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|---|---------------------------------|--|--|-----------------------------------|
| ANALYST WORKSHEET | | 1. PRODUCT Bacterial culture(s) from <i>Dog Food</i> | | 2. SAMPLE NUMBER 915750 |
| 3. SEALS <input checked="" type="checkbox"/> INTACT <input type="checkbox"/> NONE <input type="checkbox"/> BROKEN | 4. DATE REC'D 7/31/15 | 5. RECEIVED FROM <i>William K. Wilkes Sample Room Custodian</i> | 6. DISTRICT OR LABORATORY Arkansas Regional Lab | |

7. DESCRIPTION OF SAMPLE

One cardboard isolate shipping carton containing one plastic isolate shipping canister. Canister contained one *shirtpak* bag
 FDA sealed " **915750 7/29/15 Jennifer Canale - Microbiologist**".
 Bag contained **1** bacterial culture(s) on agar slant(s) in culture tube(s) id. " **915750 Comp#1 HERV isolate -
JRC 7/29/15**".

Sample received in good condition at room temperature.

| | | | |
|---|---|-------------|---|
| 8. NET CONTENTS <input checked="" type="checkbox"/> NOT APPLICABLE <input type="checkbox"/> NOT DETERMINED <input type="checkbox"/> UNITS EXAMINED | DECLARE / UNIT _____ AMOUNT FOUND _____ % OF DECLARED _____ | 9. LABELING | <input type="checkbox"/> ORIGINAL(S) SUBMITTED <input type="checkbox"/> COPIES SUBMITTED <input checked="" type="checkbox"/> NONE |
|---|---|-------------|---|

10. SUMMARY OF ANALYSIS

DATE STARTED: **8/4/15**CONTAINER: **16X125 slip-cup** culture tube(s).

LABELING: Not applicable.

CODE: None.

PRODUCT: Bacterial culture(s) on agar slant(s)

ANALYSIS: **1** culture(s) examined for *Salmonella* biochemical and serological identification.

Lab positive and negative controls also examined - Controls for biochemical testing: Positive control(s) = known *Salmonella* culture(s). Negative controls = media controls. Controls for serological testing:

Positive controls = commercial and in-house prepared QC *Salmonella* antigens - (contain antigens homologous to antisera being tested)
 Negative controls = 1. Culture and saline / formalinized saline control. 2. QC antigen and saline / formalinized saline control

METHOD: See page 2.

RESULTS:

The culture(s) submitted from this sample was / were determined to be: **915750 S. Monophasic GR B**

Positive controls positive. Negative controls negative.

11. RESERVE SAMPLE :

1 Reserve culture(s) placed in -90°C (locked ARL freezer - RT10333) in TST broth with 20% glycerol in cryogenic vial(s) identified: **915750 SLE 8/10/15 S. Monophasic GR B** on (date) **8/10/15** for long term storage. **1** bacterial culture(s) on BAB agar slant(s) in 13 x100 glass screw capped tube(s) id. **915750 SLE 8/10/15 S. Monophasic GR B** placed into *shirtpak* bag that is FDA sealed" **915750 8/10/15 Stephanie Horton**". Tube(s)

Sealed bag will be placed into isolate shipping canister and carton and sent to Denver District Lab for antibiotic resistance testing. Official seal submitted as "Attachment A" with worksheets.

| | | |
|---|-------------------------------------|--|
| 12 a. ANALYST SIGNATURE (broke seal ✓) <i>Stephanie Horton</i> | 13. WORKSHEET CHECK b. DATE | BY <i>Stephanie C. Anderson</i> b. DATE 8/10/15 |
| b. <i>Stephanie Horton</i> | | |
| c. | 14. DATE REPORTED 8/10/15 | |

| | | |
|---|--|---------------------------------|
| GENERAL CONTINUATION SHEET <i>Salmonella Serotyping Protocol</i> | PRODUCT - Bacterial cultures from : <i>Dog Food</i> | SAMPLE NO. <i>915750</i> |
| <p>Initial Set up</p> <p>1. Emulsify one loopful of culture from agar slant as received in 0.5 ml sterile 0.85% NaCl in a sterile 13 x 100 mm tube. Streak an XLD agar plate from emulsion. Inoculate TSI, MTM, LIA and Tryptose agar slants, from original slant. Incubate 18-24 hours at 35-37°C. Read XLD for purity and appearance. If observation of XLD plate appears to be mixed, pick one colony of each type and re-inoculate TSI, MTM, LIA and Tryptose agar slant</p> <p>2. Read TSI, MTM and LIA tubes and slants, record results and use growth from TSI slant diluted in 0.85% NACL to inoculate the API 20E strip. Use Tryptose agar slant for somatic (O) antigen testing. Use TST culture for flagellar (H) antigen testing.</p> <p>3. Serological Identification --Somatic Antigens Live Antigens are used for Somatic (O) serology. (caution must be exercised) Using a glass slide which has been prepared to withstand antisera, antigen and 0.85% NaCl. Growth is taken from the tryptose agar slant emulsified into one drop of 0.85% NaCl to which a specific drop of antisera is added to the appropriate well, rocking back and forth approximately one minute to observe for agglutination. First testing the O polyvalent antisera, then the groups, then the factors.</p> <p>4. Serological Identification — Flagellar (H) Antigens <ul style="list-style-type: none"> A. Use tube agglutination test explained below in steps 4.C. through step 4.E. for B-D/ Difco , SA Scientific, and California Department of Health Services antisera. B. Statens Serum Institute H antisera will be used in a slide agglutination technique as described above in #3, in the titre as received with inoculum from Tryptose agar slant. C. Dilute 10 ml TST culture with an equal volume of 0.6% formalinized saline. D. Perform first portion of Flagellar (H) antigen analysis using this suspension. Combine H antisera (diluted according to manufacturers' and CDC's Guidelines) and formalinized TST culture in a 10 x 75 mm tube. E. Incubate tube in 48-50°C bath up to one hour. Check for agglutination every 15 minutes. First test using the H polyvalent antisera and then the complex and single factor antisera. </p> <p>5. Phase Reversal of H Antigens <ul style="list-style-type: none"> A. Place one droop of concentrated H antisera, homologous to H antigen identified via step D above in a sterile 50 x 9 mm or a 50 x 11 mm Petri dish. Add 5 ml semi-solid motility test medium, mix thoroughly and allow to solidify. (This plate is known as a Gard plate.) B. Transfer small amount of growth from TSI slant to edge of Gard plate, and incubate at 35-37°C in upright position for at least 24 hours or until enough growth away form the point of inoculation is observed. C. Transfer one loopful from leading edge of motile growth on Gard plate to TST broth, and incubate 18 – 24 hours at 35 - 37°C. D. Perform second portion of Flagellar (H) antigen analysis following steps 4.A.. through 4.E. above. For Statens Serum Institute H antisera, use the Gard plate growth as source of the sample antigen for testing. </p> <p>6. Additional Biochemical Testing Additional biochemical tests such as malonate, Jordan's Tartrate, KCN broth , etc. may be used if needed to determine the subspecies and / or biotype of any <i>Salmonella</i> isolate. They are incubated at 35 – 37°C for 48 hours.</p> <p>Method References: <ol style="list-style-type: none"> 1. <u>Antigenic Formulas of the <i>Salmonella</i> Serovars</u>, 9th Edition, 2007 Patrick A.D. Grimont and Francois-Xavier Weill 2. <u>AOAC</u>, 19th Edition , 2012, Chapter 17, 3. <u>BAM</u>, Online Edition, updated May 2014 ; <i>Salmonella</i>, Chapter 5 4. <u>Identification of Enterobacteriaceae</u>, Chapter 9, The Genus <i>Salmonella</i>, P. Edwards and W. Ewing, 4th Ed., 1986 5. <u>Identification and Serotyping of <i>Salmonella</i></u>, F. Brenner and A. McWorter-Murlin, CDC, National Salmonella Reference Laboratory,1998. 6. <u>Modified Kauffmann-White Scheme</u>, Updated 1998, F. Brenner, CDC, National Salmonella Reference Laboratory. 7. Manufacturer's package inserts for <i>Salmonella</i> antisera & QC antigens from: Difco Laboratories, Statens Serum Institute, and Centers for Disease Control (CDC). </p> <p>NOTES:</p> <p><i>Stephanie Horton</i></p> | | |
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|---|-----------------------|------------------|-------------|---|-----------------------|------------------|-------------|-------------|------------------|-------------|-------------------|-------------|-------------|----------------------|----------------------------|-------------|-------------|-------------|-------------|-------------|-------------|
| GENERAL CONTINUATION SHEET | | | | PRODUCT Bacterial cultures from : Dog Food | | | | | | | | | | SAMPLE NO. 915750 | | | | | | | |
| BIOCHEMICAL IDENTIFICATION RESULTS | | | | | | | | | | | | | | | | | | | | | |
| INITIALS + DATE SLB 8/4/15 | SLB 8/5/15 8/6/15 | | | | | | | | | | | | | | | | | | | | |
| | BASIC TESTS | | | | | | | | ADDITIONAL TESTS | | | | | | | | | | | | |
| ISOLATE IDENT. # 1 | XLD | | TSI | | | | LIA | | | | Motility | Malonate | Mucate | Jordan's Tartrate | Phenol Red Broth Base with | | | | | | |
| | S L A N T | B U T T | H 2 S | G A S | S L A N T | B U T T | H 2 S | G A S | | | | | | | | | | | | | |
| 915750 | T | K | A | ++ | K | K | + | - | + | | | | | | | | | | | | |
| Media Control | NG | | | | | | | | | | | | | | | | | | | | |
| Positive Control | T | K | A | ++ | K | K | + | - | + | | | | | | | | | | | | |
| T = typical AT = atypical NG = no growth K = alkaline A = acid YB = yellow with black centers NB = no black centers | | | | | | | | | | | | | | | | | | | | | |
| API 20E Isolate Id. 915750 Initials: SLB | ON PG H | A D C | L D C | O D C | C I T | H 2 S | U R E | T D A | I N D | V P I | G E L | G L U | M A N | I N O | S O R | R H A | S A C | M E L | A M Y | A R A | O X I |
| 24 HR. Date: 8/7/15 | - | + | + | + | + | - | - | - | - | - | - | - | + | + | + | + | - | - | - | - | |
| Profile Number | 6 | 7 | 0 | 0 | 4 | 7 | 5 | 2 | | | | | | | | | | | | | |
| API 20E IDENTIFICATION FOR ISOLATE 915750 <i>Salmonella spp.</i> Ex- I.D. | | | | | | | | | | | | | | | | | | | | | |
| API 20E Isolate Id. _____ Initials: | ON PG H | A D C | L D C | O D C | C I T | H 2 S | U R E | T D A | I N D | V P I | G E L | G L U | M A N | I N O | S O R | R H A | S A C | M E L | A M Y | A R A | O X I |
| 24 HR. Date: | - | - | - | - | - | - | - | - | - | - | - | - | - | - | - | - | - | - | - | | |
| Profile Number | 6 | 7 | 0 | 0 | 4 | 7 | 5 | 2 | | | | | | | | | | | | | |
| API 20E IDENTIFICATION FOR ISOLATE : | | | | | | | | | | | | | | | | | | | | | |
| API 20E Isolate Id. _____ Media Control Initials: SLB | ON PG H | A D C | L D C | O D C | C I T | H 2 S | U R E | T D A | I N D | V P I | G E L | G L U | M A N | I N O | S O R | R H A | S A C | M E L | A M Y | A R A | O X I |
| 24 HR. Date: 8/7/15 | - | - | - | - | - | - | - | - | - | - | - | - | - | - | - | - | - | - | - | | |
| Profile Number | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | | |
| API 20E IDENTIFICATION FOR ISOLATE Media Control .85% NaCl: Negatino | | | | | | | | | | | | | | | | | | | | | |
| API 20E Isolate Id. _____ Positive Control. _____ Initials: SLB | ON PG H | A D C | L D C | O D C | C I T | H 2 S | U R E | T D A | I N D | V P I | G E L | G L U | M A N | I N O | S O R | R H A | S A C | M E L | A M Y | A R A | O X I |
| 24 HR. Date: 8/7/15 | - | + | + | + | + | - | - | - | - | - | - | - | + | + | + | + | - | - | - | | |
| Profile Number | 6 | 7 | 0 | 0 | 4 | 7 | 5 | 2 | | | | | | | | | | | | | |
| API 20E IDENTIFICATION FOR ISOLATE Positive Control S. Gaminara SEA 2575 <i>Salmonella spp.</i> Ex- I.D. | | | | | | | | | | | | | | | | | | | | | |
| ANALYST(S) Stephanie Horton | | | | | | | | | | | PAGE 3 OF 6 PAGES | | | | | | | | | | |

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|---|---------|---|---|---|---|-------------|---|-----------|--|--|--|----------------------|--|---|--------------------------|--------------|-----|-----------|-------|---|---|--|---|----|--|
| GENERAL CONTINUATION SHEET | | PRODUCT (Bacterial cultures from): <i>Dog Food</i> | | | | | | | | | | SAMPLE NO. 915750 | | | | | | | | | | | | | |
| SEROLOGICAL IDENTIFICATION RESULTS | | | | | | | | | | | | | | | | | | | | | | | | | |
| + = agglutination in tube & slide tests. - = no agglutination NA = Not Available W = Weak reaction gr = group | | | | | | | | | | | | | | | | | | | | | | | | | |
| SOMATIC (O) ANTIGEN DETERMINATION | | | | | | | | | | | | | | | | | | | | | | | | | |
| INITIALS/ DATE | | S16 8/6/15 | | | | | | | | | | | | | | | | | | | | | | | |
| ISOLATE # / ID. | O POLYS | | | | | | | O GROUPS | | | | | | | O FACTORS | | | | | | | | | | |
| | A | B | C | D | E | F | G | | | | | | | | B | | | | | | | | 4 | 5 | |
| 915750 | + | | | | | | | | | | | | | | + | | | | | | | | | ++ | |
| QC antigen + controls | + | | | | | | | | | | | | | | + | | | | | | | | | ++ | |
| Types of QC antigens | B | | | | | | | | | | | | | | B | | | | | | | | | BB | |
| | | | | | | | | | | | | | | Culture & 0.85% NaCl | | | | | | | | | | | |
| | | | | | | | | | | | | | | Negative control | | | | | | | | | | | |
| | | | | | | | | | | | | | | — | | | | | | | | | | | |
| | | | | | | | | | | | | | | Antigens & 0.85% NaCl | | | | | | | | | | | |
| FLAGELLAR H ANTIGEN DETERMINATION - PHASE (circled) | | | | | | | | | | | | | | 1 | 2 | both 1 and 2 | | | | | | | | | |
| INITIALS/ DATE | | S16 8/6/15 | | | | | | | | | | | | | | | | | | | | | | | |
| ISOLATE # / ID. | H POLYS | | | | | H COMPLEXES | | H FACTORS | | | | | | | | | | | | | | | | | |
| | A | B | C | D | E | | | | | | | | | | b | d | i | 2 | o | 2 | g | | | | |
| 915750 | +- | - | | | | | | | | | | | | * | - | + | - | - | | | | | | | |
| QC antigen + controls | +++ | + | | | | | | | | | | | | +++ | +++ | +++ | +++ | +++ | | | | | | | |
| Types of QC antigens | dehr | R | | | | | | | | | | | | b | d | i | 2 | o | 2 | g | | | | | |
| | | | | | | | | | | | | | | Culture & Formalinized 0.85% NaCl | | | | | | | | | | | |
| | | | | | | | | | | | | | | Negative control | | | | | | | | | | | |
| | | | | | | | | | | | | | | — | | | | | | | | | | | |
| | | | | | | | | | | | | | | Antigens & form. saline | | | | | | | | | | | |
| FLAGELLAR H ANTIGEN DETERMINATION - PHASE (circled) : 1 | | | | | | | | | | | | | | 2 | No 2 nd phase | | | | | | | | | | |
| INITIALS/ DATE | | S16 8/9/15 | | | | | | | | | | | | | | | | | | | | | | | |
| ISOLATE # / ID. | H POLYS | | | | | H COMPLEXES | | H FACTORS | | | | | | | | | | | | | | | | | |
| | A | B | C | D | E | | | | | | | | | | b | d | i | 2 | o | 2 | g | | | | |
| 915750 | +- | - | | | | | | | | | | | | - | - | + | - | - | | | | | | | |
| QC antigen + controls | +++ | + | | | | | | | | | | | | +++ | +++ | +++ | +++ | +++ | | | | | | | |
| Types of QC antigens | dehr | R | | | | | | | | | | | | b | d | i | 2 | o | 2 | g | | | | | |
| | | | | | | | | | | | | | | Culture & Formalinized 0.85% NaCl | | | | | | | | | | | |
| | | | | | | | | | | | | | | Negative control | | | | | | | | | | | |
| | | | | | | | | | | | | | | — | | | | | | | | | | | |
| | | | | | | | | | | | | | | Antigens & form. saline | | | | | | | | | | | |
| SEROTYPE FOR ISOLATE # / ID. | | 915750 S. Monophasic R. B | | | | | | | | | | | | 4,5:- | | | | | | | | | | | |
| SEROTYPE FOR ISOLATE # / ID. | | | | | | | | | | | | | | | | | | | | | | | | | |
| SEROTYPE FOR ISOLATE # / ID. | | | | | | | | | | | | | | | | | | | | | | | | | |
| SEROTYPE FOR ISOLATE # / ID. | | | | | | | | | | | | | | | | | | | | | | | | | |
| NEGATIVE CONTROLS RESULTS: <i>Negative</i> | | | | | | | | | | | | | | See page 1 and above for types of - controls. | | | | | | | | | | | |
| POSITIVE CONTROLS (QC ANTIGENS) RESULTS: <i>positive</i> | | | | | | | | | | | | | | | | | | | | | | | | | |
| ANALYST(S): <i>Stephanie Horton</i> | | | | | | | | | | | | | | PAGE | | 4 | OF | 6 | PAGES | | | | | | |
| Modified FORM FDA 431a | | | | | | | | | | | | | | Version 1.0 (4/2004) | | | | ARL-SER.1 | | | | | | | |

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|---|---|--|-------------------------|---------------------------------|
| Continuation Sheet | Product Bacterial Culture(s) From: <i>Dog Food</i> | | | Sample number <i>915750</i> |
| SALMONELLA MEDIA AND EQUIPMENT LOT RECORD FOR SEROTYPING | | | | |
| | Lot # | | | Lot # |
| XLD Agar (AOAC 967.25 A (d)1/2 | <i>150727-3</i> | Tryptose BAB (BAM M-166) | | <i>150624-6</i> |
| TSI (AOAC 967.25 A (g) 1/2) | <i>150623-11</i> | TST Broth (AOAC 967.25 A (s)) | | <i>150615-8</i> |
| LIA (AOAC 967.25A (m)1) | <i>150627-3</i> | Physiologic Saline (AOAC 967.25 B (f)) | | <i>150429-4</i> |
| BHI Broth | <i>150721-8</i> | Motility Test Med. (AOAC 967.25 A (n)) | | <i>150618-10</i> |
| | Lot # | | Mfr | Lot # |
| Formalized Saline (AOAC 967.25 B (g)) 0.85% NaCl | <i>150429-4</i> | w/0.6% formaldehyde | Fisher | <i>148493</i> |
| Formalized Saline (AOAC 967.25 B (g)) 0.85% NaCl | | w/0.6% formaldehyde | Baker | |
| Phenolized Saline (CDC 1998) 0.85% NaCl | <i>150429-4</i> | w/0.55 % phenol | Fisher | <i>99431</i> |
| | Lot # | | Mfr: | |
| Sterile Mineral Oil (BAM R-46) | <i>150336-2 Exp. 9-29-15</i> | | ARL | |
| API 2OE (AOAC 978.24 A (a)) | <i>1003599990 Cap. 2016-04-20</i> | | BioMerieux | |
| Ferric Chloride (BAM R-25) | <i>536564 Exp. 2016/04/27</i> | | Remel | |
| Kovacs' reagent (AOAC 967.25 B (a) | <i>519163 Exp. 2015/12/18</i> | | Remel | |
| α -Naphthol (Voges-Proskauer)(AOAC 967.25 B (b1)) | <i>150219-10 Exp. 2-19-16</i> | | ARL | |
| 40% KOH(Voges-Proskauer) (AOAC 967.25 B (b2)) | <i>493223 Exp. 2015/02/1</i> | | Remel | |
| Oxidase Reagent(BAM R-54) | <i>493788 Cap. 2015/02/25</i> | | Remel | |
| 80 % Glycerol | <i>150409-2 Cap. 4-9-2016</i> | | ARL | |
| HE Agar (AOAC 967.25A (e)1/2) | | | ARL | |
| Lysine Dec. broth (AOAC 967.25A (m)1) | | | ARL | |
| Phenol Red Broth with Dulcitol | | | Remel | |
| Phenol Red Broth with Lactose | | | Remel | |
| Phenol Red Broth with Salicin | | | Remel | |
| Phenol Red Broth with | | | | |
| Jordan's Tartrate (Edwards & Ewing, 1986) | | | Remel | |
| Mucate medium (Edwards & Ewing, 1986) | | | Remel | |
| Malonate Broth (AOAC 967.25A (L)) | | | Remel | |
| | | | | |
| Supplies | | | | |
| Sterile tubes: 13 X100 | <i>150601-9</i> | 16 x150 | 20 x150 | |
| | Mfr. | Lot # | | Mfr Lot # |
| Disposable culture tubes 10 x 75mm | <i>Fisherbrand 14258499</i> | | Sterile Cryovials | Wheaton <i>1302000572</i> |
| Sterile 1 ml Pipets | <i>Carter 16314084</i> | | Sterile 5 ml Pipets | |
| Sterile 10 ml Pipets | <i>1 28314001</i> | | | |
| Sterile Transfer pipets | <i>Fisherbrand 14447961</i> | | Sterile Transfer pipets | |
| Sterile 50 x 9 mm Petri dishes | | | Sterile swabs | |
| Sterile micropipettor tips | | | | |
| Equipment | | | | |
| Pipettors: Mfr: Eppendorf Size: 2 – 20 μ l #266750, Size: 10 – 100 μ l #3392601, Mfr: Oxford Size: 100 – 1000 μ l A94018452 Mfr: Drummond FDA #: RT11056 | | | | |
| 35°C Incubator: Forma Scientific FDA # 5022432 | | 50°C Water Bath : Thermo Scientific FDA #ARL00421 | | |
| Refrigerators GE FDA #178799 Westinghouse FDA # 182538 NC21263 | | Freezers (-90°C): Harris FDA #RT10333 & FDA #RT10866 | | |
| <u>Control Cultures:</u> | | | | |
| <input checked="" type="checkbox"/> <i>S. Gaminara</i> (Nalidixic ac.R.) SEA 2575 , | <input type="checkbox"/> | <i>E. aerogenes</i> ATCC # 13048 | | |
| <input type="checkbox"/> <i>S. subsp. diarizonae</i> ATCC# 29934 | <input type="checkbox"/> | | | |
| Analyst(s) | | | | Page <i>5</i> of <i>6</i> Pages |
| <i>Daphne Haton</i> | | | | |

| SALMONELLA ANTISERA and QC ANTIGEN LOT RECORD | | | PRODUCT: Bacterial culture(s) from: <i>Dog Food</i> | | | | | | SAMPLE NO. 915750 | |
|---|---------------|-------------------------|--|------------|---------------|-------------------------|-----|---------------|---------------------------|-----|
| O Antisera | Mfr & Lot # | rehydrated or diluted w | I N | H Antisera | Mfr & Lot # | rehydrated or diluted w | I N | H QC Antigens | Mfr & Lot # | I N |
| POLY A-I | Disco 6250603 | 14305-2* | | POLY A | Disco 3283047 | 150203-2* | | a | SAS 2122000 | |
| POLY A | SSI 5790-H1 | None | B62 | POLY B | Disco 4150864 | 141031-7* | | b | ADH S. java | |
| POLY B | SSI 580U-H1 | None | | POLY C | Disco 212446 | 130118-8* | | c | SAS 1091107 | |
| POLY C | SSI 581K-H14 | None | | POLY D | Disco 8253506 | 090217-5* | | d | SEA 2575 S. gaminara ± | |
| POLY D | Disco 1139043 | 140302-2* | | POLY E | Disco 3074210 | 141001-11* | | i | ATCC 14028 S. typhimurium | |
| POLY E | Disco 4160781 | 141031-7* | | Factor b | Disco 1015570 | 130829-8* | | Z10 | ADH S. newport | |
| POLY F | Disco 8094697 | 080312* | | Factor d | Disco 1045569 | 140305-2* | | Z29 | CFSAN S. cubana | |
| POLY G | Disco 2168340 | 140305-2* | | Factor i | Disco 1016654 | 130118-8* | | e,h | ADH S. newport | |
| Group A | Disco 8233323 | 100301-5* | | Factor z10 | Disco 3081234 | 130701-5* | | e,n,x | ATCC 9842 S. abortus equi | |
| Group B | Disco 3018191 | 140626-2* | | Factor z29 | Disco 3024368 | 140305-2* | | g,f,s | ADH S. agona | |
| Group C1 | Disco 3039184 | 140201-2* | | Factor a | Disco 8189214 | 080715-2* | | g,p,u | CDC S. rostock | |
| Group C2 | Disco 2013546 | 140305-2* | | Factor c | Disco 9020038 | 100301-5* | | g,m | ATCC 13076 S. enteritidis | |
| Group D | Disco 2251456 | 140305-2* | | Factor e,h | Disco 1041636 | 131106-6* | | g,q | CDC S. moscow | |
| Group E | Disco 2307492 | 130701-5* | | Factor h | Disco 9139719 | 140505-2* | | g,t | ADH S. budapest | |
| Group F | Disco 3109446 | 140626-2 | | en complex | Disco 0343456 | 140305-2* | | l,w | ADH S. worthington | |
| Group G | SSI 8471-H5 | Non e | | Factor x | Disco 1224754 | 140722-14* | | l,v | ADH S. bredeney | |
| Group H | Disco 9023219 | 130701-5* | | Factor z15 | Disco 1104601 | 140722-14* | | l,z28 | ATCC 10721 S. javiana | |
| | Disco 6027328 | 130701-5* | | G complex | Disco 8345581 | 140305-2* | | y | ADH S. bariilly | |
| | Disco 8114181 | 130701-5* | | Factor f | Disco 1090901 | 150203-2* | | z | ADH S. worthington | |
| Group I | SSI 600D-H7 | None | | Factor m | Disco 3193286 | 150203-2* | | r | ADH S. heidelberg | |
| Group J | SSI 601C-H8 | None | | Factor s | Disco 1126077 | 121015-1* | | z4,z23 | ATCC 10723 S. cerro | |
| Group K | SSI 602G-H6 | None | | Factor t | Disco 1120676 | 130118-8* | | z4,z24 | SAS 2122001 | |
| Group L | Disco 8345583 | 100301-5* | | Factor p | Disco 9050648 | 100301-5* | | z4,z32 | CDC S. serotype IV | |
| Group M | Disco 8331700 | 100310-5* | | Factor q | SSI 694D-S3 | None | | 2 | ATCC 1408 S. typhimurium | |
| Group N | Disco 9162587 | 100310-5* | | | CDC 98-0070 | 081015-3p | | 5 | ADH S. bareilly | |
| Group O | Disco 9156682 | 100310-5* | | Factor u | SSI 697C-S8 | None | | 6 | CFSAN S. poona | |
| Factor 1 | SSI 478G-H3 | None | | | CDC 99-0017N | 081015-3p | | 7 | SEA 2575 S. gaminara ± | |
| Factor 2 | SSI 873B-H5 | None | | L complex | Disco 1115528 | 130118-8* | | z6 | ATCC 15786 S. hoagraven | |
| Factor 4 | SSI 876E-H11 | None | | | CDC 00-0032N | 060710-4p | | z36 | CDC S. serotype II | |
| Factor 5 | SSI 782G-H2 | None | | Factor l,v | SSI 736E-S16 | None | | z39 | SAS 3013100 | |
| Factor 6 | SSI 837K-H4 | None | | Factor v | SSI 698K-S1 | None | | z42 | ATCC 15786 S. hoagraven | |
| Factor 7 | SSI 634R-H2 | 130118-8* | | | CDC 05-0035 | | | Z53 | ATCC 12325 S. diarizonae | |
| Factor 8 | Disco 9260579 | 130118-8* | | Factor w | Disco 8155842 | 081015-3* | | e,n,z15 | ADH S. braenderup | |
| Factor 14 | SSI 638F-H3 | None | | | CDC 98-0048N | 060710-4p | | k | ADH S. thompson | |
| Factor 9 | Disco 0259974 | 140305-2* | | Factor z13 | CDC 02-0028N | 060710-4p | | Z13 | CDC S. uganda | |
| Factor 46 | SSI 648L-H1 | None | | Factor z28 | Disco 9321050 | 100707-4* | | O QC Antigen | Mfr & Lot # | IN |
| Factor 10 | SSI 637R-H3 | None | | | CDC 02-0029N | | | Group A | Disco 8287460 | |
| Factor 15 | SSI 639G-H6 | None | | z4 complex | Disco 1140410 | 130102-4* | | Group B | Disco 8287461 | |
| Factor 19 | SSI 640I-H10 | None | | Factor z23 | Disco 9015612 | 100301-5* | | Group C1 | Disco 9037635 | |
| Factor 34 | Disco 9247784 | 130701-5* | | Factor z24 | SSI 701B-24 | None | | Group C2 | Disco 8189358 | |
| Factor 20 | SSI 641K-H8 | None | | | CDC 99-0009N | | | Group D | Disco 9190853 | |
| Factor 22 | SSI 642I-H3 | None | | Factor z32 | Disco 9260610 | 100707-4* | | Group D2 | SAS 2090518 | |
| | Disco 6235063 | 14305-2* | | Factor k | Disco 2342196 | 150203-2* | | Group E1 | Disco 9188690 | |
| Factor 23 | SSI 643I-H1 | Non e | | Factor r | Disco 3323320 | 140312-9* | | Group E2 | Disco 9272758 | |
| | Disco 4068832 | 14305-2* | | Factor y | Disco 3081238 | 140305-2* | | Group E3 | SAS 3081222 | |
| Factor 24/25 | SSI 874C-H1 | None | | Factor z | Disco 1104607 | 120201-1* | | Group E4 | Disco 8207552 | |
| Factor 25 | SSI 645K-SH1 | None | | Factor 2 | Disco 3150186 | 140804-7* | | Group F | Disco 9230630 | |
| Factor 27 | SSI 646H-H2 | None | | Factor 5 | Disco 3354232 | 140804-7* | | Group G1 | Disco 9167947 | |
| Group P | CDC 79-0450 | 100301-5p | | Factor 6 | Disco 3060143 | 150203-2* | | Group H | Disco 8207253 | |
| Group Q | CDC 85-0031 | 100301-5p | | Factor 7 | Disco 1126100 | 130701-5* | | Group I | Disco 8205832 | |
| Group R | CDC 01-0188N | 100301-5p | | Factor z6 | Disco 1363654 | 140804-7* | | Group J/K | SAS 2080607 / NA | |
| Group S | CDC 85-0032 | 100301-5p | | H poly a-z | Disco 8052732 | 080312-3* | | Group L/M | SAS 3021825 / NA | |
| Group T | CDC 89-0147 | 100301-5p | | | | | | Group N/O | SAS 0030201 / NA | |
| Group U | CDC 79-0453 | 100301-5p | | | | | | Group P/Q | SAS 2111810 / SAS 1091308 | |
| Group V | CDC 87-0065 | 091228-3p | | | | | | Group R/S | SAS 3202714 / SAS3031005 | |
| Group W | CDC 96-0027N | 100210-5p | | | | | | Group T/U | SAS 3022715 / SAS 208608 | |
| Group X | CDC 82-0102 | 080715-2p | | | | | | Group V/W | NA / SAS 110903 | |
| Group Y | CDC 96-0050N | 080715-2p | | | | | | Group X/Y | SAS 9022304 / SAS8081808 | |
| Group Z | CDC 96-0049N | 080715-2p | | | | | | Group Z | SAS 9110100 | |

* = 0.85% sterile saline for rehydration of antisera , p = 0.55% phenolized 0.85% sterile saline for initial dilution of antisera, CDC = Centers for Disease Control.

± = factor tested with ATCC culture is circled, IN = analyst initials for antisera/antigen used, w = with, SAS = SA Scientific/Adam Diagnostics,

Note: Working dilutions of Disco and SAS H antisera made with 0.55% phenolized 0.85% sterile saline. For this lot #: see page titled "Salmonella Media and Equipment Lot Record for Serotyping" included with this worksheet package. SSI = Statens Serum Institute -Note : SSI antisera used undiluted

NA = Not available, gp = group ADH Arkansas Department of Health CFSAN Centers for Food Safety and Nutrition

ANALYST(S)
Debra Anne Hora

LOT RECORD

Version 1.1 (5/2010)

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ARL-LOT.1

Salmonella Isolate Information Sheet

Sample Number: 915750 Date isolate shipped: 7/29/15

Product: FROZEN CHICKEN BLEND (DOG Food)

Original analyst: JENNIFER CANALE

Lab: MSB - NRL - LAB N

Total number of isolates submitted: 1

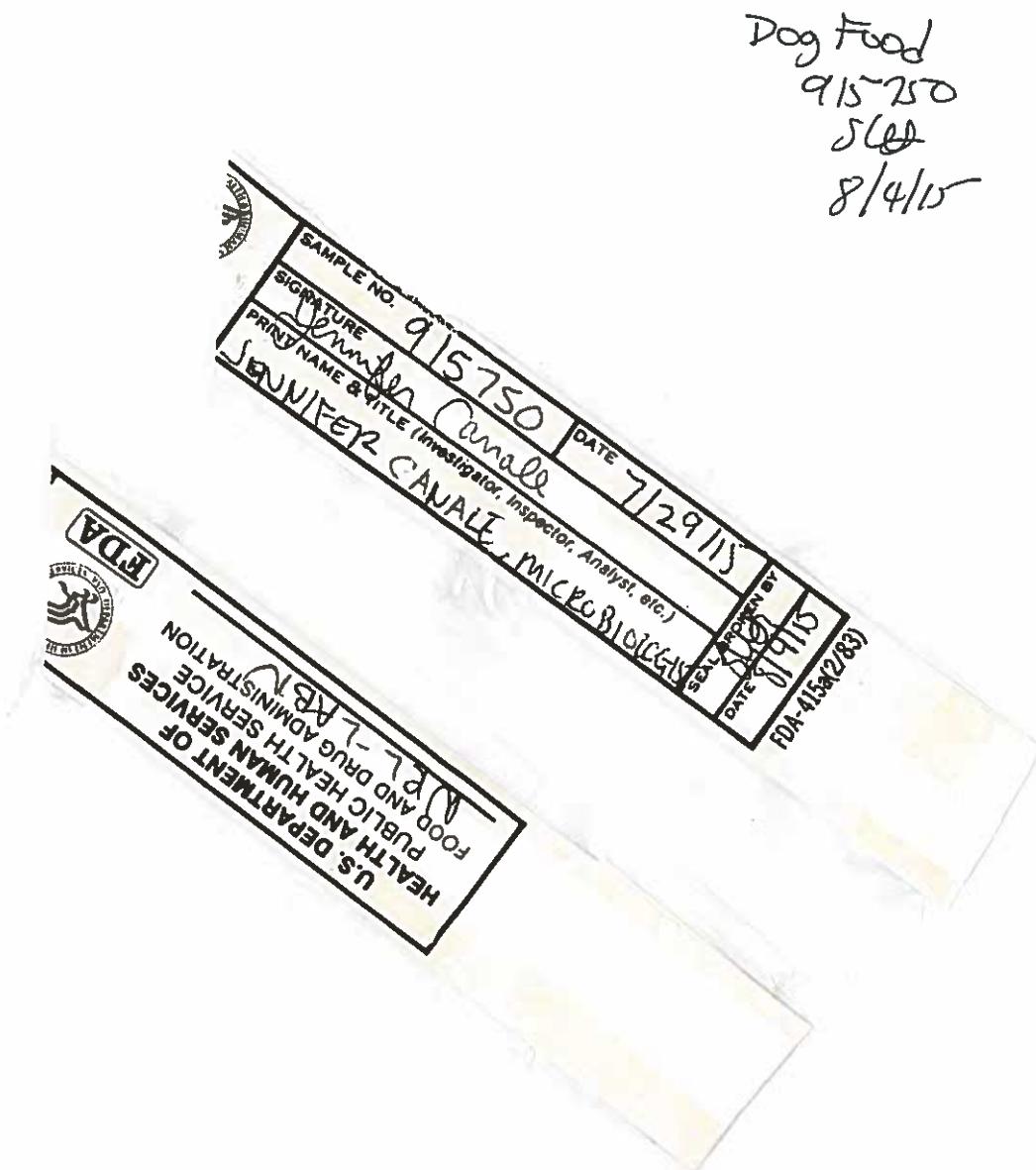
Isolates are FDA sealed: "915750 7/29/15 Jennifer Canale, microbiologist"

| Comp or Sub # | Isolate # | Tube identified as follows: "..." | TSI Result | API20E or Vitek Profile Number* | Somatic Polyvalent Result | Somatic Group Result | H Poly (a-z) Result +/- |
|---------------------|-----------------------|--|---------------|---------------------------------------|---------------------------------|----------------------------|----------------------------------|
| 1 | HERV Isolate #1 | "915750 Comp#1 HERV isolate#1 JRC 7/29/15" | KA++ KK+- | 00156104610 26200 | Poly A | Group | + |
| | | | | | Poly | Group | |
| | | | | | Poly | Group | |
| | | | | | Poly | Group | |
| | | | | | Poly | Group | |
| | | | | | Poly | Group | |

*Examples: 6704552 = an API20E profile number, 6020724533 = a Vitek profile number

Comments:

ATTACHMENT A: ANALYST'S ORIGINAL SEAL



Stephanie Hader

Attachment "A"

Version 1.0 (12/2004)

ARL-ATA.1