Blood Grouping Reagent and Anti-Human Globulin DG Gel 8 T/S Mono REF 210391 3034956

Instructions for Use

INTENDED USE

The DG Gel 8 T/S Mono card is for the determination of human A, B and D antigens on the surface of red blood cells, and for Indirect Antiglobulin Test of human blood samples. This test does not contain antibodies to complement components.

For use with the DG Gel System.

For in vitro diagnostic use.

SUMMARY AND EXPLANATION

The ABO system was the first human blood group system discovered by Landsteiner in 1900¹ and is still the most important in transfusion practice. The ABO system is defined by the presence or absence of the A and/or B antigens on human red blood cells and by the presence of antibodies in the plasma or serum corresponding to the antigen or antigens missing in the red blood cells. In the field of transfusion medicine, after A and B antigens, the most important blood group antigen is the D antigen from the Rh blood group system. The determination of RhD is defined by the presence or absence of the D antigen in the red blood cells.

Carlo Moreschi described the principle of antiglobulin technique in 1908². In 1945, Coombs and his coworkers Mourant and Race, unaware of this previous description, published and introduced the use of antihuman globulin for the detection of red blood cells coated with non-agglutinating antibodies¹. After Coombs' publication the antiglobulin test was rapidly applied in regular clinical laboratory practice and must rank as almost as important as the discovery of the ABO groups². The monospecific anti-IgG Antiglobulin Test is based on the use of anti-human globulin that allows the detection of red blood cells coated with immunoglobulin (IgG). The Indirect Antiglobulin Test allows the detection of red blood cell antibodies present in the patient's serum or plasma by *in vitro* sensitization of red blood cells. The goal of screening for unexpected antibodies is detection of clinically significant antibodies present in the donor's or patient's sample. In a positive screening of unexpected antibodies, the autocontrol will indicate whether it is due to the presence of an autoantibody, an alloantibody or both. In the antiglobulin crossmatch test the donor's red blood cells combined with the recipient's serum or plasma will show the presence or absence of unexpected antibodies in the recipient's blood that are specific to the antigens of the donor's red blood cells.

The anti-A, anti-B, anti-D and anti-IgG reagents contained in the DG Gel 8 T/S Mono card are used to perform the ABO and RhD blood group typing and the Indirect Antiglobulin Test.

PRINCIPLE OF THE TEST

The principle of the test is based on the gel technique described by Yves Lapierre³ in 1985 for detecting red blood cell agglutination reactions. The DG Gel 8 plastic cards are composed of eight microtubes. Each microtube is made of a chamber, also known as incubator chamber, at the top of a long and narrow microtube, referred to as the column. Buffered gel solution containing specific monoclonal antibodies (anti-A, anti-B and anti-D) or polyclonal anti-human globulin (anti-IgG) has been prefilled into the microtube of the

plastic card. The agglutination occurs when the red blood cell antigens react with the corresponding antibodies present in the gel solution or when the red blood cell sensitized *in vitro* by human IgG antibodies react with the antibodies anti-human globulin present in the gel solution. The gel column acts as a filter that traps agglutinated red blood cells as they pass through the gel column during the centrifugation of the card. The gel column separates agglutinated red blood cells from non-agglutinated red blood cells based on size. Any agglutinated red blood cells are captured at the top of or along the gel column, and non-agglutinated red blood cells reach the bottom of the microtube forming a pellet.

REAGENTS

Observable indications

Note: Inspect the condition of the cards before use (see Warnings and Precautions).

Cards with an alteration or change in color, trapped bubbles in the gel, cracked gel or gel with fissures, drying gel, presence of other artifacts, and opened or damaged seals may indicate an alteration of the product.

Material provided

Note: All microtubes contain sodium azide (NaN₃) as a preservative at a final concentration of 0.09%.

Each microtube of the DG Gel 8 T/S Mono card contains a gel in buffered medium with preservative. The three first microtubes contain also monoclonal antibody reagents, the fourth microtube has only the buffered medium, and microtubes from the fifth to the eighth have polyclonal antibody reagent. The different microtubes are identified on the front label of the card.

Microtube A: monoclonal antibody anti-A. IgM antibody of murine origin, clone DAM-1.

Microtube **B**: monoclonal antibody anti-B. IgM antibody of murine origin, clone 9621A8. This clone does not react with acquired B cells.

Microtube **D**: monoclonal antibody anti-D. IgM antibody of human origin, clone P3x61. This clone detects weak and partial D variants and does not detect partial DVI.

Microtube Ctl.: buffered solution without antibodies (control microtube).

Microtubes IgG: buffered low ionic strength solution (LISS) with rabbit polyclonal anti-human globulin.

Clones 9621A8, P3x61 and AHG Anti-IgG (Rabbit Polyclonal) are produced using intermediate products produced for Diagnostic Grifols S.A. in a shared manufacturing agreement with DIAGAST, Parc Eurasante, 251 av. Eugene Avinee-BP9, 59374 Loos Cedex France; US License Number 1744.

Clone DAM-1 is produced using intermediate products produced for Diagnostic Grifols S.A. in a shared manufacturing agreement with ALBA BIOSCIENCE, Ellen's Glen Road, Edinburgh, EH17 7QT United Kingdom; US License Number 1807.

Reagent preparation

DG Gel 8 T/S Mono card are supplied as ready to use. The gel card and samples to be tested should be brought to room temperature (18-25 °C) before initiating the test.

Material required but not provided

- Automatic pipettes of 10 μL, 25 μL, 50 μL and 1 mL.
- Disposable pipette tips.
- Glass or plastic test tubes.
- Grifols Diluent.

- Reagent Red Blood Cells at 0.8% from Medion Grifols Diagnostics AG or other validated RBCs at 0.8% for screening of unexpected antibodies.
- DG THERM incubator.
- DG SPIN centrifuge.

STORAGE AND STABILITY

- Do not use beyond the expiration date.
- Stored upright (as indicated by the two arrows on the outer packaging) with seal intact at 2-25 °C.
- Do not freeze.
- Do not expose cards to excessive heat.

WARNINGS AND PRECAUTIONS

- The results by themselves alone are not a clinical diagnosis. Evaluate the results together with the patient's clinical information and other data.
- If you observe microbiological contamination, alterations or changes in color, or other artifacts do not use the card.
- If you observe trapped bubbles in the gel, cracked gel or gel with fissures, drying gel, or gel without a visible fine line of supernatant do not use the card.
- Do not use the card if opened or if the aluminum film seal is damaged.
- If you identify incorrect temperature conditions during storage or shipment do not use the cards.
- If you identify improper storage or shipping conditions that results in dispersed drops observed at the top of the microtube, the card should be centrifuged with the DG SPIN before use. If after one centrifugation with the DG SPIN the drops do not descend, do not use the card.
- The product should only be used by qualified personnel.
- The use of volumes and/or red blood cell suspensions in concentrations other than those indicated in the method, may modify the reaction and lead to incorrect test results, i.e., false positive or false negative results.
- The use of diluents other than Grifols Diluent for the red blood cell suspension may modify the reaction and lead to incorrect test results.
- Do not use a centrifuge other than the DG SPIN centrifuge.
- The reagents of the DG Gel 8 T/S Mono card of human monoclonal origin are manufactured using materials that have been tested and found non-reactive for the HBs antigen, and for anti-HIV and anti-HCV antibodies. However, there is no known procedure to ensure that products of human origin will not transmit infectious agents. Human blood products and samples should be handled as if they were potentially capable of transmitting infectious diseases.
- All products with animal derived material, and human blood products and samples should be handled as if they were potentially capable of transmitting infectious diseases.
- Once used, dispose the product in containers for biological waste, according to local, state and national regulations.
- If you have any questions or need further information on the use of this product, consult the authorized distributor.

SPECIMEN COLLECTION AND PREPARATION

Blood samples collected in EDTA or sodium citrate should be used. The collection, separation and handling of the blood should be performed by qualified technical personnel according to current standards⁴⁻⁵, and following the instructions of the manufacturer of the materials used for collecting the sample.

Do not use grossly hemolyzed, cloudy or contaminated samples.

Use the red blood cells collected for the determination of the antigens of the ABO and RhD system and for crossmatch tests and autocontrol. If necessary, samples stored at 2-8 °C can be used up to 72 hours after collection.

Use serum or plasma collected for screening of unexpected antibodies, crossmatch tests and autocontrol. If necessary, samples stored at 2-8 °C can be used up to 72 hours after collection and frozen samples stored up to 5 years at -20 °C or colder may be used after thawing.

Red blood cells from bags collected in ACD, CPD, CPDA-1, CP2D, AS-1 (Adsol) or AS-3 can also be used up to 7 days after the expiration date indicated on the label of the bag, if stored at 2-8 °C. If red blood cells from the bag segment are used, it is recommended that these be washed with physiological saline solution before preparing the suspension. Do not use if clots or hemolysis are observed.

PROCEDURE

- 1. Allow DG Gel 8 T/S Mono cards, additional reagents and the samples to reach room temperature (18-25 °C).
- 2. Inspect the condition of the cards before use (see Warnings and Precautions).
- 3. Identify the cards to be used and the samples to be tested.
- Prepare a 5% red blood cell suspension in Grifols Diluent (50 μL of packed red blood cells in 1 mL of Grifols Diluent)
- 5. If you want to perform one of the following tests, prepare the red blood cell suspensions as indicated:
 - For **Antiglobulin Crossmatch tests:** Prepare a 1% donor's red blood cell suspension in Grifols Diluent (10 µL of packed red blood cells in 1 mL of Grifols Diluent).
 - For Autocontrol: Prepare a patient 1% red blood cell suspension in Grifols Diluent (10 µL of packed red blood cells in 1 mL of Grifols Diluent). Ensure the re-suspension of the red blood cells before use.
- 6. Carefully peel off the aluminum film to prevent cross-contamination of the microtube content among them.
- 7. Ensure the re-suspension of the 5% red blood cell suspension before use.
- 8. Add 10 μ L of the 5% red blood cell suspension into each of the A/B/D/Ctl microtubes.

Note: Carefully dispense the red blood cell suspension and the serum or plasma, avoiding contact of the pipette tip with the wall or the contents of the microtubes to prevent carryover.

For the **IgG microtubes**, select one or more of the following tests:

• For Screening of unexpected antibodies

- 9. Thoroughly mix the vials of Reagent Red Blood Cells for the screening of unexpected antibodies to ensure homogeneous suspension of the red blood cells before use.
- 10. Dispense 50 µL of the Reagent Red Blood Cells (as supplied) into the corresponding microtube.
- 11. Add 25 µL of serum or plasma into the IgG microtubes. Go to step 12 if you do not want to perform any of the following Antiglobulin Tests.

Note: Carefully dispense the red blood cell suspension and the serum or plasma, avoiding contact of the pipette tip with the wall or the contents of the microtubes to prevent carryover.

• For Antiglobulin Crossmatch

- 9. Ensure the re-suspension of the donor's 1% red blood cell suspension before use.
- 10. Dispense 50 µL of the donor's 1% red blood cell suspension into the corresponding IgG microtube.
- 11. Add 25 μ L of the recipient's serum or plasma into the IgG microtubes Go to step 12 if you do not want to perform the Autocontrol test

Note: Carefully dispense the red blood cell suspension and the serum or plasma, avoiding contact of the pipette tip with the wall or the contents of the microtubes to prevent carryover.

For Autocontrol

- 9. Ensure the re-suspension of the patient's 1% red blood cell suspension before use.
- 10. Dispense 50 μ L of the patient's 1% red blood cell suspension into the corresponding IgG microtube.
- 11. Add 25 µL of the patient's serum or plasma into the corresponding IgG microtube.

Note: Carefully dispense the red blood cell suspension and the serum or plasma, avoiding contact of the pipette tip with the wall or the contents of the microtubes to prevent carryover.

- 12. Incubate 15 minutes at 37 °C using DG THERM incubator.
- 13. Centrifuge the gel card in the DG SPIN centrifuge.
- 14. After centrifugation, remove the gel card from the centrifuge and read the results.

RESULTS

Report results as an agglutination grade, absence of agglutination or hemolysis.

Negative results: no agglutination and no hemolysis of red blood cells is visible in the microtube. In a negative result the red blood cells are located in the bottom of the gel column.

Positive results: agglutination and/or hemolysis of the red blood cells is visible in the microtube. In a positive result the agglutinated red blood cells may remain throughout the gel column showing different reaction grades (see Reaction Grades and Figure 1 for a picture of example of reaction grades). Some positive reactions may also form pellet in the bottom of the microtube. A very weak reaction may indicate a very weak or partial expression of the ABO and RhD antigens.

Notes:

1. Some fibrin, particulates or other artifacts may trap red blood cells at the top of the gel columns erroneously leading to an abnormal result (see limitation number 7).

2. Occasionally red blood cell retention in the incubation chamber may occur with positive 4+ samples, without interfering in the result interpretation.

Reaction Grades:

		<u></u>			
Negative	0	Well-defined pellet of non-agglutinated red blood cells at the bottom of the gel column and no visible agglutinated cells in the rest of the gel column.			
Positive	w+	Barely visible small-sized clumps of agglutinated cells in the lower part of the gel			
		column and a pellet of unagglutinated cells at the bottom.			
	1+	Some small-sized clumps of agglutinated cells most frequently in the lower half of the			
		gel column. A small pellet may also be observed at the bottom of the gel column.			
	2+	Small or medium-sized clumps of agglutinated cells throughout the gel column. A few			
		unagglutinated cells may be visible at the bottom of the gel column.			
	3+	Medium-sized clumps of agglutinated cells in the upper half of the gel column.			
	4+	A well-defined band of agglutinated red blood cells in the top part gel column. A few			
		agglutinated cells may be visible below the band.			
mf		Mixed-field. A band of red blood cells at the top part of the gel or dispersed throughout			
		the gel column, and a pellet in the bottom as a negative result.			
н		Hemolysis in the microtube with very few or no red blood cells in the gel column. Report			
		if hemolysis is present in the microtube but not in the sample.			

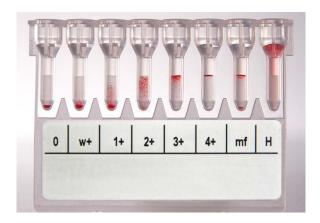


Figure 1. Picture of an example of reaction grades.

Stability of the results

After centrifuging the cards, it is recommended that the results be read immediately. Do not leave processed cards in horizontal position. If necessary, a delayed reading can be performed up to 24 hours after processing the cards if they are kept in an upright position, refrigerated (2-8 °C) and sealed with a laboratory covering film to avoid evaporation of the supernatant.

QUALITY CONTROL

Include positive and negative controls with testing on each day of use. If an unexpected control result is obtained, a complete assessment of the instrument, reagents and material used should be made.

Interpretation of the results

ABO system. The expected reaction with microtubes A and B and its interpretation are shown in the following table (+ = positive, and 0 = negative).

AE	Interpretation		
Microtube A	Microtube B	Microtube Ctl.	ABO group
0	0	0	0
+	0	0	Α
0	+	0	В
+	+	0	AB

D antigen (Rh system). A positive result in the corresponding microtube indicates the presence of antigen D (Rh system).

Indirect Antiglobulin Tests. Tests determined by the result obtained in the microtube. The interpretation of the results depends on the sample and the reagents added to the microtube.

Notes:

- 1. The acronym "Ctl" means Control.
- 2. The Ctl. microtube should be negative. If it is positive, due to the formation of rouleaux, to strong cold autoagglutinins or other causes, invalidate the test. Repeat the determination after washing the red blood cells with physiological saline solution and preparing a new suspension of the washed red blood cells. If the Ctl. microtube of the repeat test is negative, the results of the test can be interpreted; if it is positive, invalidate the test.
- 3. The ABO and RhD group obtained should be verified comparing the result obtained with a previously typed sample. If a previously typed sample is not available, it is recommended that the reverse group be determined. Forward (cell grouping) and reverse (serum grouping) discrepancies should be investigated before releasing the result.
- 4. To ensure detection of weak and partial D variants or if verification of D negative status is required, other reagents and techniques (e.g. Indirect Antiglobulin Testing) which may detect different weak and partial D variants should be used.
- 5. In Indirect Antiglobulin Test, to investigate antibodies at very low concentration (e.g. weak reaction grades) the reactivity may be enhanced by increasing the volume of serum or plasma. Increasing the serum-to-cell ratio⁶ may improve the reactivity to a high agglutination reaction grade.
- 6. Precaution should be taken in the interpretation of mixed-field events. Not all mixed-cell situations are detected. Additional information on patient history and additional testing will be necessary for resolution. Transfused patients or those subjected to bone marrow transplant may present images of mixed-field⁶. Mixed-field is also observed in some ABO subgroups (A₃), in blood group chimerism in fraternal twins, and in the very rare case of mosaicism arising from dispermy⁶.
- 7. The observation of complete or partial hemolysis (pinkish supernatant and/or gel column) in microtubes should be interpreted as a positive result, after verifying that it is not due to a problem of collection and/or handling of the sample.

LIMITATIONS OF THE PROCEDURE

- 1. Grossly hemolyzed, cloudy or contaminated samples or samples with presence of a clot, may cause false positive or false negative results.
- 2. Aged or hemolyzed specimens may cause weaker reactions compared to those obtained with a fresh sample.
- Red blood cells from individuals with A or B variants may present a weak expression of the antigens, and may not be detected. These weak antigens may be better detected using reagent anti-AB included in DG Gel 8 ABO/Rh + Kell card.
- 4. Antigen expression may be weakened in the red blood cells of persons with leukemia or other malignant diseases⁷.
- 5. Abnormal concentrations of serum proteins, the presence of infused macromolecular solutions in the serum or plasma may cause non-specific agglutination of the red blood cells⁶.
- 6. Samples with high-potency antibodies may coat the red blood cells completely, causing spontaneous agglutination⁷.
- 7. If poorly anti-coagulated plasma or incompletely clotted serum is used, fibrin residues may trap non-agglutinated red blood cells at the top of the gel, appearing as a pinkish or reddish layer. Although the results could be correctly interpreted, in a negative reaction the false appearance of a mixed filed could lead to a misinterpretation. In case of incompletely clotted serum samples, it is recommended to re-clot the serum and repeat the test⁷.
- 8. A very weak expression or variants of the D antigens may not be detected.
- 9. No single method is able to detect all unexpected antibodies. The optimum reaction conditions (e.g., sample volume, incubation times) may vary for different antibody specificities.
- 10. The presence of a high concentration of IgG paraproteins in the sample can neutralize the polyspecific anti-human globulin and lead to a false negative result in the antiglobulin test⁶.
- 11. Rare antibodies, notably some anti-Jk^a or anti-Jk^b, may be detected only when polyspecific AHG is used and when active complement is present⁶.
- 10. The Indirect Antiglobulin Test at 37 °C in gel or glass sphere techniques have been reported to show a lower level of sensitivity than results obtained with the tube technique, in the detection of weak agglutination reactions of the ABO system⁷.
- 12. Red blood cell samples with a positive Direct Antiglobulin Test should not be used for Indirect Antiglobulin Testing.
- 13. Antibody activity may decrease in the elderly, infants or persons with disease.
- 14. On occasions, unagglutinated red blood cells may be retained somewhere in the gel column with the appearance of very minute red dots or flecks. However, this nonspecific retention should not interfere with the interpretation of the result.

SPECIFIC PERFORMANCE CHARACTERISTICS

Monoclonal Blood Grouping Reagents:

Grifols DG Gel 8 Blood Grouping Reagents Anti-A, Anti-B, Anti-D meet FDA potency requirements for Blood Grouping Reagents. There is no U.S. standard of potency for the DG Gel 8 Control reagent, which contains no antibody reactivity specific for a blood group antigen.

Every lot has been tested against a panel of positive and negative samples for the relevant antigens to assure reactivity and specificity in accordance with FDA requirements. Details of specificity test results submitted to the FDA for release of product will be furnished upon request.

Anti-IgG Reagent:

Contains Anti-IgG with no anti-complement activity.

Every lot has been tested against a panel of positive and negative samples for the relevant antigens to assure Anti-IgG sensitivity and absence of contaminating antibodies. Details of specificity test results submitted to the FDA for release of the product will be furnished upon request.

The potency of Anti-IgG is verified by tests with red blood cells sensitized with Anti-D and Anti-Fy^a according to methods approved by FDA. The Anti-IgG meets FDA potency requirements.

BIBLIOGRAPHY

1. Klein HG and Anstee DJ. Mollison's Blood Transfusion in Clinical Medicine, 11th edition, Blackwell Scientific Publications, Oxford, 2005.

2. Issit PD and Anstee DJ. Applied Blood Group Serology, 4th edition, Montgomery Scientific Publications, Durham, North Carolina, 1998.

3. Lapierre Y, et al. The gel test: a new way to detect red cells antigen-antibody reactions. Transfusion, 30: 109-113, 1990.

4. CLSI H3-A6: Procedures for the collection of diagnostic blood specimens by venipuncture; Approved Standard, 6th edition, 2007.

5. CLSI H18-A3: Procedures for the handling and processing of blood specimens; Approved Guideline, 3rd edition, 2004.

6. Technical Manual, 17th edition, American Association of Blood Banks, Bethesda, Maryland, 2011.

7. Phillips P et al. An explanation and the clinical significance of the failure of microcolumn tests to detect weak ABO and other antibodies. Transfusion Medicine, 7: 47-53, 1997.

PRESENTATION

210391 DG Gel 8 T/S Mono 50 Cards

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SYMBOLS KEY

One or more of these symbols may have been used in the labeling/packaging of this product.

