COVID-19
Point of Care

Package Insert

For the qualitative detection of SARS-CoV-2 nucleic acid. For IVD Use. For Rx only. For Emergency Use Authorization Only.

PS-001541

PS-001418 Rev C07/22
**Name**
Visby Medical COVID-19 Point of Care Test

**Common or Usual Name**
Visby COVID-19 Point of Care Test

**Intended Use**
The Visby Medical COVID-19 Point of Care Test is a single-use (disposable), fully-integrated, fast, automated RT-PCR in vitro diagnostic test intended for the qualitative detection of SARS-CoV-2 nucleic acid in nasopharyngeal, anterior nasal (nasal), or mid-turbinate swabs, collected by a healthcare provider (HCP) or anterior nasal or mid-turbinate swabs self-collected (by individuals 18 years of age or older, under the supervision of an HCP) from individuals suspected of COVID-19 by their HCP. Testing is limited to laboratories certified under the Clinical Laboratory Improvement Amendments of 1988 (CLIA), 42 U.S.C. §263a, that meet requirements to perform high, moderate or waived complexity tests. The Visby COVID-19 Point of Care Test is authorized for use at the Point of Care (POC) i.e., in patient care settings operating under a CLIA Certificate of Waiver, Certificate of Compliance, or Certificate of Accreditation.

Results are for the identification of SARS-CoV-2 RNA. The SARS-CoV-2 RNA is generally detectable in respiratory specimens during the acute phase of infection. Positive results are indicative of the presence of SARS-CoV-2 RNA; clinical correlation with patient history and other diagnostic information is necessary to determine patient infection status. Positive results do not rule out bacterial infection or co-infection with other viruses. The agent detected may not be the definitive cause of disease. Laboratories within the United States and its territories are required to report all results to the appropriate public health authorities. Negative results do not preclude SARS-CoV-2 infection and should not be used as the sole basis for patient management decisions. Negative results must be combined with clinical observations, patient history, and epidemiological information.

The Visby Medical COVID-19 Point of Care Test is intended for use by laboratory personnel who have received specific training on the use of the Visby Medical COVID-19 Point of Care Test. The Visby COVID-19 Point of Care Test is only for use under the Food and Drug Administration’s Emergency Use Authorization.

**Summary and Explanation of the Procedure**
The Visby COVID-19 Point of Care Test is a fast, instrument-free, single-use (disposable) molecular in vitro diagnostic test for the qualitative detection of ribonucleic acid from the SARS-CoV-2 virus. The Visby COVID-19 Point of Care Test contains all components required to carry out an assay for SARS-CoV-2 in nasopharyngeal, anterior nasal, or mid-turbinate swabs.

**Principles of the Procedure**
The Visby COVID-19 Device is a single-use (disposable), fully-integrated, fast, compact device containing a reverse transcription-polymerase chain reaction (RT-PCR) based assay for qualitative polymerase detection of viral RNA from the SARS-CoV-2 virus. The device automatically performs all steps required to complete lysis, reverse transcription, polymerase chain reaction, and amplicon detection.

Nasopharyngeal, dual nostril mid-turbinate or dual nostril anterior nasal swabs are placed in Visby Buffer and then transferred into the sample port of the device. The sample enters a lysis module and then rehydrates the RT enzyme and RT primers. The mixture then moves through a fixed temperature module where virus is simultaneously lysed and the viral RNA reverse transcribed. The resulting fluid (containing cDNA) is then mixed with lyophilized PCR reagents containing biotinylated primers specific to the N1 gene of the SARS-CoV-2 virus and to 18S ribosomal RNA, which serves as a process control. The PCR mixture (containing cDNA template and reagents) is then thermocycled to amplify the SARS-CoV-2 (if present) and 18S targets.

After PCR, the biotinylated product is moved to the detection module, which contains covalently bound capture probes immobilized in the shape of two distinct rectangular spots along a flow channel. Detection of the target-specific PCR product is accomplished using an enzyme-linked colorimetric assay using streptavidin bound horseradish peroxidase (HRP) and a colorimetric substrate that forms a purple precipitate. The operator observes a color change at the specific locations indicating the presence of an amplified target. A purple color in the “Results Valid” spot indicates a successful internal control, and a purple color in the “Positive for SARS-CoV-2 (COVID-19)” spot indicates detection of the SARS-CoV-2 virus.

**Materials**

**Provided in Test**
- Visby COVID-19 Device
- Visby Test Tube Holder
- Visby Buffer Tube
- Visby Pastette
- Package Insert
- Quick Reference Guide
- Recommended Collection Instructions
- Biohazard Bag

**Required Accessories**
- Visby Power Adapter

**Required but Not Supplied**
- Absorbent Pads
- Hazardous Waste Disposal Bin
- Gloves
- Nasopharyngeal, Mid-turbinate or Anterior nasal swabs. Use sterile rayon, foam, polyester or flocked flexible shaft swabs

**Available but not Provided**
- SARS-CoV-2 Positive and A549 Cells Negative External Control (Swabs) by Microbiologics®
- 15 mL screw top tube, Falcon 14-959-70C, or equivalent
**Note:** This device complies with Part 15 of the FCC Rules. Operation is subject to the following conditions: (1) this device may not cause harmful interference, and (2) this device must accept any interference received, including interference that may cause undesired operation.

The Visby COVID-19 Point of Care Test has been tested and found to comply with the limits for a Class A digital device, pursuant to Part 15 of the FCC Rules. These limits are designed to provide reasonable protection against harmful interference when the equipment is operated in a commercial environment. The equipment generates, uses, and can radiate radio frequency energy and, if not installed and used in accordance with the instruction manual, may cause harmful interference to radio communications. Operation of this equipment in a residential area is likely to cause harmful interference in which case the user will be required to correct the interference at his/her own expense.

**Warnings and Precautions**

**General**

1. For in vitro diagnostic use.
2. This product has not been FDA cleared or approved, but has been authorized for emergency use by FDA under an EUA for use by laboratories certified under the Clinical Laboratory Improvement Amendments of 1988 (CLIA), 42 U.S.C. §263a, that meet the requirements to perform high, moderate, or waived complexity tests. This test is authorized for use at the Point of Care (POC), i.e., in patient care settings operating under a CLIA Certificate of Waiver, Certificate of Compliance, or Certificate of Accreditation.
3. This product has been authorized only for the detection of nucleic acid from SARS CoV-2, not for any other viruses or pathogens.
4. The emergency use of this product is only authorized for the duration of the declaration that circumstances exist justifying the authorization of emergency use of in vitro diagnostics for detection and/or diagnosis of COVID-19 under Section 564(b)(1) of the Federal Food, Drug and Cosmetic Act, 21 U.S.C. § 360bbb-3(b)(1), unless the declaration is terminated or authorization is revoked sooner.
5. This product is for single use only; do not reuse the Visby COVID-19 Point of Care Test.
6. Federal Law restricts this device for sale by or on the order of a licensed practitioner (US only).
7. While color-blind users may be unable to differentiate red, green, and white status lights, they can consult the light location and shape of the light to determine test status. When interpreting results, the purple shade may appear as a dark shade for some users.

**Visby Medical COVID-19 Testing**

1. Follow your institution’s and the CDC Interim Guidelines for Collecting, Handling, and Testing Clinical Specimens from Persons for Coronavirus Disease 2019 (COVID-19) or refer to the Recommended Collection Instructions included with the Visby COVID-19 Point of Care Test.
2. The Visby COVID-19 Point of Care Test’s control and results must be interpreted as per the instructions provided on this guide.
3. Leave the Visby COVID-19 Device sealed in the foil pouch until just before use.
4. Do not use the Visby COVID-19 Point of Care Test past its expiration date.
5. Do not use the Visby COVID-19 Device if it appears broken or has been dropped.
6. Do not shake or tilt the Visby COVID-19 Device after adding a sample.
7. Do not add excessive sample into the Device as this may result in an error.
8. Run the Device on a clean, level surface.
9. Do not touch, move, or unplug charging cable, adapter, or Device while the test is running.
10. At a low frequency, clinical samples can contain inhibitors that may generate invalid results.
11. Keep the work area clean to prevent contamination.
12. Wear gloves while handling samples and change gloves between testing each specimen. If the gloves come in contact with specimen or appear to be wet, change gloves to avoid contamination. Change gloves before leaving work area and upon entry into work areas.
13. Do not try to disassemble the Visby COVID-19 Device. In the case of a positive sample, this could lead to sample leakage and potential contamination.
14. The Visby COVID-19 Device should be placed in the biohazard bag and disposed of in the appropriate specimen waste containers according to your institution’s standard practices.
15. The results of the Visby COVID-19 Device must be read within 2 hours after the green check mark light appears. Failure to do so may yield invalid results. After 2 hours or after the Device is unplugged the green check mark will turn off indicating that the read window has expired.
16. Each button will have a different feel as it clicks into place. Push firmly to make sure all buttons are completely down or the test may yield invalid results.
17. The Visby COVID-19 Device requires a sample input of a specified volume from a fixed-volume pastette that is provided. If no sample is added into the Visby COVID-19 Device, the Results Valid spot will not be displayed.

**Visby Power Adapter**

1. Use only the supplied Visby power adapter to power the Visby COVID-19 Device. Using other power adapters to operate the Visby COVID-19 Device will void the safety protection of the Device.
2. The Visby power adapter should be replaced if an increased number of RED X errors are observed.
3. Dispose of the power adapter as per local, federal, and institutional guidelines.
Specimen and Visby Buffer

1. Follow the CDC’s guidelines and your institution’s safety procedures for working with chemicals and handling biological samples.
2. Treat all biological specimens in the Visby Buffer tube as capable of transmitting infectious agents.
3. The Visby Buffer is used to process a single specimen only. If retest is required, refer to the retesting procedure section in the Package Insert.
4. Storing the Visby Buffer above 40°C after the addition of the patient sample can lead to false negative results.
5. Mix the specimen in the Visby Buffer Tube by inverting the tube 5 times. Reducing the number of inversions may result in invalid or inaccurate results.
6. Failure to use the Visby Buffer as directed can result in inaccurate test results.
7. Do not place the swab in viral transport media, saline, water or other buffers prior to testing.
8. Do not use the Visby Buffer Tube if it appears to be leaking, damaged, or opened.
9. The Visby Buffer is a clear, colorless, and odorless solution. Do not use if the solution appears discolored, has a strong odor or has any particles in it.
10. Do not use the Visby Buffer past its expiration date.

Spills

1. If a spill occurs with the Visby COVID-19 Point of Care Test, soak up the spillage with a disposable absorbent pad. Spray the contaminated area and materials with 10% bleach. Wipe down the surface so that it is saturated with bleach and let it rest for at least 5 minutes. Once a minimum of 5 minutes has passed, spray the area with 70% ethyl or isopropyl alcohol and wipe down the surface. Dispose of affected single-use materials such as the absorbent pad, test tube holder, the Visby COVID-19 Device, and/or the COVID-19 Visby Buffer tube. Discard affected single-use materials according to your institution’s standard practices.
2. If a spill occurs on the Visby power adapter, unplug the unit and wipe it down vigorously with 70% ethyl or isopropyl alcohol. Allow the power adapter to completely dry before using it again.

Safety

1. Visby Buffer may contain irritants. Do not ingest the contents of the tube. If the contents of the tube are splashed in your eyes, flush your eyes with water. If the contents splash onto your skin, wash with soap and water. If irritation persists, notify a health care provider.
2. Follow your institution’s safety procedures for working with chemicals and handling biological samples.

Storage, Stability and Specimen Collection

Storage

Store the Visby COVID-19 Point of Care Test in a cool and dry environment (36°F-86°F). Do not freeze. In case of refrigeration or other exposure to cold temperatures, ensure that the Visby COVID-19 Device is allowed to come to its minimum operating temperature prior to use.

Specimen Collection

The Visby COVID-19 Point of Care Test is intended for testing nasopharyngeal, anterior nasal, or mid-turbinate swabs collected without transport media. Use sterile rayon, foam, polyester or flocked flexible shaft swabs. Collect samples in accordance with CDC Interim Guidelines for Collecting, Handling, and Testing Clinical Specimens from Persons for Coronavirus Disease 2019 (COVID-19) or the Recommended Collection Instructions provided with the Visby COVID-19 Point of Care Test.

WARNING: Testing samples if stored for more than one hour at room temperature can result in inaccurate test results.

Specimen Storage

Anterior Nasal, mid-turbinate, or nasopharyngeal swabs should be tested as soon as possible after collection. If immediate testing is not possible, place patient swab in a dry tube labeled with patient information, and capped tightly at room temperature for up to one (1) hour prior to testing. Ensure the swab fits securely within the tube and the cap is tightly closed.

WARNING: Testing samples if stored for more than one hour at room temperature can result in inaccurate test results.

Specimen Stability in Visby Buffer

Specimen is stable in Visby Buffer in the following conditions:

- 45 minutes at room temperature.
- 24 hours at refrigerated temperature.

WARNING: Testing samples that have exceeded these storage conditions can result in inaccurate results. Do not store above 40°C.

Visby COVID-19 Point of Care Test Instructions for Use

Please follow these instructions carefully.

Do not remove the Visby COVID-19 Device from the foil pouch until the workspace is prepared and you are ready to run the test.

Run the Visby COVID-19 Device at room temperature on a clean, level surface.

The Visby COVID-19 Device, pastettes, and Visby Buffer should be disposed of in accordance with local regulations.

Operating Conditions

66°F to 82°F (19°C to 25°C) Refrigerated
Temperature: 35°F to 46°F (2°C to 8°C)
Frozen Temperature: 5°F to -74°F (-15°C to -59°C)

Visby COVID-19
Point of Care
Visby COVID-19 Point of Care Test Procedure

**Step 1** Set Up the Workspace

**Operating Conditions:** Ensure the test is run at room temperature in a cool, dry environment. Set up a new workspace for each Visby COVID-19 Point of Care Test. Clean the workspace and use a new absorbent pad after each test. Change gloves between handling samples and setting up a new test. Place the VisbyCOVID-19 Device on a level surface.

**Note:** Leave the Visby COVID-19 Device sealed in the foil pouch until just before use. Proceed to Step 2 only when ready to run the test. Do not use the Visby COVID-19 Device if it appears broken or has been dropped. Do not use the VisbyCOVID-19 Point of Care Test past its expiration date.

Patient swab sample is stable in a dry tube for 1 hour at room temperature. Patient sample must be added to Visby Buffer within one hour of collection. Failure to add the sample within the allotted time or the use of alternate media may result in invalid or inaccurate test results.
Step 2 Add Sample to the Visby Buffer Tube

Patient sample must be used with Visby Buffer only. Place the device on a level surface.

A Open the Visby Buffer Tube.

Note: Do not use the Visby Buffer Tube past its expiration date.

B Place the Visby Buffer Tube in a tube holder.

C Take the collected patient sample.

D Place the patient swab into the Visby Buffer.

Note: If your swab still has a breakpoint - break the handle of the swab.

E Screw cap back on the Visby Buffer Tube.

Patient sample is stable in Visby Buffer at room temperature for 45 minutes and 24 hours at refrigeration.
Step 3  Load the Sample into the Device

STOP! DO NOT plug in the test until Step 4E.

A Pick up the Visby Buffer Tube.

B Mix the specimen in the Visby Buffer Tube by inverting the tube 5 times.

C Open the cap of the Visby Buffer Tube. Place cap wet side up. Take the Visby pastette.

D Squeeze the upper bulb.

E Keeping the bulb squeezed, lower the pastette tip to the bottom of the Visby Buffer Tube.

F Keep the tip fully under the fluid. Release the upper bulb.

G Fill the entire shaft with fluid. Some fluid should enter the lower bulb.

Note: Do not squeeze lower bulb or invert the pastette.

H Place the tip of the pastette into Sample Port (Button 1). Squeeze the bulb to dispense the liquid. Some fluid will remain in the lower bulb.

Note: Do not overfill.

I Discard the pastette.

Note: Store the remaining Visby Buffer for retesting if needed.
**Step 4** Run the Test

**IMPORTANT!** Each button will have a different feel as it “clicks” into place. Push firmly to make sure all buttons are completely down or the test may not work.

A After loading sample into device, close Button 1 by sliding the cap to the right.

B Push Button 1 all the way down to add the sample.

C Push Button 2 all the way down to unlock button 3.

D Push Button 3 all the way down. Use two thumbs, push firmly.

   **Note:** All buttons should be all the way down.

E Plug in the device until it clicks into place. A stable white light indicates the test is running.

   Ensure that there is no gap between the power adapter plug and device.

**WAIT 30 MINUTES!** DO NOT touch or move the charging adapter, cable or device.

DO NOT shake or tilt the Visby COVID-19 Device after adding a sample.
Step 5  Get the Results

AFTER 30 MINUTES

A Check if the results are ready. Look to the top left corner of the device.

READY
A green check \( \square \) indicates the results are ready to be read. If present, proceed to Step 5B.

ERROR
A red \( \times \) indicates that an electronic failure occurred. Repeat the test.

B If a green check appears \( \square \), confirm the results are valid. Look at the results window for a purple spot near “RESULTS VALID”.

VALID
This is the control spot. A purple spot indicates a valid test. If present, proceed to Step 5C.

INVALID
No purple spot indicates an invalid test. Repeat the test.

Instructions to repeat the test: Sample in Visby Buffer is stable for 45 minutes at room temperature and 24 hours at refrigeration. If storage conditions are exceeded, obtain new sample and repeat test with a new pastette, a new Visby Buffer Tube, and a new Visby COVID-19 Device.

C Read and record the results. Look at the results window for a purple spot near “POSITIVE for SARS-CoV-2 (COVID-19)”. Results may be read for up to 2 hours after the test is completed. The intensity of the spot in the results window may vary. Any shade of color should be considered a spot.

Positive Result

A purple spot in this location indicates a positive result.

Negative Result

No purple spot in this location indicates a negative result.

After use, the Visby COVID-19 Device should be placed in a Biohazard Bag prior to disposal. The used Device, pastette, Visby Buffer, and specimen collection kit should be disposed of in the appropriate specimen waste containers according to your institution’s standard practices.

Need Help? Call 1-833-GoVisby (1-833-468-4729)
Color Blindness Precaution

While color-blind users may be unable to differentiate red, green, and white status lights, they may observe the light location and shape of the light to determine test status.

Solid white light means test is running
Results are ready.
Electronic error, invalid test.
For retesting please refer to retest procedures section.

Quality Control

Internal Controls:
Each Visby Medical COVID-19 Device includes internal electronic and process controls.

1. Electronic Controls – The Visby COVID-19 Device will automatically detect various issues including hardware failures, invalid operating temperatures and other conditions that can lead to inaccurate test results. If one of these issues is detected, the device will display a ‘Red X’ status light and the test must be repeated. A green check mark indicates that the test was successful.

2. Process Control – The Visby COVID-19 Device includes an assay that targets 18S ribosomal RNA. This process control is carried through all stages of the testing process, including lysis, reverse transcription, PCR amplification, and colorimetric detection. Development of a purple spot in the “Results Valid” window indicates that all testing processes were successful. If a purple spot does not appear in the “Results Valid” window, the test is invalid and should be repeated.

External Positive and Negative Controls:
External controls should be used in accordance with local, state, and federal accrediting organizations as applicable. Testing of external control materials may be appropriate to train new operators or when receiving new device shipments. Visby Medical suggests the use of the Microbiologics control materials described in the figure below. Use of other commercial external control materials may be appropriate.

To run external control swabs, unwrap the swab, and gently tap the swab against the bottom of the Visby Buffer Tube 15 times. Discard the swab according to your institutions guidelines and screw the cap back onto the Visby Buffer tube. Proceed to Step 3.

Each positive or negative external control is for single use only. External control stability in the Visby Buffer has not been established.

<table>
<thead>
<tr>
<th>Product</th>
<th>Unit</th>
<th>Control Key</th>
</tr>
</thead>
<tbody>
<tr>
<td>SARS-CoV-2 Positive External Control Swab</td>
<td>Valid Positive Control Run</td>
<td></td>
</tr>
<tr>
<td>SARS-CoV-2 Positive and A549 Cells Negative External Control (Swabs) by Microbiologics*</td>
<td></td>
<td></td>
</tr>
<tr>
<td>A549 Cells Negative External Control Swab</td>
<td>Valid Negative Control Run</td>
<td></td>
</tr>
</tbody>
</table>
# Interpretation of Results

<table>
<thead>
<tr>
<th>Result</th>
<th>Interpretation</th>
<th>Next Steps</th>
</tr>
</thead>
<tbody>
<tr>
<td><img src="image" alt="Valid test" /></td>
<td>• Valid test (✔️, RESULTS VALID)</td>
<td>• Report results of patient sample</td>
</tr>
<tr>
<td><img src="image" alt="Negative for SARS-CoV-2(COVID-19)" /></td>
<td>• Negative for SARS-CoV-2(COVID-19) (❌ POSTIVE for SARS-CoV-2 (COVID-19))</td>
<td></td>
</tr>
</tbody>
</table>
| ![Invalid test; control fail](image) | • Invalid test; control fail (✔️, RESULTS VALID) | • Discard test  
• Refer to the retest procedure section below  
• If repeat test fails, contact Visby Medical Customer Support |

**Note:** Any shade of color with defined edges, as illustrated above, should be considered a spot.

- Blinking white light for 2-3 minutes, then turns to Red ❌
- Error: Invalid

- Error: Invalid

• Discard test  
• Refer to the retest procedure section below  
• Repeat the test - Do not plug in the power source before Step 4E. Push the adapter jack into the device’s charging port until you feel it click in place. Ensure that there is no gap between the adapter jack & device charging port. A stable white light will appear indicating the test is running  
• If repeat test fails, contact Visby Medical Customer Support
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Under Rare Circumstances

The following are occasionally observed and should not be confused with a positive result.

Background Staining
The background color in the results window may turn a light shade of blue or purple over time. This is a normal feature of the chemistry. This should not be considered a positive result.

Speckling and Bubbles
Samples containing blood or mucus may result in nonspecific small flakes in the results window. These are normal conditions and should not impact the interpretation of results. It is also normal for bubbles to appear in the results window during test processing.

Spot Shadow
An extremely faint spot without distinct edges may be seen in the results window. This may be a result of nonspecific binding.

Repeat the test with a new Visby Buffer tube and Visby COVID-19 Device.

If you are unsure how to interpret a result, please contact Visby Medical Customer Support at 1-833-GoVisby (1-833-468-4729) or support@visbymedical.com.

Retest Procedure

Samples stored in Visby Buffer
Obtain the leftover sample from the Visby Buffer tube. Repeat the test with a new Visby COVID-19 Device.

Note: Patient sample in Visby Buffer is stable for 45 minutes at room temperature and 24 hours at refrigeration. If storage conditions are exceeded, obtain new sample and repeat test within 48 hours.

External Controls
If the positive or negative external controls fail, repeat with a new external control and a new Visby COVID-19 Point of Care Test. If the repeat test fails, please contact Visby Medical Customer Support at 1-833-468-4729 (1-833-GoVisby).

Limitations

1. The performance of the Visby COVID-19 Point of Care Test was established using nasopharyngeal swab specimens. Midturbinate and anterior nasal swabs (collected by an HCP or self-collected under the supervision of an HCP) are considered acceptable specimen types for use with the Visby COVID-19 Point of Care Test, but performance with these specimen types has not been established.

2. Erroneous results may occur from improper specimen collection, sample dilution, technical error, sample mix-up, or if the viral load in the patient sample is below the limit of detection of the Visby COVID-19 Point of Care Test.

3. Careful compliance with the instructions in this insert and Quick Reference Guide Instructions are necessary to avoid erroneous results.

4. Because the detection of SARS-CoV-2 is dependent on the viral load present in the sample, reliable results are dependent on proper sample collection, sample processing, handling, and storage.

5. Built-in procedural controls of the Visby COVID-19 Point of Care Test cannot identify false positive results.

6. This test has been evaluated with human specimen material only.

7. The effect of interfering substances has been evaluated only for those listed within the labeling.

8. Mutations within the target region of SARS-CoV-2 could affect primer and/or probe binding, resulting in failure to detect the presence of virus.

9. This Test cannot rule out diseases caused by other bacterial or viral pathogens.

10. Performance has not been established in asymptomatic individuals.

11. Viral nucleic acid may persist in vivo, independent of virus viability. Detection of SARS-CoV2 nucleic acid does not imply that the corresponding virus is infectious or are the causative agents for clinical symptoms.

12. The performance of this test was established based on the evaluation of a limited number of clinical specimens. The clinical performance has not been established in all circulating variants but is anticipated to be reflective of the prevalent variants in circulation at the time and location of the clinical evaluation. Performance at the time of testing may vary depending on the variants circulating, including newly emerging strains of SARS-CoV-2 and their prevalence, which change over time.

Conditions of Authorization for Laboratories


However, to assist clinical laboratories and/or Point of Care Settings using the Visby COVID-19 Point of Care Test (referred to in the Letter of Authorization as “Your Product”), the relevant Conditions of Authorization are listed below:

1. Authorized laboratories* using your product must include with test result reports, all authorized Fact Sheets. Under exigent circumstances, other appropriate methods for disseminating these Fact Sheets may be used, which may include mass media.

2. Authorized laboratories using your product must use your product as outlined in the authorized labeling. Deviations from the authorized procedures, including the authorized instruments, authorized extraction methods, authorized clinical specimen types, authorized control materials, authorized ancillary reagents and authorized materials required to use your product are not permitted.

3. Authorized laboratories that receive your product must notify the relevant public health authorities of their intent to run your product prior to initiating testing.

4. Authorized laboratories using your product must have a process in place for reporting test results to healthcare providers and relevant public health authorities, as appropriate.

5. Authorized laboratories must collect information on the performance of your product and report to DMD/OHT7-OIR/OPEQ/CDRH (via email: CDRH-EUA-Reporting@fda.hhs.gov) and you (support@visbymedical.com) any suspected occurrence of false positive or false negative results and significant deviations from the established performance characteristics of your product.
of which they become aware.

6. All laboratory personnel using your product must be appropriately trained in the use of the Visby Medical COVID-19 Point of Care Test and use appropriate laboratory and personal protective equipment when handling this kit, and use your product in accordance with the authorized labeling.

7. Visby Medical, Inc., authorized distributor(s), and authorized laboratories using your product must ensure that any records associated with this EUA are maintained until otherwise notified by FDA. Such records will be made available to FDA for inspection upon request.

* The letter of authorization refers to “authorized laboratories” as “laboratories certified under CLIA that meet requirements to perform high, moderate, or waived complexity tests. The Visby Medical COVID-19 Point of Care test is authorized for use at the Point of Care (POC), i.e., in patient care settings operating under a CLIA Certificate of Waiver, Certificate of Compliance, or Certificate of Accreditation.”

### Analytical Performance

**Analytical Sensitivity (Limit of Detection)**

The limit of detection (LoD) is the lowest concentration of viral nucleic acid that is reliably detected by the Visby COVID-19 Point of Care Test. The LoD was first estimated by preparing 3-fold serial dilutions of inactivated SARS-CoV-2 virus (USA WA1/2020 strain) into negative clinical matrix. The dilutions were then transferred onto nasopharyngeal swabs and four replicates of five different concentrations were tested. The lowest concentration that had 100% detection was estimated to be the LoD. The LoD was then confirmed by preparing and testing 40 replicates at the concentration (435 copies/swab). The LoD was confirmed when 39/40 replicates gave positive test results (Table 01). One sample was excluded due to an invalid test result.

<table>
<thead>
<tr>
<th>Concentration (genomic copies/swab)</th>
<th>Detection Rate (# positive for SARS-CoV-2 / # total tests)</th>
</tr>
</thead>
<tbody>
<tr>
<td>LoD Serial Dilution</td>
<td></td>
</tr>
<tr>
<td>145</td>
<td>1/4</td>
</tr>
<tr>
<td>435</td>
<td>4/4</td>
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<td>11745</td>
<td>4/4</td>
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<tr>
<td>LoD confirmation</td>
<td>435</td>
</tr>
<tr>
<td></td>
<td>39/39*</td>
</tr>
</tbody>
</table>

*One result was excluded due to an invalid result

### Analytical Reactivity (Inclusivity)

Visby Medical follows FDA policy to routinely monitor SARS-CoV-2 sequences to determine if there is any impact to the Visby Medical COVID-19 test performance. As of June 2022, 11,322,345 SARS-CoV-2 sequences submitted to the GISAID database have been analyzed, including sequences from Alpha (B.1.1.7), Beta (B.1.351), Gamma (P.1), Delta (B.1.617.2) and Omicron (B.1.1.529 and BA) variants. To date, this period process has identified one mismatch that occurred in >5% of sequences, however, in-silico assessment indicates that this variant has no impact on the performance of the Visby test.

### Analytical Specificity/Exclusivity (Cross-Reactivity and Microbial Interference)

An *in silico* study was performed to assess potential cross-reactivity with related pathogens and normal or pathogenic flora that are reasonably likely to be encountered in clinical specimens. This assessment showed no sequence homology with SARS coronavirus and Bat SARS-like coronavirus genome for the forward and reverse primers; high sequence homology with SARS coronavirus and Bat SARS-like coronavirus genome was identified with the probe sequence. However, there are no significant homologies with the human genome, other coronaviruses, or normal or pathogenic flora that would predict potential false positive results when combining primers and probes. In addition, wet testing was also performed to evaluate the Visby COVID-19 Test performance when in the presence of 31 viral and bacterial organisms. Each organism was individually seeded into an artificial nasal matrix and tested on three devices with both COVID-19 negative samples and COVID-19 positive samples at 2x the LoD. The expected results were achieved 100% of the time. The organisms, concentrations and results are listed below. None of the 31 organisms demonstrated cross-reactivity with the Visby COVID-19 Test at the concentrations in Table 02. Testing was repeated with the Visby COVID-19 Point of Care test using a direct swab specimen for influenza A, influenza B and RSV using clinical matrix. Assessments with the other organisms were not repeated with the Visby COVID-19 Point of Care Test as the device including primer and probe sequences is unchanged and testing with the Visby COVID-19 Test was performed at high organism concentrations.
### Table 02: Summary of performance for organisms tested on the Visby COVID-19 Test (Cross-Reactivity and Microbial Interference)

<table>
<thead>
<tr>
<th>Organism</th>
<th>Concentration Tested</th>
<th>Units</th>
<th>Negative Samples (# of Valid Devices) Negative for SARS-CoV-2</th>
<th>Positive Samples (# of Valid Devices) Positive for SARS-CoV-2</th>
</tr>
</thead>
<tbody>
<tr>
<td>Human Coronavirus 229E</td>
<td>$1.1 \times 10^9$</td>
<td>genomic copies/mL</td>
<td>$8/9$ (1)</td>
<td>$3/3$</td>
</tr>
<tr>
<td>Human Coronavirus OC43</td>
<td>$1.1 \times 10^9$</td>
<td>genomic copies/mL</td>
<td>$3/3$</td>
<td>$3/3$</td>
</tr>
<tr>
<td>Human Coronavirus HKU1</td>
<td>$1.1 \times 10^9$</td>
<td>genomic copies/mL</td>
<td>$3/3$</td>
<td>$3/3$</td>
</tr>
<tr>
<td>Human Coronavirus NL63</td>
<td>$1.1 \times 10^9$</td>
<td>genomic copies/mL</td>
<td>$3/3$</td>
<td>$3/3$</td>
</tr>
<tr>
<td>SARS-CoVoronavirus (2003)</td>
<td>$1.1 \times 10^9$</td>
<td>genomic copies/mL</td>
<td>$3/3$</td>
<td>$3/3$</td>
</tr>
<tr>
<td>MERS-CoVoronavirus</td>
<td>$1.1 \times 10^9$</td>
<td>genomic copies/mL</td>
<td>$3/3$</td>
<td>$3/3$</td>
</tr>
<tr>
<td>Adenovirus, C1 Ad 71</td>
<td>$2.5 \times 10^4$</td>
<td>ng/μL</td>
<td>$3/3$</td>
<td>$3/3$</td>
</tr>
<tr>
<td>Human metapneumovirus (hMPV)</td>
<td>$1.1 \times 10^7$</td>
<td>genomic copies/mL</td>
<td>$3/3$</td>
<td>$3/3$</td>
</tr>
<tr>
<td>Human parainfluenza virus 1</td>
<td>$2.5 \times 10^3$</td>
<td>ng/μL</td>
<td>$3/3$</td>
<td>$3/3$</td>
</tr>
<tr>
<td>Human parainfluenza virus 2</td>
<td>$2.5 \times 10^3$</td>
<td>ng/μL</td>
<td>$3/3$</td>
<td>$3/3$</td>
</tr>
<tr>
<td>Human parainfluenza virus 3</td>
<td>$2.5 \times 10^3$</td>
<td>ng/μL</td>
<td>$3/3$</td>
<td>$8/9$ (2)</td>
</tr>
<tr>
<td>Human parainfluenza virus 4b</td>
<td>$2.5 \times 10^3$</td>
<td>ng/μL</td>
<td>$3/3$</td>
<td>$3/3$</td>
</tr>
<tr>
<td>Influenza A</td>
<td>$1.1 \times 10^6$</td>
<td>CEID₅₀/mL</td>
<td>$3/3$</td>
<td>$6/6$ (4)</td>
</tr>
<tr>
<td>Influenza B</td>
<td>$1.1 \times 10^6$</td>
<td>CEID₅₀/mL</td>
<td>$3/3$</td>
<td>$6/6$ (4)</td>
</tr>
<tr>
<td>Enterovirus 68</td>
<td>$1.1 \times 10^6$</td>
<td>genomic copies/mL</td>
<td>$3/3$</td>
<td>$3/3$</td>
</tr>
<tr>
<td>Respiratory syncytial virus</td>
<td>$1.1 \times 10^6$</td>
<td>genomic copies/mL</td>
<td>$3/3$</td>
<td>$6/6$ (4)</td>
</tr>
<tr>
<td>Human rhinovirus 17 (strain 33342)</td>
<td>$1.1 \times 10^6$</td>
<td>genomic copies/mL</td>
<td>$3/3$</td>
<td>$3/3$</td>
</tr>
<tr>
<td>Chlamydia pneumoniae</td>
<td>$1.1 \times 10^6$</td>
<td>IFU/mL</td>
<td>$3/3$</td>
<td>$3/3$</td>
</tr>
<tr>
<td>Haemophilus influenza</td>
<td>$1.1 \times 10^6$</td>
<td>genomic copies/mL</td>
<td>$3/3$</td>
<td>$3/3$</td>
</tr>
<tr>
<td>Legionella pneumophila</td>
<td>$1.1 \times 10^6$</td>
<td>genomic copies/mL</td>
<td>$3/3$</td>
<td>$3/3$</td>
</tr>
<tr>
<td>Mycobacterium tuberculosis</td>
<td>$1.1 \times 10^6$</td>
<td>genomic copies/mL</td>
<td>$3/3$</td>
<td>$3/3$</td>
</tr>
<tr>
<td>Streptococcus pneumoniae</td>
<td>$1.1 \times 10^6$</td>
<td>genomic copies/mL</td>
<td>$3/3$</td>
<td>$3/3$</td>
</tr>
<tr>
<td>Streptococcus pyogenes</td>
<td>$1.1 \times 10^6$</td>
<td>genomic copies/mL</td>
<td>$3/3$</td>
<td>$3/3$</td>
</tr>
<tr>
<td>Bordetella parapertussis</td>
<td>$1.1 \times 10^6$</td>
<td>genomic copies/mL</td>
<td>$3/3$</td>
<td>$3/3$</td>
</tr>
<tr>
<td>Mycoplasma pneumoniae</td>
<td>$1.1 \times 10^6$</td>
<td>genomic copies/mL</td>
<td>$3/3$</td>
<td>$3/3$</td>
</tr>
<tr>
<td>Pneumocystis jirovecii (PJP), also called: Pneumocystis carinii Delanoë and Delanoë</td>
<td>$1.1 \times 10^6$</td>
<td>nuclei/mL</td>
<td>$3/3$</td>
<td>$3/3$</td>
</tr>
<tr>
<td>Candida albicans</td>
<td>$1.1 \times 10^6$</td>
<td>genomic copies/mL</td>
<td>$3/3$</td>
<td>$3/3$</td>
</tr>
<tr>
<td>Pseudomonas aeruginosa</td>
<td>$1.1 \times 10^6$</td>
<td>genomic copies/mL</td>
<td>$3/3$</td>
<td>$3/3$</td>
</tr>
<tr>
<td>Staphylococcus epidermis</td>
<td>$1.1 \times 10^6$</td>
<td>genomic copies/mL</td>
<td>$3/3$</td>
<td>$3/3$</td>
</tr>
<tr>
<td>Streptococcus salivarius</td>
<td>$1.1 \times 10^6$</td>
<td>genomic copies/mL</td>
<td>$3/3$</td>
<td>$3/3$</td>
</tr>
<tr>
<td>Pooled human nasal wash</td>
<td>10%</td>
<td>percent of total volume</td>
<td>$3/3$</td>
<td>$3/3$</td>
</tr>
</tbody>
</table>

(1) A fresh sample was retested for the potential cross-reactive organism and tested with twice the number of devices; the expected results were achieved in all cases. As the contrived positive SARS-CoV-2 samples were prepared in the same lab space as the negative samples, this is the suspected root cause for the observed false positive result.

(2) A fresh sample was retested for potential microbial interference with twice the number of devices, and expected results were achieved in all cases.

(3) Testing was performed with the Visby COVID-19 Test and the Visby COVID-19 Point of Care Test.
Analytical Specificity (Interfering Substances)

A study was executed to determine the effect of potentially interfering endogenous and exogenous substances that may be present in a clinical sample on the performance of the Visby COVID-19 Point of Care Test. Each potential interfering substance was seeded into negative clinical matrix and then transferred to a nasopharyngeal swab. For each substance, additional negative matrix was transferred to three swabs to create a negative sample, and matrix with inactivated SARS-CoV-2 virus (USA WA1/2020 strain) was transferred to three swabs to create a positive (2X LoD) sample. Both the negative and positive samples were tested in triplicate. The substances, concentrations, and results are listed below (Table 03). None of the substances tested for interference impacted the performance or results of the Visby COVID-19 Point of Care Test.

Table 03: Summary of valid device performance for each interfering substance

<table>
<thead>
<tr>
<th>Interfering Substance</th>
<th>Assay Interference Limit</th>
<th>Negative Samples (Number of Valid Devices Negative for SARS-CoV-2)</th>
<th>Low Positive Samples (2X LoD) (Number of Valid Devices Positive for SARS-CoV-2)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Afrin</td>
<td>25% (v/v)</td>
<td>3/3</td>
<td>3/3</td>
</tr>
<tr>
<td>Biotin</td>
<td>3.5 µg/mL</td>
<td>3/3</td>
<td>3/3</td>
</tr>
<tr>
<td>Fresh Whole Blood Pooled Human Donors</td>
<td>5% (v/v)</td>
<td>3/3</td>
<td>3/3</td>
</tr>
<tr>
<td>Flonase</td>
<td>25% (v/v)</td>
<td>3/3</td>
<td>3/3</td>
</tr>
<tr>
<td>Mucin</td>
<td>1% (w/v)</td>
<td>3/3</td>
<td>3/3</td>
</tr>
<tr>
<td>Mupirocin</td>
<td>12 mg/mL</td>
<td>3/3</td>
<td>3/3</td>
</tr>
<tr>
<td>Nasacort</td>
<td>25% (v/v)</td>
<td>3/3</td>
<td>3/3</td>
</tr>
<tr>
<td>NeoSynephrine Cold &amp; Sinus Extra Strength Spray</td>
<td>25% (v/v)</td>
<td>3/3</td>
<td>3/3</td>
</tr>
<tr>
<td>Nasal Saline Spray</td>
<td>25% (v/v)</td>
<td>3/3</td>
<td>3/3</td>
</tr>
<tr>
<td>Tobramycin</td>
<td>2.43 mg/mL</td>
<td>3/3</td>
<td>3/3</td>
</tr>
<tr>
<td>Zanamivir (Relenza)</td>
<td>500 ng/mL, 5 mg/mL</td>
<td>3/3</td>
<td>3/3</td>
</tr>
<tr>
<td>Zicam Allergy Relief</td>
<td>25% (v/v)</td>
<td>3/3</td>
<td>3/3</td>
</tr>
</tbody>
</table>

Clinical Performance – Point of Care

Clinical Study Performance

The clinical performance of the Visby Medical COVID-19 Point of Care Test was established in a single center prospective clinical study conducted in a typical point of care (POC) setting. Five operators representing typical POC users tested specimens from 96 study participants over a 4-week period.

Study participants were consented, and two nasopharyngeal swab (NPS) samples were collected. One NPS was placed in universal transport media (UTM) and sent to a reference laboratory for comparator testing using a EUA COVID-19 Test. The other NPS sample was not placed in any transport media and was tested on-site using the Visby COVID-19 Point of Care Test. All study participants were symptomatic with the exception of two. The average age among study subjects was 31 with a range between 9 and 72. Of the 96 specimens tested, 11 yielded initial invalid results (initial invalid rate 11.5% (11/96)). For one subject, the Visby COVID-19 Point of Care Test didn’t yield a valid result during retest. The overall valid rate of the Visby COVID-19 Point of Care Test was 99.0% (95/96).

Positive percent agreement (PPA) was calculated as 100% x (TP / TP + FN). True positive (TP) indicates that both the Visby and comparator method had a positive result for SARS-CoV-2, and false negative (FN) indicates that the Visby result was negative while the comparator result was positive. Negative percent agreement (NPA) was calculated as 100% x (TN / TN+ FP). True negative (TN) indicates that both the Visby and the comparator method had negative results, and a false positive (FP) indicates that the Visby result was positive, but the comparator result was negative. The exact binomial two-sided 95% confidence interval was calculated. The results are summarized in Table 04.
**Table 04: Visby COVID-19 Point of Care Test vs EUA Comparator Assay**

<table>
<thead>
<tr>
<th>Visby COVID-19 Point of Care Test</th>
<th>EUA COVID-19 Test</th>
<th></th>
<th></th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Positive</td>
<td>Negative</td>
<td>Totals</td>
</tr>
<tr>
<td>Positive</td>
<td>31</td>
<td>3</td>
<td>34</td>
</tr>
<tr>
<td>Negative</td>
<td>0</td>
<td>61</td>
<td>61</td>
</tr>
<tr>
<td>Totals</td>
<td>31</td>
<td>64</td>
<td>95</td>
</tr>
</tbody>
</table>

* One of three false positive results was positive when tested with another EUA COVID-19 assay.

PPA 100.0% (95% CI: 89.0%-100.0%)

NPA 95.3% (95% CI: 87.1%-98.4%)

---

**Second Prospective Clinical Study**

A second two-armed prospective study was performed in a typical POC setting using a different molecular EUA COVID-19 Test as the comparator assay. As in the previous study, subjects were consented and two NPS samples were collected. One swab was placed in UTM and tested at a reference laboratory using a molecular EUA COVID-19 test while a second swab was placed directly into Visby Buffer and tested on-site with the Visby Medical COVID-19 Point of Care Test.

In the first arm of the study, subjects suspected of COVID-19 by their HCP were enrolled without regard to the results of standard of care test results and the Visby testing was performed by 2 typical POC personnel. A total of 95 subject were enrolled over 12 non-consecutive days. Eight (8) tests (8%, 8/95) had an initial invalid test result, of which all provided valid results upon retesting. Of the 95 subjects with valid test result, 85 were symptomatic and 10 were asymptomatic. The PPA and NPA for the Visby Medical COVID-19 POC Test for the first arm of the study is shown in Table 05.

**Table 05: Visby COVID-19 Point of Care Test vs EUA Comparator Assay**

<table>
<thead>
<tr>
<th>Visby COVID-19 Point of Care Test</th>
<th>EUA COVID-19 Test</th>
<th></th>
<th></th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Positive</td>
<td>Negative</td>
<td>Totals</td>
</tr>
<tr>
<td>Positive</td>
<td>40</td>
<td>3</td>
<td>43</td>
</tr>
<tr>
<td>Negative</td>
<td>2</td>
<td>50</td>
<td>52</td>
</tr>
<tr>
<td>Totals</td>
<td>42</td>
<td>53</td>
<td>95</td>
</tr>
</tbody>
</table>

PPA 95.2% (95% CI: 84.2%-98.7%)

NPA 94.3% (95% CI: 84.6%-98.1%)

a. Retesting of one specimen with the Visby test gave a positive result.

b. One of the three false positive sample were positive when tested with an alternate EUA molecular assay.

In the second arm of study, subjects with positive results by standard of care testing were selectively enrolled and tested. Testing was performed by one typical POC personnel. A total of 17 subjects were enrolled over 13 non-consecutive days. Three (3) tests (16.7%, 3/18) had an initial invalid test result, of which one was invalid upon retesting and was excluded from data analysis. Of the 16 subjects with valid test results, all were symptomatic. The PPA and NPA for the Visby Medical COVID-19 POC Test, for this arm of the study, is shown in Table 06.

Between the two study arms, the Visby test detected 100% (12/12) samples with low viral loads (as determined by the Ct value of the comparator assay).

**Table 06: Visby COVID-19 Point of Care Test vs EUA Comparator Assay with standard of care positive samples**

<table>
<thead>
<tr>
<th>Visby COVID-19 Point of Care Test</th>
<th>EUA COVID-19 Test</th>
<th></th>
<th></th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Positive</td>
<td>Negative</td>
<td>Totals</td>
</tr>
<tr>
<td>Positive</td>
<td>13</td>
<td>1</td>
<td>14</td>
</tr>
<tr>
<td>Negative</td>
<td>0</td>
<td>2</td>
<td>2</td>
</tr>
<tr>
<td>Totals</td>
<td>13</td>
<td>3</td>
<td>16</td>
</tr>
</tbody>
</table>

PPA 100% (95% CI: 77.2%-100%)

NPA 66.7% (95% CI: 20.8%-93.9%)

a. The false positive sample was positive when tested with an alternate EUA molecular assay.
Contrived Specimen Performance

Contrived direct swab samples were used to evaluate performance of the test near the assay LoD in a POC setting. Testing was integrated into the workflow of the prospective study over a 2-day period. Each operator tested 3 low positive (<2xLoD) and 3 negative specimens. Results are summarized in Table 07 by operator, by sample type, and overall. Of the 30 tested specimens, 3 (10%) yielded invalid results during the initial test. Study operators were able to perform the test correctly with an overall agreement rate of 100.0%.

Table 07: Contrived Specimen Results

<table>
<thead>
<tr>
<th>Operator</th>
<th>1</th>
<th>2</th>
<th>3</th>
<th>4</th>
<th>5</th>
<th>Total</th>
<th>95% CI</th>
</tr>
</thead>
<tbody>
<tr>
<td>Positive</td>
<td>100.0% (3/3)</td>
<td>100.0% (3/3)</td>
<td>100.0% (3/3)</td>
<td>100.0% (3/3)</td>
<td>100.0% (3/3)</td>
<td>100.0% (15/15)</td>
<td>79.6%-100.0%</td>
</tr>
<tr>
<td>Negative</td>
<td>100.0% (3/3)</td>
<td>100.0% (3/3)</td>
<td>100.0% (3/3)</td>
<td>100.0% (2/2)*</td>
<td>100.0% (3/3)</td>
<td>100.0% (14/14)</td>
<td>78.5%-100.0%</td>
</tr>
</tbody>
</table>

*Initial and retest yielded invalid results
References


5. FDA The GISAID Initiative, which promotes the rapid sharing of data from all influenza viruses and the coronavirus causing COVID-19: https://www.gisaid.org/

Note: Safety Data Sheets (SDS) are available at Visby Medical Customer Support 1-833-GoVisby (1-833-468-4729) or support@visbymedical.com.

Note: For information on how to obtain additional materials, contact Visby Medical Customer Support at 1-833-GoVisby(1-833-468-4729) or support@visbymedical.com.

Index of Symbols

<table>
<thead>
<tr>
<th>Symbol/Reference number</th>
<th>Title</th>
<th>Symbol/Reference number</th>
<th>Title</th>
</tr>
</thead>
<tbody>
<tr>
<td>5.1.6</td>
<td>Catalog number</td>
<td>5.3.7</td>
<td>Temperature limit</td>
</tr>
<tr>
<td>5.4.2 REF</td>
<td>Do not re-use</td>
<td>5.3.8</td>
<td>Humidity limitation</td>
</tr>
<tr>
<td>5.3.1</td>
<td>Fragile, handle with care</td>
<td>5.4.1</td>
<td>Biological risks</td>
</tr>
<tr>
<td>5.1.5 LOT</td>
<td>Batch code</td>
<td>5.5.1</td>
<td>In vitro diagnostic medical device</td>
</tr>
<tr>
<td>5.4.4 Caution</td>
<td>Do not use if package is damaged</td>
<td></td>
<td></td>
</tr>
<tr>
<td>5.4.3 Consult instructions for use</td>
<td>5.5.3</td>
<td>Negative control</td>
<td></td>
</tr>
<tr>
<td>5.1.1 Manufacturer</td>
<td>5.5.4 Positive control</td>
<td></td>
<td></td>
</tr>
<tr>
<td>5.5.5 Contains sufficient for m/qtests</td>
<td>21CFR801.109</td>
<td>For prescription use only</td>
<td></td>
</tr>
<tr>
<td>5.1.4 Use-by date</td>
<td>Nemko 61010</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Power supply</td>
<td>Waste container</td>
<td></td>
<td></td>
</tr>
</tbody>
</table>
Materials Provided and Required

Visby COVID-19 Device
- Upper Bulb
- Lower Bulb
- Shaft
- Status Lights
- Power Connection
- Button 1 (Sample Port)
- Button 2
- Button 3

Visby Pastette Biohazard Bag

Visby COVID-19 Device

Materials Required but not Supplied
- Gloves
- Nasopharyngeal, Mid-turbinate or Anterior Nasal Specimen Collection Swabs
- Disposal Bin
- Absorbent Pads

Materials Required and Supplied
- Visby Buffer Tube
- Test Tube Holder
- Visby Power Adapter

Storage Specifications

Store the Visby COVID-19 Point of Care Test in a cool and dry environment (36°F-86°F). Do not freeze.

Ensure Visby COVID-19 Device comes to minimum operating temperature before use.

Patient swab sample is stable in dry tube for 1 hour at room temperature.

Patient sample is stable in Visby Buffer for 24 hours at refrigerated temperature and 45 minutes at room temperature. Ensure the sample comes to minimum operating temperature before adding to the Visby COVID-19 Device.

Quality Control

External controls should be used in accordance with local, state, and federal accrediting organizations as applicable. Visby Medical suggests the use of the following control materials, however other commercial external control materials may be appropriate.

External Positive and Negative Controls

SARS-CoV-2 Positive and A549 Cells Negative External Control (Swabs) by Microbiologics®

To run external control swabs, unwrap the swab, and gently tap the swab against the bottom of the Visby Buffer Tube 15 times. Discard the swabs according to your institutions guidelines and screw the cap back onto the Visby Buffer Tube. Proceed to Step 3.

External control stability in the Visby Buffer has not been established. Each positive or negative external control is for single use only.

If the positive or negative external controls fail, repeat with new external control and a new Visby COVID-19 Point of Care Test. If the repeat test fails, please contact Visby Medical Customer Support at 1-833-468-4729 (1-833-GoVisby).

Operating Conditions

Temperature Definitions

Room Temperature: 66°F - 82°F
Refrigerated Temperature: 35°F - 46°F

Caution

Keep the work area clean to prevent contamination.
Wear gloves while handling samples and change gloves between testing each specimen.

Color Blindness Precaution

While color-blind users may be unable to differentiate red, green, and white status lights, they may observe the light location and shape of the light to determine test status.

For the qualitative detection of SARS-CoV-2 nucleic acid. For IVU use. For Rx only. For Emergency Use Authorization Only.

For urgent assistance, visit our website or call our Customer Support team.

For help, email us at support@visbymedical.com or call us at 1-833-GoVisby (1-833-468-4729).

Need Help?

Email Us
support@visbymedical.com

Call Us
1-833-GoVisby
(1-833-468-4729)

www.visbymedical.com
Step 1  Set Up the Workspace

Operating Conditions: Ensure the test is run at room temperature in a cool, dry environment. Set up a new workspace for each Visby COVID-19 Point of Care Test. Clean the workspace and use a new absorbent pad after each test. Change gloves between handling samples and setting up a new test. Place the Visby COVID-19 Device on a level surface.

Note: Leave the Visby COVID-19 Device seated in the foil pouch until just before use. Please proceed to Step 2 only when ready to complete the test. Do not use the Visby COVID-19 Device if it appears broken or has been dropped. Do not use the Visby COVID-19 Point of Care Test past its expiration date.

Step 2  Add Sample to the Visby Buffer Tube

Patient sample must be used with Visby Buffer only. Place the device on a level surface.

A. Open the Visby Buffer Tube. Note: Do not use the Visby Buffer Tube past its expiration date.

B. Place the Visby Buffer Tube in a tube holder.

C. Take the collected patient sample.

D. Place the patient swab into the Visby Buffer Tube. Note: If your test has a breakpoint - break the handle of the swab.

E. Screw cap back on the Visby Buffer Tube.

Note: Patient sample is stable in a dry tube for 1 hour at room temperature. Patient sample must be added to Visby Buffer when ready to run the test. Do not use the Visby COVID-19 Device after adding a sample.

Step 3  Load the Sample into the Device

A. Stop! Do not plug in the test until Step 4E.

B. Mix the specimen in the Visby Buffer Tube by inverting the tube 5 times.

C. Open the cap of the Visby Buffer Tube. Place cap and swab on the Visby palette.

D. Squeeze the upper bulb.

E. Keep the tip fully under the fluid. Release the upper bulb.

F. Fill the extra shaft with fluid. Some fluid should enter the lower bulb.

G. Gently press the tiny shaft until fluid is present in the lower bulb.

H. Squeeze the tip to drain off fluid. Some fluid will remain in the lower bulb.

I. Place the tip of the palette into the sample port (Button 1). Squeeze the bulb to drain off fluid.

J. Some fluid will remain in the lower bulb. Note: Do not overfill.

Step 4  Run the Test

A. IMPORTANT! Each button will have a different test and is “clicks” into place. Push firmly to make sure all buttons are completely down or the test may not work.

B. Push Button 1 all the way down to load the sample.

C. Push Button 2 all the way down to unlock button 3.

D. Push Button 3 all the way down to add the sample.

Note: All buttons should be all the way down.

E. Plug in the device until it clicks into place. A stable white light indicates the test is running. Ensure that there is no gap between the power adapter plug and device.

F. WAIT 3 MINUTES! Do not touch or move the charging adapter, cable, or device. Do NOT shake or tilt the Visby COVID-19 Device after adding a sample.

Step 5  Get the Results

A. After 30 minutes.

B. If a green check mark appears, confirm the results are valid. Look at the results window for a purple spot near “RESULTS VALID.”

C. Read and record the results. Look at the results window for a purple spot near “POSITIVE for SARS-CoV-2 (COVID-19).” Results may be read for up to 2 hours after the test is completed. The intensity of the spot in the results window may vary. Any shade of color should be considered a spot.

Note: Visby Buffer, and specimen collection kit should be disposed of in the appropriate specimen waste containers according to your institution’s standard practices.

Positive Result

Negative Result

POSITIVE for SARS-CoV-2 (COVID-19)

NEGATIVE for SARS-CoV-2 (COVID-19)

Need Help? Call 1-833-GoVisby (1-833-468-4729)