
Population Pharmacokinetics Guidance for Industry

DRAFT GUIDANCE

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**U.S. Department of Health and Human Services
Food and Drug Administration
Center for Drug Evaluation and Research (CDER)
Center for Biologics Evaluation and Research (CBER)**

**July 2019
Clinical Pharmacology**

Population Pharmacokinetics Guidance for Industry

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**Population Pharmacokinetics
Guidance for Industry¹**

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I. INTRODUCTION

This guidance is intended to assist sponsors of new drug applications (NDAs) and biologics license applications (BLAs) in the application of population pharmacokinetic (population PK) analysis. Population PK analysis is frequently used to guide drug² development and inform recommendations on therapeutic individualization (e.g., through tailored dosing) (Marshall et al. 2015; Lee et al. 2011; Bhattaram et al. 2005). Adequate population PK data collection and analyses submitted in marketing applications have in some cases alleviated the need for postmarketing requirements (PMRs) or postmarketing commitments (PMCs).

This guidance includes common applications of population PK analysis to inform drug development and drug use. This list of applications is not meant to be comprehensive, but rather provides illustrative examples. This guidance also includes the FDA’s current thinking on the data and model requirements needed to support regulatory decisions, recommendations to sponsors on drug labeling based on population PK analysis, and the general expectations regarding the format and content for population PK reports submitted to the Agency.

In general, FDA’s guidance documents do not establish legally enforceable responsibilities. Instead, guidances describe the Agency’s current thinking on a topic and should be viewed only as recommendations, unless specific regulatory or statutory requirements are cited. The use of the word *should* in Agency guidances means that something is suggested or recommended, but not required.

¹ This guidance has been prepared by Office of Clinical Pharmacology in the Center for Drug Evaluation (CDER) and Research and the Center for Biologics Evaluation and Research (CBER) at the Food and Drug Administration.

² For purposes of this guidance, references to *drugs* and *drug and biological products* includes drugs approved under section 505 of the Federal Food, Drug, and Cosmetic Act (the FD&C Act or Act) (21 U.S.C. 355) and biological products licensed under 351 of the Public Health Service Act (PHSA) (42 U.S.C. 262) that are drugs.

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38 **II. BACKGROUND**

39
40 Population PK analysis is a well-established, quantitative method that can quantify and explain
41 the variability in drug concentrations among individuals (Sheiner, Rosenberg, and Marathe
42 1977; Grasela Jr and Sheiner 1991). Drug concentrations can vary significantly among
43 individuals who are following the same dosing regimen. Variability can be attributed to intrinsic
44 patient factors, such as the presence and extent of liver or renal impairment or the presence of
45 genetic polymorphisms, or to extrinsic patient factors, such as food consumption or concomitant
46 medications that may interact with the administered drug. In some cases, intrinsic or extrinsic factors
47 lead to clinically relevant changes in drug concentrations that require clinical management strategies,
48 such as a change in the dose or dosing regimen.

49
50 Intrinsic and extrinsic factors that commonly influence drug exposure are often investigated in stand-
51 alone clinical pharmacology studies.³ Stand-alone studies are well controlled and provide a robust
52 assessment of these interactions. However, stand-alone studies are usually designed to focus on
53 intrinsic and extrinsic factors with the highest potential to affect drug exposure, leaving many
54 possible interactions unstudied. Population PK analysis typically includes data directly collected
55 from patients, allowing an assessment of multiple intrinsic and extrinsic factors that are not
56 otherwise evaluated in healthy volunteers. In addition, the relatively large numbers of patients
57 included in population PK analysis may improve the precision of the estimated effect of the
58 factors that affect drug exposures and confirm which factors do not change drug exposures.

59
60 Population PK analysis integrates all relevant PK information across a range of doses to identify
61 factors that can affect a drug's exposure. Such information can come from studies with rich PK
62 sampling or sparse PK sampling, after a single dose or at steady state, and from healthy
63 individuals or the patient population. These analyses, in turn, can inform strategies to manage
64 dosing and administration for a given subpopulation, plan subsequent studies, or support
65 labeling.

66
67

68 **III. APPLICATION OF POPULATION PK ANALYSIS**

69
70 Sponsors seeking advice on the use of population PK analysis for drug development decisions or
71 to answer regulatory questions are encouraged to do so at appropriate milestone meetings with
72 the Agency. Sponsors should contact the Office of Clinical Pharmacology to discuss novel
73 methodologies and applications of population PK analyses to inform drug development and use.

74
75 Confidence in a given population PK analysis to support an intended objective is increased by
76 the following:

- 77
78
 - Understanding of the drug's PK properties

79

³ See IX. References for specific FDA guidances for industry that address stand-alone clinical pharmacology studies.

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- 80 • Prespecified questions in the study protocol or in the data analysis plan that will be
81 addressed by a population PK analysis
- 82
- 83 • PK data of sufficient quantity and quality that represents the indicated population and
84 relevant subpopulations of interest
- 85
- 86 • Good model performance (i.e., the model should describe the data with acceptable bias
87 and precision) and valid for the intended purpose
- 88
- 89

A. The Application of Population PK Analysis in Drug Development

1. Selecting Dosing Regimens To Be Tested in Clinical Trials

94 Population PK analyses can identify covariates that significantly impact PK variability and
95 inform dosing regimens for testing in clinical trials to help minimize the variability of treatment
96 response in patients. For example, an observed strong relationship between body weight and
97 drug exposure can provide support for body weight-based dosing schemes (e.g., mg/kg dosing or
98 categorical dosing based on body weight cutpoints). Such analyses should be combined with a
99 robust understanding of the relationship between drug exposure and drug effect (e.g., by using
100 pharmacodynamic biomarkers or clinical endpoints), target engagement (e.g., receptor
101 occupancy), or drug toxicity to inform and further refine dosing.

102

103 Population PK models can also be used to simulate drug exposures that are expected to occur
104 following doses or dosing regimens that have not been directly investigated in prior clinical
105 studies (see section V.D for a discussion on simulation strategies). For example, a population
106 PK analysis can be used to predict PK changes resulting from the inclusion of a loading dose,
107 changing the dose, or altering the dosing frequency of a dosing regimen for later trials in the drug
108 development program. In rare instances and when appropriately justified, such analyses, in
109 conjunction with exposure-response data, have been used to approve dosing regimens that have
110 not been directly evaluated in the clinical trials (Kimko and Peck 2010). Sponsors are
111 recommended to seek Agency input for such applications.

2. Deriving Sample Size and Sampling Scheme Requirements to Facilitate the Reliable Estimation of Covariate Effects

112

113 Simulations with population PK models can help determine the number of patients in a
114 subpopulation that is needed to achieve sufficient power to detect a significant covariate given a
115 defined covariate effect size (e.g., the number of patients receiving a concomitant medication
116 that need to be included in an analysis to detect a significant drug-drug interaction; see section
117 III.B.2). Simulations and optimal design methods can maximize the utility of such analyses. For
118 example, the study sample size and PK sampling schedule can be optimized so that the PK
119 parameters can be estimated with a defined degree of precision (see section IV.B for a discussion
120 on various sampling schedules).

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3. Deriving Exposure Metrics for Conducting Exposure-Response Analysis

The importance and application of exposure-response (E-R) relationships for new drugs are outlined in the 2003 FDA guidance for industry entitled *Exposure-Response Relationships — Study Design, Data Analysis, and Regulatory Applications*.⁴ Population PK analysis can be used to derive patient PK exposure metrics that can be used to conduct sequential E-R analyses. Derived exposure metrics (e.g., the area under the curve (AUC), the minimum drug concentration (C_{\min})) can be used as a measure of average drug exposures for a patient at steady state. The derivation of exposure metrics should account for: (1) dose interruptions or modifications; and (2) variations in a drug's pharmacokinetics with time, the disease state, or the severity of the disease.

Population PK models can predict individual patient exposures at specific time points regardless of the spread in sampling times (e.g., trough concentrations can be predicted for all subjects). When PK data are missing in a small number of subjects, population PK models can predict the most likely concentration-time profile based on the subject's individual covariates (e.g., body weight, genetic polymorphism, sex). Such predictions are useful, assuming that the residual error and the between-subject variability are low, and the effect of observed covariates on the PK properties of the drug is large (see section V.D).

Individual PK patient exposure metrics are generated based on Empirical Bayes Estimates (EBEs). When individual data are sparse or uninformative, and parameter shrinkage is high (i.e., usually greater than 20 to 30 percent), EBEs are considered less reliable (Savic and Karlsson 2009). In addition to parameter shrinkage, the reliability of individual PK patient exposure metrics is dependent on the nature of the collected PK data and the validity of model assumptions (e.g., time-invariant pharmacokinetics, model structure, dose-proportional pharmacokinetics). See section V.C for discussions on model validation and section IV for discussions regarding the adequacy of the data used for population PK analysis.

4. Pediatric Study Designs

The use of modeling and simulation to inform study designs and to optimize dose selection for pediatric patients has been outlined in the 2014 draft FDA guidance for industry entitled *General Clinical Pharmacology Considerations for Pediatric Studies for Drugs and Biological Products*.⁵ Additional sample size considerations for pediatric studies have been presented in the literature (Wang et al. 2012). Population PK analysis is especially appropriate in children because it allows the use of infrequent (i.e., sparse) sampling compared to the rich sampling associated with traditional PK analyses, thus minimizing the total volume of blood sampled. Sampling windows in pediatric studies are generally expected to be wider than those of adult studies, because of the limited number of blood samples obtained from pediatric patients.

⁴ We update guidances periodically. For the most recent version of a guidance, check the FDA guidance web page at <https://www.fda.gov/RegulatoryInformation/Guidances/default.htm>.

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165
166 Selecting a dosing regimen for pediatric studies can be justified through simulations using a
167 population PK model developed with adult PK data and incorporating: (1) principles of
168 allometry; (2) knowledge of developmental changes that can influence drug pharmacokinetics
169 (ontogeny); and (3) data on the bioavailability of the pediatric formulation (Holford, Heo, and
170 Anderson 2013; Barbour, Fossler, and Barrett 2014; Zhang et al. 2015; Mahmood I 2014).
171 Including the latest understanding in the population PK model on the maturation of physiology in
172 pediatric patients across various ages, especially in those less than 2 years old, may further
173 improve the ability to identify appropriate pediatric dosing. It should be noted that dose
174 selection will also require understanding of disease similarity and E-R relationships in adults and
175 pediatrics (see the 2014 draft FDA guidance for industry entitled *General Clinical*
176 *Pharmacology Considerations for Pediatric Studies for Drugs and Biological Products*⁶).

B. Application of Population PK Analysis to Inform Drug Use

177
178
179
180 Population PK analysis using data from late-stage clinical trials, together with results from stand-
181 alone clinical pharmacology studies, can be used to support the absorption, distribution,
182 metabolism, and excretion (ADME) information in the drug's labeling. In certain cases,
183 traditional PK data analysis using non-compartmental analysis (NCA) methods is not adequate.
184 For example, it can be difficult to design a study for drugs with a long half-life that would allow
185 the area under the concentration-time curve from zero to infinity ($AUC_{0-∞}$) to be estimated with
186 less than 20 percent extrapolation from the area under the concentration-time curve from zero to
187 a definite time point (AUC_{0-t}). Such studies should be analyzed with population PK methods
188 (Svensson et al. 2016).

189
190 In addition, population PK analysis, together with relevant information from stand-alone studies,
191 nested studies, or other sources, is routinely used to evaluate the effects of covariates on the PK
192 parameters of a drug and its relevant metabolites to support dosing recommendations. The need
193 for dose adjustments due to a covariate's effect on the drug's exposure should be interpreted in
194 the context of the known E-R relationships for efficacy and safety. The E-R relationships should
195 be used to establish concentration boundaries between which dose adjustments are not clinically
196 relevant (i.e., covariate effects within these boundaries may not warrant a dose alteration;
197 however, covariate effects outside these boundaries may require a dose modification to optimize
198 the benefit-risk profile of the drug).

1. Specific Populations

199
200
201
202 Results from population PK analyses may be incorporated into drug product labeling to describe
203 the PK properties in general patient populations or specific populations. Labeling in specific
204 populations based on results from population PK analysis typically includes language describing
205 the magnitude of the covariate effect, an assessment of the clinical relevance of the changes, and
206 may include a recommendation on the need, or lack thereof, for dosage adjustments (see section
207 VI for more information). Whether a covariate analysis supports the labeling statements depends
208 on multiple factors, including the number of subjects with the covariate included in the analysis,

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209 the range of the covariate (for continuous covariates), and the extent of available PK data (see
210 section IV.A).

211
212 Some examples of these situations are listed below:

- 213
- 214 • Stand-alone renal or hepatic impairment studies in patients without the medical condition
215 of interest may not be ethical with some drugs (e.g., highly toxic agents). In such cases,
216 adequate representation of patients in clinical trials with the intrinsic factor of interest, as
217 well as adequate PK sampling to reliably characterize the intrinsic factor effects, can
218 allow for the use of population PK analysis to inform labeling for such patients.
219
 - 220 • In situations where specific populations (e.g., patients with varying degrees of renal
221 impairment) can safely be included in late-stage clinical trials, population PK analysis
222 can be used to characterize the drug's exposure and its relationship to response and derive
223 dosing recommendations in that population.
224
 - 225 • Some defined specific populations are traditionally not studied in stand-alone, clinical
226 pharmacology studies because of the lack of an a priori hypothesis of a large effect on a
227 drug's pharmacokinetics. Instead, factors such as the influence of sex, age, body weight,
228 or race on the pharmacokinetics of the investigational drug are often studied without
229 stand-alone trials. Population PK analysis could conceivably be used to describe a drug's
230 pharmacokinetics in these subgroups.
231
 - 232 • Simulations from population PK models established from pediatric and adult data can be
233 used to compare a drug's exposure in pediatric patients and adults to derive
234 recommended pediatric doses for labeling.
235

2. *Drug-Drug Interactions*

236
237
238 Clinical DDIs (e.g., nested studies as part of a phase 3 study) may be evaluated using population
239 PK analysis. General design considerations for nested DDI studies are found in the 2017 FDA
240 draft guidance for industry entitled *Clinical Drug Interaction Studies - Study Design, Data*
241 *Analysis, Dosing Implications, and Labeling Recommendations*.⁷ Using population PK
242 approaches to characterize the DDI potential of a drug is not without limitations, and
243 methodological considerations for using population PK analysis to evaluate DDIs have been
244 described elsewhere (Bonate et al. 2016; Wang et al. 2017). Specific considerations for the use
245 of a population PK approach to evaluate DDIs include the following:

- 246
- 247 • DDIs should be characterized for individual compounds and not for therapeutic classes of
248 drugs, although exceptions are possible (e.g., multiple proton pump inhibitors can be
249 pooled if the interaction mechanism is pH-dependent).
250

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- 251 • Multiple perpetrators can be pooled to form one covariate category provided that the
252 perpetrators belong to the same class of index inhibitors/inducers (i.e., weak, moderate,
253 or strong) as identified by the Agency. The pooled perpetrators should all have the same
254 mechanism of interaction with similar specificity for their respective metabolizing
255 enzymes.
256
- 257 • An adequate number of subjects with the concomitant medication should be included in
258 the study. The PK sampling schedule should appropriately characterize the PK
259 parameters of interest. Simulations can determine the number of subjects needed to
260 detect an interaction of a defined magnitude within a given study design.
261
- 262 • Interactions should be investigated on all physiologically plausible structural elements of
263 the PK model (e.g., clearance (CL/F), relative bioavailability (F_{rel}), rate of absorption).
264
265

IV. DATA USED FOR POPULATION PK ANALYSIS

266 Sponsors are encouraged to use all available data to support the population PK model. The
267 sponsor should justify the omission of data from certain studies, certain individuals, or certain
268 time points and prespecify such omissions in the data analysis plan or study protocol. The
269 sponsor should demonstrate that the data are adequate for the purpose of the analysis in their
270 report. For example, sponsors can conduct model-based power analyses showing that the given
271 study design has sufficient power to detect clinically meaningful covariate relationships under
272 the proposed population PK model (see section III.B.2).
273
274
275

A. Study Subjects and Covariates

276 Determining if the data are adequate to address the intended study question is a vital step in any
277 population PK analysis (see section V.C). The dataset should include a sufficient number of
278 subjects with an adequate number of PK samples at informative time points. Covariates cannot
279 be claimed to be influential or not influential on a drug's exposure if the covariate distribution is
280 narrow (for continuous covariates) or if the number of subjects in the category is inadequate (for
281 categorical covariates). Many continuous covariates have established cutoff values that define
282 category levels. If dosing recommendations are proposed for these categories, the range of the
283 continuous covariates should preferably span the entire category and not just the upper or lower
284 ends. An increase in the range and frequency of the covariates tends to increase the likelihood of
285 finding a true clinically significant covariate relationship and decreases the likelihood of finding
286 false-positive relationships (Wählby et al. 2002; Wählby, Jonsson, and Karlsson 2001).
287
288
289

B. PK Sampling Schedule

290 The precision and bias of model-derived PK parameters are dependent on multiple factors,
291 including the number of subjects, the number of samples per subject, and the sampling schedule.
292 As the number of samples per subject decreases, the importance of the timing of PK samples
293 increases. For example, if the purpose of an analysis is to match C_{max} observations across
294 populations or dosage forms, then a sufficient number of PK samples should be collected in the
295
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297 absorption phase. Sponsors are encouraged to prospectively plan the PK sampling schedule so
298 that the population PK model is maximally informative.

299
300 The methodology and available software for the optimal design of the population PK model are
301 covered in multiple reviews in the literature (Dodds, Hooker, and Vicini 2005; Nyberg et al.
302 2015; Ogungbenro and Aarons 2007; Ogungbenro and Aarons 2008). The list below shows
303 some examples of sampling schedules. Depending on the purpose of the analysis, one or a
304 combination of the listed strategies may be adequate:

- 305
306 • Patients are randomly assigned sampling windows that are derived based on optimal
307 design methods. The number of samples per patient and the number of sampling
308 windows are also determined based on optimal design methods.
- 309
310 • Patients randomly contribute two or more samples that, when combined, cover the entire
311 dosing interval.
- 312
313 • Most patients contribute one sample at a specified time point, often immediately before
314 the next dose.

315
316 Sponsors are encouraged to collect PK data from all patients. However, the extent of sampling
317 per individual patient and the percentage of sampled patients should ultimately depend on the
318 intended use of the data. For example, if the C_{\max} will be used in subsequent E-R analyses, then
319 there should be adequate sampling around the time to the maximum concentration (T_{\max}) (see
320 section III.A.3). In any situation, it is important to verify that patients with missing PK data do
321 not differ from other patients. For example, patients with missing PK data should not have
322 higher dropout rates because of a lack of effect or adverse events. If between-occasion
323 variability will be estimated, multiple samples per individual at more than one occasion are
324 needed. Ignoring large between-occasion variability can lead to biased population parameter
325 estimates (Karlsson and Sheiner 1993).

326

327

V. DATA ANALYSIS

328

329
330 This section of the document provides some guiding principles on methodological aspects of
331 population PK analysis. Further methodological considerations and good practices, including
332 topics not covered here, have been described by others (Ette and Williams 2007; Bonate and
333 Steimer 2013; Mould and Upton 2013; Byon et al. 2013; Tatarinova et al. 2013; Lunn et al.
334 2002; Schmidt and Radivojevic 2014).

335

A. Preliminary Examination of the Data

336

337
338 All population analyses should begin with an examination of the observed data. Preliminary
339 examination of the data isolates and reveals patterns and features in the population dataset using
340 graphical and statistical techniques and can provide powerful diagnostic tools for confirming
341 assumptions or, when the assumptions are not met, for suggesting corrective actions (Tukey
342 1977; Ette and Ludden 1995). For example, correlations between highly correlated covariates

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343 may not provide unique information about the population. This scenario is often the case with,
344 for example, body weight and creatinine clearance as calculated by the Cockcroft-Gault
345 Equation. Relevant preliminary examination of the data should be concisely described in the
346 population PK analysis report.

347

B. Model Development

348

349 Model development methods and best practice recommendations are constantly evolving.

350 Specific advice on how to develop population PK models is beyond the scope of this guidance.

351 However, to facilitate the regulatory review of population PK models, sponsors should explicitly

352 describe their model development procedures (see section VII for more discussion on population

353 PK reporting). Some aspects of model development that are important for regulatory review are

354 provided below:

355

356

- 357 • Model development issues can be addressed through several valid approaches, each with
358 its own benefits and drawbacks. For example, covariate analysis can be performed based
359 on several approaches or their possible combinations (e.g., stepwise covariate analysis,
360 full covariate model approach, the Lasso) (Wählby, Jonsson, and Karlsson 2002;
361 Gastonguay 2004; Ribbing et al. 2007). In such cases, sponsors should justify why a
362 particular approach was used.

363

- 364 • Covariate-parameter relationships can be established based on current knowledge of
365 biology, physiology, or allometric principles.

366

- 367 • Issues regarding missing data, including missing covariates and data below the limit of
368 quantification (LOQ), should be addressed with appropriate analysis methods (Beal 2001;
369 Bergstrand and Karlsson 2009; Johansson and Karlsson 2013; Keizer et al. 2015). The
370 sponsor should justify their methodological approach with regard to missing data and
371 outliers and provide a sensitivity analysis.

372

- 373 • The sponsor should distinguish between outlying individuals and outlier data points.
374 Individual data points that are suspected outliers could be omitted during the model
375 development process. However, the sponsor should investigate the influence of the
376 outliers on the final parameter estimates by refitting the final model to the complete
377 dataset. Removal of suspected outlying individuals is generally discouraged unless the
378 reason for the outlier is a protocol violation or other human error. Sponsors should
379 specify how outliers are identified and handled in the analysis. The reasons for declaring
380 a data point to be an outlier should be prespecified in the data analysis plan. For
381 example, data points with a weighted residual greater than five could be considered
382 outliers in some cases. The number of excluded outliers should be kept at a minimum
383 and clearly documented in reports and datasets.

384

C. Model Validation

385

386 Model validation is a critical step for any population PK analysis and should be conducted to
387 examine whether the developed model can sufficiently characterize the observed data and
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389 generate reliable modeling and simulation results to meet the needs of the analysis. No single
390 model validation method is generally sufficient to evaluate all components of a model. Several
391 methods are generally needed so that the relative strengths and weaknesses of each method can
392 complement each other. In general, models need to describe the data with an acceptable level of
393 bias and an acceptable degree of precision. An appropriate model should be biologically
394 plausible, consistent with current knowledge, and have mathematically identifiable parameters.
395

396 Model validation depends on the objective of the analysis and should follow a *fit-for-purpose*
397 approach. In some cases, a model may be valid for one purpose but not for another. For
398 example, a model with high shrinkage on CL/F may not be useful to derive individual drug
399 exposure levels to be used in a sequential E-R analysis (see section III.A.3). However, such a
400 model can still be useful for a covariate analysis provided that the method for covariate modeling
401 is insensitive to shrinkage.
402

403 Submissions to the Agency should contain both a detailed description of the model validation
404 methods used and an explanation of why those methods were selected (see section VII.A).
405

1. Common Approaches for Model Validation

406
407
408 Procedures for conducting a thorough model validation are continuously evolving, and the
409 Agency welcomes innovations in this field. Several common methods of model validation are
410 reported in literature (Karlsson and Savic 2007; Byon et al. 2013). Some of these methods are
411 discussed in the sections below.
412

413 Basic goodness-of-fit (GOF) plots illustrate how well the model describes the observed data.
414 GOF plots are also used to evaluate model assumptions (e.g. normality of the random effects)
415 and to guide model development. Although GOF plots can show that the overall fitting is
416 acceptable, additional evaluation of the model in subgroups of patients is often necessary. For
417 example, if the model will be used to predict drug exposures in pediatric patients, model
418 validation should be conducted for all age groups. GOF plots that are stratified for important
419 patient characteristics (e.g., stratified by age group or CYP polymorphisms), study design (e.g.,
420 dose or formulation), or other important variables are often more convincing of the adequate
421 performance of a model than GOF plots of the full dataset.
422

423 The following is a list of some of the GOF plots that are considered informative:
424

- 425 • The dependent variable (DV) versus the individual predictions (IPRED)
- 426
- 427 • The DV versus population predictions (PRED)
- 428
- 429 • The absolute individual weighted residuals (|IWRES|) versus IPRED or time⁸
- 430

⁸ Time should be evaluated as continuous time and as time after dose.

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- 431 • The conditional weighted residuals (CWRES) versus PRED or time⁷
- 432
- 433 • A representative sample of IPRED, PRED and observations versus time (one plot per
- 434 subject)⁷
- 435
- 436 • A histogram or Quantile-Quantile (Q-Q) plot of random effects
- 437
- 438 • The correlations between random effects
- 439
- 440 • The random effects versus covariates⁹
- 441

442 Individual parameters, IPRED and IWRES, tend to shrink towards the population estimate in
443 individuals with few observations. When shrinkage is high (usually greater than 20-30 percent),
444 diagnostic plots that rely on EBEs, IPRED, or IWRES can become uninformative, and
445 correlations between random effects and covariates can be obscured (Savic and Karlsson 2009).
446 Furthermore, high shrinkage may limit the value of using individual post hoc estimates for E-R
447 analysis (see section III.A.3). Simulation-based, diagnostic plots are not affected by shrinkage in
448 a similar manner and can be more informative for diagnostic purposes when shrinkage is high.
449 There are several available simulation-based diagnostics, including, but not limited to, Visual
450 Predictive Check (VPC), the prediction corrected VPC (pcVPC), and the Numerical Predictive
451 Check (NPC) (Bergstrand et al. 2011).

452
453 GOF criteria can also be reflected by some numerical metrics, such as the estimate of a
454 parameter's precision. Estimates of parameter precision can provide valuable information
455 regarding the adequacy of the data to support that parameter. Parameter uncertainty can be
456 estimated through several methods, including bootstrap procedures, log-likelihood profiling, or
457 using the asymptotic standard errors of parameter estimates. In addition to parameter
458 uncertainty, it is important to compare parameter point estimates with previous analyses and to
459 evaluate the physiological plausibility of the point estimates. Another useful numerical metric to
460 evaluate models is the condition value. A condition number (the ratio of the largest and the
461 smallest eigenvalue) over 1000 indicates that the observed data cannot support the estimation of
462 one or several parameters (Montgomery, Vining, and Peck 2012). Over-parameterized models
463 can be simplified and updated as additional data becomes available.

464
465 Performance of the model can be checked against a set of test data, either internal or external.
466 The validation approach mentioned above relies on the data used for model building, hence is
467 considered as internal validation. External validation, on the other hand, relies on the data not
468 used for model building. Data collected in a drug development program can be split into a model
469 building dataset and a testing dataset. The testing dataset is commonly used for external
470 validation. Data splitting is a powerful method for model evaluation. However, before deciding
471 on a data splitting approach, sponsors should consider the potential impact of the loss of data on
472 the model's power to detect covariate relationships and estimate parameters with an acceptable
473 degree of precision.

⁹ This plot is most informative when it is compared before and after the inclusion of covariates.

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474
475 There is an additional level of uncertainty when the purpose of the model is to simulate PK
476 profiles for scenarios that are beyond those that have been clinically studied. Such uncertainty
477 can be addressed by a sensitivity analysis of the parameter estimates and their impact on the
478 metric used to inform a decision (Kimko and Peck 2010).

D. Simulations Based on Population PK Models

481
482 Simulations should be based on protocols that outline the simulation study to be performed. The
483 level of detail in the protocol should correspond to the complexity and impact of the question the
484 simulation addresses. Models used for simulations should be validated to address the specific
485 question the simulation study is trying to answer. Although a model can have been previously
486 used and validated, it needs to be revalidated if the new purpose is different from the original
487 purpose. Depending on the purpose of the model, various levels of uncertainty and variability
488 can be added to the simulations. Some examples are discussed below.

1. Simulations Based on Fixed-Effect Estimates

491
492 In their simplest form, simulations are used to illustrate the drug-concentration profile for a
493 typical individual. Such simulations are based on the typical estimates of the fixed-effect
494 parameters. Note that predictions based on typical parameter estimates do not correspond to
495 mean predictions. Mean predictions are obtained by calculating the mean concentration-time
496 profiles based on simulations with interindividual variability in population PK parameters and
497 residual error.

2. Simulations Based on Uncertainty of Fixed-Effect Estimates

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500
501 Uncertainty in parameter estimates can be accounted for if the desire is to illustrate the
502 probability of the typical subject's drug exposure to reach or stay above a specific cutoff point or
503 if one wishes to illustrate the effect of covariates. For example, forest plots that illustrate the
504 effect of covariates on AUC or other parameters can be generated based on simulations with
505 uncertainty in fixed-effect parameters, thereby facilitating the interpretation of the relative
506 importance of covariates on exposure. These types of simulations are also useful to evaluate the
507 performance of new dosing regimens for testing in future trials. Additionally, simulations with
508 parameter uncertainty can be used to graphically illustrate the effect of parameter precision on
509 PK profiles.

3. Simulations Based on Estimates of Between-Subject Variability

511
512
513 Between-subject variability (BSV) in PK parameters is accounted for in simulations when the
514 purpose is to show the range of individual predictions of concentrations in the studied
515 population. Residual error is added to the individual prediction when the range of observed
516 concentrations is the main interest.

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518 If the purpose is to predict the observed concentration range of a future population, then
519 uncertainty in the fixed-effect parameters should be accounted for in addition to the residual
520 error and the BSV.

521
522 The correlation between random effects should be accounted for to avoid unrealistic parameter
523 combinations in subjects. Simulations that account for BSV and include covariate effects should
524 be conducted in a population with realistic demographic variables. Demographic variables can
525 be obtained from databases, generated by resampling with replacement of the individuals in the
526 original study or by sampling the covariate distributions and their correlations in the target
527 population.

528

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VI. LABELING BASED ON THE RESULTS OF POPULATION PK ANALYSIS

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532 Results from population PK analysis may be presented in the CLINICAL PHARMACOLGY
533 section and summarized in other sections of labeling, as appropriate. Other relevant labeling
534 sections should not repeat this detailed information but rather provide a succinct description or
535 recommendation based upon these results followed by a cross-reference to the CLINICAL
536 PHARMACOLGY section, as appropriate. In general, there is no need to explicitly state that the
537 information is based on population PK analysis. Recommendations for developing the
538 CLINICAL PHARMACOLOGY section are discussed in the FDA guidance for industry entitled
539 *Clinical Pharmacology Section of Labeling for Human Prescription Drug and Biological*
540 *Products — Content and Format.*

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VII. POPULATION PK STUDY REPORTING

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545 This section outlines the recommended format and content for a sponsor to submit a population
546 PK analysis in support of its drug's clinical pharmacology program. Population PK analyses can
547 be appropriate at multiple points in the drug development process, for example in the
548 investigational new drug application (IND), NDA, or postmarketing stages. The depth and
549 breadth of population PK analyses at each stage can vary because of the availability or quality of
550 clinical data.

551

552 Population PK study reports that are important for regulatory decisions should be included in the
553 electronic common technical document (eCTD) module 5.3.3.5 and the corresponding programs,
554 datasets and define files under the *datasets* folder within eCTD module 5.

555

A. Format and Content of the Population PK Report

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558 To enable the efficient and consistent review of population PK analyses, the FDA recommends
559 that the results from population PK analyses should be accompanied with a structured population
560 PK report. The report should contain the following sections: (1) an executive summary, (2) a
561 synopsis, (3) an introduction, (4) data, (5) the methods, (6) the results, (7) a discussion, (8)
562 conclusions, and (9) an appendix (if applicable). Expected information/elements to be included
563 in these sections is discussed briefly in Table 1. Sections of the annotated labeling that contain

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564 information based on population PK analysis should include hyperlinks to the population PK
565 report. Additional discussion regarding reporting is available in the scientific literature (Dykstra
566 et al. 2015).

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610 **Table 1: Expected Content in Each Section of the Population PK Study Report**

Sections	Expected Content
Executive Summary	<ul style="list-style-type: none"> • The purpose of the analysis • The key findings that affect approval or labeling decisions • Any other recommendations based on the population PK analysis
Synopsis	<ul style="list-style-type: none"> • A plain language summary of objectives, data, methodology, and conclusions • A brief explanation on the sufficiency of the data to evaluate different drug exposures in relevant subpopulations • Results presented as their effect on clinically relevant drug exposures, not PK variables • Visual presentations of relevant exposure metrics to illustrate how identified subpopulations differ from the typical population
Introduction	<ul style="list-style-type: none"> • The background to place the population PK study in the context of the overall clinical development program • The objectives for the population PK analysis • The PK characteristics of the drug and their relevance to the final model
Data	<ul style="list-style-type: none"> • A description of the studies and the study data included in the population PK analysis with information on the dose, including the frequency and duration of dosing, as well as the number of subjects, number of samples, and number of LOQ samples • A distinction between all available data versus the final data used for model building and evaluation • Reasons for not including studies with available and potentially informative data, if applicable • References to the original study reports of data used for the analysis and the bioanalytical evaluation reports for each study • The LOQ should be reported for each study and bioanalytical method (if different) • Detailed demographic information • Information about the investigated covariates in tables, histograms, or matrix plots to illustrate correlations

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612 **Table 1 continued: Expected Content in Each Section of the Population PK Study Report**

Sections	Expected Content
Methods	<ul style="list-style-type: none"> • Criteria and procedures used for model building and evaluation as well as a description of the handling of outliers and missing data • Equations for all tested covariate relationships • The choice and justification of the model-estimation method • Equations for parameter transformations, if parameters are presented in dimensions other than those in the model output file • The method for incorporating variability or parameter uncertainty or for deriving confidence intervals or prediction intervals • Detailed descriptions of simulations used to support conclusions and recommendations • Sufficient information to reproduce the analysis and to conduct supplemental analysis, when necessary • Software and electronic files used for the analysis and simulation* • Deviations from standard procedures, if applicable <p>* Note: The FDA does not recommend or require the use of particular software for conducting population PK analysis. The sponsor should report the software used for the analysis and submit the electronic files supporting the analysis and simulations (see section VII.B below). If necessary, consult the FDA regarding the feasibility of submitting certain types of electronic files.</p>
Results	<ul style="list-style-type: none"> • A description of the final model, model building steps, qualification assessments, and final parameter estimates • Application of the model to objectives, including the relevance of covariates on PK parameters and simulations of alternative dosing, if applicable • Accompanying tables and figures to place findings into context • Typically, tables should include: <ul style="list-style-type: none"> ○ Key modeling building steps, including a description of the structural and covariate models and objective function changes ○ A comparison of parameter estimates from the base to the final model ○ Parameter estimates and their associated uncertainty, with variability reported as the CV% and precision reported as the percent relative standard error (RSE%) or the 95 percent confidence interval

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613 **Table 1 continued: Expected Content in Each Section of the Population PK Study Report**

Results <i>continued</i>	<ul style="list-style-type: none">• Typically, figures should include:<ul style="list-style-type: none">○ Diagnostic plots○ GOF plots stratified by the relevant covariates to illustrate model performance in specific subgroups
Discussion	<ul style="list-style-type: none">• An interpretation of the modeling results, including discussions on:<ul style="list-style-type: none">○ The adequacy or limitation of the data to support modeling conclusions○ The rationale for the modeling approach, assumption verification, and assessment of uncertainty○ Consistency or inconsistency between the results from population PK and stand-alone clinical pharmacology studies• An assessment of the clinical relevance of the results, including physiological plausibility and clinical significance of the identified relationships• An assessment of any alternative dosing regimens in the context of exposure-response analyses for safety and efficacy• Discussion of the drug development and regulatory decisions based on the results of the model
Conclusions	<ul style="list-style-type: none">• A short summary of major findings from the analysis written in plain language
Appendix	<ul style="list-style-type: none">• A run record describing the steps undertaken for the analysis• Methods and codes for generating the key figures (i.e., for figures other than standard diagnostic plots)

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B. Submitting Electronic Files to the Agency

Sponsors should refer to the FDA Web site¹⁰ for general advice on submitting data and related files (e.g., coding scripts). It is critical that all datasets and model files submitted for the base, final, and key intermediate models are the same as those used for generating the model outputs in the appendices of the report. In addition, FDA staff should be able to identify the source data for any dataset constructed using output files or postprocessed results from population PK analyses. For example, if exposure metrics from population PK analyses are included in datasets for E-R analyses, the sponsor should ensure the traceability between the population PK model output and

¹⁰ The FDA Web Site on submitting data can be accessed at <http://www.fda.gov/AboutFDA/CentersOffices/OfficeofMedicalProductsandTobacco/CDER/ucm180482.htm>.

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624 any postprocessing steps involved in the construction of the dataset. This can be accomplished
625 by providing definition files, reviewer guides, or codes utilized for dataset assembly.

626
627 All conclusions made from the population PK analysis should be reproducible by the Agency
628 with the available codes and data. It is also important to include the unique subject identifier
629 information for each subject in the population PK dataset and ensure that the identifiers are the
630 same in the individual clinical study report datasets. This information is vital if data integration
631 is required between the individual level outputs (e.g. individual, post hoc estimates for CL or V_d)
632 generated from the population PK model and the efficacy or safety datasets from the individual
633 clinical study reports.

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634	VIII. GLOSSARY OF SELECTED TERMS	
635		
	BSV	Between-subject variability, a measure of PK variability between subjects
	Covariate	An observed factor that correlates with drug exposure in patients (e.g., renal function, body weight, age, sex, genetic polymorphism)
	CWRES	Conditional Weighted Residuals, a type of diagnostic
	DV	Dependent variable (e.g., drug plasma concentrations)
	EBE	Empirical Bayes Estimates, or individual parameter estimates in a mixed-effects model
	GOF	Goodness of fit, a collection of diagnostic criteria used to evaluate model performance
	IPRED	Individual Predicted Data, based on Individual Empirical Bayes parameter estimates
	IWRES	Individual Weighted Residuals, a type of residual
	Model evaluation	The use of various methods to evaluate model performance
	Model qualification	The use of various methods to evaluate model performance for a specific purpose
	NPC	Numerical Predictive Check, a GOF method related to VPC
	PRED	Predicted data, based on population parameter estimates
	prVPC	Prediction corrected VPC, a GOF plot related to VPC
	QQ	Quantile-quantile, a type of GOF plot
	Residual	The difference between the predicted and the observed value
	Residual error	An estimate of the remaining unexplained variability
	Shrinkage	A measure of the extent to which EBE depends on the population parameters versus the individual observed data
	VPC	Visual predictive check, a type of GOF plot

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¹¹ When final, this guidance will represent the FDA's current thinking on this topic