Elecsys Anti-SARS-CoV-2

- For in vitro diagnostic and Laboratory Professional use. For use under the Emergency Use Authorization (EUA) only

**English**

**System information**
- For Cobas e 411 analyzer: test number 3000
- For Cobas e 601 and Cobas e 602 analyzers: Application Code Number 737
- For Cobas e 801 analyzer: Application Code Number 10226

**Warning**
- Not for screening of donated blood.

**Intended use**
Elecsys Anti-SARS-CoV-2 is an immunoassay intended for qualitative detection of antibodies to SARS-CoV-2 in human serum and plasma (K2, EDTA, K3, EDTA, Li-heparin). The test is intended for use as an aid in identifying individuals with an adaptive immune response to SARS-CoV-2, indicating recent or prior infection. At this time, it is unknown for how long antibodies persist following infection and if the presence of antibodies confers protective immunity. The Elecsys Anti-SARS-CoV-2 assay should not be used to diagnose acute SARS-CoV-2 infection. Testing is limited to laboratories certified under the Clinical Laboratory Improvement Amendments of 1988 (CLIA), 42 U.S.C 263a, to perform moderate and high complexity tests.

Results are for the detection of SARS-CoV-2 antibodies. Antibodies to SARS-CoV-2 are generally detectable in blood several days after initial infection, although the duration of time antibodies are present post-infection is not well characterized. Patients may have detectable virus present for several weeks following seroconversion.

Laboratories within the United States and its territories are required to report all positive results to the appropriate public health authorities.

The sensitivity of the Elecsys Anti-SARS-CoV-2 assay early after infection is unknown. Negative results do not preclude acute SARS-CoV-2 infection. If acute infection is suspected, direct testing for SARS-CoV-2 is necessary. False positive results for the Elecsys Anti-SARS-CoV-2 assay may occur due to cross reactivity from pre-existing antibodies or other possible causes.

The Elecsys Anti-SARS-CoV-2 assay is only for use under the Food and Drug Administration’s Emergency Use Authorization.

The electrochemiluminescence immunoassay “ECLIA” is intended for use on Cobas e immunoassay analyzers.

**Summary**
SARS-CoV-2 is an enveloped, single-stranded RNA virus of the family Coronaviridae, genus Beta-coronavirus. All coronaviruses share similarities in the organization and expression of their genome, which encodes 16 nonstructural proteins and the 4 structural proteins: spike (S), envelope (E), membrane (M), and nucleocapsid (N). Viruses of this family are of zoonotic origin. They cause disease with symptoms ranging from those of a mild common cold to more severe ones such as the Severe Acute Respiratory Syndrome (SARS), Middle East Respiratory Syndrome (MERS) and Coronavirus Disease 2019 (COVID-19). Other coronaviruses known to infect people include 229E, NL63, OC43 and HKU1. The latter are ubiquitous and infection typically causes common cold or flu-like symptoms.

The Elecsys Anti-SARS-CoV-2 assay uses a recombinant protein representing the nucleocapsid (N) antigen for the determination of antibodies against SARS-CoV-2.

**Test principle**
Sandwich principle. Total duration of assay: 18 minutes.

**Reagents - working solutions**
- Cobas e 411, Cobas e 601, and Cobas e 602 analyzers:
  - The reagent rackpack (M, R1, R2) is labeled as ACOV2.
  - M Streptavidin-coated microcarriers (transparent cap), 1 bottle, 12 mL: Streptavidin-coated microcarriers 0.72 mg/mL; preservative.
  - R1 SARS-CoV-2-Ag-biotin, (gray cap), 1 bottle, 16 mL: Biotinylated SARS-CoV-2-specific recombinant antigen (E. coli); preservative.
  - R2 SARS-CoV-2 Ag-Ru(bpy)3+ (black cap), 1 bottle, 16 mL: SARS-CoV-2-specific recombinant antigen labeled with ruthenium complex; preservative.
- ACOV2 Cal1 Negative calibrator 1 (white cap), 1 bottle of 0.67 mL: Human serum, non-reactive for anti-SARS-CoV-2 antibodies; buffer; preservative.
- ACOV2 Cal2 Positive calibrator 2 (black cap), 1 bottle of 0.67 mL: Human serum, reactive for anti-SARS-CoV-2 antibodies; buffer; preservative.

**Cobas e 801 analyzer**
- The Cobas e pack (M, R1, R2) is labeled as ACOV2.
  - M Streptavidin-coated microcarriers, 1 bottle, 16 mL: Streptavidin-coated microcarriers 0.72 mg/mL; preservative.
  - R1 SARS-CoV-2-Ag-biotin, 1 bottle, 18.8 mL: Biotinylated SARS-CoV-2-specific recombinant antigen (E. coli); preservative.
  - R2 SARS-CoV-2-Ag-Ru(bpy)3+ 1 bottle, 18.8 mL: SARS-CoV-2-specific recombinant antigen labeled with ruthenium complex; preservative.

**ACOV2 Cal1** Negative calibrator 1, 1 bottle of 0.67 mL: Human serum, non-reactive for anti-SARS-CoV-2 antibodies; buffer; preservative.

**ACOV2 Cal2** Positive calibrator 2, 1 bottle of 0.67 mL: Human serum, reactive for anti-SARS-CoV-2 antibodies; buffer; preservative.

**ACOV2 Cal1** Negative calibrator 1, 1 bottle of 0.67 mL: Human serum, non-reactive for anti-SARS-CoV-2 antibodies; buffer; preservative.
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ACOV2 Cal2 Positive calibrator 2, 1 bottle of 0.67 mL:
Human serum, reactive for anti-SARS-CoV-2 antibodies; buffer, preservative.

Precautions and warnings
For Emergency Use Authorization only.
For in vitro diagnostic use.
Exercise the normal precautions required for handling all laboratory reagents.
Disposal of all waste material should be in accordance with local guidelines.
Safety data sheet available for professional user on request.
For USA: Caution: Federal law restricts this device to sale by or on the order of a physician.
This kit contains components classified as follows in accordance with the Regulation (EC) No. 1272/2008:

Warning
H317 May cause an allergic skin reaction.

Prevention:
P261 Avoid breathing dust/fume/gas/mist/vapours/spray.
P272 Contaminated work clothing should not be allowed out of the workplace.
P280 Wear protective gloves.

Response:
P333 + P313 If skin irritation or rash occurs: Get medical advice/attention.
P362 + P364 Take off contaminated clothing and wash it before reuse.

Disposal:
P501 Dispose of contents/container to an approved waste disposal plant.

Product safety labeling follows EU GHS guidance.
Contact phone: 1-866-987-6243

All human material should be considered potentially infectious.
All products derived from human blood are prepared exclusively from the blood of donors tested individually and shown to be free from HBsAg and antibodies to HCV and HIV.
The testing methods used assays approved by the FDA or cleared in compliance with the European Directive 98/79/EC, Annex II, List A.
The serum containing anti-SARS-CoV-2 (ACOV2 Cal2) was heat-inactivated for 30 minutes at 56 °C.
However, as no inactivation or testing method can rule out the potential risk of infection with absolute certainty, the material should be handled with the same level of care as a patient specimen.
In the event of exposure, the directives of the responsible health authorities should be followed.
Avoid foam formation in all reagents and sample types (specimens, calibrators and controls).

Reagent handling
The reagents in the kit are ready-for-use and are supplied in bottles compatible with the system.

Calibrators:
The calibrators are supplied ready-for-use in bottles compatible with the system.

cobas e 411 analyzer: The calibrators should only be left on the analyzer during calibration at 20-25 °C. After use, close the bottles as soon as possible and store upright at 2-8 °C.
Due to possible evaporation effects, not more than 4 calibration procedures per calibrator bottle set should be performed.

cobas e 801 analyzer: Store the calibrators at 2-8 °C for later use.

cobas e 601, cobas e 602 and cobas e 801 analyzers:
Perform only one calibration procedure per bottle.

cobas e 411, cobas e 601 and cobas e 602 analyzers:
All information required for correct operation is read in from the respective reagent barcodes.

cobas e 801 analyzers:
All information required for correct operation is available via the cobas link.
Please note for cobas e 602 analyzers: Both the vial labels contain 2 different barcodes. The barcode between the yellow markers is for cobas 8000 systems only. If using a cobas 8000 system, please turn the vial cap 180° into the correct position so that the barcode between the yellow markers can be read by the system. Place the vial on the analyzer as usual.

Storage and stability
Store at 2-8 °C.
Do not freeze.

Store the Elecsys reagent kit / cobas e pack upright in order to ensure complete availability of the microparticles during automatic mixing prior to use.

<table>
<thead>
<tr>
<th>Stability of the reagent rack pack</th>
<th>up to the stated expiration date</th>
</tr>
</thead>
<tbody>
<tr>
<td>unopened at 2-8 °C</td>
<td>up to the stated expiration date</td>
</tr>
<tr>
<td>after opening at 2-8 °C</td>
<td>72 hours</td>
</tr>
<tr>
<td>on the cobas e 411, cobas e 601</td>
<td>72 hours</td>
</tr>
<tr>
<td>and cobas e 602 analyzers</td>
<td></td>
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</tbody>
</table>

<table>
<thead>
<tr>
<th>Stability of the cobas e pack</th>
<th>up to the stated expiration date</th>
</tr>
</thead>
<tbody>
<tr>
<td>unopened at 2-8 °C</td>
<td>up to the stated expiration date</td>
</tr>
<tr>
<td>after opening at 2-8 °C</td>
<td>72 hours</td>
</tr>
<tr>
<td>on the cobas e 801 analyzer</td>
<td>72 hours</td>
</tr>
</tbody>
</table>

<table>
<thead>
<tr>
<th>Stability of the calibrators</th>
<th></th>
</tr>
</thead>
<tbody>
<tr>
<td>unopened at 2-8 °C</td>
<td>up to the stated expiration date</td>
</tr>
<tr>
<td>after opening at 2-8 °C</td>
<td>72 hours</td>
</tr>
<tr>
<td>on cobas e 411 at 20-25 °C</td>
<td>up to 3 hours</td>
</tr>
<tr>
<td>on cobas e 601, cobas e 602, and</td>
<td>use only once</td>
</tr>
<tr>
<td>cobas e 801 analyzers at 20-25 °C</td>
<td></td>
</tr>
</tbody>
</table>

Store calibrators upright in order to prevent the calibrator solution from adhering to the snap-cap.

Specimen collection and preparation
Only the specimens listed below were tested and found acceptable.
Serum collected using standard sampling tubes or tubes containing separating gel.
Li-heparin, K2-EDTA and K2-EDTA plasma.
Plasma tubes containing separating gel can be used.
Criterion: Absolute deviation of negative samples ± 0.3 COI (cutoff index) from serum value; reactive samples: recovery within 70-130 % of serum value.
Stable for 3 days at 15-25 °C, 7 days at 2-8 °C, 28 days at -20 °C (± 5 °C).
The samples may be frozen once.
The sample types listed were tested with a selection of sample collection tubes or systems that were commercially available at the time of testing, i.e. not all available tubes of all manufacturers were tested. Sample collection systems from various manufacturers may contain differing materials which could affect the test results in some cases. When processing samples in primary tubes (sample collection systems), follow the instructions of the tube/collection system manufacturer.
Specimens should not be subsequently altered with additives (e.g. biocides, anti-oxidants or substances that could possibly change the pH or ionic strength of the sample) in order to avoid erroneous findings.
Pooled samples and other artificial material may have different effects on different assays and thus may lead to discrepant findings.

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Centrifuge samples containing precipitates and thawed samples before performing the assay. Do not use heat-inactivated samples. Do not use samples and controls stabilized with azide. Ensure the samples, calibrators and controls are at 20-25 °C prior to measurement. Due to possible evaporation effects, samples, calibrators and controls on the analyzers should be analyzed/measured within 2 hours. The performance of the Elecsys Anti-SARS-CoV-2 assay has not been established with cadaveric samples or body fluids other than serum and plasma. Sample stability claims were established by experimental data by the manufacturer or based on reference literature and only for the temperatures/lime frames as stated in the method sheet. It is the responsibility of the individual laboratory to use all available references and/or its own studies to determine specific stability criteria for its laboratory.

Materials provided
See “Reagents – working solutions” section for reagents.

Materials required (but not provided)
- 03609987190, Diluent MultiAssay, 2 x 16 mL sample diluent
- 07299010190, Diluent MultiAssay, 45.2 mL sample diluent
- General laboratory equipment
- cobas e analyzer

Additional materials for the cobas e 411 analyzer:
- 11662988122, ProCell, 6 x 380 mL system buffer
- 11662970122, CleanCell, 6 x 380 mL measuring cell cleaning solution
- 11930346122, Elecsys SysWash, 1 x 500 mL washwater additive
- 11933159001, Adapter for SysClean
- 11706802001, AssayCup, 60 x 60 reaction cups
- 11706799001, AssayTip, 30 x 120 pipette tips
- 11800507001, Clean-Liner

Additional materials for cobas e 601 and cobas e 602 analyzers:
- 04880340190, ProCell M, 2 x 2 L system buffer
- 04880293190, CleanCell M, 2 x 2 L measuring cell cleaning solution
- 03023141001, PC/CC-Cups, 12 cups to prewarm ProCell M and CleanCell M before use
- 03005712190, ProbeWash M, 12 x 70 mL cleaning solution for run initialization and rinsing during reagent change
- 12102137001, AssayTip/AssayCup, 48 magazines x 84 reaction cups or pipette tips, waste bags
- 03023150001, WasteLiner, waste bags
- 03027651001, SysClean Adapter M

Additional materials for the cobas e 601 analyzer:
- 06908799190, ProCell II M, 2 x 2 L system solution
- 04880293190, CleanCell M, 2 x 2 L measuring cell cleaning solution
- 07448540901, Reservoir Cup, 8 cups to supply ProCell II M and CleanCell M
- 05694302001, Assay Tip/Assay Cup tray, 6 magazines x 6 magazine stacks x 105 assay tips and 105 assay cups, 3 wasteliners
- 07448542501, Liquid Flow Cleaning Cup, 2 adaptor cups to supply ISE Cleaning Solution/Elecsys SysClean for Liquid Flow Cleaning Detection Unit
- 07448543300, PreWash Liquid Flow Cleaning Cup, 1 adaptor cup to supply ISE Cleaning Solution/Elecsys SysClean for Liquid Flow Cleaning PreWash Unit

Additional materials for all analyzers:
- 11298500160, ISE Cleaning Solution/Elecsys SysClean, 5 x 100 mL system cleaning solution

Assay
For optimum performance of the assay follow the directions given in this document for the analyzer concerned. Refer to the appropriate operator’s manual for analyzer-specific assay instructions. Resuspension of the microparticles takes place automatically prior to use.

cobas e 411, cobas e 601, and cobas e 602 analyzers:
Read in the test-specific parameters via the reagent barcode. If in exceptional cases the barcode cannot be read, enter the 15-digit sequence of numbers. Bring the cooled reagents to approximately 20 °C and place on the reagent disk (20 °C) of the analyzer.

cobas e 801 analyzer:
Place the cooled (stored at 2-8 °C) cobas e pack on the reagent manager. Avoid foam formation. The system automatically regulates the temperature of the reagents and the opening/closing of the bottles and cobas e pack.

Calibrators:
Place the calibrators in the sample zone.

cobas e 411, cobas e 601, and cobas e 602 analyzers:
All the information necessary for calibrating the assay is automatically read into the analyzer.

cobas e 801 analyzers:
Read in all the information necessary for calibrating the assay. After calibration has been performed, store the calibrators at 2-8 °C or discard (cobas e 601, cobas e 602 and cobas e 801 analyzers).

Calibration
No international standard is available for Anti-SARS-CoV-2.

Calibration frequency: Calibration must be performed once per reagent lot using ACOV2 Cal1, ACOV2 Cal2 and fresh reagent (i.e., not more than 24 hours since the reagent kit / cobas e pack was registered on the analyzer).

Calibration interval may be extended based on acceptable verification of calibration by the laboratory.

Renewed calibration is recommended as follows:
- after 3 days when using the same reagent lot
- after 3 days when using the same reagent kit / cobas e pack on the analyzer
- as required: e.g. quality control findings outside the defined limits

Quality control
For quality control, use controls prepared as follows:

Negative control: Determine the COI of ACOV2 Cal1 by measuring it as a routine sample. Pool serum samples with a COI result of ≤ 150 % compared to the COI result of ACOV2 Cal1 (pooling of ≥ 5 non-reactive samples in this range is recommended). Mix carefully, avoiding foam formation. Prepare aliquots of at least 250 µL from this sample pool and store frozen at -20 °C (± 5 °C) or colder. Use these aliquots to perform regular quality control.

This negative control has a target value range of COI < 0.8 (qualitative assay result “non-reactive”).

Positive control: Determine the COI of ACOV2 Cal2 by measuring it as a routine sample. Pool serum samples with a COI result that is higher than the COI result of ACOV2 Cal2 (pooling of ≥ 3 reactive samples in this range is recommended). Dilute the sample pool by adding pooled negative serum (for pooling criterion see negative control) or Diluent MultiAssay to obtain a COI between 3 and 15. Mix carefully, avoiding foam formation. It is recommended to confirm calculated reactivity after dilution by a measurement. Prepare aliquots of at least 250 µL from this sample pool and store frozen at -20 °C (± 5 °C) or colder. Use these aliquots to perform regular quality control. Upon first use of this control, determine the COI of the control by measurement of the control in triplicate and using a freshly opened and calibrated reagent rack pack / cobas e pack.

The obtained median of these measurements serves as target value for this positive control. Subsequent measurements of all aliquots of this control material must match this target value ± 45 % (3SD = 45 %, 1SD = 15 %; qualitative assay result “reactive”).
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The target value of the positive control is lot specific and target value assessment as described above has to be performed for every assay lot. After measurement, discard aliquots with a remaining volume of 250 µL or less. Aliquots with a remaining volume of more than 250 µL can be re-used if sealed tightly and stored immediately at 2-8 °C for a maximum of 3 days. In case quality control fails for any reason, thaw a new control aliquot and re-assess the performance of the assay.

Pools of plasma samples with similar reactivity can be used, however fibrin clots frequently occur with plasma after thawing. If this occurs, either discard or centrifuge the aliquot before use. Do not mix serum samples and plasma samples to prepare a sample pool.

The control intervals and limits should be adapted to each laboratory’s individual requirements. Values obtained should fall within the defined limits. Each laboratory should establish corrective measures to be taken if values fall outside the defined limits.

Controls for the various concentration ranges should be run individually at least once every 24 hours when the test is in use, once per reagent kit/cobas e pack, and following each calibration.

If necessary, repeat the measurement of the samples concerned.

Follow the applicable government regulations and local guidelines for quality control.

Note: The controls should be run like external controls. All values and ranges have to be entered manually. Please refer to the section “QC” in the operator’s manual or to the online help of the instrument software. Only one target value and range for each control level can be entered in the analyzer. The reagent lot-specific target values have to be re-entered each time a specific reagent lot with different control target values and ranges is used. Two reagent lots with different control target values and ranges cannot be used in parallel in the same run.

Calculation

The analyzer automatically calculates the cutoff based on the measurement of ACOV2 Cat1 and ACOV2 Cat2. The result of a sample is given either as reactive or non-reactive as well as in the form of a cutoff index (COI; signal sample/cutoff).

Interpretation of the results

Results obtained with the Elecsys Anti-SARS-CoV-2 assay can be interpreted as follows:

<table>
<thead>
<tr>
<th>Numeric result</th>
<th>Result message</th>
<th>Interpretation</th>
</tr>
</thead>
<tbody>
<tr>
<td>COI &lt; 1.0</td>
<td>Non-reactive</td>
<td>Negative for anti-SARS-CoV-2 antibodies</td>
</tr>
<tr>
<td>COI ≥ 1.0</td>
<td>Reactive</td>
<td>Positive for anti-SARS-CoV-2 antibodies</td>
</tr>
</tbody>
</table>

The magnitude of the measured result above the cutoff is not indicative of the total amount of antibody present in the sample. The individual immune response following SARS-CoV-2 infection varies considerably and might give different results with assays from different manufacturers. Results of assays from different manufacturers should not be used interchangeably.

Limitations - interference

The effect of the following pharmaceutical compound on assay performance was tested. Interference was tested up to the listed concentration and no impact on results was observed.

Endogenous substance

<table>
<thead>
<tr>
<th>Compound</th>
<th>Concentration tested</th>
</tr>
</thead>
<tbody>
<tr>
<td>Biotin</td>
<td>≤ 4912 nmol/L or ≤ 1200 ng/mL</td>
</tr>
</tbody>
</table>

This assay has no biotin interference in serum concentrations up to 1200 mg/mL. Some studies have shown that serum concentrations of biotin can reach up to 355 mg/mL within the first hour after biotin ingestion for subjects consuming supplements of 20 mg biotin per day and up to 1160 ng/mL for subjects after a single dose of 300 mg biotin.

Potential endogenous interferences e.g. hemolysis, bilirubin, rheumatoid factors and pharmaceutical compounds other than biotin have not been tested and an interference cannot be excluded.

In rare cases, interference due to extremely high titers of antibodies to analyte-specific antibodies, streptavidin or ruthenium can occur. These effects are minimized by suitable test design.

The results should always be assessed in conjunction with the patient’s medical history, clinical examination and other findings.

A negative test result does not rule out the possibility of an infection with SARS-CoV-2. Serum or plasma samples from the early (pre-seroconversion) phase of illness can yield negative findings. Therefore, this test cannot be used to diagnose an acute infection. Also, over time, titers may decline and eventually become negative.

Testing with a molecular diagnostic should be performed to evaluate for active infection in symptomatic individuals.

It is not known at this time if the presence of antibodies to SARS-CoV-2 confers immunity to reinfection.

Conditions of Authorization for the Laboratory

The Elecsys Anti-SARS-CoV-2 assay Letter of Authorization, along with the authorized Fact Sheet for Healthcare Providers, the authorized Fact Sheet for Patients, and authorized labeling are available on the FDA website: https://www.fda.gov/medical-devices/emergency-situations-medical-devices/emergency-use-authorization/covid19ld. However, to assist clinical laboratories using the Elecsys Anti-SARS-CoV-2 (“your product” in the conditions below), the relevant Conditions of Authorization are listed below:

- Authorized laboratories using your product will include with result reports of your product, all authorized Fact Sheets. Under exigent circumstances, other appropriate methods for disseminating these Fact Sheets may be used, which may include mass media.
- Authorized laboratories using your product will use your product as outlined in the instructions for Use. Deviations from the authorized procedures, including the authorized instruments, authorized clinical specimen types, authorized control materials, authorized other ancillary reagents and authorized materials required to use your product are not permitted.
- Authorized laboratories that receive your product will notify the relevant public health authorities of their intent to run your product prior to initiating testing.
- Authorized laboratories using your product will have a process in place for reporting test results to healthcare providers and relevant public health authorities, as appropriate.
- Authorized laboratories will collect information on the performance of your product and report to DMD/DHT/7-QIR/OPED/CDRH (via email: CDRH-EUA-Reporting@fas.hhs.gov) and Roche (1-866-987-6243) any suspected occurrence of false reactive or false non-reactive results and significant deviations from the established performance characteristics of your product of which they become aware.
- All laboratory personnel using your product must be appropriately trained in automated immunosassay techniques and use appropriate laboratory and personal protective equipment when handling this kit, and use your product in accordance with the authorized labeling. All laboratory personnel using the assay must also be trained in and be familiar with the interpretation of results of the product.
- Roche, authorized distributors, and authorized laboratories using your product will ensure that any records associated with this EUA are maintained until otherwise notified by FDA. Such records will be made available to FDA upon request.

The letter of authorization refers to the “Laboratories certified under the Clinical Laboratory Improvement Amendments of 1988 (CLIA), 42 U.S.C. §263a, to perform moderate or high complexity tests” as “authorized laboratories”.

Specific performance data

Representative performance data on the analyzers are given below. Results obtained in individual laboratories may differ.

Cross-reactivity

A study was conducted to evaluate the Elecsys Anti-SARS-CoV-2 assay for potential cross-reactivity in specimens obtained before December 2019. The following results were obtained:

- 10 HJV, 16 Anti-HCV, 7 Anti-HBs, 8 Anti-HBC IgM, 9 ANA, 10 Human coronavirus 229E, 10 Human coronavirus OC43, 10 Human coronavirus HKU1, 10 Human coronavirus NL63 and 40 Common cold samples were measured.
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All sample results were non-reactive. In addition, due to prevalence of immunization and/or exposure to common viral agents in the general population, the Specificity section below indicates no major cross-reactivity to antibodies for Hepatitis B, Influenza A/B, Haemophilus, common colds (i.e. Rhinovirus), Respiratory Syncytial Virus, or the Coronavirus strains listed.

Specificity
A total of 5272 samples were tested with the Elecsys Anti-SARS-CoV-2 assay. All samples were obtained before December 2019. 10 false positive samples were detected. The resulting overall specificity in the internal study was 99.81%. The 95% lower confidence limit was 99.65%.

<table>
<thead>
<tr>
<th>Cohort</th>
<th>N</th>
<th>Non-reactive</th>
<th>Reactive</th>
<th>Specificity, % (95% CI)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Diagnostic routine</td>
<td>3420</td>
<td>3413</td>
<td>7</td>
<td>99.80 (99.58-99.92)</td>
</tr>
<tr>
<td>Blood donors</td>
<td>1772</td>
<td>1769</td>
<td>3</td>
<td>99.83 (99.51-99.97)</td>
</tr>
<tr>
<td>Common cold panel</td>
<td>40</td>
<td>40</td>
<td>0</td>
<td>100 (91.19-100)</td>
</tr>
<tr>
<td>Coronavirus panel(2)</td>
<td>40</td>
<td>40</td>
<td>0</td>
<td>100 (91.19-100)</td>
</tr>
<tr>
<td>Overall</td>
<td>5272</td>
<td>5262</td>
<td>10</td>
<td>99.81 (99.65-99.91)</td>
</tr>
</tbody>
</table>

c) CI = confidence interval
d) 40 potentially cross-reactive samples from individuals following an infection with Coronavirus HKU1, NL63, 229E or OC43, confirmed via PCR

Sensitivity
A total of 204 samples from 69 symptomatic patients with a PCR confirmed SARS-CoV-2 infection were tested with the Elecsys Anti-SARS-CoV-2 assay. 1 or more consecutive specimens from these patients were collected after PCR confirmation at various time points.

<table>
<thead>
<tr>
<th>Days post PCR confirmation</th>
<th>N</th>
<th>Reactive</th>
<th>Non-reactive</th>
<th>Sensitivity, % (95% CI)</th>
</tr>
</thead>
<tbody>
<tr>
<td>G-6</td>
<td>116</td>
<td>76</td>
<td>40</td>
<td>65.5 (56.1-74.1)</td>
</tr>
<tr>
<td>7-13</td>
<td>59</td>
<td>52</td>
<td>7</td>
<td>88.1 (77.1-95.1)</td>
</tr>
<tr>
<td>≥ 14</td>
<td>29</td>
<td>29</td>
<td>0</td>
<td>100 (88.1-100)</td>
</tr>
</tbody>
</table>

After recovery from infection, confirmed by a negative PCR result, 26 consecutive samples from 5 individuals were tested with the Elecsys Anti-SARS-CoV-2 assay.

<table>
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<th></th>
<th></th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>9</td>
<td>COI</td>
<td>24.7</td>
<td>-</td>
<td>27.4</td>
<td>31.7</td>
<td>38.9</td>
<td>58.0</td>
<td>-</td>
<td></td>
</tr>
<tr>
<td>2</td>
<td>12</td>
<td></td>
<td>25.8</td>
<td>25.8</td>
<td>30.6</td>
<td>32.7</td>
<td>35.7</td>
<td>-</td>
<td>-</td>
<td></td>
</tr>
<tr>
<td>3</td>
<td>17</td>
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* Day 0 represents initial positive PCR

References


For further information, please refer to the appropriate operator’s manual for the analyzer concerned, the respective application sheets, the product information and the Method Sheets of all necessary components (if available in your country).

Symbols
Roche Diagnostics uses the following symbols and signs in addition to those listed in the ISO 15223-1 standard (for USA: see dialog.roche.com for definition of symbols used):