



Biomarker Qualification Letter of Intent (LOI) Template

Administrative Information

Requesting Organization

Name: _____

Address: _____

Phone: _____

Website: _____

Primary Contact

Name: _____

Address: _____

Phone: _____

Email: _____

Alternate Contact

Name: _____

Address: _____

Phone: _____

Email: _____

Submission Date (MM/DD/YYYY):

If there is a prior, current, or planned submission to other regulatory agencies, list the agencies and dates as appropriate.

Context of Use

Proposed Context of Use (COU) (limited to 500 characters)

Drug Development Need

Describe the drug development need that the biomarker is intended to address, including (if applicable) the proposed benefit over currently used biomarkers for similar COUs (limited to 1,500 characters).

Biomarker Information

Biomarker name and description. If composite, please list the biomarker components.

Type of Biomarker

- Molecular
- Histologic
- Other (please describe)
- Radiologic
- Physiologic characteristic

Biomarker Information

For molecular biomarkers, please provide a unique ID.

Scheme:

ID:

Matrix (e.g., blood) or modality (e.g., MRI):

Primary biomarker category  (see [BEST Glossary](#)):

Describe the mechanistic rationale or biologic plausibility to support the biomarker and its associated COU (limited to 1,500 characters).



If biomarker is an index/scoring system, please provide information on how the index is derived (e.g., algorithm), the biologic rationale for inclusion of each of the components, the rationale for any differential weighting of the elements, and the meaning/interpretation of the index/score (limited to 1,500 characters).

Biomarker Measurement Information

Provide a general description of what aspect of the biomarker is being measured and by what methodology (e.g., radiologic findings such as lesion number, specific measure of organ size, serum level of an analyte, change in the biomarker level relative to a reference such as baseline) (limited to 1,500 characters).

Is the biomarker test/assay currently available for public use? Yes No

Indicate whether the biomarker test/assay is one or more of the following:

- Laboratory Developed Test (LDT)
- Research Use Only (RUO)
- FDA Cleared/Approved. Provide 510(k)/PMA Number:

If the biomarker is qualified, will the test/assay be performed in a [Clinical Laboratory Improvement Amendments \(CLIA\)](#)–certified laboratory? Yes No

Is the biomarker test currently under review by the [Center for Devices and Radiological Health](#) or the [Center for Biologics Evaluation and Research](#)? Yes
 No
 Don't Know

Is there a standard operating procedure (SOP) for sample collection and storage? Yes No

Is there a laboratory SOP for the test/assay methodology? Yes No

Biomarker Measurement Information

Describe the extent of analytical validation that has been performed (e.g., sensitivity, specificity, accuracy, and/or precision of the assay or method) (limited to 1,500 characters).

Additional Considerations for Radiographic Biomarkers

How has the method for image acquisition, analysis, and integration of the data been optimized? (Limited to 1,000 characters.)

Does data currently exist to support the proposed cut point(s), if imaging results are not reported as a continuous variable? Yes No

Provide the name and version of the software package to be used for image acquisition and analysis (limited to 500 characters).

Supporting Information

Please summarize existing preclinical or clinical data to support the biomarker in its COU (e.g., summaries of literature findings, previously conducted studies) (limited to 2,000 characters).

Please summarize any planned studies to support the biomarker and COU. How will these studies address any current knowledge gaps? (Limited to 2,000 characters.)

Previous Regulatory Interactions

- None
- Letter of Support (LOS) issued for this biomarker on date: _____
- Discussed in a Critical Path Innovation Meeting (CPIM) on date: _____
- Previous FDA Qualification given to this biomarker with DDT Tracking Record Number: _____

Attachments

Please provide a list of publications relevant to this biomarker development proposal.

Optional* – If this biomarker development effort is part of a longer-term goal, please summarize your long-term objectives.*

Optional* – If you have other supporting information you would like to provide, please submit as attachment(s).

*Optional information will not be posted publicly.

Please refer to the [Biomarker Qualification Contacts and Submitting Procedures](#) for the mailing address and other important submission-related instructions. If you have any questions about submission procedures, please contact CDERBiomarkerQualificationProgram@fda.hhs.gov.

ATTACHMENT A – Biomarker Measurement Information

Table 1: Urine Biomarker Assay Method and Manufacturer used by PBI

Assay	OPN	KIM-1	CysC	Clusterin	NGAL	NAG	Total Protein	Albumin	Cr
Method	ELISA	ELISA	ELISA	ELISA	ELISA	Colorimetric	Turbidimetric	Immuno-turbidimetric	Enzymatic
Manufacturer	R&D Systems	R&D Systems	R&D Systems	R&D Systems	BioPorto Diagnostics	Roche Diagnostics	Roche Diagnostics	Roche Diagnostics	Roche Diagnostics

Table 2: Proposed Statistically and Medically Significant Thresholds for Use in the Prospective Studies

Biomarker	ULN	ULN FC from BL	Statistically Significant Thresholds			Medically Significant Thresholds		
			T _{SS}	Specificity (%)	Sensitivity (%)	T _{MS}	Specificity (%)	Sensitivity (%)
Clusterin	301 ng/mg uCr	2.3	FC > 2.6	94.9	95.0	FC > 6.0	77.1	60.0
OPN	2.10 µg/mg uCr	2.4	FC > 3.1	94.9	100.0	FC > 5.3	44.4	70.0
Albumin	30 µg/mg uCr	NA	> 1.0x ULN	97.5	100.0	> 10.0x ULN	97.1	40.0
Total Protein	0.20 mg/mg uCr	NA	> 1.0x ULN	96.2	100.0	>3.1x ULN	77.1	70.0
NAG	3.54 mU/mg uCr	2.1	FC > 3.9	100.0	100.0	FC > 8.7	65.7	55.0
KIM-1	1.19 ng/mg uCr	2.2	FC > 3.1	99.2	100.0	FC > 7.6	48.6	60.0
CysC	0.052 µg/mg uCr	2.2	FC > 2.2	96.6	95.0	FC > 4.5	71.4	70.0
NGAL	87.6 ng/mg uCr	3.2	FC > 3.5	95.9	95.0	FC > 9.6	59.5	60.0

ULNs = median of estimated ULNs based on Horn and Pesce method using PSTC NHV Study of Healthy Volunteers data; T_{SS} = Statistically significant threshold; T_{MS} = Medically significant threshold; Specificity/Sensitivity of Statistically Significant Thresholds based on Controls = NHV and Cases = mesothelioma patients with medically relevant increases in sCr; Specificity/Sensitivity of Medically Significant Thresholds based on Controls = mesothelioma patients without medically relevant increases in sCr and Cases = mesothelioma patients with medically relevant increases in sCr

ATTACHMENT B – Assay Validation Performance Characteristics

Table 1: Urine Biomarker Assay Method and Manufacturer

Assay	CLU	CysC	KIM-1	NGAL	NAG	OPN	Cr
Method	ELISA	ELISA	ELISA	ELISA	Colori-metric	ELISA	Enzymatic
Manu- facturer	R&D Systems	R&D Systems	R&D Systems	BioPorto Diagnostics	Roche Diagnostics	R&D Systems	Roche Diagnostics

Accuracy: The determination of accuracy was made using recovery experiments in which manufacturer supplied standards that consisted of recombinant human proteins or natural animal proteins were added (“spiked”) to individual donor human urine samples in various concentrations to test the range of the standard curve. As the proteins being measured were all endogenous molecules, urine that had low concentrations of the analyte were used. These “spiked” samples were assayed and the measured values were compared to the calculated values. In all assays, the recovery was within the established criteria for recovery of 80-120%.

Precision: Precision was determined using standard CAP compliant methods. In general, within-run precision was evaluated using low, medium and high in-house control urine samples assayed 16 to 20 times in one analytical run. Between-run precision was evaluated using the same control samples used for within-run precision in three separate analytical runs.

Dilutional Linearity: Linearity was determined using standard CAP compliant methods. Verification of manufacturer’s linearity range was performed using assay specific calibrator diluent. Mean recoveries for each sample were compared to target values using linear regression analysis.

Dynamic Range: PBI used the upper limit of linearity and the lower limit of sensitivity (usually lower limit of quantification [LLOQ]) for determining the dynamic range of each assay. When the sensitivity of the assay was below the lowest standard in an ELISA, for example, the lowest standard for the low end of the dynamic range was used. Limit of Detection (LOD) is generally a method for determining assay sensitivity that is based on the ability of an assay to distinguish the presence of analyte from instrument noise (no analyte present). Thus, LOD is lower than the LLOQ, which is the lowest level of analyte with a coefficient of variation (CV) of 20% or less. PBI used the more restrictive standard for sensitivity and this measure exceeds the LOD standard.

The upper limit of quantification (ULOQ) is defined as the highest level of analyte with a CV of 20% or less. PBI used the highest reference standard that met this criteria as the ULOQ. That number was then multiplied by the maximum dilution where assay linearity was still maintained to obtain the upper reportable limit (URL).

Interference: PBI conducted a series of studies to evaluate the potential interference of high concentrations of albumin, hemoglobin, and blood contamination on the measurement of urinary CLU, CysC, KIM-1, NAG, NGAL, OPN and uCr. Albumin (5 mg/mL), hemoglobin (~30 mg/dL), and blood (0.2%) did not interfere with the determination of urinary biomarker concentrations.

Albumin concentration greater than 3.0 mg/mL did interfere with the determination of urinary KIM-1. Thus, samples with greater than acceptable albumin will be diluted if KIM-1 levels are not already observed to be markedly elevated. In the original validation, linearity for KIM-1 was acceptable up to a 32-fold dilution. Furthermore, if a sample is observed to have moderate hemolysis, prior to the analysis for total protein, NGAL, NAG and KIM-1, these samples will be diluted until the measured hemoglobin concentration is 30 mg/dL or less. In the original validation, linearity for total protein, NGAL, NAG, and KIM-1 was acceptable up to a 54-fold, 64-fold, 3-fold, and 32-fold dilution, respectively.

Recovery: Recovery data were generated by PBI using either recombinant proteins or assay standards as outlined in the validation documents. Testing samples were prepared by the Merck Clinical Development Laboratory for all the biomarkers by adding known quantities of all proteins, except OPN, to urine. These samples were submitted to PBI for analysis to evaluate claimed recovery, precision and stability. The observed results were within or better than assay specifications stated in the vendor's validation documents.

Hook effect: To eliminate the possibility of a hook effect with values reported from the albumin assay, total protein concentration was evaluated prior to the analysis for albumin and samples with high values were appropriately diluted prior to performing the albumin assay. Hook effects may be dismissed by demonstrating linear dilution behavior throughout the expected maximum biological range of the analyte. In addition, hook effects are generally only of concern in homogeneous immunoassays. In a two-step ELISA (heterogeneous immunoassay), the hook effect is generally not of concern due to inclusion of wash steps to eliminate unbound excess analyte. This permits signal complexes to form at the maximum range of the assay yielding an over range condition (>ULOQ) rather than a falsely depressed value resulting from a hook effect ([Ermens 2000](#)).

Reagent Stability: PBI relied on individual manufacturer's claims of reagent stability.

Analyte Stability: Aliquots of urine were frozen at -70°C and used to examine stability (both 2-8°C storage and freeze/thaw cycle).

Bridging Studies: Due to a slight difference in the urine matrix used in the original assay validation and those collected in the studies to support qualification, PBI conducted a series of bridging studies to evaluate potential differences in the methods for measuring urinary CLU, CysC, KIM-1, NAG, NGAL, OPN and uCr. From these studies it was determined that the difference in urine matrices did not significantly affect the quantitation of the biomarkers and it was concluded that including a centrifugation step in urine sample processing is preferred to reduce the potential contribution of biomarker activity from cellular contamination.

ATTACHMENT C – Assay Performance Summary

Table 1: PSTC Kidney Safety Project Biomarker Qualification Assay Performance Summary

Biomarker	Albumin	Clusterin	Creatinine _e	Creatinine	Cystatin-C	KIM-1	NAG	NGAL	Osteopontin	Protein (Total)
Platform	Roche Modular P	R&D ELISA	Modified Jaffé	Roche Modular P	R&D ELISA	R&D ELISA	Roche Modular P	BioPorto ELISA	R&D ELISA	Roche Modular P
Detection	Turbidometric	Colorimetric	Colorimetric	Colorimetric	Colorimetric	Colorimetric	Enzymatic colorimetric	Colorimetric	Colorimetric	Turbidometric
Precision WIR – L	2.6 %	10.5 %	0.7 %	1.0 %	3.9 %	8.3 %	4.3 %	5.4 %*	3.2 %	6.9 %
Precision WIR – M	1.4 %	6.3 %	1.0 %	2.4 %	3.0 %	1.1 %	2.5 %	8.8 %*	3.0 %	0.9 %
Precision WIR – H	1.1 %	5.8 %	0.8 %	0.7 %	4.3 %	5.3 %	1.9 %	8.7 %*	3.9 %	1.2 %
Precision BTR – L	4.6 %	14.5 %	2.2 %	0.9 %	1.3 %	11.3 %	5.6 %	-	12.5 %	8.5 %
Precision BTR – M	2.2 %	4.3 %	2.7 %	2.9 %	6.7 %	14.6 %	4.5 %	-	7.7 %	1.6 %
Precision BTR – H	1.5 %	4.7 %	2.2 %	1.5 %	5.8 %	15.8 %	2.5 %	-	10.0 %	1.4 %
Precision BTR – Mean (CV)	<5%	< 15%	-	< 3%	< 12%	< 16%	< 6%	< 7%	< 13%	<9%
Units	mg/L	ng/mL	mg/dL	mg/dL	ng/mL	pg/mL	U/L	ng/mL	ng/mL	mg/dL
Precision WIR – L Sample Value	5.64	16	-	49.2	14.2	169	0.86	4.2	807	6.9
Precision WIR – M Sample Value	31.53	105	-	97.4	27.2	579	2.41	20	1775	53.2
Precision WIR – H Sample Value	105.09	238	-	146.1	79.4	1161	9.9	44.5	4524	181.3
LLOQ	3.0	10 (Incl. PAD)	0.8	3.6	1.31	11.6	0.31	0.004 (Incl. PAD)	44 (Incl. PAD)	3.5
ULOQ	400	800	600	900	100	2000	55.25	100	8,800	200
Upper reportable limit	4,400	3,200	19,200	16,864	6,400	64,000	2,210 U/L	6,400	281,600	10,800
Recovery range	ND	90-107.5%	-	103.5-107.9%	83.8-104.2%	96.6-118%	99.1-104.5%	93.3-109.4%	97.9-101.5%	104.1-118.8%
Reference interval (normalized to uCr)	ND	35-383 ng/mg	-	40.0-278 mg/dL (M); 29.0-226 mg/dL (F)	0.014-0.058 µg/mg	<1.191 ng/mg	<0.78 U/mmol	<41.8 ng/mg	495-2029 ng/mg	1.3 – 10.1 mg/mg (x100)
Dilutional range	≤11-fold	≤4-fold (Pre-Diluted)	≤32-fold	≤32-fold	≤64-fold	≤32-fold	≤40-fold	≤64-fold (Pre-Diluted)	≤32-fold (Pre-Diluted)	≤54-fold
Dilutional linearity	±13.8%	±20%	±8.3	±4.9 %	±19.6%	±18.0%	±12.1%	ND	±8.3%	±20.3%
Procedural Dilution_a	-	4	-	-	-	-	-	100	440	-
Interferences – no effect	-	-	Icterus <70 mg/dL Hb <966 mg/dL ascorbic acid <300 mg/L glucose <2000 mg/dL	Bilirubin <50 mg/dL Hb <1100 mg/dL Glucose <2100 mg/dL urobilinogen <40 mg/dL	-	-	-	-	Calcium <200 µg/mL	Icterus <36 mg/dL

Biomarker	Albumin	Clusterin		Creatinine _e	Creatinine	Cystatin-C		KIM-1		NAG	NGAL		Osteopontin	Protein (Total)	
				urobilinogen <40 mg/dL											
Interferences – False Negative	-	-	-	calcium dobesilate (Dexium) α-methyl dopa	-	-	-	-	-	-	-	-	-	Many	
Interferences – False Positive	-	-	-	-	cephalosporin	-	-	-	-	-	-	-	-	Many	
Interferences - Quantitative measured as percentages	-	223 ng/ml H	110.8 ng/ml L	110.8 ng/ml		43.2 ng/ml H	18.1 ng/ml L	1.020 ng/ml H	0.358 ng/ml L	5.5 ng/ml	23.1 ng/ml H	11.4 ng/ml L	2325 ng/ml H	817n g/ml L	-
LLN/ULN	-	64.8 / 107.2		-	13.4 / 43.2		0.239/0.496		-	6.3/13.6		608/1106		-	
Albumin	NA	7.5	21.9	-0.7	3.8	28.2	26.9	64.1	4.5	3.2	2.6	9.1	21.2	NA	
Blood 1%	62.3	31.4	87.7	5.4	7.4	34.5	46.2	112.9	133.4	54.2	108.2	4.0	-12.7	94.9	
Blood 0.8%	393.3	521.9	1392.7	-0.8	10.2	64.6	15.7	62.1	2.3	7.5	43.5	4.7	24.0	230.2	
Blood 0.2%	94.3	103.5	218.8	0.2	-0.8	25.7	5.9	51.4	-0.2	3.0	7.4	2.6	21.5	54.3	
HGB 0.8%	4.7	11.2	21	2.7	6.3	32.7	25.0	83.5	52.1	32.1	52	-1.5	1.6	31.9	
HGB 0.2%	0.9	9.5	22.7	1.7	0.6	24.8	11.8	62.1	9.9	8.0	16.7	0.7	6.1	12.9	
Stability	-	-	-	-	-	-	-	-	-	-	-	-	-	-	
Ambient	N/A	N/A	-	1 day	Not stable for times tested		1 hour	N/A	3 hours	At least 2 hours	6 hours				
2 – 8 C	N/A	N/A	-	7 day	1 day	3 days	5 days	3 days	6 days	2 days					
Freeze/Thaw	N/A	N/A	--	3	1	2	3	3	3	2					

Biomarker	Albumin	Clusterin	Creatinine ^e	Creatinine	Cystatin-C	KIM-1	NAG	NGAL	Osteopontin	Protein (Total)
Long Term (-70C)	N/A	N/A	-	N/A	N/A	1 year c	1 year c	At least 3 months (PBI) 1 year c	N/A	N/A

BTR – Between Run

WIR – Within Run

PAD – Preanalytical Dilutions

ND – Not Determined

a information is for urine with no preservatives added

b *The stability method reflects the number of Freeze/Thaw cycles after initial freeze*

c *Han WK et al. Urinary biomarkers in early detection of acute kidney injury after cardiac surgery. Clin J Am Soc Nephrol 4:873-882, 2009.*

d *Procedural dilution mandated by the manufacturer's protocol, performed on every sample including standards and controls, thus factor out.*

LLN and ULN – Upper Limit of Normal and Lower Limit of Normal as determined in this study

NA – Not Applicable

Based on the previously submitted PBI document, 0.8% washed blood is reported as 1.2 g/L Hbg. From Tietz, Mean Corpuscular Hemoglobin (MCH) 95% normal range for adults (male and female) is 31 ± 4 pg Hbg per erythrocyte. Thus,

0.8 % washed blood = $(1.2 \text{ g/L Hbg}) / (3.1 \cdot 10^{12} \text{ g Hbg/Ery}) = 0.387 \cdot 10^6 \text{ Ery}/\mu\text{L}$

0.2 % washed blood = $(0.3 \text{ g/L Hbg}) / (3.1 \cdot 10^{12} \text{ g Hbg/Ery}) = 0.096 \cdot 10^6 \text{ Ery}/\mu\text{L}$

e Two creatinine columns are listed due to two creatinine assay platforms being employed

ATTACHMENT D – Publications Relevant to Proposed Biomarkers

- Agarwal, Rajiv. 2004. “Statin Induced Proteinuria: Renal Injury or Renoprotection?” *Journal of the American Society of Nephrology: JASN* 15 (9): 2502–3. doi:10.1097/01.ASN.0000143720.71748.79.
- Amin, Rupesh P., Alison E. Vickers, Frank Sistare, Karol L. Thompson, Richard J. Roman, Michael Lawton, Jeffrey Kramer, et al. 2004. “Identification of Putative Gene Based Markers of Renal Toxicity.” *Environmental Health Perspectives* 112 (4): 465–79.
- Ariza, Xavier, Elsa Solà, Chiara Elia, Rogelio Barreto, Rebeca Moreira, Manuel Morales-Ruiz, Isabel Graupera, et al. 2015. “Analysis of a Urinary Biomarker Panel for Clinical Outcomes Assessment in Cirrhosis.” *PloS One* 10 (6): e0128145. doi:10.1371/journal.pone.0128145.
- Ascione, R., C. T. Lloyd, M. J. Underwood, W. J. Gomes, and G. D. Angelini. 1999. “On-Pump versus off-Pump Coronary Revascularization: Evaluation of Renal Function.” *The Annals of Thoracic Surgery* 68 (2): 493–98.
- Bailly, Veronique, Zhiwei Zhang, Werner Meier, Richard Cate, Michele Sanicola, and Joseph V. Bonventre. 2002. “Shedding of Kidney Injury Molecule-1, a Putative Adhesion Protein Involved in Renal Regeneration.” *The Journal of Biological Chemistry* 277 (42): 39739–48. doi:10.1074/jbc.M200562200.
- Bakris, George L. 2004. “Implications of Albuminuria on Kidney Disease Progression.” *Journal of Clinical Hypertension (Greenwich, Conn.)* 6 (11 Suppl 3): 18–22.
- Berg, José G. van den, Marius A. van den Bergh Weerman, Karel J. M. Assmann, Jan J. Weening, and Sandrine Florquin. 2004. “Podocyte Foot Process Effacement Is Not Correlated with the Level of Proteinuria in Human Glomerulopathies.” *Kidney International* 66 (5): 1901–6. doi:10.1111/j.1523-1755.2004.00964.x.
- Betton, Graham R., Daniela Ennulat, David Hoffman, Jean-Charles Gautier, Ernie Harpur, and Syril Pettit. 2012. “Biomarkers of Collecting Duct Injury in Han-Wistar and Sprague-Dawley Rats Treated with N-Phenylanthranilic Acid.” *Toxicologic Pathology* 40 (4): 682–94. doi:10.1177/0192623311436174.
- Biomarker Assay Collaborative Evidentiary Considerations Writing Group, Critical Path Institute (C-Path). 2016. “Points to Consider Document: Scientific and Regulatory Considerations for the Analytical Validation of Assays Used in the Qualification of Biomarkers in Biological Matrices.” https://healthpolicy.duke.edu/sites/default/files/atoms/files/white_paper_draft_11_4_16.pdf.
- Bolignano, Davide, Valentina Donato, Giuseppe Coppolino, Susanna Campo, Antoine Buemi, Antonio Lacquaniti, and Michele Buemi. 2008. “Neutrophil Gelatinase-Associated Lipocalin (NGAL) as a Marker of Kidney Damage.” *American Journal of Kidney Diseases* 52 (3): 595–605. doi:10.1053/j.ajkd.2008.01.020.
- Cassidy, Hilary, Jennifer Slyne, Patrick O’Kelly, Carol Traynor, Peter J. Conlon, Olwyn Johnston, Craig Slattery, Michael P. Ryan, and Tara McMorrow. 2015. “Urinary Biomarkers of Chronic Allograft Nephropathy.” *Proteomics. Clinical Applications* 9 (5–6): 574–85. doi:10.1002/prca.201400200.

- Cho, Yeoungjee, David W. Johnson, David A. Vesey, Carmel M. Hawley, Margaret Clarke, Nicholas Topley, and balANZ Trial Investigators. 2015. "Utility of Urinary Biomarkers in Predicting Loss of Residual Renal Function: The balANZ Trial." *Peritoneal Dialysis International: Journal of the International Society for Peritoneal Dialysis* 35 (2): 159–71. doi:10.3747/pdi.2013.00170.
- Christensen, E. I., H. Birn, B. Rippe, and A. B. Maunsbach. 2007. "Controversies in Nephrology: Renal Albumin Handling, Facts, and Artifacts!" *Kidney International* 72 (10): 1192–94. doi:10.1038/sj.ki.5002526.
- Collé, A., C. Tavera, P. Laurent, J. Leung-Tack, and J. P. Girolami. 1990. "Direct Radioimmunoassay of Rat Cystatin C: Increased Urinary Excretion of This Cysteine Proteases Inhibitor during Chromate Nephropathy." *Journal of Immunoassay* 11 (2): 199–214. doi:10.1080/01971529008053269.
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- Devarajan, P. 2014. "NGAL for the Detection of Acute Kidney Injury in the Emergency Room." *Biomarkers in Medicine* 8 (2): 217–19. doi:10.2217/bmm.13.149.
- Dieterle, Frank, Frank Sistare, Federico Goodsaid, Marisa Papaluca, Josef S Ozer, Craig P Webb, William Baer, et al. 2010. "Renal Biomarker Qualification Submission: A Dialog between the FDA-EMEA and Predictive Safety Testing Consortium." *Nature Biotechnology* 28 (5): 455–62. doi:10.1038/nbt.1625.
- Eddy, Allison A. 2004. "Proteinuria and Interstitial Injury." *Nephrology, Dialysis, Transplantation: Official Publication of the European Dialysis and Transplant Association - European Renal Association* 19 (2): 277–81.
- Eknoyan, Garabed, Thomas Hostetter, George L. Bakris, Lee Hebert, Andrew S. Levey, Hans-Henrik Parving, Michael W. Steffes, and Robert Toto. 2003. "Proteinuria and Other Markers of Chronic Kidney Disease: A Position Statement of the National Kidney Foundation (NKF) and the National Institute of Diabetes and Digestive and Kidney Diseases (NIDDK)." *American Journal of Kidney Diseases: The Official Journal of the National Kidney Foundation* 42 (4): 617–22.
- Emeigh Hart, Susan G. 2005. "Assessment of Renal Injury in Vivo." *Journal of Pharmacological and Toxicological Methods* 52 (1): 30–45. doi:10.1016/j.vascn.2005.04.006.

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