

M13A BIOEQUIVALENCE FOR IMMEDIATE- RELEASE SOLID ORAL DOSAGE FORMS

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FOREWORD

The International Council for Harmonisation of Technical Requirements for Pharmaceuticals for Human Use (ICH) has the mission of achieving greater regulatory harmonization worldwide to ensure that safe, effective, and high-quality medicines are developed, registered, and maintained in the most resource-efficient manner. By harmonizing the regulatory expectations in regions around the world, ICH guidelines have substantially reduced duplicative clinical studies, prevented unnecessary animal studies, standardized safety reporting and marketing application submissions, and contributed to many other improvements in the quality of global drug development and manufacturing and the products available to patients.

ICH is a consensus-driven process that involves technical experts from regulatory authorities and industry parties in detailed technical and science-based harmonization work that results in the development of ICH guidelines. The commitment to consistent adoption of these consensus-based guidelines by regulators around the globe is critical to realizing the benefits of safe, effective, and high-quality medicines for patients as well as for industry. As a Founding Regulatory Member of ICH, the Food and Drug Administration (FDA) plays a major role in the development of each of the ICH guidelines, which FDA then adopts and issues as guidance to industry.

INTERNATIONAL COUNCIL FOR HARMONISATION OF TECHNICAL
REQUIREMENTS FOR PHARMACEUTICALS FOR HUMAN USE

ICH HARMONISED GUIDELINE

**BIOEQUIVALENCE FOR IMMEDIATE-RELEASE
SOLID ORAL DOSAGE FORMS
M13A**

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At Step 2 of the ICH Process, a consensus draft text or guideline, agreed by the appropriate ICH Expert Working Group, is transmitted by the ICH Assembly to the regulatory authorities of the ICH regions for internal and external consultation, according to national or regional procedures.

M13A
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ICH HARMONISED GUIDELINE
BIOEQUIVALENCE FOR IMMEDIATE-RELEASE
SOLID ORAL DOSAGE FORMS

M13A

ICH Consensus Guideline

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1 **1 INTRODUCTION**

2 **1.1 Objective**

3 This guideline is intended to provide recommendations on conducting bioequivalence (BE) studies
4 during both development and post approval phases for orally administered immediate-release (IR)
5 solid oral dosage forms designed to deliver drugs to the systemic circulation, such as tablets,
6 capsules, and granules/powders for oral suspension.

7 Deviations from the recommendations in this guideline may be acceptable if appropriate scientific
8 justification is provided. Applicants are encouraged to consult the regulatory authority(ies) when
9 an alternate approach is proposed or taken.

10 **1.2 Background**

11 **1.2.1 Bioequivalence**

12 BE for IR solid oral dosage forms with systemic action is largely established via clinical
13 pharmacokinetic (PK) BE studies or comparative *in vitro* dissolution studies. In addition to the
14 oral dosage forms stated above, the PK principles of this guideline are generally applicable to non-
15 orally administered drug products with immediate action in which reliance on systemic exposure
16 measures is suitable for establishing BE, e.g., certain rectal, inhalation, and nasal drug products.

17 BE assessment for these oral dosage forms is important for establishing therapeutic equivalence
18 for generic drug products to their respective comparator products. In addition, there may be
19 situations in new (innovator) drug development when demonstration of BE may be critical for
20 approval decisions. Furthermore, BE studies are used by innovator and generic product developers
21 for supporting post-approval formulation and/or manufacturing process changes.

22 Two drug products containing the same drug substance(s) are considered bioequivalent if their
23 relative bioavailability (BA) (rate and extent of drug absorption) after administration in the same
24 molar dose lies within acceptable predefined limits. These limits are set to ensure comparable *in*
25 *vivo* performance, i.e., similarity in terms of safety and efficacy.

26 The Biopharmaceutics Classification System (BCS)-based biowaiver may be used to waive *in vivo*
27 BE studies for certain orally administered IR solid oral dosage forms as delineated in ICH M9,

28 *Biopharmaceutics Classification System-Based Biowaivers.*

29 **1.2.2 Data Integrity**

30 BE studies should be conducted according to the principles and recommendations in ICH E6, *Good*
31 *Clinical Practice*. In conducting BE studies, sponsors, study investigators, and service providers,
32 e.g., contract research organisations or laboratories, should ensure that the data generated are
33 attributable, legible, contemporaneously documented, original (or a certified copy),
34 accurate, complete, and traceable. The ultimate responsibility for the quality and integrity of the
35 study data submitted to a regulatory authority lies with the applicant.

36 **1.3 Scope**

37 M13A is the first guideline in the series to describe the scientific and technical aspects of study
38 design and data analysis to support BE assessment for orally administered IR solid oral dosage
39 forms. How regulatory decisions may be made based on BE assessment is out of the scope of this
40 guideline.

41 Acceptance of comparator products across regulatory jurisdictions could reduce the burden of
42 multiple clinical trials demonstrating BE against local comparator products. However, in many
43 regions this is governed by local laws rather than scientific guidelines. Therefore, the acceptance
44 of comparator products across regions is not in the scope of M13A. However, study designs
45 containing multiple comparator products or test products are included in M13A to take some initial
46 steps to reduce the associated burden without prejudice to regional legal requirements.

47 The second guideline in the series, M13B, will describe biowaiver considerations for additional
48 strengths not investigated in BE studies.

49 The third guideline in the series, M13C, will include data analysis and BE assessment for 1) highly
50 variable drugs, 2) drugs with narrow therapeutic index, and 3) complex BE study design and data
51 analysis considerations, e.g., adaptive BE study design.

52 These guidelines do not cover PK study design or data analysis to support BA assessment for new
53 drug development in support of intended use or dosing recommendations in drug labelling, e.g.,
54 relative BA assessment, food effect, drug-drug interactions, special population studies, bridging

55 formulations without the necessity to demonstrate BE, and studies to support changes in dosing
56 regimens or routes of administration. In such cases, study design and decision criteria may be
57 based on the objective of the study and availability of other information including exposure-
58 response and proposed labelling.

59 **2 GENERAL PRINCIPLES IN ESTABLISHING BIOEQUIVALENCE**

60 **2.1 Study Design for Pharmacokinetic Endpoint Bioequivalence Studies**

61 ***2.1.1 Study Population***

62 The subject population for BE studies should be selected with the aim of permitting detection of
63 differences in the *in vivo* release characteristics between pharmaceutical products. In order to
64 reduce variability not related to differences between products, the studies should normally be
65 performed in healthy subjects unless the drug carries safety concerns that make this approach
66 unethical. Conducting BE studies in healthy subjects is regarded as adequate in most instances to
67 detect formulation differences and to allow extrapolation of the results to populations for which
68 the product is intended.

69 The subject inclusion and exclusion criteria should be clearly stated in the study protocol. Subjects
70 should be at least 18 years of age and preferably have a Body Mass Index between 18.5 and 30.0
71 kg/m². If a drug product is intended for use in both sexes, it is recommended the study include
72 male and female subjects.

73 Subjects should be screened for suitability by means of clinical laboratory tests, a medical history,
74 and a physical examination. Depending on the drug's therapeutic class and safety profile, special
75 medical investigations and precautions may have to be carried out before, during, and after the
76 completion of the BE study. The risk to women of childbearing potential should be considered,
77 and the investigators should ensure that female subjects are not pregnant or lactating during the
78 BE study and the follow-up. Subjects should preferably be non-nicotine users and without a history
79 of alcohol or drug abuse. Phenotyping and/or genotyping of subjects may be considered for safety
80 or PK reasons.

81 If the investigated active substance is known to have adverse effects and the pharmacological
82 effects or risks are considered unacceptable for healthy subjects, the study may instead be

83 conducted in a targeted patient population under suitable precautions and supervision.

84 **2.1.2 Study Design**

85 A randomised, single-dose, two-period, two-sequence crossover study design is recommended
86 when comparing two formulations, as single-dose studies provide the most sensitive conditions to
87 detect differences in the rate and extent of absorption. Treatment periods should be separated by a
88 sufficiently long washout period, e.g., at least 5 elimination half-lives. In general, the highest to-
89 be-marketed strength should be used in a BE study. If the highest strength of a product cannot be
90 administered to healthy subjects for safety and/or tolerability reasons, a single-dose study
91 conducted in healthy subjects using a lower strength may be possible (see Section 2.1.6) or
92 alternatively, if feasible given the drug product under investigation, a single-dose study conducted
93 in patients using the highest proposed strength could be considered.

94 A multiple-dose study may be conducted in patients if a single-dose study cannot be conducted in
95 either healthy subjects for safety and/or tolerability reasons or in patients for ethical reasons. For
96 a multiple-dose study, the study protocol should include an appropriate number of dosage
97 administrations to reach steady-state, which could be justified using an appropriate sampling
98 scheme, i.e., concentrations at the end of the dosing interval should be sampled sequentially until
99 C_{tau} is stable. The washout of the last dose of the first treatment period can overlap with the
100 accumulation of the second treatment. The accumulation period should be sufficiently long to
101 reach the new steady-state after switching and allow the elimination of the drug from the previous
102 treatment, e.g., at least 5 elimination half-lives.

103 For drugs with long elimination half-lives, a parallel design may be employed when a crossover
104 design is impractical due to the need for a prolonged washout period. In this situation, special care
105 should be taken to ensure similar subject demographics in each of the treatment groups.

106 Alternative study designs are acceptable, if scientifically justified.

107 **2.1.3 Sample Size for Bioequivalence Studies**

108 The number of subjects to be included in the BE study should be based on an appropriate sample
109 size calculation to achieve a pre-specified power and pre-specified type 1 error. A sufficient
110 number of subjects should be enrolled in the BE study to account for possible dropouts and/or

111 withdrawals. The use of “spare” subjects is not acceptable. Additional cohort(s) of subjects may
112 be added to the study, e.g., if the number of evaluable subjects falls below the calculated sample
113 size; however, this should be specified in the study protocol and done prior to any bioanalysis. The
114 number of evaluable subjects in a pivotal BE study should not be less than 12 for a crossover
115 design or 12 per treatment group for a parallel design.

116 **2.1.4 Comparator and Test Products**

117 A comparator product is the drug product accepted by regulatory agencies that an applicant can
118 use to compare against the test product in conducting a BE study.

119 The selection of the batch of the comparator product used in the BE study should be based on assay
120 content. It is advisable to investigate more than one batch of the comparator product when selecting
121 the batch of comparator product for use in the BE study.

122 The test product used in the BE study should be representative of the product to be marketed and
123 this should be discussed and justified by the applicant.

124 For pivotal BE studies, the test product used should meet the following criteria:

- 125 a) The production of batches used should provide a high level of assurance that the product
126 and process will be feasible on a commercial scale. The test product should usually
127 originate from a batch of at least 1/10 of production scale or 100,000 units, whichever is
128 greater, unless otherwise justified. In case of a production batch smaller than 100,000 units,
129 a full production batch is required.
- 130 b) Unless otherwise justified, the assayed content of the batch used as test product should not
131 differ by more than 5% from that of the batch used as comparator product, as determined
132 with the test procedure proposed for routine quality testing of the test product.

133 **2.1.5 Fasting and Fed Study Conditions**

134 BE studies should be conducted under standardised conditions that minimise variability to better
135 detect potential PK differences between drug products. For IR solid oral dosage forms, single-dose
136 BE studies conducted under fasting conditions typically provide greater discrimination between
137 the PK profiles of two products. Therefore, for the majority of these drug products, BE may be
138 demonstrated in a single study conducted under fasting conditions.

139 However, food can have a differential, formulation-dependent impact on the absorption of drug
140 substances from drug products that are of high risk (see “High-risk products” section below),
141 which would preclude the extrapolation of BE under fasting conditions to fed conditions. In such
142 cases, BE under fed conditions also needs to be demonstrated.

143 The design of a BE study with regard to the use of fasting and/or fed conditions depends on the
144 dosing instructions of the comparator product as well as the properties of the drug substance and
145 product formulation. A rationale should be provided for the selection of the type of BE study(ies)
146 (fasting or fed or both) and meal type, e.g., fat and calorie content, based on the understanding of
147 the comparator product and the test product (high or non-high risk) as described below. The
148 rationale can be supported by modelling, e.g., appropriately validated/qualified physiologically-
149 based pharmacokinetic (PBPK) modelling or semi-mechanistic absorption models.

150 In addition, safety-related aspects need to be considered when selecting the appropriate condition
151 for a BE study regarding food intake. If safety concerns make it unethical to administer a single
152 dose of the drug product under either fed or fasted conditions, the BE study should be conducted
153 under the condition with less safety concerns.

154 For non-high-risk products, the following is recommended:

- 155 • For a product that is labelled to be taken only under fasting conditions or can be taken
156 under fasting or fed conditions i.e., without regard to food, a single BE study conducted
157 under fasting conditions is recommended to demonstrate bioequivalence.
- 158 • For a product that is labelled to be taken only with food due to PK reasons, e.g., enhancing
159 absorption or reducing variability, a single BE study conducted under fed conditions is
160 recommended to demonstrate bioequivalence.
- 161 • For a product that is labelled to be taken only with food due to tolerability reasons, e.g.,
162 stomach irritation, a single BE study conducted under either fasting or fed conditions is
163 acceptable.

164

165 High-risk products:

166 High-risk products are those where the complexity of the formulation design or manufacturing

167 process leads to an increased likelihood that *in vivo* performance will be impacted differently by
168 varying gastrointestinal (GI) conditions between the fasted and fed states. For these products,
169 performance differences related to differences in formulation and/or manufacturing process may
170 not be detected with a single BE study, i.e., results from a fasting BE study may not be extrapolated
171 to predict fed BE study outcome or vice versa, thus both fasting and fed BE studies should be
172 conducted. For example, some drug products containing low solubility drug substances (as defined
173 by the BCS low solubility criterion described in ICH M9) have complex formulation and/or
174 manufacturing methods (such as solid dispersions, microemulsions, lipid-based formulations,
175 nanotechnologies, or other specialised technologies) to ensure sufficient solubility of the drug
176 substance and dissolution of the drug products or to manage the impact of food. For these high-
177 risk products, BE studies should be conducted under both fasting and fed conditions, irrespective
178 of the product labelling with regard to food intake, except when safety concerns make it unethical
179 to administer a single dose of the drug product under either fed or fasted conditions. Then the BE
180 study should be conducted under the condition with less safety concerns.

181 Especially for low solubility drug substances, the comparator product may be the result of an
182 extensive formulation and/or manufacturing process development program, obtaining for instance
183 a specific formulation without a food effect. If the test product uses a substantially different
184 manufacturing technology or particle size control method from the comparator, or if substantially
185 different excipients are used in the test and comparator that are likely to impact dissolution,
186 solubility, or permeability, this may warrant the need for BE studies under fasting and fed
187 conditions.

188 The above principles with regard to fasting and fed study conditions also apply when BE studies
189 are deemed necessary to bridge formulation and/or manufacturing process changes during pre- or
190 post-marketing phases.

191 Standardisation with regard to meals and water:

192 For studies conducted under fasting conditions, subjects should be fasted for at least 10 hours
193 before drug administration. Subjects should be allowed water as desired, except for 1 hour before
194 and 1 hour after drug administration. The dose should be administered with a standardised volume
195 of water, in the range of 150 to 250 millilitres (ml). No food should be allowed for at least 4 hours

196 post-dose on each day of drug administration and meals taken should be standardised with respect
197 to composition and timing.

198 In the case of studies conducted under fed conditions, the same controls should be employed with
199 the exception that a pre-dose meal should be provided. For a fed BE study, it is recommended that
200 subjects start the meal 30 minutes before administration of the drug product and consume the meal
201 within 30 minutes.

202 If BE studies are conducted under both fasting and fed conditions, i.e., for high-risk products, the
203 BE study conducted under fed conditions should be conducted using a meal that has the potential
204 to cause the greatest effect on GI physiology. The meal should be a high-fat (approximately 50%
205 of total caloric content of the meal) and high-calorie (approximately 800 to 1000 kcal) meal, which
206 should derive approximately 150, 250, and 500-600 kcal from protein, carbohydrate, and fat,
207 respectively. It is recognised that there may be situations where it is appropriate to administer a
208 pre-dose meal with a different caloric/fat content from these recommendations, e.g., for studies
209 performed in patient populations who cannot tolerate the recommended meal composition.

210 If, however, only one BE study conducted under fed conditions is needed for a non-high-risk
211 product, either a high-fat, high-calorie meal or a low-fat, low-calorie meal, e.g., a meal of
212 approximately 500 kcal with approximately 25% of calories from fat, may be administered. If the
213 type of meal to be consumed at the time of drug product administration is clearly specified in the
214 comparator product labelling, then this meal should be employed in the BE study.

215 The composition of the meal to be administered should be described with regard to protein,
216 carbohydrate, and fat content (specified in grams, kcal, and relative caloric content (%)) in the
217 study protocol.

218 In all situations, subjects should abstain from foods and drinks that may interact with circulatory,
219 GI transporter, GI enzymatic, hepatic, or renal function, e.g., alcoholic or caffeinated drinks, or
220 certain fruit juices such as grapefruit juice, during a suitable period before and during the study.

221 ***2.1.6 Dose or Strength to be Studied***

222 In case of an application with multiple strengths, the strength to be used in the BE study depends
223 on the dose proportionality in PK and solubility of the analyte. Generally, the highest to-be-

224 marketed strength can be administered as a single unit. Selection of a lower strength may also be
225 accepted if the highest strength cannot be administered to healthy subjects for safety and/or
226 tolerability reasons and dose proportional PK, i.e., area under the concentration vs time curve
227 (AUC) and C_{max} , has been documented over the range of strengths. If warranted to achieve
228 sufficient bioanalytical sensitivity, multiple units of the highest strength can be administered,
229 provided the total single-dose remains within the labelled dose range and the total dose is safe for
230 administration to the study subjects.

231 For non-proportional increases in AUC and/or C_{max} with increased dose there may be a difference
232 between different strengths in the sensitivity to detect potential differences between formulations.
233 To assess dose proportionality, the applicant should consider all available data regarding dose
234 proportionality. Assessment of dose proportionality should consider single-dose studies only.

235 For drugs with a more than proportional increase in AUC and/or C_{max} with increasing dose over
236 the therapeutic dose range, the BE study should in general be conducted at the highest strength.

237 For drugs with a less than proportional increase in AUC and/or C_{max} with increasing dose over the
238 therapeutic dose range, BE should be established at the lowest strength if this situation is due to
239 saturation of absorption. If the less than proportional increase in AUC and/or C_{max} with increasing
240 dose is due to limited drug solubility, BE studies should be conducted at both the lowest and highest
241 strengths. If the reason for non-dose proportionality is unknown, BE studies should generally be
242 conducted at both the lowest and highest strengths.

243 ***2.1.7 Moieties to be Measured***

244 ***2.1.7.1 Parent versus Metabolite***

245 Demonstration of BE should be based on the analysis of the parent drug because the concentration-
246 time profile of the parent drug is usually considered more sensitive to detect a difference between
247 formulations than metabolite data. This also applies to prodrugs. However, some prodrugs are
248 rapidly eliminated resulting in difficulties in demonstrating BE based on parent drug data, because
249 the parent drug levels are too low to allow reliable bioanalytical measurement. In this situation, it
250 is acceptable to demonstrate BE based on a primary metabolite, i.e., a first-step metabolite of the
251 parent drug, without measurement of the parent compound.

252 In rare cases, demonstration of BE based on the parent drug alone may not be sufficient and the
253 primary active metabolite should also be considered, e.g., drugs that have metabolites formed
254 through gut wall or gut lumen metabolism that contribute to efficacy or safety. This is intended to
255 address situations in which the formation of the metabolite could be influenced by formulation
256 differences, which may not be detectable when measuring systemic levels of the parent drug.

257 **2.1.7.2 Enantiomers versus Racemates**

258 The use of an achiral bioanalytical assay to measure the racemate is generally acceptable.
259 However, a stereoselective assay measuring individual enantiomers in BE studies should be
260 employed when it is known that all of the following conditions have been met:

- 261 a) the enantiomers exhibit different pharmacodynamic properties,
- 262 b) the enantiomers exhibit different PK properties, and
- 263 c) the exposure (AUC) ratio of enantiomers is modified by a difference in the rate of
264 absorption.

265 It is sufficient to demonstrate BE for only the active enantiomer in cases where one enantiomer is
266 inactive (or makes a low contribution) with respect to both safety and efficacy.

267 **2.1.8 Sampling**

268 The sampling schedule in a BE study should cover the concentration-time curve, including a pre-
269 dose sample, samples in the absorption phase, frequent samples around the expected T_{max} , and
270 sufficient samples after T_{max} to ensure a reliable estimate of the extent of exposure, which is
271 achieved when $AUC_{(0-t)}$ covers at least 80% of $AUC_{(0-inf)}$. This period is usually at least three times
272 the terminal half-life of the drug, unless a suitable truncated AUC, e.g., $AUC_{(0-72h)}$, is used. To
273 permit calculation of the relevant PK parameters, a sufficient number of samples should be
274 collected per subject per period, distributed across all phases of disposition.

275 The exact times at which the samples are taken should be recorded to obtain the elapsed time
276 relative to drug administration and sampling should be spaced such that C_{max} , $AUC_{(0-t)}$, and k_d can
277 be estimated accurately.

278 There may be considerable inaccuracies in the estimates of k_{el} if the constant is estimated from
279 linear regression based on a small number of data points. To reduce these inaccuracies, it is

280 recommended that three or more data points in the terminal log-linear phase of the concentration-
281 time curve be used to estimate k_{el} .

282 In multiple-dose studies, the pre-dose sample should be taken immediately before dosing, i.e.,
283 within 5 minutes of dosing, and the last sample is recommended to be taken within 10 minutes of
284 the nominal time for the dosage interval to ensure an accurate determination of $AUC_{(0-\tau_{SS})}$.

285 **2.1.8.1 First Point C_{max}**

286 The sampling schedule should include frequent sampling around the anticipated T_{max} to provide a
287 reliable estimate of C_{max} . In particular, the occurrence of C_{max} at the first post-dose sampling time
288 point should be avoided by careful consideration of the known pharmacokinetic properties of the
289 drug and selection of a suitable early sampling schedule. Datasets where C_{max} occurs at the first
290 post-dose sampling time may result in exclusion of the data from affected subjects from the
291 analysis.

292 **2.1.8.2 Long Half-life Drugs and Truncated AUC Considerations**

293 Truncating AUC for orally administered IR drug products known to exhibit longer elimination
294 half-lives, i.e., 24 hours or longer, mitigates the clinical challenge of prolonged sampling and
295 follow-up. For such products, $AUC_{(0-72h)}$ may be used in place of $AUC_{(0-t)}$ for comparison of the
296 extent of absorption. Seventy-two hours is considered to be adequate to ensure completion of GI
297 transit of the drug product and absorption of the drug substance.

298 **2.1.8.3 Early Exposure**

299 For orally administered IR drug products, BE can generally be demonstrated by measurement of
300 rate and extent of absorption, i.e., C_{max} and $AUC_{(0-t)}$. However, in some situations, C_{max} and
301 $AUC_{(0-t)}$ may be insufficient to adequately assess the BE between two products, e.g., when the
302 early onset of action is clinically relevant. In these cases, an additional PK parameter, such as area
303 under the concentration vs. time curve between two specific time points (pAUC), may be applied.
304 This pAUC is typically evaluated from the time of drug administration until a predetermined time-
305 point that is related to a clinically relevant pharmacodynamic measure. Samples should be spaced
306 such that the pAUC can be estimated accurately.

307 **2.2 Data Analysis for Non-Replicate Study Design**

308 **2.2.1 Considerations for the Bioequivalence Analysis Population**

309 It is imperative that all criteria for study subject inclusion into the BE analysis population be clearly
310 defined in the study protocol. Any exclusions from the BE analysis population should be
311 documented prior to bioanalytical analysis, e.g., subjects that are withdrawn from the study, have
312 protocol violations, or experience GI disturbances potentially affecting absorption.

313 **2.2.1.1 Removal of Data Due to Low Exposure**

314 BE studies are studies with a smaller number of subjects compared to other clinical trials. An
315 extreme value in the dataset can have a large impact on the outcome of the BE study. Although
316 statistical tests may identify extreme values in the PK variables, such data should not be removed
317 from the statistical analysis of BE studies solely on this basis. Data should only be removed from
318 the statistical analysis based on protocol violations which are contemporaneously documented. A
319 prospective plan should be included in the study protocol for removing data from the BE statistical
320 analysis.

321 An exception to the above can be made for a subject without measurable concentrations or only
322 very low concentrations following either comparator or test product administration. A subject is
323 considered to have very low concentrations if the AUC for that period is less than 5% of the
324 geometric mean AUC of the product in question, which should be calculated without inclusion of
325 data from the subject. These very low concentrations are considered the result of subject non-
326 compliance and should, as far as possible, be avoided by documenting mouth check of subjects
327 after administration of study medication to ensure the subjects have swallowed the drug product.
328 The exclusion of data for this reason will only be accepted in exceptional cases (in general with
329 no more than 1 subject in each study) and may bring the reliability of dose administration into
330 question.

331 Data from redosing studies are not considered evidence to support removal of extreme values from
332 the statistical analysis.

333 Note that all subject data should be submitted and potential extreme values flagged with
334 appropriate documentation as part of the application.

335 2.2.2 Presentation of Data**336 2.2.2.1 Concentration Time Data**

337 For both the test and comparator products, the drug concentration in a suitable biological fluid,
338 e.g., plasma, serum or blood, determined at each sampling time point should be tabulated for each
339 subject participating in the study, along with descriptive statistics. These data should be presented
340 on the original scale, i.e., as unadjusted, measured drug concentrations. Deviations from the
341 protocol, e.g., missed samples or samples with significant time deviation, should be clearly
342 identified. Drug concentrations in study samples should be measured in accordance with ICH M10,
343 *Bioanalytical Method Validation and Study Sample Analysis*.

344 Two concentration-time graphs (linear and log-linear) should be provided for both the test and
345 comparator products for each individual subject. In addition, two concentration-time graphs (linear
346 and log-linear) should be provided for both the test and comparator products for the mean drug
347 concentrations of all subjects. For the individual subject concentration-time graphs, the drug
348 concentrations should be plotted against time using the actual sampling times. For the mean
349 concentration-time graphs the drug concentrations should be plotted using the nominal sampling
350 times.

351 2.2.2.2 Pharmacokinetic Analysis

352 For single-dose studies, the following PK parameters should be tabulated for each subject-
353 formulation combination: 1) primary parameters for analysis: $AUC_{(0-t)}$, C_{max} , and, where
354 applicable, $pAUC$, and 2) additional parameters for analysis to assess the acceptability of the
355 bioequivalence study: $AUC_{(0-inf)}$, $AUC_{(0-t)}/AUC_{(0-inf)}$, T_{max} , k_{el} , and $t_{1/2}$. For single-dose studies,
356 $AUC_{(0-t)}$ should cover at least 80% of $AUC_{(0-inf)}$. If the $AUC_{(0-t)}/AUC_{(0-inf)}$ percentage is less than
357 80% in more than 20% of the observations, then the validity of the study may need to be discussed
358 in the submission. If the AUC is truncated at 72 hours for long half-life drugs, the primary AUC
359 parameter for analysis is $AUC_{(0-72h)}$ and the following additional parameters are not required:
360 $AUC_{(0-inf)}$, $AUC_{(0-t)}/AUC_{(0-inf)}$, k_{el} , and $t_{1/2}$.

361 Summary statistics to be reported include geometric mean, median, arithmetic mean, standard
362 deviation, coefficient of variation, number of observations, minimum, and maximum. Each

363 variable should be computed using the actual time of sampling for each concentration data point.
364 The non-compartmental methods used to derive the PK parameters from the raw data should be
365 reported, e.g., linear trapezoidal method for AUC and the number of data points of the terminal
366 log-linear phase used to estimate the terminal elimination rate constant (k_{el}).

367 For multiple-dose studies, applicants should document appropriate dosage administration and
368 sampling to demonstrate the attainment of steady-state. For steady-state studies, the following PK
369 parameters should be tabulated: 1) primary parameters for analysis: C_{maxSS} and $AUC_{(0-tauSS)}$, and 2)
370 additional parameters for analysis: C_{tauSS} , C_{minSS} , C_{avSS} , degree of fluctuation, swing, and T_{max} .

371 Any concentration reported as below the lower limit of quantification (LLOQ) should be treated
372 as zero in PK parameter calculations. Values below the LLOQ are to be omitted from the
373 calculation of K_{el} and $t_{1/2}$.

374 ***2.2.2.3 Potency Differences in Lots***

375 The results from the potency assay of the test and comparator products should be submitted and
376 the test product batch should be within 5% of the comparator product batch. In exceptional cases
377 where a comparator product batch with a measured drug content within 5% of a test product batch
378 cannot be obtained, a potency correction may be accepted with supporting justification, e.g.,
379 potency data from multiple lots of comparator product, pending market availability, and
380 considering the totality of evidence. If potency correction is to be used, this intention should be
381 pre-specified in the study protocol. Analysis should be provided for both uncorrected data and for
382 potency-corrected data. If the potency correction is justifiable, the applicable BE standards should
383 be met on potency-corrected data.

384 ***2.2.3 Statistical Analysis***

385 ***2.2.3.1 General Considerations***

386 The statistical analyses should include all data for all subjects who provide evaluable data for the
387 products being compared. Decisions made to exclude subjects from the BE analysis population,
388 e.g., due to incomplete sampling or protocol violation, should be documented at the end of the
389 clinical blood sampling portion of the study and prior to subject sample analysis. A study will not
390 be considered acceptable if there are fewer than 12 evaluable subjects for a crossover analysis or

391 for each treatment arm for a parallel analysis.

392 The assessment of BE is based on 90% confidence intervals for the geometric mean ratios
393 (test/comparator) for the primary PK parameters under consideration. This method is equivalent
394 to two one-sided t-tests with the null hypotheses of bioinequivalence at the 5% significance level.
395 The data should be transformed prior to analysis using a logarithmic transformation.

396 The model to be used for the analysis should be pre-specified in the study protocol. The statistical
397 analysis should take into account sources of variation that can be reasonably assumed to have an
398 effect on the response variable.

399 The report on the data analysis should be sufficiently detailed to enable the PK and the statistical
400 analyses to be repeated, e.g., data on actual time of blood sampling after dose, drug concentrations,
401 the values of the PK parameters for each subject in each period, and the randomisation scheme
402 should be provided.

403 ***2.2.3.2 Crossover Design Studies***

404 Conventional two-treatment, two-period, two-sequence randomised crossover design studies
405 should be analysed using an appropriate parametric method, e.g., ANOVA. The tables resulting
406 from such analyses including the appropriate statistical tests of all effects in the model should be
407 submitted, e.g., a summary of the testing of Sequence, Subject within Sequence, Period, and
408 Formulation effects should be presented. In general, the primary analyses should include all data
409 for all subjects who provide evaluable data for both the test and comparator products.

410 ***2.2.3.3 Carry-over***

411 A test for carry-over is not considered relevant and no decisions regarding the analysis, e.g.,
412 analysis of the first period only, should be made on the basis of such a test. In crossover studies,
413 the potential for carry-over can be directly addressed by examination of the pre-treatment plasma
414 concentrations in period 2 and beyond if applicable, e.g., period 3 in a 3-period study.

415 If there are subjects for whom the pre-dose concentration is greater than 5% of the C_{\max} value for
416 the subject in that period, then the pivotal statistical analysis should be performed excluding the
417 data from that subject.

418 2.2.3.4 Parallel Design Studies

419 The statistical analysis for parallel design studies should reflect independent samples.
420 Demographic characteristics or other relevant covariates known to affect the PK should be
421 balanced across groups, to the extent possible. The use of stratification in the randomisation
422 procedure based on a limited number of known relevant factors is therefore recommended. Those
423 factors are also recommended to be accounted for in the pre-defined primary statistical analysis.
424 Post hoc and data-driven adjustments are not acceptable for the primary statistical analysis.

425 2.2.3.5 Multi-Group Design Studies

426 Sample size requirements and/or study logistics may necessitate studies to be conducted with
427 groups of subjects. The BE study should be designed to minimise the group effect in the study.
428 The combination of multiple factors may complicate the designation of group.

429 BE should be determined based on the overall treatment effect in the whole study population. In
430 general, the assessment of BE in the whole study population should be done without including the
431 Group by Treatment interaction term in the model, but applicants may also use other pre-specified
432 models, as appropriate. However, the appropriateness of the statistical model should be evaluated
433 to account for the multi-group nature of the BE study. Applicants should evaluate potential for
434 heterogeneity of treatment effect across groups, i.e., Group by Treatment interaction. If the Group
435 by Treatment interaction is significant, this should be reported and the root cause of the Group by
436 Treatment interaction should be investigated to the extent possible. Substantial differences in the
437 treatment effect for PK parameters across groups should be evaluated. Further analysis and
438 interpretation may be warranted in case heterogeneity across groups is observed.

439 In multicentre BE studies, when there are very few subjects in some sites, these subjects may be
440 pooled into one group for consideration in the statistical analysis. Rules for pooling subjects into
441 one group should be pre-specified and a sensitivity analysis is recommended.

442 Statistical methods and models should be fully pre-specified. Data-driven post hoc analysis is
443 highly discouraged but could be considered only in very rare cases where a very robust scientific
444 justification is provided.

445 2.2.4 Bioequivalence Criteria

446 For the majority of drug products, the PK parameters to demonstrate BE include C_{\max} and $AUC_{(0-t)}$.

447 For drugs with a long elimination half-life, $AUC_{(0-72h)}$ may be used in place of $AUC_{(0-t)}$ (see Section
448 2.1.8.2). For drugs where it is clinically relevant to assess the early exposure or early onset of
449 action, an additional PK parameter, pAUC, may be used to establish BE (see Section 2.1.8.3).

450 The 90% confidence interval for the geometric mean ratio of these PK parameters used to establish
451 BE should lie within a range of 80.00 - 125.00%.

452 2.2.5 Multiple Comparator and Multiple Test Product Studies**453 2.2.5.1 Multiple Comparator Products**

454 It may be necessary to demonstrate BE between a test product and multiple comparator products
455 to meet requirements from multiple jurisdictions. In such case, including comparator products
456 from different regions in one trial is acceptable to streamline the BE demonstration by conducting
457 one single higher-order crossover BE study with multiple comparator products.

458 Although there are multiple comparator products tested, multiplicity correction, i.e., alpha
459 adjustment, is not needed because comparator products are considered independent and region-
460 specific. Decisions will be made independently about a test product relative to a single comparator
461 product within a single jurisdiction. It is preferred for the statistical analysis to only test two at a
462 time and not all at once, making pairwise comparison within the analysis.

463 It is possible that the results meet the BE acceptance criteria with one region-specific comparator
464 product but not meet BE acceptance criteria with the other region-specific comparator product. In
465 such case, BE is demonstrated with one comparator product and not demonstrated with the other
466 comparator product. The protocol should specify the main objectives of the study and which
467 comparisons are to be performed.

468 Complete study results from all comparisons performed should be included in the clinical study
469 report.

470 2.2.5.2 Multiple Test Products

471 It may be necessary to demonstrate BE between multiple test products and the comparator product,

472 e.g., in order to include different test formulations that may be required due to drug development
473 needs. To streamline the demonstration of BE, it is permitted to conduct one single crossover BE
474 study with multiple test products.

475 The need to apply multiplicity correction in pivotal trials depends on the underlying objectives of
476 the trial:

477 a) If the objective is to achieve BE for all test formulations versus the comparator product,
478 then no alpha adjustment is needed.

479 b) If the objective is to show BE for any of the test formulations, then multiplicity adjustment
480 may be needed.

481 The objective of the trial and method for multiplicity correction should be pre-specified in the
482 study protocol.

483 **3 SPECIFIC TOPICS**

484 **3.1 Endogenous Compounds**

485 As endogenous compounds are identical to the drug that is being administered it can be challenging
486 to determine the amount of drug released from the dosage form and absorbed for BE assessment.
487 Therefore, in most cases, it is important to measure the baseline endogenous concentrations in
488 biological matrices, e.g., blood, plasma, or urine, and subtract these concentrations from the total
489 concentrations measured from each subject after the drug product is administered.

490 When the endogenous concentrations are influenced by diet, restricting or standardising the dietary
491 intake of the substance before and during the study should be considered.

492 The exact method for baseline correction should be pre-specified and justified in the study
493 protocol. Multiple baseline endogenous concentrations should be measured from each subject in
494 the time period before administration of the study drug. The time-averaged baseline or time-
495 matched baseline concentrations are subtracted from post-dose concentrations for those subjects
496 in an appropriate manner consistent with the PK properties of the drug. For the time-averaged
497 method, either the mean or median value may be used.

498 Baseline concentrations should be determined for each period and baseline correction should be

499 period specific. It should be ensured that the washout period is of an adequate duration because
500 carry-over effects cannot be readily detected. If a baseline correction results in a negative
501 concentration value, the value should be set equal to zero.

502 PK and statistical analyses should be performed on both baseline uncorrected and baseline
503 corrected data. In general, determination of BE should be based on the baseline corrected data.

504 Alternatively, the need for baseline correction may be avoided by enrolling study subjects with
505 low or no production of the endogenous compounds.

506 **3.2 Other Immediate-Release Dosage Forms**

507 ***3.2.1 Orally Disintegrating Tablets***

508 Orally Disintegrating Tablets (ODTs) should be administered in BE studies according to the
509 comparator product labelling with regard to intake of water.

510 If the comparator product labelling states that the ODT can be taken with or without water, the test
511 and comparator products should be administered in the BE study without water, as this is
512 considered to be the more discriminating scenario. BE of the test and comparator ODT products
513 taken with water can then be inferred.

514 For new intended label use/instructions, e.g., ODT as an extension to another orally administered
515 IR drug product, BE studies may be conducted to determine whether the ODT is BE to the
516 comparator product. In this scenario, the ODT product should be administered according to its
517 intended labelling and compared with the comparator product administered as per its labelling.

518 If the new intended label use/instructions states that the ODT can be taken with or without water,
519 a 3-arm BE study is recommended to determine BE of the ODT administered with and without
520 water compared to the comparator product administered as per its labelling.

521 In studies evaluating ODTs without water, it is recommended to wet the mouth by swallowing a
522 small amount of water, e.g., 20 ml, directly before applying the ODT on the tongue. It is
523 recommended not to allow fluid intake earlier than 1 hour after administration.

524 Other oral formulations such as orodispersible films, buccal tablets or films, and sublingual tablets

525 may be handled in a similar way to that described above for ODTs.

526 **3.2.2 Chewable Tablets**

527 Chewable tablets should be administered in BE studies according to the comparator product
528 labelling with regard to intake of water.

529 If the comparator product labelling states that the chewable tablets can be taken with or without
530 water, the test and comparator products should be administered in the BE study without water, as
531 this is considered to be the more discriminating scenario. BE of the test and comparator chewable
532 tablet products taken with water can then be inferred.

533 For new intended label use/instructions, e.g., chewable tablets as an extension to another orally
534 administered IR drug product, BE studies may be conducted to determine whether the chewable
535 tablet is BE to the comparator product. In this scenario, the chewable tablet product should be
536 administered according to its intended labelling and compared with the comparator product
537 administered as per its labelling.

538 If the new intended label use/instructions state that the chewable tablets can be taken with or
539 without water, a 3-arm BE study is recommended to determine BE of the chewable tablets
540 administered with and without water compared to the comparator product administered as per its
541 labelling.

542 **3.2.3 Oral Suspensions**

543 For tablets, granules, and powders labelled as being only intended to be dispersed in a liquid before
544 administration as an oral suspension, BE studies should be conducted according to the comparator
545 product labelling.

546 For new intended label use/instructions (not included in the comparator product labelling), the test
547 product should be administered according to its intended labelling and compared with the
548 comparator product administered as per its labelling.

549 **3.3 Fixed Dose Combination**

550 The BE study design for fixed-dose combination products should follow the principles described
551 in this guideline. BE should be determined using a PK sampling scheme suitable for the

552 determination of the PK parameters of the individual components (drugs) and employing
553 bioanalytical methods validated for the determination of the individual drugs in the presence of
554 the other component(s) in the combination product. PK parameters to be assessed and reported are
555 those that would normally be required for each drug if it were in the formulation as a single entity.
556 BE should be demonstrated for all components (drugs) in the fixed-dose combination product
557 according to the principles described in this guideline. Failure to demonstrate BE for one
558 component of the fixed-dose combination results in failure to demonstrate BE for the proposed
559 fixed-dose combination product as a whole.

560 **3.4 pH-Dependency**

561 The absorption of drug substances with pH-dependent solubility may be influenced by the gastric
562 pH. This impact on drug absorption can be altered due to the use of, for instance, pH stabilising
563 excipients or a specific salt-form in the formulation. Moreover, the formulation of the final
564 marketed comparator product may be the result of an extensive formulation development program,
565 obtaining for instance a specific formulation without an effect on drug absorption due to gastric
566 pH differences. This is especially relevant in cases where it is foreseen that the product will be
567 taken with acid reducing drug products, e.g., proton pump inhibitors, or is going to be used in
568 certain populations, e.g., patients with achlorhydria. Therefore when, relative to the comparator
569 product, there are qualitative or quantitative differences in the pH stabilising excipient(s),
570 significant differences in manufacturing process, or differences in salt form that possess a different
571 pH dependent solubility, BE under normal fasting conditions between the two products may not
572 ensure BE of the two products in a gastric pH-altered situation, e.g., in the presence of a pH-
573 modifying drug product. In such a situation, an additional BE study with concomitant treatment of
574 a pH-modifying drug product would generally be necessary to demonstrate BE.

575 Applicants may provide a scientific justification to demonstrate that a BE study in a gastric pH-
576 altered situation may not be needed. Such a justification should be based on the totality of evidence
577 referring to the pH-solubility profile of the drug substance, impact of excipients, formulation and
578 manufacturing design, e.g., formulation designed to overcome pH effects, extent of the differences
579 between the test and comparator products, and comparative dissolution testing at multiple pHs.
580 Modelling, e.g., appropriately validated/qualified PBPK modelling or semi-mechanistic
581 absorption models, and virtual BE simulation may be used to further assess the risk of

582 bioinequivalence.

583 **4 DOCUMENTATION**

584 The report of the BE study should include the complete documentation of its protocol, conduct,
585 and evaluation. It should be written in accordance with ICH E3, *Structure and Content of Clinical*
586 *Study Reports*.

587 Names and affiliations of the responsible investigator(s), the site of the study, and the period of its
588 execution should be stated.

589 Listing of inspection history for BE studies conducted at the relevant clinical and bioanalytical
590 site(s) for the 5 years preceding completion of the study should also be provided in the study report
591 (but may alternatively be provided elsewhere in the Common Technical Document (CTD)).

592 Comparator product name, strength, pharmaceutical form, batch number, manufacturer, expiration
593 date, and country of purchase should be stated.

594 Certificates of analysis, or equivalent documents, of comparator and test batches used in the study
595 should be included in an appendix to the study report.

596 The identity of the of the test product(s) used in the study should be provided, i.e., pharmaceutical
597 form, strength, batch number, and measured content (% of label claim). The batch size,
598 manufacturing date (and, if available, the expiry date) as well as the qualitative and quantitative
599 composition of the test product should also be indicated (but may alternatively be provided
600 elsewhere in the CTD).

601 Concentrations and PK data and statistical analyses should be presented in the level of detail
602 described in this guideline (see Section 2.2). The reporting format should include tabular and
603 graphical presentations showing individual and mean results and summary statistics.

604 Information on bioanalytical method validation and study sample analysis according to ICH M10
605 should be included in the appropriate section of Module 5 of the CTD.

606 The data generated should be properly documented and available for audit and inspection.

607 Essential documents should be archived in accordance with ICH E6 and applicable regulatory
608 requirements.

609 Data in a suitable electronic format should be submitted to enable the PK and the statistical
610 analyses to be repeated, e.g., data on actual times of blood sampling, drug concentrations, the
611 values of the PK parameters for each subject in each period, and the randomisation scheme.

612 Module 2.7.1 of the CTD should list all relevant BE studies conducted regardless of the study
613 outcome. Full study reports should be provided for the BE study(ies) upon which the applicant
614 relies for approval. For all other studies, synopses of the study reports (in accordance with ICH
615 E3) are sufficient. However, complete study reports for these studies should be available upon
616 request.

617 **5 GLOSSARY**

618 **Applicant:**

619 The entity submitting the application for marketing authorisation to the relevant regulatory
620 authority.

621 **AUC:**

622 Area under the concentration vs. time curve

623 **AUC_(0-inf):**

624 Area under the concentration vs. time curve extrapolated to infinity

625 **AUC_(0-t):**

626 Area under the concentration vs. time curve from time zero to the time of last quantifiable
627 concentration

628 **AUC_(0-tauSS):**

629 Area under the concentration vs. time curve for one dosing interval at steady state

630 **AUC_(0-72h):**

631 Area under the concentration vs. time curve from time 0 to 72 hours

632 **Batch (or Lot):**

633 A specific quantity of material produced in a process or series of processes so that it is expected to
634 be homogeneous within specified limits. In the case of continuous production, a batch may
635 correspond to a defined fraction of the production. The batch size can be defined either by a fixed
636 quantity or by the amount produced in a fixed time interval.

637 **Batch Number (or Lot Number):**

638 A unique combination of numbers, letters, and/or symbols that identifies a batch (or lot) and from
639 which the production and distribution history can be determined.

640 **C_{avSS}:**

641 Average concentration observed during dosing interval at steady state ($AUC_{0-\tau}/\tau$)

642 **Chewable Tablets:**

643 An oral dosage form designed to facilitate chewing and swallowing by the patient rather than
644 swallowing a whole tablet. They must be chewed or crushed before swallowing.

645 **C_{max}:**

646 Maximum concentration after dosing

647 **C_{maxSS}:**

648 Maximum concentration observed during dosing interval at steady state

649 **C_{minSS}:**

650 Minimum concentration observed during dosing interval at steady state

651 **Comparator (Product):**

652 An investigational or marketed product, i.e., active control, or placebo, used as a reference in a
653 clinical trial. In the context of this guidance, a comparator product is the drug product accepted by
654 regulatory agencies that an applicant can use to compare against the test product in conducting a
655 BE study.

656 **C_{tau}:**

657 Concentration observed at end of dosing interval

658 **C_{tauSS}:**

659 Concentration observed at end of dosing interval at steady state

660 **Enantiomers:**

661 Compounds with the same molecular formula as the drug substance, which differ in the spatial
662 arrangement of atoms within the molecule and are nonsuperimposable mirror images.

663 **Endogenous Compounds:**

664 Compounds already present in the body either because the body produces them or because they
665 are present in a normal diet.

666 **Fluctuation:**

667 $[(C_{\max SS} - C_{\min SS}) / C_{\text{avSS}}]$

668 **Immediate-Release:**

669 Allows the drug to dissolve in the GI contents, with no intention of delaying or prolonging the
670 dissolution or absorption of the drug.

671 **kcal:**

672 A unit used to describe amount of energy in relation to food or energy burned with exercise. When
673 it comes to nutrition and exercise, kilocalories (kcal) and calories equal the same amount of energy.
674 One kcal (kilocalorie) equals 1 calorie in nutrition.

675 **k_{el} :**

676 The apparent terminal elimination rate constant of the drug.

677 **Orally Disintegrating Tablet:**

678 A solid dosage form which is designed to disintegrate and dissolve rapidly on contact with saliva
679 when placed on the tongue or in the oral cavity, thus eliminating the need to chew the tablet,
680 swallow an intact tablet, or take the tablet with water.

681 **pAUC:**

682 Area under the concentration vs. time curve between two specific time points

683 **Protocol:**

684 A document that describes the objective(s), design, methodology, statistical considerations, and
685 organisation of a trial. The protocol usually also gives the background and rationale for the trial,
686 but these could be provided in other protocol referenced documents. Throughout ICH E6, *Good*
687 *Clinical Practice*, the term protocol refers to protocol and protocol amendments.

688 **Racemate:**

689 A composite (solid, liquid, gaseous, or in solution) of equimolar quantities of two enantiomeric
690 species. It is devoid of optical activity.

691 **Spare Subject:**

692 A study subject that is included in the drug administration and sample collection regimens of a
693 study but, as per study protocol, whose data will only be included in the PK and statistical analysis
694 if the number of evaluable study subjects drops below a pre-specified number due to subject
695 dropouts and/or withdrawals (use of spare subjects is not acceptable).

696 **Sponsor:**

697 An individual, company, institution, or organisation which takes responsibility for the initiation,
698 management, and/or financing of a clinical trial.

699 **Swing:**

700 $[(C_{\max SS} - C_{\min SS}) / C_{\min SS}]$

701 **Tau:**

702 Dosing Interval

703 **T_{max}:**

704 Time to maximum observed concentration

705 **t_{1/2}:**

706 Half-life