shipper). It is recommended to keep
388 the canister inside the dry shipper for short-term storage (up to 48 hours) or transfer it into a
389 liquid nitrogen (LN2)-cooled storage device at the Transplant Center for storage greater than 48
390 hours.

391

392 **I MATERIALS**

393

Equipment:

394

- Refrigerated centrifuge

395

- Plasma extractor/expressor

396

- Biological safety cabinet

397

- Scale

398

- Tube sealer compatible with polyvinyl chloride plastic

399

- Water bath

400

- Canister-opening tool (supplied by LifeSouth Community Blood Centers)

401

- LN2 storage freezer at -150° C or colder

402

- Sterile docker

403

- Automated cell counter and/or microscope and cell count chamber for cell count

404

- and viability determination (optional)

405

406 **Personal Protective Equipment:**

407

- Gloves (sterile preferred)

408

- Protective cryogloves

409

- Safety goggles or face shield

410

- Lab coat

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412 **Reagents:**

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- 10% Dextran 40 in Sodium Chloride Injection, USP

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- 5% Albumin (Human), USP

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416 **Supplies:**

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- Sterile disposable syringes - (2) 30-mL, (8) 60-mL

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- 18-gauge injection needles

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- Hemostat

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- Small plastic zip-lock bags (sterile preferred)

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- Alcohol prep pads

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- (3) sampling site couplers

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- (4) 300-mL transfer bags

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- Sterile pipettes

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- Sterile sampling cups

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- Bacterial culture bottles (aerobic and anaerobic) or other supplies for conducting

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- sterility testing

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- Tubes for cell counts, progenitor assays (optional)

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430 **Forms:**

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- *Receipt of Cord Blood Unit form*

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- *Umbilical Cord Blood Cryopreserved Transfer Report form*

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434 **II PRODUCT IDENTITY VERIFICATION**

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1. Apply personal protective equipment.
2. Open the dry shipper lid upon receipt using scissors to remove the tie-tag from the outside of the shipper.
3. Verify that the National Marrow Donor Program (NMDP) number on the tie-tag matches the NMDP number on the *Umbilical Cord Blood Cryopreserved Unit Transfer Report*. If the NMDP numbers do not match, contact LifeSouth Community Blood Centers (LifeSouth) at 1-888-795-2707.
4. Remove the canister from the dry shipper and the canister-opening tool from the shipment documentation packet (see Figure 1).

Figure 1:



5. Compare product bar code information on bar-coded label on the side of the canister (see Figure 1) with the product identification (ID) information included in the packaging. Verify this information as soon as the shipment arrives and before administering the HPC, Cord Blood unit. If the bar-coded label is not found on the outside of the canister, the product bar code information can be found on the frozen cord blood unit enclosed in the canister.
6. Using protective cryogloves and the canister-opening tool, open the canister at top and bottom using the following steps:
 - a. Avoid damaging the frozen cord blood bag.
 - b. Align the canister opening tool with the slot in the bottom of the canister (see Figure 2).
 - c. Turn the canister opening tool clockwise to open the bottom of the canister (see Figure 3).

Figure 2:

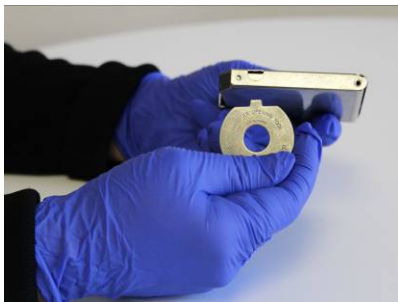


Figure 3:

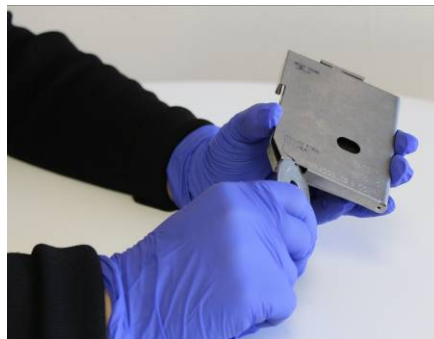


Figure 4:



- d. Locate the opening at the top of the canister and use the tool in a counterclockwise motion to open the top of the canister (see Figure 4).

- 486 e. Open the canister hinges so the HPC, Cord Blood unit can be removed (see
487 Figure 5).
488

489 Figure 5:



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7. Once the canister is open, compare the product bar code information with your records.
 8. Close canister after verification is complete. Using protective cryogloves, return the canister to the dry shipper for short term storage (up to 48 hours) or to LN2-cooled storage device for storage greater than 48 hours.
 9. Once records are all verified, indicate acceptance by initial and date on the indicated space on the *Receipt of Cord Blood Unit* and *Umbilical Cord Blood Cryopreserved Transfer Report* forms.

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For Incorrect Information:

1. If any information is incorrect or cannot be verified, close the canister and return the frozen unit to the dry shipper for short term storage (up to 48 hours) or into a LN2-cooled storage device for storage greater than 48 hours.
2. Report the discrepancy immediately to LifeSouth at 1-888-795-2707 and to the transplant physician.
3. Perform a thorough investigation, keeping the HPC, Cord Blood unit frozen at or below -150° C until all discrepancies are resolved.
4. Once records are all verified, indicate acceptance by initial and date on the indicated space on the *Receipt of Cord Blood Unit* and *Umbilical Cord Blood Cryopreserved Transfer Report* forms.

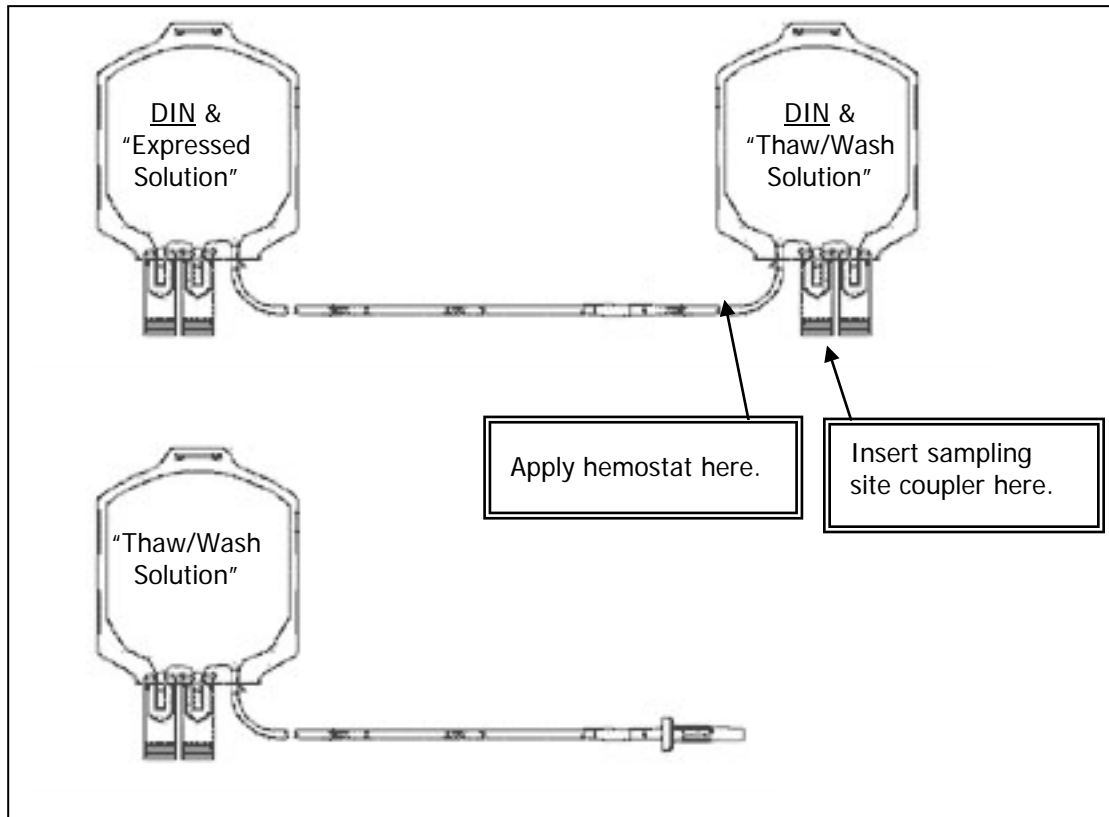
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526 **III PREPARATION**

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528 **A Prepare Thawing Solution**

1. Label a 300-mL transfer bag "Thaw/Wash Solution" (see Figure 6 below).
2. Clamp the "Thaw/Wash Solution" bag with a hemostat to prevent leaking.
3. Refrigerate the 10% Dextran 40 in Sodium Chloride Injection, USP and the 5% Albumin (Human), USP at 2-6°C.
4. Complete the following steps inside a biological safety cabinet:
 - i. Add 72.5 mL of refrigerated 10% Dextran 40 in Sodium Chloride Injection, USP to the 300-mL transfer bag labeled Thaw/Wash Solution using two 60-mL sterile syringes.
 - ii. Add 72.5 mL of refrigerated 5% Albumin (Human), USP to the same 300-mL transfer bag labeled Thaw/Wash Solution using two 60-mL syringes.
 - iii. Mix by inverting the bag at least 10 times.

- iv. Label a 60-mL sterile syringe "Resuspension Solution"; fill with 50 mL of the solution from transfer bag labeled Thaw/Wash Solution (1:1 ratio of 10% Dextran 40 in Sodium Chloride Injection, USP / 5% Albumin (Human), USP) for final end-product re-suspension. Refrigerate until ready for use.
- v. Label two 60-mL sterile syringes and two 30-mL sterile syringes as "Thaw/Wash Solution."
 1. For the two 60-mL syringes draw 20 mL of thaw/wash solution into each syringe; refrigerate until ready for use.
 2. For the two 30-mL syringes draw 5 mL of thaw/wash solution into each syringe; refrigerate until ready for use.
- vi. Using a sterile docker, connect two 300-mL transfer bags into a transfer set (see Figure 6 below).
 1. Label one transfer bag with the HPC, Cord Blood unit Donation Identification Number (DIN) and "Thaw/Wash Solution."
 2. Label the second transfer bag with the HPC, Cord Blood unit DIN and "Expressed Solution."
- vii. Insert a sampling site coupler into the port closest to the tubing on the DIN and Thaw/Wash Solution labeled transfer bag (see Figure 6).
- viii. Attach a hemostat to the tubing approximately one inch above the sampling site coupler on the DIN and Thaw/Wash Solution transfer bag (see Figure 6).

Figure 6: Transfer Set Diagram



B Thaw HPC, Cord Blood Unit

595 Schedule the infusion time of the transplant with the transplant team in advance of the
596 procedure. Re-confirm on the day of infusion with the transplant team so that the start
597 time for the thawing procedure can be adjusted to have the unit ready for infusion at a
598 time the patient can receive the infusion.

599
600 If canister is stored in LN2, wear protective cryogloves, lift canister containing the HPC,
601 Cord Blood unit from the liquid phase of the LN2 container, and rest canister in the vapor
602 phase within the container for five to ten minutes before proceeding.

603
604 **Note:** Carefully check the identity of the unit to be thawed.

- 605
606 1. Open canister with the canister opening tool (refer to section **II PRODUCT**
607 **IDENTITY VERIFICATION**). Avoid damage to the plastic bag containing
608 the frozen HPC, Cord Blood unit. Carefully examine the plastic bag for
609 breaks or cracks.
610 2. Remove the HPC, Cord Blood unit bag from the canister.
611 3. Place the HPC, Cord Blood unit bag into a clean plastic zip-lock bag. Let the
612 air out, and close it tightly. Use a sterile plastic zip-lock bag if available.
613 4. Place the plastic zip-lock bag containing the frozen cord blood unit into a
614 warm water bath at a temperature of 37° C +/- 1° C. Gently and carefully
615 agitate the bag in the water bath to accelerate thawing and resuspension of the
616 cells. Use your fingers to massage the bag to ensure equitable distribution of
617 heat (see Figure 7).

618
619 Figure 7:



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637 5. Watch closely for any cracks or breaks, as shown by red cells leaking from the
638 cord blood bag into the plastic zip-lock bag.
639 6. If any leakage is seen, keep the leakage site upright to prevent further leaking
640 while continuing to gently agitate the bag until the product is slushy. (See
641 section **V EMERGENCY RECOVERY PROCEDURE IN THE EVENT**
642 **OF A CONTAINER FAILURE** for procedures for emergency recovery of
643 the thawed cord blood cells).
644 7. If no leakage is seen, remove the plastic zip-lock bag from water bath. Dry
645 the outside of the bag, disinfect it with alcohol, and place it inside a biological
646 safety cabinet.
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648 **C Transfer the Thawed HPC, Cord Blood Unit to Transfer Set**

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Complete the following steps inside a biological safety cabinet:

1. Obtain pre-labeled transfer set (see section IIIA, step 4vi).
2. Obtain HPC, Cord Blood unit; clean both sealed ports with alcohol.
3. Obtain scissors; disinfect port covers and cut off top of port to create an opening for insertion of sampling site coupler.
4. Disinfect cut port-cover surfaces with alcohol and attach one sample site coupler to each port.
5. Obtain one of the prepared 30-mL syringes containing 5 mL of thaw/wash solution (see section IIIA, step 4v).
6. Connect the 30-mL syringe into the 5-mL cord blood compartment port; slowly dispense thaw/wash solution into this compartment.
7. Slowly pull back and push in the syringe plunger to mix the cord blood and thaw/wash solution; repeat three to four times. After the last one, draw all the fluid from the compartment into the syringe.
8. Disperse all the fluid in the syringe obtained in step 7 into the labeled DIN and Thaw/Wash Solution bag.
9. Repeat steps 5 through 8 with the second prepared 30-mL syringe containing 5 mL of thaw/wash solution to wash any remaining HPC, Cord Blood from the 5-mL compartment. The DIN and Thaw/Wash Solution Bag should contain 15 mL of HPC, Cord Blood and solution.
10. Obtain one of the prepared 60-mL syringes containing 20 mL of thaw/wash solution (see section IIIA, step 4v).
11. Connect the 60-mL syringe into the larger 20-mL cord blood compartment port; slowly dispense the thaw/wash solution into this compartment.
12. Slowly pull back and push in the syringe plunger to mix the cord blood and thaw/wash solution; repeat three to four times. After the last one, draw all the fluid from the compartment into the syringe.
13. Disperse all the fluid in the syringe obtained in step 12 into the labeled DIN and Thaw/Wash Solution bag.
14. Repeat steps 10 through 14 using the second prepared 60-mL syringe containing 20 mL of thaw/wash solution to wash the remaining contents from the 20-mL compartment. The DIN and Thaw/Wash Solution bag should contain 75 mL of HPC, Cord Blood and solution.
15. Allow five minutes for the mixture to equilibrate.
16. Draw any remaining thaw/wash solution from the Thaw/Wash Solution transfer bag into a 60-mL syringe and add solution into the DIN and Thaw/Wash Solution transfer bag. The DIN and Thaw/Wash Solution bag should contain approximately 120 mL of HPC, Cord Blood and solution.
17. Mix the DIN and Thaw/Wash Solution transfer bag well by inverting the transfer bag 180° 10 to 15 times.
18. Discard the empty Thaw/Wash Solution bag.

D Wash the Thawed HPC, Cord Blood Unit

1. Place the DIN and Thaw/Wash Solution transfer bag containing the hemostat, sampling site coupler, and DIN and Expressed Solution transfer bag into refrigerated centrifuge in upright position. Do not allow bags to crease. Inserts may be used to achieve upright position and to prevent damage as a result of centrifugation with a sampling site coupler.

2. Balance the centrifuge before beginning centrifugation cycle.
3. Centrifuge at 400 g for 20 minutes at 10°C.
4. After centrifugation, look for clear separation of red blood cell (RBC) pellet.
5. Place transfer bag labeled DIN and Thaw/Wash Solution into a plasma extractor (see Figure 8) and allow the supernatant to flow into the transfer bag labeled DIN and Expressed Solution by removing the hemostat from the tubing. Do not disturb the cells. If cells transfer, restore everything back into the primary transfer bag, re-centrifuge, and repeat extraction procedure.

Figure 8:



6. Allow all supernatant to leave bag but do not allow the RBC pellet to escape (see Figure 9). Hemostat the tubing after expressing to close tubing on the DIN and Thaw/Wash Solution transfer bag. Seal tubing with heat sealer on DIN and Thaw/Wash Solution transfer bag where the tubing begins. Make two seals; cut in between the two seals. Discard the line and DIN and Express Solution transfer bag.

Figure 9:



7. Obtain the prepared 60-mL “Resuspension Solution” syringe containing 50 mL thaw/wash solution (see section IIIA, step 4iv). Slowly add 50 mL of thaw/wash solution to the DIN and Thaw/Wash Solution transfer bag.
8. Obtain an unlabeled 300 mL transfer bag. Seal the tubing of the transfer bag approximately one inch above the introduction of the tubing into the bag.

- 756 Remove the excess tubing. Weigh the bag to obtain a tare weight of the
757 transfer bag.
- 758 9. Weigh the DIN and Thaw/Wash Solution bag. Calculate volume based on
759 weight of bag minus tare weight obtained in step 8.
- 760 10. Complete sampling for CFU, TNC, CD34+, viability, and ABO/Rh and HLA,
761 if necessary.
- 762 11. Label DIN and Thaw/Wash Solution transfer bag with the expiration time and
763 the time of completion of washing. The recommended expiration time is 2
764 hours after the completion of wash until infusion, if stored at room
765 temperature (19-25°C) or 4 °C.
- 766 12. Notify the Transplant Center that the HPC, Cord Blood unit is thawed,
767 washed, and available for infusion.
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769 **IV ADMINISTRATIVE REQUIREMENTS**

- 770 1. Prepare a written summary of the procedure, including:
- 771 a. HPC, Cord Blood ID number
 - 772 b. Date of receipt of HPC, Cord Blood unit
 - 773 c. Liquid nitrogen storage temperature
 - 774 d. Date of thaw, including whether and at what stage leaks or cracks
775 occurred
 - 776 e. Date and time HPC, Cord Blood unit removed from liquid nitrogen
777 storage
 - 778 f. Volume of final product
 - 779 g. TNC (Total nucleated cell) count, CD34+ count
 - 780 h. Viability of recovered cells (TNC or CD34+) plus name of method used
 - 781 i. Results of bacterial and fungal cultures
- 782 2. Make a copy of the report for your records.
- 783 3. Fax a copy of the report to LifeSouth at 352-224-1650.
- 784 4. Return the dry shipper to LifeSouth. The return address is:

785
786 LifeSouth Community Blood Centers, Inc.
787 LifeCord Cord Blood Bank
788 4039 Newberry Road
789 Gainesville, FL 32607

790
791 Phone: 888-795-2707 x 41738
792 Fax: 352-224-1650
793

794 **V EMERGENCY RECOVERY PROCEDURE IN THE EVENT OF A CONTAINER**
795 **FAILURE**

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- 797 1. Handle the frozen cord blood bag with extreme care at every step including
798 opening the metal containers, inspecting, thawing and/or washing.
 - 799 2. Use standard procedures and competent personnel to perform post-thaw
800 sampling and/or bag rescue.
 - 801 3. Perform all steps on lab benches, under biological safety cabinet, or another
802 surface to prevent inadvertent drop of the frozen unit.
 - 803 4. To facilitate thawing, gradually remove the HPC, Cord Blood unit from the
804 liquid phase of the LN2 storage area, suspending in the vapor phase for at
805 least five minutes prior to bringing the container to room temperature.

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5. Put the frozen bag inside a zip-lock bag prior to initiating the thaw to facilitate salvage of the product and to reduce the possibility of contamination.
 6. If the HPC, Cord Blood unit is seen to be cracked when removed from the LN2 storage container, or if cracks or leaks occur during thawing, immediately notify LifeSouth at 1-888-795-2707. Notify the transplant physician/team and the laboratory director as soon as possible.
 7. The transplant physician or team will determine whether to use or discard the HPC, Cord Blood product and whether any additional units should be requested.
 8. If the transplant physician or team decides that the product in the leaking unit could be used, the HPC, Cord Blood unit may be recovered as follows:
 - a. Obtain sterile sampling cups and sterile pipettes.
 - b. Open a sterile sampling cup and set cup in working space to receive contents of zip-lock bag.
 - c. If any contents remain in the broken HPC, Cord Blood unit bag, remove the contents from the bag using the syringes prepared in section IIIA.
 - d. Wash all contents out of the HPC, Cord Blood bag and place contents in the DIN and Thaw/Wash Solution labeled transfer bag.
 - e. Using a sterile syringe, transfer 20 mL from the Thaw/Wash Solution bag into a sterile sample cup.
 - f. Using a sterile pipette, obtain 3 mL of thaw/wash solution from the sample cup and inject into the zip-lock bag containing the remaining HPC, Cord Blood unit contents that leaked when thawing.
 - g. Using a different sterile pipette, remove the HPC, Cord Blood and thaw/wash solution from the zip-lock bag and place in a sterile sample cup.
 - h. Repeat steps f through g until all remaining HPC, Cord Blood unit is transferred to the sterile sample cup.
 - i. Using a sterile 20-mL syringe, draw the contents from the sterile sampling cup into the syringe. Inject the solution into the transfer bag labeled DIN and Thaw/Wash Solution.
 - j. Repeat until all of the contents from the sample cup is transferred into the DIN and Thaw/Wash Solution transfer bag.
 - k. After all contents have been transferred from the sample cup and are contained in the DIN and Thaw/Wash Solution transfer bag, mix well by inverting 180° 10 to 15 times.
 - l. Continue with Section **D Wash the Thawed HPC, Cord Blood Unit**.

845 Distributed by:

846
847 LifeSouth Community Blood Centers, Inc.
848 LifeCord Cord Blood Bank
849 4039 Newberry Road
850 Gainesville, FL 32607
851 License No. XXXX