



FDA Briefing Document

Pediatric Oncology Subcommittee of the Oncologic Drugs Advisory Committee (ODAC)

June 20, 2018

DISCLAIMER STATEMENT

The attached package contains background information prepared by the Food and Drug Administration (FDA) for the panel members of the advisory committee. The FDA background package often contains assessments and/or conclusions and recommendations written by individual FDA reviewers. Such conclusions and recommendations do not necessarily represent the final position of the individual reviewers, nor do they necessarily represent the final position of the Review Division or Office.

We have brought the following issues to this Advisory Committee in order to gain the Committee's insights and opinions, and the background package will not include issues relevant to any final regulatory recommendation and instead is intended to focus on issues identified by the Agency for discussion by the advisory committee.

The issues under consideration are- Topic 1: Target List, Topic 2: FDARA Implementation, and Topic 3: Mechanisms to Assure Efficiency and to Enhance Global Coordination Through International Collaboration.

The FDA will not issue a final determination on the issues at hand until input from the advisory committee process has been considered and all reviews have been finalized. The final determination may be affected by issues not discussed at the advisory committee meeting.

Memorandum

Date: June 5, 2018

To: Pediatric Oncology Subcommittee of the Oncologic Drugs Advisory Committee (ODAC) Members, Consultants, and Guests

From: Gregory Reaman, MD
Associate Director, Office of Hematology and Oncology Products, CDER, and
Associate Director for Pediatric Oncology, Acting, Oncology Center of Excellence,
FDA

Subject: FDA Background Package for June 20, 2018 Meeting

Thank you for agreeing to participate in the upcoming Pediatric Oncology Subcommittee of the ODAC. The Subcommittee will hear about the review and discussion of a list of molecular targets for which evidence and/or biologic rationale exist to determine their potential relevance to the growth or progression of one or more pediatric cancers and a list of those targets deemed unlikely to be associated with the growth or progression of pediatric tumors. These lists are expected to fulfill the statutory obligation of the Food and Drug Administration Reauthorization Act (FDARA) and provide some guidance to industry in planning for initial Pediatric Study Plan submissions for new drug and/or biologic products in development for cancer in accordance with the amended provisions of the Pediatric Research Equity Act. The Subcommittee will review and discuss considerations other than scientific relevance that FDA will include in decision making with respect to the need and timing of pediatric evaluation of specific new drug and biologic products. The Subcommittee will discuss possible criteria and mechanisms for the prioritization by sponsors and the clinical investigator community of select targeted new agents for pediatric evaluation especially in the setting of multiple same in class agents. Preliminary discussion will focus on approaches to coordination of and collaboration in pediatric clinical investigations of new agents that might be pursued to efficiently accommodate international regulatory requirements and global pediatric product development. The open public hearing sessions are- Topic 1: Target List, Topic 2: FDARA Implementation, and Topic 3: Mechanisms to Assure Efficiency and to Enhance Global Coordination Through International Collaboration.

We believe that this focused discussion will utilize the expertise of the Pediatric Oncology Subcommittee in guiding the Agency's decisions related to the potential relevance of new drugs and biologics that require early pediatric evaluation to assure timely development of and access to safe and effective new therapies for children with cancer. As well, the expertise of the subcommittee is expected to inform the FDA and its development of a guidance on the implementation of the FDARA 2017 pediatric provisions.

As always, we appreciate your time and commitment and look forward to an informative meeting on June 20, 2018.

REFERENCE:

1. **Food and Drug Administration Safety and Innovation Act of 2012 (FDASIA):**
Title V – Pediatric Drugs and Devices (pages 47-58).

FDASIA legislation is available at: <http://www.gpo.gov/fdsys/pkg/BILLS-112s3187enr/pdf/BILLS-112s3187enr.pdf>

2. **FDA Reauthorization Act of 2017 (FDARA):**
Title V-Pediatric Drugs and Devices (Section 501-505)

FDARA legislation is available at: <https://www.congress.gov/115/plaws/publ52/PLAW-115publ52.pdf>

**Pediatric Oncology Subcommittee of the
Oncologic Drugs Advisory Committee (ODAC)
June 21, 2017
FDA Briefing Document**

TABLE OF CONTENTS

1. Pediatric Initiatives.....	4
2. Executive Summaries	
First Session: Target List	11
Second Session: FDARA Implementation.....	12
Third Session: Mechanisms to Assure Efficiency and to Enhance Global Coordination Through International Collaboration.....	13
3. Pediatric Molecular Target List.....	14

Relevant Molecular Targets in Pediatric Cancers: Applicability to Pediatric Therapeutic Investigations Required Under FDARA 2017

Pediatric cancer drug development typically leverages adult cancer drug discovery but has lagged far behind development of cancer drugs for adults. To date, the Pediatric Research Equity Act (PREA) has not been an effective mechanism to support the development of drugs for pediatric cancers because the requirement for conduct of pediatric studies is linked to the indication sought in adults and most adult cancers occur rarely, if ever, in children (e.g., cancers of the lung, prostate and breast). Therefore, sponsors obtain waivers for conducting assessments of these drugs in pediatric patients because studies would be infeasible or highly impracticable. Additionally, drugs developed for those rare cancer indications which may occur in both adult and pediatric populations which are frequently granted orphan designation are exempt from the requirements of PREA.

While there has been limited obligation to study investigational cancer therapies in children, incentives exist to promote the development of oncology products for pediatric cancer when these agents are in development or already approved for adult use. The Best Pharmaceuticals for Children Act (BPCA) is a voluntary mechanism which provides incentives in the form of 6 months of exclusivity for marketing to sponsors upon the completion and submission of pediatric studies that meet the terms of a written request from FDA (FD&C Act Sec. 505A, 21 USC 355a, FDA Amendments Act (FDAAA) Public Law 110-85). BPCA has been the sole legislative mechanism available to evaluate those oncology products of interest for their potential role in the treatment of malignancies in children and adolescents.

The 2017 enactment of Title V of the Food and Drug Administration Reauthorization Act (FDARA) (FD&C Act Sec. 505B (a)(3), 21 USC 355c (a)(3), Public Law 115-52), which amends the requirement for pediatric assessment of new drugs under the Pediatric Research Equity Act (PREA) based on molecular mechanism of action rather than clinical indication, may dramatically change the landscape for pediatric cancer drug development. FDARA has now created a mechanism to expedite the evaluation of novel medicines with the potential to address the unmet need in the pediatric population.

Molecularly targeted agents have advanced the concept of Precision Medicine in oncology. As malignancies occurring in children and adolescents can harbor molecular abnormalities similar to those found in adult cancers, these agents may be relevant to the treatment of pediatric patients with cancer. Although large scale sequencing efforts, such as TARGET, the Pediatric Cancer Genome Project, and the International Cancer Genome Consortium's Pedbrain Tumor and MMML-seq projects provide evidence that the genetic and epigenetic repertoires of driver gene aberrations often differ between adult and pediatric cancers, a growing body of evidence suggests that genetic and other molecular biological vulnerabilities of certain adult cancers may also occur in pediatric cancers, thereby providing opportunities for the use of targeted therapies in select pediatric tumors; up to 50% of pediatric cancers harbor a potentially druggable event. Timely investigation of the antitumor activity of potentially useful targeted drugs and biologics

under development in adults, and of their toxicities relative to the unique growth and developmental considerations of pediatric patients, is warranted for pediatric populations with cancer.

Title V of FDARA has amended PREA to support early evaluation of such drugs by requiring pediatric investigation of appropriate new drugs intended for adults with cancer. The investigations that FDA may require by statute are referred to as **molecularly targeted pediatric cancer investigations**. These investigations may include clinical studies designed to yield clinically meaningful pediatric study data, gathered using appropriate formulations for each age group for which the study is required, regarding dosing, safety and preliminary efficacy to inform potential pediatric labeling [FDARA Title V Sec 504 (a)(3)(A), FD&C Act Sec. 505B (a)(3)(A), 21 USC 355c(a)(3)(A)]. Importantly, Title V of FDARA also specifies that the requirement for early pediatric investigations of drugs directed at molecular targets considered substantially relevant to the growth or progression of a pediatric cancer be applied, even when the adult indication has received an orphan designation, or when the adult cancer indication does not occur or is biologically different in the pediatric population.

The statute also directs the FDA, in collaboration with the National Cancer Institute (NCI), to establish, publish, and regularly update a list of molecular targets considered, on the basis of data the Agency determines to be adequate, to be substantially relevant to the growth or progression of pediatric cancers, and that may trigger the requirement for pediatric investigations [21 USC 355c (m)(1)(A)]. Molecular targets that are considered “not relevant” to the growth or progression of pediatric cancers will be placed on a second list [21 USC 355c (m)(1)(B)]. The statute does not stipulate that a molecular target to which a specific drug is directed must appear on the relevant target list to require a clinical evaluation of the drug in the pediatric population. Furthermore, presence of a target on the relevant target list does not in itself constitute a requirement for a clinical study.

The FDA is mandated to convene a public meeting no later than 1 year after the date of the enactment of FDARA to solicit views of physicians, academic researchers (including pediatric oncologists and rare disease specialists), patient advocates, industry and other stakeholders for the establishment of the molecular targets lists [21 USC 355c (m)(2)(1)]. This meeting of the Pediatric Subcommittee of the Oncologic Drugs Advisory Committee (ODAC) on June 20, 2018 is one such meeting.

A Friends of Cancer Research (FOCR)-sponsored multi-stakeholder public workshop held on February 20, 2018, discussed approaches for developing, updating and applying the molecular target list. This meeting included discussions of a framework to assess factors to define molecular targets as substantially relevant or not relevant to the growth or progression of one or more pediatric cancers; proposed processes and timelines for regularly updating the lists of molecular targets; and additional considerations for the application of the molecular target lists to decision-making regarding pediatric evaluation of specific drugs.

Although there may be differences in the way “molecular target” is defined, for the purposes of establishing a list of molecular targets considered to be substantially relevant to the growth or progression of pediatric cancer, a molecular target is defined **as a molecule in human cells that**

is intrinsically associated with a particular disease process, such as etiology, progression, or drug resistance. To be referred to as a target, there must be evidence that by engaging the target, either with a targeted small molecule, biologic product, or other treatment intervention, a desired therapeutic effect is produced that results in the alteration of the disease process. In other words, a molecule would not be referred to as a molecular target if there is no evidence to inform the hypothesis that its modulation (i.e., inhibition or activation) alters the disease.

The proposed framework for assessing those factors that may guide the definition of molecular targets as substantially relevant or not relevant to the growth or progression of one or more pediatric cancers includes a classification system to facilitate an organized approach to determination of relevance. Targets may represent the result of specific gene abnormalities, are present in a critical biologically-related pathway of a gene abnormality or exhibit a synthetic lethal relationship to a gene abnormality (**gene abnormality-based targets**). Targets can be intrinsic to the cancer cell lineage or developmental stage (**cancer cell lineage-based targets**), or they may be identified in non-cancer cells, such as normal immune cells or supporting cells contributing to the tumor micro-environment (**non-cancer cell targets**). Targets may exist as essential elements of cancer cells as well as some non-cancer cells and are not caused by a specific genetic aberration, such as tubulin or heat-shock proteins (**other targets**).

Evidence of effectiveness for a drug or biologic directed at a molecular target in an adult cancer and identification of the target as substantially relevant for the growth or progression of a pediatric cancer provide a rationale for the agent's evaluation in the pediatric cancer population, regardless of similarity to the histologically-defined cancer found in the adult. Although not a requirement, it is beneficial for sponsors of such an agent to develop *in vitro* and/or *in vivo* data using pediatric non-clinical models to provide increased confidence in either establishing or refuting potential relevance.

Two specific elements of a framework were identified, one that outlines factors that may be useful when determining whether a target is substantially relevant in pediatric cancer and may trigger the requirement for pediatric investigations. The second outlines factors to consider when assessing the available data that may help determine there is insufficient evidence of relevance, and that the target is hence "not relevant". (See Tables 1 and 2)

The FDA, in collaboration with the NCI, is tasked with determining whether a molecular target is or is not considered substantially relevant to the growth or progression of pediatric cancer. FDA has the regulatory authority to determine whether adequate evidence is available to define a target as substantially relevant to require pediatric investigation of a drug directed at that target. Defining a specific evidence standard for determination of target relevance is not feasible for several reasons, including the different classes of targets, variability in evidence base that may exist among targets and between specific target classes, and the fact that emerging science evolves making pre-defined qualifications based on peer-reviewed publications or publicly available registry data difficult. Several factors may support a scientifically-based and data-driven decision-making approach. These factors are not meant to be either all-inclusive or prescriptive, as there may be additional factors for some specific targets and some factors may not be required for all targets within a class. Table 1 characterizes the factors used in the

determination of whether a molecular target is substantially relevant to the growth or progression of pediatric cancers. The framework is not meant to be interpreted as a checklist, and it is important to note that the totality of evidence available may be considered when guiding discussions to determine target relevance. Additionally, the presence of a single factor or a combination of factors may not be sufficient to define relevance.

Table 1: Framework of factors and characteristics that may guide the determination of whether molecular targets are substantially relevant in the growth or progression of pediatric cancer

Factors	Considerations
Presence of target	The target has been identified in at least one case of a pediatric cancer
Target class: Gene abnormality	The gene abnormality has been identified in at least one case of a pediatric cancer
Target class: Cancer cell lineage	The target is intrinsically and differentially expressed in the cancer of interest compared to normal site-specific tissues.
Function/Mechanism	The biological function of the target is relevant to the etiology and growth of the childhood cancer
Target class: Gene abnormality	Modulation of the affected gene product or of a critical downstream pathway or correction/deletion of the affected gene defect adversely affects cancer cells
Target class: Cancer cell lineage	The presence of the gene abnormality creates a synthetic lethal relationship with another cellular pathway
Target class: Cancer cell lineage	The target is associated to cancer cell development, growth and survival
Non-clinical evidence	Non-clinical evidence supports relevance of target in one or more pediatric cancers
<i>In vitro</i> activity	Target modulation shows <i>in vitro</i> selectivity for cancer cell lines containing/expressing the molecular target (pediatric or adult cell lines if target is known to be shared by multiple cancer types regardless of patient population) compared to the sensitivity of cell lines not containing/expressing the target
<i>In vivo</i> activity ¹	Target modulation shows <i>in vivo</i> activity manifested as tumor stabilization or regression in models of pediatric cancers with the molecular target of interest (or adult cancer models containing/expressing the target)
Lack of <i>in vitro</i> or <i>in vivo</i> activity	For targets for which target modulation does not show <i>in vivo</i> or <i>in vitro</i> activity, support for relevance may be found in evidence for supra-additive or synergistic activity when target modulation is used in biologically rational combinations
Adult clinical experience	Target modulation by investigational agents known to affect the target, shows clinical activity in specific cancers in adults
Predictive biomarkers	Biomarkers that predict responses to target modulation may be useful in the selection of appropriate pediatric study populations
Location	For immunotherapy targets, the target is expressed on the cell surface (excepting immunotherapies that target intracellular antigens that are displayed as peptides by MHC proteins on the cell surface)
Agent under development	There is an agent in development or proceeding to development that addresses the specific target

¹The *in vivo* activity should be observed at drug exposures that are relevant to the clinical setting if there is clinical experience with the agent. Prolonged stable disease may be relevant, particularly for agents that induce their anticancer effect through mechanisms other than cancer cell apoptosis.

Because of the potential importance of non-clinical evaluation in contributing to the evidence base for relevance of a molecular target, every effort should be made to ensure sponsors expedite early non-clinical investigation, which could be in collaboration with academic research teams with pediatric expertise in non-clinical testing. The creation of these collaborations and/or partnerships, ideally international in scope, should be explored further as they will be crucial for early testing of non-clinical models, such as patient-derived xenograft models.

Biomarkers that are identified as predictive for the activity of adult cancer targeted agents should also be evaluated for prevalence of expression and potential utility across pediatric cancers. Sponsors are strongly encouraged to test samples from pediatric cancers to determine prevalence, especially when an assay to identify a biomarker is developed in conjunction with the investigational agent and may not be available for use on patient samples by investigators.

There may be evidence available that demonstrates a molecular target is not relevant in pediatric cancers that would prevent it from being added to the substantially relevant molecular target list. The factors listed in Table 2 highlight considerations that may guide the determination of whether a molecular target is not relevant to the growth or progression of pediatric cancer. Again, it is solely the FDA’s responsibility to determine what evidence is necessary to determine whether a molecular target is considered not relevant in pediatric cancer, and thus this document does not attempt to define what “adequate evidence” refers to in this context.

Table 2: Framework of factors and characteristics to consider that may guide the determination of whether molecular targets are *not relevant* to the growth or progression of pediatric cancer¹

Factors	Considerations
Biologically implausible	Molecular targets for which available evidence supports no role for the targets in pediatric cancers (e.g. endocrine/autocrine sex steroid hormonal pathways that are known to be drivers of specific adult cancer types but are very rarely to never observed in pediatric cancers)
Non-clinical evidence	Evidence of lack of activity of an agent in development against a specific target in non-clinical systems could be a component of the evidence base used to determine that a specific molecular target may not be relevant to the growth or progression of a pediatric cancer.
Adult clinical evidence	Evidence of lack of clinical activity of an agent in development against a specific target could be a component of the evidence base used to determine that a specific molecular target may not be relevant to the growth or progression of a pediatric cancer.

¹There may be agents that are relevant to the growth or progression of disease but that would not be considered for development because of their association with developmental processes such that their inhibition would raise concerns about irreversibly deleterious developmental effects and subsequent growth-related toxicities.

Molecular targets which lack sufficient evidence to make a determination of “substantially relevant” or “not relevant” will not be included in either list. Decisions regarding relevance of these targets to the growth or progression of pediatric cancers will be made when there is an adequate evidence base to make such a determination. Sponsors and investigators are strongly encouraged to investigate the potential relevance of new and currently unlisted targets as expeditiously as possible, especially when there are early non-clinical or clinical signals of activity.

To ensure molecular targets lists are updated in a timely fashion using the most relevant evidence available in light of the rapid pace of scientific advances, three distinct suggestions emerged:

- The first suggestion includes a semi-annual public workshops at which all stakeholders could discuss potential changes to the molecular targets lists. The FDA is responsible for

convening and presiding over these workshops , which may occur following a national or international scientific meeting. These meetings will seek input from individual stakeholders on advances in relevant scientific evidence that may impact the inclusion of one or more molecular targets on the current published lists, including potential relevance of unlisted targets. Decisions related to the lists will also reflect input from the Pediatric Subcommittee of ODAC.

- The second mechanism is a transparent nomination process open to sponsors and academic investigators to occur during or prior to meetings of the Pediatric Subcommittee of the ODAC. The details of such a transparent electronic nomination process, coordinated by the FDA's Oncology Center of Excellence will be determined.
- The third proposed mechanism is for clinical investigators or sponsors to request a meeting with the Pediatric Oncology Program of the FDA's Oncology Center of Excellence to discuss new scientific data related to a new or existing molecular target which may warrant a change in that target's status as relevant or non-relevant.

Information from these sources could then be assessed by the FDA, with possible input from the Pediatric Subcommittee of the ODAC, to determine whether there is sufficient new evidence to support changing the relevance status of the target of interest.

Additional factors that may require consideration when seeking to utilize the list of molecular targets for decisions regarding pediatric evaluation include analysis of clinical benefit and risk, the availability of pediatric formulations, and the adequacy of patient populations when planning clinical trials. These factors may vary with each targeted product under consideration for pediatric study and will be assessed and subject to multi-stakeholder discussion.

The preliminary list of relevant and non-relevant targets (attached) has been constructed in collaboration with the NCI and with input from recognized, international, multi-disciplinary experts in translational and clinical research in pediatric cancer from academia and the pharmaceutical industry and reviewed publicly on April 20, 2018. The FDA welcomes discussion of the proposed lists and input on the additional considerations and factors to guide pediatric clinical investigations with the expectation of expediting the development of safe and effective therapies for children with cancer.

First Session

Topic 1: Target List

Issues Relating to the Development of the Target List

1. Title V of the FDA Reauthorization Act (FDARA) 2017 assigns FDA to establish, publish and regularly update a list of molecular targets considered on the basis of data the FDA determines to be adequate to be substantially relevant to the growth or progression of pediatric cancers. New drug products directed at these targets may trigger the requirement for pediatric investigations [21 USC 355c(m)(1)(A)]. As well, a list of targets considered “not relevant” [21 USC 355c(m)(1)(B)] has been developed. Comment on the process utilized to construct the list, the classification of molecular targets, the factors utilized to designate a target as relevant or non-relevant and indicate your concurrence with the lists as currently presented.
2. Please comment on the process proposed for formally updating the lists at semi-annual public workshops, the methods for nominating potential future candidate targets, and the required transparency in multi-stakeholder discussions to determine relevance. Comment on additional measures to assure timely discussion of emerging science and its clinical translation which has the potential to expedite drug development to improve the care and outcome of children with cancer.

Second Session

Topic 2: FDARA Implementation

Issues Relating to FDARA Implementation

1. Please comment on the proposed additional considerations for which the FDA might engage with industry, clinical investigators, and advocates when making decisions regarding the requirement for pediatric studies of new drug and biologic products based on molecular mechanism of action and their timing.

Third Session

Topic 3: Mechanisms to Assure Efficiency and to Enhance Global Coordination Through International Collaboration

Issues Relating to the Mechanisms to Assure Efficiency and to Enhance Global Coordination Through International Collaboration

1. Please discuss transparent mechanisms for industry, advocates, and the academic investigator community to communicate and provide input to the FDA for purposes of eliminating unnecessary duplication of clinical trials in rare pediatric cancer populations of same in class agents.
2. Please comment on process development aimed at enhancing international collaboration between clinical trial networks to facilitate global cancer drug development for children in light of currently non-aligned regulatory requirements.

Pediatric Molecular Target List

Please see next page.

Target Symbol	Gene Abnormality	Citation(1)	Link(1)	Citation(2)	Link(2)	Citation(3)	Link(3)
ABL1/2	ABL1/2 gene fusions (BCR-ABL1, etc.)	Greuber, E. K., Smith-Pearson, P., Wang, J., & Pengdast, A. M. (2013). Role of ABL family kinases in cancer. <i>From leukaemia to solid tumours. Nature Reviews Cancer</i> , 13(8), 559-571. doi:10.1038/nrc3563	https://www.ncbi.nlm.nih.gov/pmc/articles/PMC3935732/				
ACVR1	ACVR1	Taylor, K. R., Vinci, M., Bullock, A. N., & Jones, C. (2014). ACVR1 mutations in DIPG: lessons learned from FOP. <i>Cancer Research</i> , 74(17), 4565-4570. http://doi.org/10.1158/0008-5472.CAN-14-1298	https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4154859/				
ALK	ALK and ALK gene fusions	Holla, V. R., Elamin, Y. Y., Bailey, A. M., Johnson, A. M., Litzemberger, B. C., Khotskaya, Y. B., ... Simon, G. R. (2017). ALK a tyrosine kinase target for cancer therapy. <i>Cold Spring Harbor Molecular Case Studies</i> , 3(1), a001115. http://doi.org/10.1101/mcs.a001115	https://www.ncbi.nlm.nih.gov/pmc/articles/PMC5171696/				
ASCL1	ASCL1 gene	Kasim, M., Benko, E., Winkelmann, A., Mrowka, R., Staudacher, J. J., Persson, P. B., ... Fahlring, M. (2014). Shutdown of Achaete-scute Homolog-1 Expression by Heterogeneous Nuclear Ribonucleoprotein (hnRNP-A2/B1) in Hypoxia. <i>Journal of Biological Chemistry</i> , 289(39), 26973-26988. doi:10.1074/jbc.m114.579391	http://www.jbc.org/content/289/39/26973.full				
BRAF	BRAF	Kieran, M. W. (2014). Targeting BRAF in Pediatric Brain Tumors. <i>American Society of Clinical Oncology Educational Book</i> , 34. doi:10.14694/edbook.am.2014.34.e436	https://meetinglibrary.asco.org/record/890/39/edbook#fulltext	Dahiya S, Emmett RJ, Haydon DH, et al. BRAF-V600E mutation in pediatric and adult glioblastoma. <i>Neuro Oncol</i> . 2014;16:318-319.	https://www.ncbi.nlm.nih.gov/pmc/articles/PMC3895374/		
CDK12	EWSR1-FLI1	Iniguez, A. B., Stolte, B., Wang, E. J., Conway, A. S., Alexe, G., Dharria, N. V., ... Stegmaier, K. (2018). EWS/FLI Confers Tumor Cell Synthetic Lethality to CDK12 Inhibition in Ewing Sarcoma. <i>Cancer Cell</i> , 33(2). doi:10.1016/j.ccell.2017.12.009	https://www.ncbi.nlm.nih.gov/pubmed/29358035				
CSF1R	CSF1R gene fusions	Rovida, E., & Sbarba, P. D. (2015). Colony-Stimulating Factor-1 Receptor in the Polarization of Macrophages: A Target for Turning Bad to Good Ones? <i>Journal of Clinical & Cellular Immunology</i> , 06(06). doi:10.4172/2155-9899.1000379	https://www.omicsonline.org/open-access/colonystimulating-factor1-receptor-in-the-polarization-of-macrophages-target-for-turning-bad-to-good-ones-2155-9899-1000379.pdf	Butowski, N., Colman, H., Groot, J. F., Omuro, A. M., Nayak, L., Wen, P. Y., ... Prados, M. (2015). Orally administered colony stimulating factor 1 receptor inhibitor PLX3397 in recurrent glioblastoma: An Ivy Foundation Early Phase Clinical Trials Consortium phase II study. <i>Neuro-Oncology</i> , 18(4), 557-564. doi:10.1093/neuonc/nov245	https://academic.oup.com/neuro-oncology/article/18/4/557/2509330		
CTNNB1 (β-catenin)	CTNNB1	Shukla, N., Ameer, N., Yilmaz, I., Nafa, K., Lau, C., Marchetti, A., ... Ladanyi, M. (2011). Oncogene Mutation Profiling of Pediatric Solid Tumors Reveals Significant Subsets of Embryonal Rhabdomyosarcoma and Neuroblastoma with Mutated Genes in Growth Signaling Pathways. <i>Clinical Cancer Research</i> , 18(3), 748-757. doi:10.1158/1078-0432.ccr-11-2056	https://www.ncbi.nlm.nih.gov/pmc/articles/PMC3271129/				
DDX3X	DDX3X	Epling, L. B., Grace, C. R., Lowe, B. R., Partridge, J. F., & Enemark, E. J. (2015). Cancer-associated mutants of RNA helicase DDX3X are defective in RNA-stimulated ATP hydrolysis. <i>Journal of Molecular Biology</i> , 427(9), 1779-1796. http://doi.org/10.1016/j.jmb.2015.02.015	https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4402148/				
DOT1L	MLL gene fusions	Wong, M., Tee, A., Milazzo, G., Bell, J., Hüttemaier, S., Polly, P., ... Liu, T. (2017). Abstract LB-080 The histone methyltransferase DOT1L promotes neuroblastoma by regulating gene transcription. <i>Cancer Research</i> , 77(13 Supplement). doi:10.1158/1538-7445.am2017-lb-080	https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4633909/				
EGFR	EGFR	Lee, J. A., Ko, Y., Kim, D. H., Lim, J. S., Kong, C.-B., Cho, W. H., ... Koh, J.-S. (2012). Epidermal Growth Factor Receptor: Is It a Feasible Target for the Treatment of Osteosarcoma? <i>Cancer Research and Treatment: Official Journal of Korean Cancer Association</i> , 44(3), 202-209. http://doi.org/10.4143/crt.2012.44.3.202	https://www.ncbi.nlm.nih.gov/pmc/articles/PMC3467424/				
ERK	BRAF, MAP2K1	Tp, T. A. (2015). Targeted Therapy for MAPK Alterations in Pediatric Gliomas. <i>Brain Disorders & Therapy</i> , S2. doi:10.4172/2168-975x.s2-005	https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4627711/	Knight, T., & Irving, J. A. (2014). Ras/Raf/MEK/ERK Pathway Activation in Childhood Acute Lymphoblastic Leukemia and Its Therapeutic Targeting. <i>Frontiers in Oncology</i> , 4. doi:10.3389/fonc.2014.00160	https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4067595/		
EWSR1-FLI1	EWSR1-FLI1	Gamberi, G., Cocchi, S., Benini, S., Magagnoli, G., Morandi, L., Kreshak, J., ... Alberghini, M. (2011). Molecular Diagnosis in Ewing Family Tumors. <i>The Journal of Molecular Diagnostics</i> , 13(3), 313-324. doi:10.1016/j.jmoldx.2011.01.004	https://www.ncbi.nlm.nih.gov/pmc/articles/PMC3077725/				
EZH2	SMARCB1, SMARCA4	D'Angelo, V., Iannotta, A., Ramaglia, M., Lombardi, A., Zarone, M. R., Desiderio, V., ... Caraglia, M. (2015). EZH2 is increased in paediatric T-cell acute lymphoblastic leukemia and is a suitable molecular target in combination treatment approaches. <i>Journal of Experimental & Clinical Cancer Research</i> , 34(1). doi:10.1186/s13046-015-0191-0	https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4335295/	Chang, C., & Hung, M. (2011). The role of EZH2 in tumour progression. <i>British Journal of Cancer</i> , 106(2), 243-247. doi:10.1038/bjc.2011.551	https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4132442/		
FGFR	FGFR and FGFR gene fusions	Venneti, S., & Huse, J. T. (2015). The Evolving Molecular Genetics of Low-grade Glioma. <i>Advances In Anatomic Pathology</i> , 22(2), 94-101. doi:10.1097/pap.0000000000000049	https://www.ncbi.nlm.nih.gov/pmc/articles/PMC466750/	Porta, R., Borea, R., Coelho, A., Khan, S., Araújo, A., Reclusa, P., ... Rolfo, C. (2017). FGFR a promising druggable target in cancer: Molecular biology and new drugs. <i>Critical Reviews in Oncology/Hematology</i> , 113, 256-267. doi:10.1016/j.critrevonc.2017.02.018	http://www.croh-online.com/article/S1040-8428(17)30085-9/fulltext	Linzey, J. R., Marini, B., Mcfadden, K., Lorenzana, A., Mody, R., Roberson, P. L., & Koschmann, C. (2017). Identification and targeting of an FGFR fusion in a pediatric thalamic "central oligodendroglioma". <i>Npj Precision Oncology</i> , 1(1). doi:10.1038/s41698-017-0036-8	https://www.nature.com/articles/s41698-017-0036-8
FLT3	FLK2, STK1, CD135	Grafone, T., Palmisano, M., Nicci, C., & Storti, S. (2012). An overview on the role of FLT3-tyrosine kinase receptor in acute myeloid leukemia. <i>Biology and treatment. Oncology Reviews</i> , 6(1), 8. doi:10.4081/oncol.2012.e8	https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4419636/	Levis, M. (2013). FLT3 mutations in acute myeloid leukemia: what is the best approach in 2013? <i>Hematology / the Education Program of the American Society of Hematology. American Society of Hematology. Education Program</i> , 2013, 220-226. http://doi.org/10.1182/asheducation-2013.1.220	https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4714709/		
Gamma secretase	NOTCH1 and FBXW7	Kolb, E. A., Gorlick, R., Keir, S. T., Maris, J. M., Lock, R., Carol, H., ... Smith, M. A. (2011). Initial testing (stage 1) by the pediatric preclinical testing program of RO4929097, a γ-secretase inhibitor targeting notch signaling. <i>Pediatric Blood & Cancer</i> , 58(5), 815-818. doi:10.1002/pbc.25290	https://www.ncbi.nlm.nih.gov/pmc/articles/PMC3276746/				

GF1I	GF1I	Hönes, J. M., Botezatu, L., Helness, A., Vadnais, C., Vassen, L., Robert, F., . . . Khandanpour, C. (2016). GF1I as a novel prognostic and therapeutic factor for AML/MDS. <i>Leukemia</i> , 30(6), 1237-1245. doi 10.1038/leu.2016.11	https://www.nature.com/articles/leu201611
GF1IB	GF1IB	Stevenson, W. S., Morel-Kopp, M., Chen, Q., Liang, H. P., Bromhead, C. J., Wright, S., . . . Ward, C. M. (2013). GF1I mutation causes a bleeding disorder with abnormal platelet function. <i>Journal of Thrombosis and Haemostasis</i> , 11(11), 2039-2047. doi 10.1111/jth.12368	https://onlinelibrary.wiley.com/doi/full/10.1111/jth.12368
Histone 3 G34R/V	Histone 3 G34R/V	Yuen, B., & Knopfle, P. (2013). Histone H3.3 Mutations A Variant Path to Cancer. <i>Cancer Cell</i> , 24(5), 567-574. doi 10.1016/j.ccr.2013.09.015	https://www.ncbi.nlm.nih.gov/pmc/articles/PMC3882088/
Histone 3 K27M	Histone 3 K27M	Yuen, B., & Knopfle, P. (2013). Histone H3.3 Mutations A Variant Path to Cancer. <i>Cancer Cell</i> , 24(5), 567-574. doi 10.1016/j.ccr.2013.09.015	https://www.ncbi.nlm.nih.gov/pmc/articles/PMC3882088/
IDH1 and IDH2	IDH1 and IDH2	Yang, H., Ye, D., Guan, K., & Xiong, Y. (2012). IDH1 and IDH2 Mutations in Tumorigenesis Mechanistic Insights and Clinical Perspectives. <i>Clinical Cancer Research</i> , 18(20), 5562-5571. doi 10.1158/1078-0432.ccr-12-1773	https://www.ncbi.nlm.nih.gov/pmc/articles/PMC3897211/
JAK1, 2, and 3	JAK1, 2, and 3	Yang, H., Ye, D., Guan, K., & Xiong, Y. (2012). IDH1 and IDH2 Mutations in Tumorigenesis Mechanistic Insights and Clinical Perspectives. <i>Clinical Cancer Research</i> , 18(20), 5562-5571. doi 10.1158/1078-0432.ccr-12-1773	https://www.ncbi.nlm.nih.gov/pmc/articles/PMC3897211/
LIN28B	LIN28B	Balzeau, J., Menezes, M. R., Cao, S., & Hagan, J. P. (2017). The LIN28let-7 Pathway in Cancer. <i>Frontiers in Genetics</i> , 8, 31. http://doi.org/10.3389/fgene.2017.00031	https://www.ncbi.nlm.nih.gov/pmc/articles/PMC5368188/
MDM2	MDM2, TP53	Barone, G., Tweddle, D., Shohet, J., Chesler, L., Moreno, L., Pearson, A., & Maerken, T. (2014). MDM2-p53 Interaction in Paediatric Solid Tumours Preclinical Rationale, Biomarkers and Resistance. <i>Current Drug Targets</i> , 15(1), 114-123. doi 10.2174/13894501113149990194	https://www.ncbi.nlm.nih.gov/pubmed/24387312
MEK	BRAF and BRAF gene fusions, MAP2K1, NF1	Ciccarelli, C., Vulcano, F., Milazzo, L., Gravina, G. L., Marampon, F., Macioce, G., . . . Zani, B. M. (2016). Key role of MEK/ERK pathway in sustaining tumorigenicity and in vitro radioresistance of embryonal rhabdomyosarcoma stem-like cell population. <i>Molecular Cancer</i> , 15(1), doi 10.1186/s12943-016-0501-y	https://molecular-cancer.biomedcentral.com/articles/10.1186/s12943-016-0501-y
Menin	MLL gene fusions	Slany, R. K. (2016). The molecular mechanics of mixed lineage leukemia. <i>Oncogene</i> , 35(40), 5215-5223. doi 10.1038/onc.2016.30	https://www.ncbi.nlm.nih.gov/pmc/articles/PMC2704309/
MET	MET	Boufflet, E. (2007). Faculty of 1000 evaluation for Phase 2 study of temozolomide in children and adolescents with recurrent central nervous system tumors A report from the Childrens Oncology Group. F1000 - Post-publication Peer Review of the Biomedical Literature. doi 10.3410/f.1098180.554184	https://www.ncbi.nlm.nih.gov/pmc/articles/PMC3123765/
MLL	MLL gene fusions	Winters, A. C., & Bernt, K. M. (2017). MLL-Rearranged Leukemias—An Update on Science and Clinical Approaches. <i>Frontiers in Pediatrics</i> , 5. doi 10.3389/fped.2017.00004	https://www.ncbi.nlm.nih.gov/pmc/articles/PMC5299633/
mTOR	TSC1, TSC2	Barrett, D., Brown, V. I., Grupp, S. A., & Teachey, D. T. (2012). Targeting the PI3K/AKT/mTOR Signaling Axis in Children with Hematologic Malignancies. <i>Pediatric Drugs</i> , 14(5), 299-316. doi 10.1007/bf03262236	https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4214862/
MYC	MYC translocations and amplification	Hutter, S., Bolin, S., Weishaupt, H., & Swartling, F. (2017). Modeling and Targeting MYC Genes in Childhood Brain Tumors. <i>Genes</i> , 8(4), 107. doi 10.3390/genes8040107	https://www.ncbi.nlm.nih.gov/pmc/articles/PMC5406854/
MYCN	MYCN amplification	Sala, A. (2015). Editorial Targeting MYCN in Pediatric Cancers. <i>Frontiers in Oncology</i> , 4. doi 10.3389/fonc.2014.00330	https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4429566/
Neoantigens	MSH2, MLH1, MSH6, PMS2 POLE, and POLD1	Schumacher, T. N., & Schreiber, R. D. (2015). Neoantigens in cancer immunotherapy. <i>Science</i> , 348(6230), 69-74. doi 10.1126/science.aaa4971	http://science.sciencemag.org/content/348/6230/69/tab-pdf
NFKappaB	RELA fusion	Cahill, K. E., Morshed, R. A., & Yamini, B. (2015). Nuclear factor-κB in glioblastoma Insights into regulators and targeted therapy. <i>Neuro-Oncology</i> , 18(3), 329-339. doi 10.1093/neuonc/nov265	https://academic.oup.com/neuro-oncology/article/18/3/329/2509337
NOTCH1	NOTCH1, FBXW7	Zage, P. E., Nolo, R., Fang, W., Stewart, J., Garcia-Manero, G., & Zweidler-Mckay, P. A. (2011). Notch pathway activation induces neuroblastoma tumor cell growth arrest. <i>Pediatric Blood & Cancer</i> , 58(5), 682-689. doi 10.1002/pbc.23202	https://www.ncbi.nlm.nih.gov/pubmed/21744479
NTSC2	NTSC2	Meyer, J. A., Wang, J., Hogan, L. E., Yang, J. J., Dandekar, S., Patel, J. P., . . . Carroll, W. L. (2013). Relapse specific mutations in NTSC2 in childhood acute lymphoblastic leukemia. <i>Nature Genetics</i> , 45(3), 290-294. http://doi.org/10.1038/ng.2558	https://www.ncbi.nlm.nih.gov/pmc/articles/PMC3681285/
NTRK	NTRK gene fusions	Prasad, M. L., Vyas, M., Home, M. J., Virk, R. K., Morotti, R., Liu, Z., . . . Nikiforov, Y. E. (2016). NTRK fusion oncogenes in pediatric papillary thyroid carcinoma in northeast United States. <i>Cancer</i> , 122(7), 1097-1107. doi 10.1002/cncr.29887	https://onlinelibrary.wiley.com/doi/pdf/10.1002/cncr.29887
ODC1	MYC target gene	Schultz, C. R., Geerts, D., Mooney, M., El-Khawaja, R., Koster, J., & Bachmann, A. S. (2018). Synergistic drug combination GC7/DFMO suppresses hypusine/spermidine-dependent eIF5A activation and induces apoptotic cell death in neuroblastoma. <i>Biochemical Journal</i> , 475(2), 531-545. doi 10.1042/bj20170597	http://www.biochemj.org/content/ppbiochem/early/2018/01/02/BJ20170597.full.pdf
PARP	BRCA1/2, PALB2, ATM, BRIP1, CHEK2, RAD51, etc.	Ricks, T. K., Chiu, H.-J., Ison, G., Kim, G., McKee, A. E., Kluetz, P., & Puzdur, R. (2015). Successes and Challenges of PARP Inhibitors in Cancer Therapy. <i>Frontiers in Oncology</i> , 5, 222. http://doi.org/10.3389/fonc.2015.00222	https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4604313/
PAX-FOXO1	PAX-FOXO1	Linardic, C. M. (2008). PAX3-FOXO1 fusion gene in rhabdomyosarcoma. <i>Cancer Letters</i> , 270(1), 10-18. doi 10.1016/j.canlet.2008.03.035	https://www.ncbi.nlm.nih.gov/pmc/articles/PMC2575376/
PDGFRA/B	PDGFRA/B gene fusions	Heldin, C. (2013). Targeting the PDGF signaling pathway in tumor treatment. <i>Cell Communication and Signaling</i> , 11(1), 97. doi 10.1186/1478-811x-11-97	https://www.ncbi.nlm.nih.gov/pmc/articles/PMC3878225/
		Goethem, A. V., Yigit, N., Moreno-Smith, M., Vasudevan, S. A., Barbieri, E., Speleman, F., . . . Maerken, T. V. (2017). Dual targeting of MDM2 and BCL2 as a therapeutic strategy in neuroblastoma. <i>Oncotarget</i> , 8(34). doi 10.18632/oncotarget.18982	https://www.ncbi.nlm.nih.gov/pmc/articles/PMC5593624/
		Ferrando, A. A. (2009). The role of NOTCH1 signaling in T-ALL. <i>Hematology</i> , 2009(1), 353-361. doi 10.1182/asheducation-2009.1.353	https://www.ncbi.nlm.nih.gov/pmc/articles/PMC2847371/

PI3K α	PIK3CA	Khan, K. H., Yap, T. A., Yan, L., & Cunningham, D. (2013). Targeting the PI3K-AKT-mTOR signaling network in cancer. <i>Chinese Journal of Cancer</i> , 32(5), 253-265. doi:10.5732/cjc.013.10057	https://www.ncbi.nlm.nih.gov/pmc/articles/PMC3845556/	
PPM1D (WIP1)	PPM1D (WIP1)	Milosevic, J., Eissler, N., Treis, D., Wickström, M., Fransson, S., Sveinbjornsson, B., . . . Kogner, P. (2017). Abstract 1945 PPM1D/Wip1, promising new target in childhood cancers neuroblastoma and medulloblastoma. <i>Cancer Research</i> , 77(13 Supplement), 1945-1945. doi:10.1158/1538-7445.am2017-1945	http://cancerres.aacrjournals.org/content/77/13/Supplement/1945	
RAS	RAS	Ward, A. F., Braun, B. S., & Shannon, K. M. (2012). Targeting oncogenic Ras signaling in hematologic malignancies. <i>Blood</i> , 120(17), 3397-3406. doi:10.1182/blood-2012-05-378596	https://www.ncbi.nlm.nih.gov/pmc/articles/PMC3309527/	
RET	RET	Levy, A. S., Roh, M., Patterson, N., Scott, E., Quispe-Tintaya, W., Ewart, M. R., . . . Montagna, C. (2016). Abstract 15 Target next sequencing profiling of pediatric solid tumors Potential use for the identification of actionable mutations. <i>Clinical Cancer Research</i> , 22(1 Supplement), 15-15. doi:10.1158/1557-3265.pmsclingen15-15	http://clincancerres.aacrjournals.org/content/22/1/Supplement/15	Dupain, C., Harttrampf, A. C., Urbinati, G., Geogerger, B., & Massaad-Massade, L. (2017). Relevance of Fusion Genes in Pediatric Cancers Toward Precision Medicine. <i>Molecular Therapy - Nucleic Acids</i> , 6, 315-326. doi:10.1016/j.omtn.2017.01.005
SH2B3	SH2B3	Perez-Garcia, A., Ambesi-Impiomato, A., Hadler, M., Rigo, L., LeDuc, C. A., Kelly, K., . . . Ferrando, A. A. (2013). Genetic loss of SH2B3 in acute lymphoblastic leukemia. <i>Blood</i> , 122(14), 2425-2432. http://doi.org/10.1182/blood-2013-05-500850	https://www.ncbi.nlm.nih.gov/pmc/articles/PMC3790510/	
SHP2	SHP2	Liu, X., Zheng, H., Li, X., Wang, S., Meyerson, H. J., Yang, W., . . . Qu, C.-K. (2016). Gain-of-function mutations of Ptpn11 (Shp2) cause aberrant mitosis and increase susceptibility to DNA damage-induced malignancies. <i>Proceedings of the National Academy of Sciences of the United States of America</i> , 113(4), 984-989. http://doi.org/10.1073/pnas.1508535113	https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4743778/	
Smoothed	PATCH1, SMO	Rimkus, T., Carpenter, R., Qasem, S., Chan, M., & Lo, H. (2016). Targeting the Sonic Hedgehog Signaling Pathway Review of Smoothed and GLI Inhibitors. <i>Cancers</i> , 8(2), 22. doi:10.3390/cancers8020022	http://www.mdpi.com/2072-6694/8/2/22	
SYT-SSX	SYT-SSX	Stegmaier, S., Leuschner, I., Poremba, C., Ladenstein, R., Kazanowska, B., Ljungman, G., . . . Koscielniak, E. (2016). The prognostic impact of SYT-SSX fusion type and histological grade in pediatric patients with synovial sarcoma treated according to the CWS (Cooperative Weichteilsarkom Studie) trials. <i>Pediatric Blood & Cancer</i> , 64(1), 89-95. doi:10.1002/pbc.26306	https://www.ncbi.nlm.nih.gov/pubmed/27621063	
TERT	TERT	Reitman, Z. J., Pirozzi, C. J., & Yan, H. (2013). Promoting a new brain tumor mutation TERT promoter mutations in CNS tumors. <i>Acta Neuropathologica</i> , 126(6), 789-792. http://doi.org/10.1007/s00401-013-1207-5	https://www.ncbi.nlm.nih.gov/pmc/articles/PMC3888653/	
TORC1/2 as distinct from mTOR	TORC1/2	Foster, K. G., & Fingar, D. C. (2010). Mammalian Target of Rapamycin (mTOR) Conducting the Cellular Signaling Symphony. <i>Journal of Biological Chemistry</i> , 285(19), 14071-14077. doi:10.1074/jbc.r109.094003	http://www.jbc.org/content/285/19/14071/full	
TrkB	TrkB	Amatu, A., Sartore-Bianchi, A., & Siena, S. (2016). NTRK gene fusions as novel targets of cancer therapy across multiple tumour types. <i>ESMO Open</i> , 1(2), e000023. http://doi.org/10.1136/esmoopen-2015-000023	https://www.ncbi.nlm.nih.gov/pmc/articles/PMC5070277/	
TP53	TP53	Rausch, T., Jones, D., Zapatka, M., Stütz, A., Zichner, T., Weischenfeldt, J., . . . Korbel, J. (2012). Genome Sequencing of Pediatric Medulloblastoma Links Catastrophic DNA Rearrangements with TP53 Mutations. <i>Cell</i> , 148(1-2), 59-71. doi:10.1016/j.cell.2011.12.013	https://www.nature.com/articles/nature25480	
TYK2	TYK2	Waanders, E., Scheijen, B., Jongmans, M. C., Venselaar, H., Reijmersdal, S. V., Dijk, A. H., . . . Kuiper, R. P. (2016). Germline activating TYK2 mutations in pediatric patients with two primary acute lymphoblastic leukemia occurrences. <i>Leukemia</i> , 31(4), 821-828. doi:10.1038/leu.2016.277	https://www.nature.com/articles/leu2016277	

Target Symbol (Cell Lineage)	Citation(1)	Link(1)	Citation(2)	Link(2)	Citation(3)	Link(3)
AKR1C3	Liu C, Hsu Y, Pan P, Wu M, Ho C, Su L, ... Christiani D. C. (2008). Maternal and offspring genetic variants of AKR1C3 and the risk of childhood leukemia. <i>Carcinogenesis</i> 29(5): 98-99. doi:10.1093/carcin/bgn071	c/download/doi/10.1158/2963 https://doi.org/10.1158/2963				
BCOR	Chen X, Pappo A, & Dyer M.A. (2015). Pediatric solid tumor genomics and the developmental plan. <i>Oncogene</i> 34(1): 5207-5215. doi:10.1038/nco.2014.7	https://www.nature.com/articles/nco201447				
BTK	Uckan F. & D. (2013). No el Bruton's tyrosine kinase inhibitors current in de elopment. <i>Oncotargets and Therapy</i> 161. doi:10.21773/onco.33373	https://www.ncbi.nlm.nih.gov/pmc/articles/PMC3591036/				
CD7	Azad V.F., Ash A.A., Tashighi M., Mofrad N.N., Haghighi M., & Mehrzad A. (2015). CD7 aberrant expression led to a molecular switch at relapsed childhood acute pre-B lymphoblastic leukemia. <i>Medical Molecular Morphology</i> 9(1): 53-56. doi:10.1007/s00795-015-0117-0	https://doi.org/10.1007/s00795-015-0117-0				
CD19	Shalabi H., Angiolillo A., & Fry T.J. (2015). Beyond CD19: Open avenues for Future De elopment of Targeted Immunotherapy in Pediatric Relapsed/Refractory Acute Leukemia. <i>Frontiers in Pediatrics</i> 3: 80. doi:10.3389/fped.2015.00080	https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4589638/				
CD20	Dworak M., N. Schumich A., Prinz D., Pöschger U., Husak Z., Anbaruchi A., ... Gubler H. (2008). CD20 up-regulation in pediatric B-cell precursor acute lymphoblastic leukemia during induction treatment setting the stage for anti-CD20 directed immunotherapy. <i>Blood</i> 112(10): 3982-3988. doi:10.1182/blood-2008-06-16129	https://www.ncbi.nlm.nih.gov/pmc/articles/PMC2581996/				
CD22	Sun W., Gaynon P.S., Spoto R., & Wayne A.S. (2015). Improving Access To No of Agents For Childhood Leukemia. <i>Cancer</i> 121(12): 1927-1936. doi:10.1002/cncr.29267	https://www.ncbi.nlm.nih.gov/pmc/articles/PMC457590/				
CD30	Nagpal P., Aki M.R., Ayoub N.M., Tomiyama T., Coates T., Tai B., ... Suh K.S. (2016). Pediatric Hodgkin lymphoma - biomarkers drugs and clinical trials for translational science and medicine. <i>Oncotarget</i> 7(1): 67551-67573. doi:10.18632/oncotarget.11509	https://www.ncbi.nlm.nih.gov/pmc/articles/PMC5311896/				
CD33	O'Hear C., Heiber J.F., Schubert I., Fey G., & Geiger T.L. (2015). Anti-CD33 chimeric antigen receptor targeting of acute myeloid leukemia. <i>Haematologica</i> 100(3): 336-337. doi:10.3324/haematol.2014.11278	https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4319272/				
CD37	De Winde C.M., Veenbergen S., Young K.H., Xu-Monette Z.Y., Wang X., Xia Y., ... & Spriel A.B. (2016). Tetraspanin CD37 protects against the de elopment of B cell lymphoma. <i>The Journal of Clinical Investigation</i> 126(2): 653-666. doi:10.1172/JCI81011	https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4731177/				
CD38	Jiang Z., Wu D., Lin S., & Li P. (2016). CD38 and CD38 are prognostic biomarkers for acute B lymphoblastic leukemia. <i>Biomarker Research</i> 4: 23. doi:10.1186/s12874-016-0080-5	https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4915997/				
CD56	Aref S., Army E., El-Bakry K., Ibrahim L., & Mabed M. (2017). Prognostic impact of CD200 and CD56 expression in adult acute lymphoblastic leukemia patients. <i>Hematology</i> 1-8. doi:10.1080/10781332.2017.1301276	https://www.ncbi.nlm.nih.gov/pmc/articles/PMC5491828/	Neff J. & Chen D. (2017). Pediatric Philadelphia-positive B lymphoblastic leukemia with CD56 expression and L2 morphology. Case report and review of the literature. <i>Human Pathology Case Reports</i> 8: 9-12. doi:10.1016/j.hpcr.2016.12.002	https://www.sciencedirect.com/science/article/pii/S2211330016300801		
CD70	Shaffer D.R., Suda B., Wu Z., Chow K.H., Kakara S., Spencer D.M., ... Gotschalk S. (2011). T cells redirected against CD70 for the immunotherapy of CD70-positive malignancies. <i>Blood</i> 117(16): 30-31. doi:10.1182/blood-2010-04-278218	https://www.ncbi.nlm.nih.gov/pmc/articles/PMC3087180/				
CD79b	Gordon M.S., Kato R.M., Lansigan F., Thompson A.A., Wall R., & Rawlings D.J. (2000). Aberrant B cell receptor signaling from CD79b (CD79b) gene mutations of chronic lymphocytic leukemia B cells. <i>Proceedings of the National Academy of Sciences of the United States of America</i> 97(10): 550-5509.	https://www.ncbi.nlm.nih.gov/pmc/articles/PMC258585/				
CD123/IL3RA	Testa U., Pelosi E., & Frankel A. (2011). CD123 is a membrane biomarker and a therapeutic target in hematologic malignancies. <i>Biomarker Research</i> 2: 1. doi:10.1186/2050-7717-2-1	https://www.ncbi.nlm.nih.gov/pmc/articles/PMC3226610/	Bonifant C.L., Saver A., Torres D., Joseph N., Velasquez M.P., Iwabuchi K., ... Gotschalk S. (2016). CD123-Engager T Cells as a No of Immunotherapeutic for Acute Myeloid Leukemia. <i>Molecular Therapy</i> 29(9): 1615-1626. doi:10.1038/mt.2016.116	https://www.ncbi.nlm.nih.gov/pmc/articles/PMC5113997/		
CD276 (B7-1H3)	Zhou Z., Luther N., Ibrahim G.M., Hawkins C., Vishwakar R., Handler M.H., & Souweidane M.M. (2013). B7-1H3 a potential therapeutic target is expressed in diffuse intrinsic pontine glioma. <i>Journal of Neuro-Oncology</i> 111(3): 257-265. doi:10.1007/s11060-012-1021-2	https://www.ncbi.nlm.nih.gov/pmc/articles/PMC3700828/				
Cereblin CBL (E3 Ubiquitin protein ligase)	Morrow J.K., Lin H.-K., Sun S.-C., & Zhang S. (2015). Targeting ubiquitination for cancer therapies. <i>Future Medicinal Chemistry</i> 7(17): 2333-2350. doi:10.1515/fmc-2015-1518	https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4497684/				
DL1	Zhao X., Arca D.D., Lim W.K., Brnhamachary M., Carro M.S., Ludwig T., ... Lasonella A. (2009). The N-Myc-DLL3 cascade is suppressed by the ubiquitin ligase Hw1 to inhibit proliferation and cell motility in neuroblastoma. <i>Developmental Cell</i> 17(2): 210-221. doi:10.1016/j.devcel.2009.07.009	https://www.ncbi.nlm.nih.gov/pmc/articles/PMC2769070/	Saunders L.R., Rankoich A.J., Anderson W.C., Anuj M.A., Bheekah S., Black K., ... Dyba S.J. (2015). A DLL3-targeted antibody-drug conjugate eradicates high-grade pulmonary neuroendocrine tumor-initiating cells in mice. <i>Science Translational Medicine</i> 7(302): 302ra36. doi:10.1126/scitranslmed.309599	https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4943437/	Rudin C.M., Petanz M.C., Bauer T.M., Realy N., Mogensen D., Gleason B.S., ... Spigel D.R. (2017). Rovalpituzumab, a DLL3-targeted antibody-drug conjugate, in recurrent small-cell lung cancer: A first-in-human first-in-class 1L open-label phase 1 study. <i>The Lancet Oncology</i> 18(1): 2-51. doi:10.1016/S1473-0165(16)30565-4	https://www.thelancet.com/jou/soils/annex1/a/PIIS1473-0165(16)30565-4/fulltext
DLK1	Felix F.A., Aronson D.C., Lamers W.H., & Gaemers I.C. (2012). Possible roles of DLK1 in the Notch pathway during de elopment and disease. <i>Biochimica et Biophysica Acta (BBA) - Molecular Basis of Disease</i> 1822(6): 988-995. doi:10.1016/j.bbadis.2012.02.003	https://www.sciencedirect.com/science/article/pii/S092554331200312				
EGFR III	Li G., Mitra S.S., Monge M., Henrich K.N., Bangs C.D., Nitta K.T., & Wong A.J. (2012). Expression of epidermal growth factor variant III (EGFR-III) in pediatric diffuse intrinsic pontine gliomas. <i>Journal of Neuro-Oncology</i> 108(3): 395-402. doi:10.1007/s11060-012-0428-2	https://www.ncbi.nlm.nih.gov/pmc/articles/PMC3480992/				
EPHA2	Chow K.K., Naik S., Kakara S., Barlow V.S., Shaffer D.R., Yi Z., ... Gotschalk S. (2013). T Cells Redirected to EphA2 for the Immunotherapy of Glioblastoma. <i>Molecular Therapy</i> 21(3): 629-637. doi:10.1038/mt.2012.210	https://www.ncbi.nlm.nih.gov/pmc/articles/PMC389170/				
GD2	Cepelin C.M., Ono M., DeSmet K.B., & Sondel P.M. (2011). Immunotherapy in pediatric malignancies: current status and future perspectives. <i>Future Oncology (London, England)</i> 07(9): 1659-1678. doi:10.1080/17447011.2011.61252	https://www.ncbi.nlm.nih.gov/pmc/articles/PMC3179370/				
GPC2	Bosse K.R., Raman P., Zhu Z., Lane M., Martinez D., Heitzmeider S., ... Maris J.M. (2017). Identification of GPC2 as an Oncoprotein and Candidate Immunotherapeutic Target in High-Risk Neuroblastoma. <i>Cancer Cell</i> 32(3). doi:10.1016/j.ccr.2017.08.003	https://www.cell.com/cancer-cell/abstract/S1535-6108(17)30146X				
GPC3	Tanaka S., Soutaki R., Miyoshi K., Kohashi K., Oda Y., Nakamura T., ... Kinoshita Y. (2011). Glypican 3 Expression in Pediatric Malignant Solid Tumors. <i>European Journal of Pediatric Surgery</i> 25(01): 138-141. doi:10.1055/s-003-1393961	https://www.ncbi.nlm.nih.gov/pmc/articles/PMC2931910/				
GPNMB	Roth M., Barris D.M., Pardi S., Kuo V., Erens S., Geller D., ... Grottel T. (2015). Targeting Glypican 3 with Antibody-Drug Conjugates: Glioma Immunohistochemical and Preclinical Studies. <i>Pediatric Blood & Cancer</i> 63(1): 32-38. doi:10.1002/pbc.25688	https://www.ncbi.nlm.nih.gov/pmc/articles/PMC42630518/				
HERB2 (HER2/Neu)	Orenta R.J., Lee D.W., & Mackall C. (2012). Immunotherapy Targets in Pediatric Cancer. <i>Frontiers in Oncology</i> 2: 3. doi:10.3389/fonc.2012.00003	https://www.ncbi.nlm.nih.gov/pmc/articles/PMC315558/	Gibberton R.J. (2005). ERB2 in Pediatric Cancer: Immortin Pro on Gality. <i>The Oncologist</i> 10(7): 508-517. doi:10.1634/theoncologist.10-7-508	https://theoncologist.alphamedpress.org/content/10/7/508.full		
IL6	Egler R.A., Burlingame S.M., Nuchtern J.G., & Russell H.V. (2008). Interleukin-6 and soluble IL-6 receptor are markers of disease extent and prognosis in neuroblastoma. <i>Clinical Cancer Research: An Official Journal of the American Association for Cancer Research</i> 14(12): 7028-7033. doi:10.1158/1078-0432.CCR.07-5017	https://www.ncbi.nlm.nih.gov/pmc/articles/PMC2613272/				
IL13RA2	Deng H., Zeng J., Zhang T., Gong L., Zhang H., Cheng E., ... Li G. (2018). Histone H3.3K27M Mobilizes Multiple Cancer/Transit (CT) Antigens in Pediatric Glioma. <i>Molecular Cancer Research</i> 16(1): 623-633. doi:10.1158/1538-7465.2017-060	https://mcr.aacrjournals.org/content/16/1/623.full.pdf				
LRRC15	Reynolds P.A., Smolken G.A., Palmer R.E., Sgani D., Yajnik V., Gerald W.L., & Haber D.A. (2003). Identification of a DNA-binding site and transcriptional target for the EWS-WT1 (KTS) oncoprotein. <i>Genes & De elopment</i> 17(17): 209-2107. doi:10.1101/gad.111073	https://www.ncbi.nlm.nih.gov/pmc/articles/PMC196192/				
MAGE-A3	Jacobs J.F., Bissener F., Kain C.A., Raki M.W., Figlar C.G., Adema G.J., ... Vries I.J. (2006). Cancer germline gene expression in pediatric solid tumors using quantitative real-time PCR. <i>International Journal of Cancer</i> 120(1): 67-71. doi:10.1002/ijc.22118	https://onlinelibrary.wiley.com/doi/10.1002/ijc.22118				
MSLN (mesothelin)	Steinbach D. (2006). Identification of a Set of Serine Genes for the Monitoring of Minimal Residual Disease in Pediatric Acute Myeloid Leukemia. <i>Clinical Cancer Research</i> 12(8): 23-32. doi:10.1158/1078-0432.ccr-05-2552	https://www.ncbi.nlm.nih.gov/pmc/articles/PMC146192/				

NR5A1 (Steroidogenic factor 1)	<p>Ferraz-de-Souza B, Lin L, & Achemann J.C. (2011). Steroidogenic factor-1 (SF-1/NR5A1) and human disease. <i>Molecular and Cellular Endocrinology</i> 336(1-2): 198-205. http://doi.org/10.1016/j.mce.2010.11.006</p> <p>Singh N, Kuliko skaya I, Barrett D.M, Bieder-Scholl G, Jakobsen B, Martinez D, ... Grupp S.A. (2016). T cells targeting NY-ESO-1 demonstrate efficacy against disseminated neuroblastoma. <i>Oncotimmunology</i> 5(1): e10 0216. http://doi.org/10.1080/2162 02X.2015.10 0216</p>	<p>https://www.ncbi.nlm.nih.gov/pmc/articles/PMC3057017/</p>
NY-ESO-1	<p>Kapp R, Shayer L, Ten A-C, Szabo E, Sami N, Rowitch D.H, & Mehta S. (2016). Lineage-restricted OLG2-RTK signaling governs the molecular subtype of glioma stem-like cells. <i>Cell Reports</i> 16(11): 2838-28 5. http://doi.org/10.1016/j.celrep.2016.08.0 0</p>	<p>https://www.ncbi.nlm.nih.gov/pmc/articles/PMC492 710/</p>
Olig2	<p>Khan K.H, Yap T.A, Yan L, & Cunningham D. (2013). Targeting the PI3K-AKT-mTOR signaling network in cancer. <i>Chinese Journal of Cancer</i> 32(5): 253-265. doi: 10.5732/cjcc.013.10057</p>	<p>https://www.ncbi.nlm.nih.gov/pmc/articles/PMC3639 8/</p>
PIK3CD (PI3 kinase delta)	<p>Oberhaer A. (200). The Tumor-Associated Antigen PRAME Is Universally Expressed in High-Stage Neuroblastoma and Associated with Poor Outcome. <i>Clinical Cancer Research</i> 10(13): 307- 313. doi: 10.1158/1078-0 32.ccr-03-0813</p>	<p>https://www.ncbi.nlm.nih.gov/pubmed/152 0616</p>
PRAME	<p>Cain C. (2012). SYK inhibitors on retinoblastoma. <i>Science-Business EXchange</i> 5(7). doi: 10.1038/scsb.2012.168</p>	<p>https://www.nature.com/scsb/exchange/5/7/full/scsb.2012.168.html</p>
SYK	<p>Noronha S, A. Farrar J.E, Alonzo T.A, Gerbing R.B, Lacayo N.J, Dahl G.V, ... Loeb D.M. (2009). WT1 expression at diagnosis does not predict survival in pediatric AML: A report from the Children's Oncology Group. <i>Pediatric Blood & Cancer</i> 53(6): 1136-1139. http://doi.org/10.1002/pbc.221 2</p>	<p>https://www.ncbi.nlm.nih.gov/pmc/articles/PMC2926132/</p>
WT1	<p>Abdouch, R.H, Bueno, A.C, Leal, L.F., Cavalcanti, M.M., Gomes, D.C, Bandalise, S.R., ... Antonin, S.R. (2016). Unveiling the expression of the oncogene YAP1, a Wnt/beta-catenin target, in adult neuroblastoma and its association with poor outcome in pediatric patients. <i>Oncotarget</i>, 7(51), 84634-84644. http://doi.org/10.18632/oncotarget.12382</p>	<p>https://www.ncbi.nlm.nih.gov/pmc/articles/PMC456687/</p>
YAP1		

Target Symbol (Tumor MicroENVT&ImmunoTherapy)	Citation(1)	Link(1)	Citation(2)	Link(2)	Citation(3)	Link(3)	Comments
	Theruvath, J., Heitzinger, S., Majner, R., Cui, K., Nallas, A., Graf, C. M., ... Mina, S. S. (2017). Intra-45 Checkpoint Molecule B7-H3 Is Highly Expressed On Medulloblastoma And Proves To Be A Promising Candidate For Car T Cell Immunotherapy. <i>Neuro-Oncology</i> , 19(Suppl_6), vi122-vi122. doi:10.1093/neuonc/nwz168.503	https://academic.oup.com/neuro-oncology/article/abstract/19/suppl_6/vi122/4590631?redirectedFrom=abstract					
B7H3	Petrov, I., Sutssova, M., Matorova, O., et al. Molecular pathway activation features of pediatric acute myeloid leukemia (AML) and acute lymphoblast leukemia (ALL) cells. <i>Ageing (Abing NY)</i> , 2016;8(11):2936-2946. doi:10.18632/aging.101102.	https://www.ncbi.nlm.nih.gov/pmc/articles/PMC5152073/	Vonderheide, R. H. (2007). Prospect of Targeting the CD40 Pathway for Cancer Therapy. <i>Clinical Cancer Research</i> , 13(6), 1083-1088. doi:10.1158/1078-0432.ccr-06-1893			http://clincancerres.aacrjournals.org/content/13/6/1083	
CD40	An Anti-CD47 Antibody Is Effective in Pediatric Brain Tumor Models. (2017). <i>Cancer Discovery</i> , 7(5). doi:10.1158/2159-8290.cd-rw2017-057	http://cancerdiscovery.aacrjournals.org/content/7/5/453.2.full-text.pdf					
CD47	Angiolillo, A. L., Yu, A. L., Reaman, G., Ingle, A. M., Secola, R., & Adamson, P. C. (2009). A Phase II Study of Campath-1H in Children with Relapsed or Refractory Acute Lymphoblastic Leukemia: A Children's Oncology Group Report. <i>Pediatric Blood & Cancer</i> , 53(6), 978-983. http://doi.org/10.1002/pbc.22209	https://www.ncbi.nlm.nih.gov/pmc/articles/PMC3129889/					
CD52	Matsuo, H., Nakamura, N., Tomizawa, D., Saito, A. M., Kiyokawa, N., Horibe, K., ... Adachi, S. (2016). CXCR4 Overexpression is a Poor Prognostic Factor in Pediatric Acute Myeloid Leukemia With Low Risk: A Report From the Japanese Pediatric Leukemia Lymphoma Study Group. <i>Pediatric Blood & Cancer</i> , 63(8), 1394-1399. doi:10.1002/pbc.26035	https://www.ncbi.nlm.nih.gov/pubmed/27135782					
CXCR4	Lumardi, S., Lim, S. Y., Muschel, R. J., & Brunner, T. B. (2015). IP-10/CXCL10 attracts regulatory T cells: Implication for pancreatic cancer. <i>Oncotarget</i> , 4(9). doi:10.1080/2162402.2015.1027473	https://www.tandfonline.com/doi/full/10.1080/2162402.2015.1027473					
CXCR10	Merchant, M. S., Wright, M., Baird, K., Weider, L. H., Rodriguez-Gallardo, C., Bernstein, D., ... Mackall, C. L. (2016). Phase 1 Clinical Trial of Iplimimab In Pediatric Patients With Advanced Solid Tumors. <i>Clinical Cancer Research: An Official Journal of the American Association for Cancer Research</i> , 22(6), 1364-1370. http://doi.org/10.1158/1078-0432.CCR-15-0491	https://www.ncbi.nlm.nih.gov/pmc/articles/PMC5027963/					
CTLA4	Alper, A. M., Frieden-Korovina, V. P., Buzdin, A., Roumiantsev, S. A., & Zhavoronkov, A. (2014). A role for G-CSF and GM-CSF in nonmyeloid cancers. <i>Cancer Medicine</i> , 3(4), 737-746. http://doi.org/10.1002/cam4.239	https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4303143/					
GM-CSF	Folgiere, V., Goffredo, B. M., Fippini, P., Masetti, R., Bonanno, G., Caruso, R., ... Ruella, S. (2013). Indoleamine 2,3-dioxygenase 1 (IDO1) activity in leukemia blasts correlates with poor outcome in childhood acute myeloid leukemia. <i>Oncotarget</i> , 5(8). doi:10.18632/oncotarget.1504	https://review.ncbi.nlm.nih.gov/pmc/articles/PMC4050144/					
IDO1	Reid GSD, Shan X, Coughlin CM, et al. Interferon-gamma dependent infiltration of human T cells into neuroblastoma tumors in vivo. <i>Clinical cancer research: an official journal of the American Association for Cancer Research</i> . 2009;15(21):6602-6608. doi:10.1158/1078-0432.CCR-09-0829	https://www.ncbi.nlm.nih.gov/pmc/articles/PMC2783677/					
IFN-gamma	Capitini CM, Mackall CL, Wayne AS. Immune-based Therapeutics for Pediatric Cancer. <i>Expert opinion on biological therapy</i> . 2010;10(2):163-178. doi:10.1517/14712590903431022	https://www.ncbi.nlm.nih.gov/pmc/articles/PMC2809045/					
IL-2	Birley, K., Chester, K., & Anderson, J. (2018). Ant body based therapy for childhood solid cancers. <i>Current Opinion in Chemical Engineering</i> , 19, 153-162. doi:10.1016/j.coche.2018.01.005	https://www.sciencedirect.com/science/article/pii/S2211339817300603					
LAG3	Reu, E. D., Staegs, M. S., Kithanol, C. D., & Föll, J. (2015). Immunostimulation by OX40 Ligand Transgenic Ewing Sarcoma Cells. <i>Frontiers in Oncology</i> , 5, 242. http://doi.org/10.3389/fonc.2015.00242	https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4621427/					
OX40	Chen, L., & Han, X. (2015). Anti-PD-1/PD-L1 therapy of human cancer: past, present, and future. <i>The Journal of Clinical Investigation</i> , 125(9), 3384-3391. http://doi.org/10.1172/JCI89011	https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4588282/	Alison M. Martin et al. PD-L1 expression in medulloblastoma: an evaluation by subgroup. <i>Oncotarget</i> (2018).	http://www.oncotarget.com/index.php?journal=oncotarget&page=fulltext&ogview=&path[]=24951&path[]=72227			Low level PD-L1 expression in small cohort of medulloblastoma. Clinical significance not clear.
PD-1/PD-L1	Dupuis, C., Hartmann, A. C., Urbain, G., Georger, B., & Massaad-Massad, L. (2017). Relevance of Fusion Genes in Pediatric Cancers: Toward Precision Medicine. <i>Molecular Therapy. Nucleic Acids</i> , 6, 315-326. http://doi.org/10.1016/j.omtn.2017.01.005	https://www.ncbi.nlm.nih.gov/pmc/articles/PMC63514/					
RELA	Kaneda, Y. (2013). The RIG-1/MAVS signaling pathway in cancer cell-selective apoptosis. <i>Oncotarget</i> , 2(4), e23566. http://doi.org/10.4161/onc.23566	https://www.ncbi.nlm.nih.gov/pubmed/23743213					
RIG-1	Grunewald, T. G., Diebold, I., Esposi, O., Plehm, S., Hauer, K., Thiel, U., ... Burdach, S. (2011). STEAP1 Is Associated with the Invasive and Oxidative Stress Phenotype of Ewing Tumors. <i>Molecular Cancer Research</i> , 10(1), 52-65. doi:10.1158/1541-7786.mcr-11-0524	https://www.ncbi.nlm.nih.gov/pubmed/22080479					
STEAP1	Lemos, H., Mohamed, E., Huang, L., Ou, R., Pacholczyk, G., Arbab, A. S., ... Mellor, A. L. (2016). STING promotes the growth of tumors characterized by low antigenicity via IDO activation. <i>Cancer Research</i> , 76(8), 2076-2081. http://doi.org/10.1158/0008-5472.CAN-15-1456	https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4873329/					
STING	Williams, K. M., Grant, M., Isma, I. M., Hoq, F., Martin-Masso, M., Hoovec, J., ... Bolland, C. (2017). Complete remissions post infusion of multiple tumor antigen specific T cells for the treatment of high risk leukemia and lymphoma patients after HCT. <i>Cytophera</i> , 19(5). doi:10.1016/j.jcyt.2017.03.013	https://online.liebertpub.com/doi/full/10.1002/cyt.26772					
TIM3/TIM4	Gilde Bender, J., Yamashiro, D. J., & Fox, E. (2011). Clinical Development of VEGF Signaling Pathway Inhibitors in Childhood Solid Tumors. <i>The Oncologist</i> , 16(11), 1614-1625. http://doi.org/10.1634/theoncologist.2011-0148	https://www.ncbi.nlm.nih.gov/pmc/articles/PMC3233297/					
VEGF and VEGFR							

Target Symbol (Others)	Citation(1)	Link(1)	Citation(2)	Link(2)
AKT	Barrett, D., Brown, V. I., Grupp, S. A., & Teachey, D. T. (2012). Targeting the PI3K/AKT/mTOR Signaling Axis in Children with Hematologic Malignancies. <i>Pediatric Drugs</i> , 14(5), 299-316. doi 10.1007/bf03262236	https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4214862/		
ATM	Takagi, M., Yoshida, M., Nemoto, Y., Tamaichi, H., Tsuchida, R., Seki, M., ... Takita, J. (2017). Loss of DNA Damage Response in Neuroblastoma and Utility of a PARP Inhibitor. <i>JNCI Journal of the National Cancer Institute</i> , 109(11). doi 10.1093/jnci/djx062	https://academic.oup.com/jnci/article/109/11/djx062/4096548 https://www.ncbi.nlm.nih.gov/pmc/articles/PMC5066844/ [Erratum of the research article] https://www.sciencedirect.com		
ATR	Weber, A. M., & Ryan, A. J. (2015). ATM and ATR as therapeutic targets in cancer. <i>Pharmacology & Therapeutics</i> , 149, 124-138. doi 10.1016/j.pharmthera.2014.12.001	https://www.sciencedirect.com		
ATRX	Koschmann, C., Calinescu, A.-A., Nunez, F. J., Mackay, A., Fazal-Salom, J., Thomas, D., ... Castro, M. G. (2016). ATRX Loss Promotes Tumor Growth and Impairs Non-Homologous End Joining DNA Repair in Glioma. <i>Science Translational Medicine</i> , 8(35), 4078-4085.	https://www.ncbi.nlm.nih.gov/pmc/articles/PMC5381643/		
AURKA (Aurora kinase A)	Wetmore C, Boyett J, Li S, et al. Alisertib is active as single agent in recurrent atypical teratoid rhabdoid tumors in 4 children. <i>Neuro-Oncology</i> , 2015;17(6) 882-888. doi 10.1093/neuonc/nov017.	https://preview.ncbi.nlm.nih.gov/pmc/articles/PMC4483126/		
AURKB (Aurora kinase B)	Bavetsias, V., & Linardopoulos, S. (2015). Aurora Kinase Inhibitors: Current Status and Outlook. <i>Frontiers in Oncology</i> , 5, 278. http://doi.org/10.3389/fonc.2015.00278	https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4685048/		
AXL	Huey, M. G., Minson, K. A., Earp, H. S., DeRyckere, D., & Graham, D. K. (2016). Targeting the TAM Receptors in Leukemia. <i>Cancers</i> , 8(11), 101. http://doi.org/10.3390/cancers8110101	https://www.ncbi.nlm.nih.gov/pmc/articles/PMC5126761/		
A1/BFL	Huhn, A. J., Guerra, R. M., Harvey, E. P., Bird, G. H., & Walensky, L. D. (2016). Selective Covalent Targeting of Anti-Apoptotic BCL-1 by Cysteine-Reactive Stapled Peptide Inhibitors. <i>Cell Chemical Biology</i> , 23(9), 1123-1134. http://doi.org/10.1016/j.chembiol.2016.07.022	https://www.ncbi.nlm.nih.gov/pmc/articles/PMC5055752/		
BAK	Katz, S. G., Fisher, J. K., Correll, M., Bronson, R. T., Ligon, K. L., & Walensky, L. D. (2013). Brain and Testicular Tumors in Mice with Progenitor Cells Lacking BAX and BAK. <i>Oncogene</i> , 32(35), 4078-4085. http://doi.org/10.1038/ncr.2012.421	https://www.ncbi.nlm.nih.gov/pmc/articles/PMC3529761/	Westhoff, M., Marschall, N., Grunert, M., Karpel-Massler, G., Burdach, S., & Debatin, K. (2018). Cell death-based treatment of childhood cancer. <i>Cell Death & Disease</i> , 9(2). doi 10.1038/s41419-017-0062-z	https://www.nature.com/articles/s41419-017-0062-z
BAX	Kaparou, M., Choumerianou, D., Perdikiogianni, C., Martimianaki, G., Kalmanti, M., & Stiakaki, E. (2013). Enhanced levels of the apoptotic BAX/BCL-2 ratio in children with acute lymphoblastic leukemia and high-risk features. <i>Genetics and Molecular Biology</i> , 36(1), 7-11. http://doi.org/10.1590/S1415-47572013005000003	https://www.ncbi.nlm.nih.gov/pmc/articles/PMC3615527/		
BCL2 family members (Bcl-2, Bcl-XL, Mcl-1, A1/BFL, BAK, BAX)	Chaber, R., Fiszler-Maliszewska, L., Noworolska-Sauren, D., Kwasnicka, J., Wrobel, G., & Chybicka, A. (2013). The BCL-2 Protein in Precursor B Acute Lymphoblastic Leukemia in Children. <i>Journal of Pediatric Hematology/Oncology</i> , 35(3), 180-187. doi 10.1097/MPH.0b013e318286d29b	https://www.ncbi.nlm.nih.gov/pubmed/23511489		
BET bromodomain family	Wadhwa E, Nicolaidis T (May 21, 2016) Bromodomain Inhibitor Review Bromodomain and Extra-terminal Family Protein Inhibitors as a Potential New Therapy in Central Nervous System Tumors. <i>Cureus</i> 8(5) e620. http://doi.org/10.7759/cureus.620	https://www.ncbi.nlm.nih.gov/pubmed/27382528	Hensel, T., Giorgi, C., Schmidt, O., Calzadawack, J., Neff, F., Buch, T., ... Richter, G. H. (2015). Targeting the EWS-ETS transcriptional program by BET bromodomain inhibition in Ewing sarcoma. <i>Oncotarget</i> , 7(2). doi 10.18632/oncotarget.6385	https://mediatum.ub.tum.de/doc/1398856/1398856.pdf
BMPR	Liu, S., Tian, Z., Yin, F., Zhang, P., Wu, Y., Ding, X., ... Fan, M. (2009). Expression and Functional Roles of Smad1 and BMPR-1B in Glioma Development. <i>Cancer Investigation</i> , 27(7), 734-740. doi 10.1080/07357900802620786	https://www.semanticscholar.org/paper/Expression-and-functional-roles-of-Smad1-and-in-Liu-Tian/d4d4dea12c9e329a45478a76e18ab423c255215		
Brd1	Herz, H.-M., Morgan, M., Gao, X., Jackson, J., Rickels, R., Swanson, S. K., ... Shilatifard, A. (2014). Histone H3 lysine-to-methionine mutants as a paradigm to study chromatin signaling. <i>Science (New York, N.Y.)</i> , 345(6200), 1065-1070. http://doi.org/10.1126/science.1255104	https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4508193/	Long, W., Yi, Y., Chen, S., Cao, Q., Zhao, W., & Liu, Q. (2017). Potential New Therapies for Pediatric Diffuse Intrinsic Pontine Glioma. <i>Frontiers in Pharmacology</i> , 8. doi 10.3389/fphar.2017.00495	https://www.frontiersin.org/articles/10.3389/fphar.2017.00495/full
Brd4	Herz, H.-M., Morgan, M., Gao, X., Jackson, J., Rickels, R., Swanson, S. K., ... Shilatifard, A. (2014). Histone H3 lysine-to-methionine mutants as a paradigm to study chromatin signaling. <i>Science (New York, N.Y.)</i> , 345(6200), 1065-1070. http://doi.org/10.1126/science.1255104	https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4508193/		
CDK4/6	Hamilton, E., & Infante, J. R. (2016). Targeting CDK4/6 in patients with cancer. <i>Cancer Treatment Reviews</i> , 45, 129-138. doi 10.1016/j.ctrv.2016.03.002	https://www.deepdyve.com/lp/elsevier/targeting-cdk4-6-in-patients-with-cancer-bnyBj02		
CHK1	Prince, E. W., Balakrishnan, I., Shah, M., Mulcahy Levy, J. M., Griesinger, A. M., Alimova, I., ... Vibhakkar, R. (2016). Checkpoint kinase 1 expression is an adverse prognostic marker and therapeutic target in MYC-driven medulloblastoma. <i>Oncotarget</i> , 7(33), 53881-53894. http://doi.org/10.18632/oncotarget.10692	https://www.ncbi.nlm.nih.gov/pmc/articles/PMC5288228/	Lowery, C. D., Vanwye, A. B., Dowless, M., Blosser, W., Falcon, B. L., Stewart, J., ... Stancato, L. F. (2017). The Checkpoint Kinase 1 Inhibitor Praxasertib Induces Regression of Preclinical Models of Human Neuroblastoma. <i>Clinical Cancer Research</i> , 23(15), 4354-4363. doi 10.1158/1078-0432.ccr-16-2876	http://clincancerres.aacrjournals.org/content/early/2017/03/07/1078-0432.CCR-16-2876
CDK2	Chen, Z., Wang, Z., Pang, J. C., Yu, Y., Bieerkehazhi, S., Lu, J., ... Yang, J. (2016). Multiple CDK inhibitor dinaciclib suppresses neuroblastoma growth via inhibiting CDK2 and CDK9 activity. <i>Scientific Reports</i> , 6, 29090. http://doi.org/10.1038/srep29090	https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4932496/		
CDK7	Kwiatkowski, N., Zhang, T., Rahl, P. B., Abraham, B. J., Reddy, J., Ficarro, S. B., ... Gray, N. S. (2014). Targeting transcription regulation in cancer with a covalent CDK7 inhibitor. <i>Nature</i> , 511(7511), 616-620. http://doi.org/10.1038/nature13393	https://www.ncbi.nlm.nih.gov/pmc/articles/PMC424910/		
CDK9	Moreno, N., Holsten, T., Mertins, J., Zhoghi, A., Johann, P., Kool, M., ... Kerl, K. (2017). Combined BRD4 and CDK9 inhibition as a new therapeutic approach in malignant rhabdoid tumors. <i>Oncotarget</i> , 8(49), 84986-84995. http://doi.org/10.18632/oncotarget.18583	https://www.ncbi.nlm.nih.gov/pmc/articles/PMC5689588/		
CKI	Richter, J., Ullah, K., Xu, P., Alschner, V., Blatz, A., Peifer, C., ... Knippschild, U. (2014). Effects of altered expression and activity levels of CK1δ and ε on tumor growth and survival of colorectal cancer patients. <i>International Journal of Cancer</i> , 136(12), 2799-2810. doi 10.1002/ijc.29346	https://onlinelibrary.wiley.com/doi/10.1002/ijc.29346		

CK2 (casein kinase 2)	Chua, M. M. J., Ortega, C. E., Sheikh, A., Lee, M., Abdur-Rassoul, H., Hartshorn, K. L., & Dominguez, I. (2017). CK2 in Cancer: Cellular and Biochemical Mechanisms and Potential Therapeutic Target. <i>Pharmaceuticals</i> , 10(1), 18. https://doi.org/10.3390/ph10010018	Buontempo, F., Mccubrey, J. A., Orsini, E., Ruzzene, M., Cappellini, A., Lonetti, A., ... Martelli, A. M. (2017). Therapeutic targeting of CK2 in acute and chronic leukemias. <i>Leukemia</i> , 32(1), 1-10. doi 10.1038/leu.2017.301 https://www.nature.com/articles/leu2017301
CREBBP/EP300	Mullighan, C. G., Zhang, J., Kasper, L. H., Lerach, S., Payne-Turner, D., Phillips, L. A., ... Downing, J. R. (2011). CREBBP mutations in relapsed acute lymphoblastic leukaemia. <i>Nature</i> , 471(7337), 235-239. https://doi.org/10.1038/nature09727	
DNA (alkylators)	Bobola, M. S. (2005). O6-Methylguanine-DNA Methyltransferase, O6-Benzylguanine, and Resistance to Clinical Alkylators in Pediatric Primary Brain Tumor Cell Lines. <i>Clinical Cancer Research</i> , 11(7), 2747-2755. doi 10.1158/1078-0432.ccr-04-2045 https://www.ncbi.nlm.nih.gov/pubmed/15814657	
DNA-PK	Dolman, M. E. M., van der Ploeg, I., Koster, J., Bate-Eya, L. T., Versteeg, R., Caron, H. N., & Molenaar, J. J. (2015). DNA-Dependent Protein Kinase As Molecular Target for Radiosensitization of Neuroblastoma Cells. <i>PLoS ONE</i> , 10(12), e0145744. https://doi.org/10.1371/journal.pone.0145744	Becher, O. J., Peterson, K. M., Khatua, S., Santi, M. R., & MacDonald, T. J. (2008). IGF1BP2 is Overexpressed by Pediatric Malignant Astrocytomas and Induces the Repair Enzyme DNA-PK. <i>Journal of Child Neurology</i> , 23(10), 1205-1213. https://doi.org/10.1177/0883073808321766
DNMT (DNA methyl transferase)	Diede, S. J., Guenthoer, J., Geng, L. N., Mahoney, S. E., Marotta, M., Olson, J. M., ... Tapscott, S. J. (2009). DNA methylation of developmental genes in pediatric medulloblastomas identified by denaturation analysis of methylation differences. <i>Proceedings of the National Academy of Sciences</i> , 107(1), 234-239. doi 10.1073/pnas.0907606106 http://www.pnas.org/content/pnas/early/2009/11/30/0907606106.full.pdf	
FAK	Waters, A. M., Stafan, L. L., Garner, E. F., Mruthyunjappa, S., Stewart, J. E., Mroczek-Musulman, E., & Beierle, E. A. (2016). Targeting Focal Adhesion Kinase Suppresses the Malignant Phenotype in Rhabdomyosarcoma Cells. <i>Translational Oncology</i> , 9(4), 263-273. https://doi.org/10.1016/j.tranon.2016.06.001	
FOLR1 (folate receptor 1)	Orentas, R. J., Lee, D. W., & Mackall, C. (2012). Immunotherapy Targets in Pediatric Cancer. <i>Frontiers in Oncology</i> , 2, 3. https://doi.org/10.3389/fonc.2012.00003	
GSK-3	Hu, Y., Gu, X., Li, R., Luo, Q., & Xu, Y. (2010). Glycogen synthase kinase-3 β inhibition induces nuclear factor- κ B-mediated apoptosis in pediatric acute lymphocyte leukemia cells. <i>Journal of Experimental & Clinical Cancer Research</i> CR, 29(1), 154. http://doi.org/10.1186/1756-9966-29-154	Mills, C. N., Nowsheen, S., Bonner, J. A., & Yang, E. S. (2011). Emerging Roles of Glycogen Synthase Kinase 3 in the Treatment of Brain Tumors. <i>Frontiers in Molecular Neuroscience</i> , 4, 47. http://doi.org/10.3389/fnmol.2011.00047
HDAC	West, A. C., & Johnstone, R. W. (2014). New and emerging HDAC inhibitors for cancer treatment. <i>The Journal of Clinical Investigation</i> , 124(1), 30-39. https://doi.org/10.1172/JCI69738	
HIF1A	Cruzeiro, G. A., Reis, M. B., Silveira, V. S., Lira, R. C., Jr, C. G., Neder, L., ... Valera, E. T. (2018). HIF1A is Overexpressed in Medulloblastoma and its Inhibition Reduces Proliferation and Increases EPAS1 and ATG16L1 Methylation. <i>Current Cancer Drug Targets</i> , 18(3), 287-294. doi 10.2174/1568009617666170315162525 https://www.ncbi.nlm.nih.gov/pubmed/28302031/	
Hippo pathway (YAP, TAZ, TEADs)	Ahmed, A. A., Mohamed, A. D., Gener, M., Li, W., & Taboada, E. (2017). YAP and the Hippo pathway in pediatric cancer. <i>Molecular & Cellular Oncology</i> , 4(3), e1295127. https://doi.org/10.1080/23723556.2017.1295127	
Hsp90	Li, W., Tsen, F., Sahu, D., Bhatia, A., Chen, M., Multhoff, G., & Woodley, D. T. (2013). Extracellular Hsp90 (eHsp90) as the Actual Target in Clinical Trials: Intentionally or Unintentionally. <i>International Review of Cell and Molecular Biology</i> , 303, 203-235. https://doi.org/10.1016/B978-0-12-407697-6.00005-2	
IAPs (inhibitor-of-apoptosis)	Tyner, J. W., Jemal, A. M., Thayer, M., Druker, B. J., & Chang, B. H. (2012). Targeting survivin and p53 in pediatric acute lymphoblastic leukemia. <i>Leukemia</i> , 26(4), 623-632. https://doi.org/10.1038/leu.2011.249	
IGFR-1	BADR, M., HASSAN, T., TARHONY, S. E., & METWALLY, W. (2010). Insulin-like growth factor-1 and childhood cancer risk. <i>Oncology Letters</i> , 1(6), 1055-1059. https://doi.org/10.3892/ol.2010.169	
KDM4A	D Oto, A., Tian, Q., Davidoff, A. M., & Yang, J. (2016). Histone demethylases and their roles in cancer epigenetics. <i>Journal of Medical Oncology and Therapeutics</i> , 1(2), 34-40. https://www.ncbi.nlm.nih.gov/pubmed/279889/	
LSD1	Theisen, E. R., Pishas, K. I., Saund, R. S., & Lessnick, S. L. (2016). Therapeutic opportunities in Ewing sarcoma EWS-FLI1 inhibition via LSD1 targeting. <i>Oncotarget</i> , 7(14), 17616-17630. https://doi.org/10.18632/oncotarget.7124	
MCL1	Luedtke, D. A., Niu, X., Pan, Y., Zhao, J., Liu, S., Edwards, H., ... Ge, Y. (2017). Inhibition of Mcl-1 enhances cell death induced by the Bcl-2-selective inhibitor ABT-199 in acute myeloid leukemia cells. <i>Signal Transduction and Targeted Therapy</i> , 2, 17012. doi 10.1038/sigtrans.2017.12 https://www.nature.com/articles/sigtrans201712	
MCT1 (monocarboxyate transporter 1)	Noble, R. A., Bell, N., Blair, H., Sikka, A., Thomas, H., Phillips, N., ... Wedge, S. R. (2017). Inhibition of monocarboxyate transporter 1 by AZD3965 as a novel therapeutic approach for diffuse large B-cell lymphoma and Burkitt lymphoma. <i>Haematologica</i> , 102(7), 1247-1257. https://doi.org/10.3324/haematol.2016.163030	
MEK	Dupain, C., Harttrampf, A. C., Urbinati, G., Geogger, B., & Massaad-Massade, L. (2017). Relevance of Fusion Genes in Pediatric Cancers: Toward Precision Medicine. <i>Molecular Therapy. Nucleic Acids</i> , 6, 315-326. https://doi.org/10.1016/j.omtn.2017.01.005	
MIZ1	Vo, B. T., Wolf, E., Kawachi, D., Gebhardt, A., Rehg, J. E., Finkelstein, D., ... Roussel, M. F. (2016). The interaction of Mye with Miz1 defines medulloblastoma subgroup identity. <i>Cancer Cell</i> , 29(1), 5-16. https://doi.org/10.1016/j.ccr.2015.12.003	

MGMT	Saletta, F., Wadham, C., Ziegler, D. S., Marshall, G. M., Haber, M., Mccowage, G., . . . Byrne, J. A. (2014). Molecular profiling of childhood cancer Biomarkers and novel therapies. <i>BBA Clinical</i> , 1, 59-77. doi 10.1016/j.bbaci.2014.06.003	https://www.sciencedirect.com/science/article/pii/S2214647414000105	
MLL5	Gallo, M., Coutinho, F. J., Vanner, R. J., Gayden, T., Mack, S. C., Murison, A., . . . Dirks, P. B. (2015). MLL5 Orchestrates a Cancer Self-Renewal State by Repressing the Histone Variant H3.3 and Globally Reorganizing Chromatin. <i>Cancer Cell</i> , 28(6), 715-729. doi 10.1016/j.ccr.2015.10.005	https://www.cell.com/cancer-cell/abstract/S1535-6108(15)00382-7	
MYST3 (MYST histone acetyltransferase (monocytic leukemia))	Andrade, F. G., Noronha, E. P., Baseggio, R. M., Fonseca, T. C. C., Freire, B. M. R., Quezado Magalhaes, I. M., . . . Pombode-Oliveira, M. S. (2016). Identification of the MYST3-CREBBP fusion gene in infants with acute myeloid leukemia and hemophagocytosis. <i>Revista Brasileira de Hematologia E Hemoterapia</i> , 38(4), 291-297. http://doi.org/10.1016/j.bjh.2016.06.005	https://www.ncbi.nlm.nih.gov/pmc/articles/PMC5119666/	
NAMPT	Heske, C. M., Davis, M. I., Baumgart, J. T., Wilson, K., Gormally, M. V., Chen, L., . . . Thomas, C. J. (2017). Matrix Screen Identifies Synergistic Combination of PARP Inhibitors and Nicotinamide Phosphoribosyltransferase (NAMPT) Inhibitors in Ewing Sarcoma. <i>Clinical Cancer Research</i> , 23(23), 7301-7311. doi 10.1158/1078-0432.ccr-17-1121	https://www.ncbi.nlm.nih.gov/pubmed/28899971	
NEDD8 activating enzyme (NAE)	Bhatia, S., Pavlick, A. C., Boasberg, P., Thompson, J. A., Mulligan, G., Pickard, M. D., . . . Hamid, O. (2016). A phase I study of the investigational NEDD8-activating enzyme inhibitor pevonedistat (TAK-924/MLN4924) in patients with metastatic melanoma. <i>Investigational New Drugs</i> , 34, 439-449. http://doi.org/10.1007/s10637-016-0348-5	https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4919369/	Mansouri, S., & Zadeh, G. (2015). Neddylation in glioblastomas. <i>Neuro-Oncology</i> , 17(10), 1305-1306. http://doi.org/10.1093/neuonc/nov165
PARP	Ricks, T. K., Chiu, H.-J., Ison, G., Kim, G., McKee, A. E., Kluetz, P., & Pazdur, R. (2015). Successes and Challenges of PARP Inhibitors in Cancer Therapy. <i>Frontiers in Oncology</i> , 5, 222. http://doi.org/10.3389/fonc.2015.00222	https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4604313/	
PDK-1 (3-phosphoinositide-dependent protein kinase 1)	Velpula, K. K., Guda, M. R., Sahu, K., Tuszynski, J., Asuthkar, S., Bach, S. E., . . . Tsung, A. J. (2017). Metabolic targeting of EGFRvIII/PDK1 axis in temozolomide resistant glioblastoma. <i>Oncotarget</i> , 8(22), 35639-35655. http://doi.org/10.18632/oncotarget.16767	https://www.ncbi.nlm.nih.gov/pmc/articles/PMC5482605/	
PI3Kdelta	Smith, M. A., & Reaman, G. H. (2015). Remaining challenges in childhood cancer and newer targeted therapeutics. <i>Pediatric Clinics of North America</i> , 62(1), 301-312. http://doi.org/10.1016/j.pcl.2014.09.018	https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4336187/	
PIMI	Padi, S. K. R., Luevano, L. A., An, N., Pandey, R., Singh, N., Song, J. H., . . . Kraft, A. S. (2017). Targeting the PIM protein kinases for the treatment of a T-cell acute lymphoblastic leukemia subset. <i>Oncotarget</i> , 8(18), 30199-30216. http://doi.org/10.18632/oncotarget.16320	https://www.ncbi.nlm.nih.gov/pmc/articles/PMC5444737/	
PKA	Huang, S. Y., & Yang, J.-Y. (2015). Targeting the Hedgehog Pathway in Pediatric Medulloblastoma. <i>Cancers</i> , 7(4), 2110-2123. http://doi.org/10.3390/cancers7040880	https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4695880/	
PKC	Kikuchi, K., Soundararajan, A., Zarzabal, L. A., Weems, C. R., Nelson, L. D., Hampton, S. T., . . . Keller, C. (2013). Protein Kinase C iota as a Therapeutic Target in Alveolar Rhabdomyosarcoma. <i>Oncogene</i> , 32(3), 286-295. http://doi.org/10.1038/nc.2012.46	https://www.ncbi.nlm.nih.gov/pmc/articles/PMC3360112/	
PLK1	Hartsink-Segers, S. A., Exalto, C., Clifford, S. C., Caron, H. N., Pieters, R., & Den Boer, M. L. (2012). Polo-Like Kinase 1 (PLK1) Inhibition Reduces Cell Proliferation and Induces Apoptosis in Childhood Acute Lymphoblastic Leukemia. <i>Blood</i> , 120(21), 3529. Accessed March 26, 2018. Retrieved from http://www.bloodjournal.org/content/120/21/3529	http://www.bloodjournal.org/content/120/21/3529?so-checked=true	
POL1	Jones, L., Carol, H., Evans, K., Richmond, J., Houghton, P. J., Smith, M. A., & Lock, R. B. (2016). A review of new agents evaluated against pediatric acute lymphoblastic leukemia by the Pediatric Preclinical Testing Program. <i>Leukemia</i> , 30(11), 2133-2141. doi 10.1038/leu.2016.192	https://www.nature.com/articles/leu2016192.pdf	
PRDM1	Cubedo, E., Maurin, M., Jiang, X., Lossos, I. S., & Wright, K. L. (2011). PRDM1/Blimp1 Down-regulates Expression of Germinal Center Genes LMO2 and HGAL. <i>The FEBS Journal</i> , 278(17), 3065-3075. http://doi.org/10.1111/j.1742-4658.2011.08227.x	https://www.ncbi.nlm.nih.gov/pmc/articles/PMC3158840/	
PRDM8	Weidner, C. I., Lin, Q., Birkhofer, C., Gerstenmaier, U., Kaiflie, A., Kirschner, M., . . . Wagner, W. (2016). DNA methylation in PRDM8 is indicative for dyskeratosis congenita. <i>Oncotarget</i> , 7(10), 10765-10772. http://doi.org/10.18632/oncotarget.7458	https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4905437/	
PRDM10	Hofvander, J., Tayehwa, J., Nilsson, J., Magnusson, L., Brosjo, O., Larsson, O., . . . Mertens, F. (2014). Recurrent PRDM10 Gene Fusions in Undifferentiated Pleomorphic Sarcoma. <i>Clinical Cancer Research</i> , 21(4), 864-869. doi 10.1158/1078-0432.ccr-14-2399	https://www.ncbi.nlm.nih.gov/pubmed/25516889	
PRMT2	Zhong, J., Cao, R., Zu, X., Hong, T., Yang, J., Liu, L., . . . Wen, G. (2011). Identification and characterization of novel spliced variants of PRMT2 in breast carcinoma. <i>FEBS Journal</i> , 279(2), 316-335. doi 10.1111/j.1742-4658.2011.08426.x	https://febs.onlinelibrary.wiley.com/doi/full/10.1111/j.1742-4658.2011.08426.x	
PRMT5	Jin, Y., Zhou, J., Xu, F., Jin, B., Cui, L., Wang, Y., . . . Pan, J. (2016). Targeting methyltransferase PRMT5 eliminates leukemia stem cells in chronic myelogenous leukemia. <i>The Journal of Clinical Investigation</i> , 126(10), 3961-3980. http://doi.org/10.1172/JCI85239	https://www.ncbi.nlm.nih.gov/pmc/articles/PMC5096815/	
Proteasome	Cloos, J., Roeten, M. S., Franke, N. E., Meerloo, J. V., Zweegman, S., Kaspers, G. J., & Jansen, G. (2017). (Immuno)proteasomes as therapeutic target in acute leukemia. <i>Cancer and Metastasis Reviews</i> , 36(4), 599-615. doi 10.1007/s10555-017-9699-4	https://www.ncbi.nlm.nih.gov/pubmed/29071527	
PTPN (protein tyrosine phosphatase)	Lazo, J. (2018). Faculty of 1000 evaluation for In vivo CRISPR screening identifies Ptpn2 as a cancer immunotherapy target. F1000 - Post-publication Peer Review of the Biomedical Literature. doi 10.3410/f.727819093.793543387	https://www.ncbi.nlm.nih.gov/pmc/articles/PMC5924693/	

RPA3	Zuber, J., Shi, J., Wang, E., Rappaport, A. R., Herrmann, H., Sison, E. A., ... Vakoc, C. R. (2011). RNAi screen identifies Brd4 as a therapeutic target in acute myeloid leukaemia. <i>Nature</i> , 478(7370), 524–528. http://doi.org/10.1038/nature10334	https://www.ncbi.nlm.nih.gov/pmc/articles/PMC3328300/
SHP2	Zhang, R.-Y., Yu, Z.-H., Zeng, L., Zhang, S., Bai, Y., Miao, J., ... Zhang, Z.-Y. (2016). SHP2 phosphatase as a novel therapeutic target for melanoma treatment. <i>Oncotarget</i> , 7(45), 73817–73829. http://doi.org/10.18632/oncotarget.12074	https://www.ncbi.nlm.nih.gov/pmc/articles/PMC5342016/
SMYD3	Mazur, P. K., Reynoird, N., Khatri, P., Jansen, P. W. T. C., Wilkinson, A., Liu, S., ... Gozani, O. (2014). SMYD3 links lysine methylation of MAP3K2 to Ras-driven cancer. <i>Nature</i> , 510(7504), 283–287. http://doi.org/10.1038/nature13320	https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4122675/
Somatostatin Receptor	Khanna, G., Bushnell, D., & Odoriso, M. S. (2008). Utility of Radiolabeled Somatostatin Receptor Analogues for Staging/Restaging and Treatment of Somatostatin Receptor-Positive Pediatric Tumors. <i>The Oncologist</i> , 13(4), 382–389. doi 10.1634/theoncologist.2007-0175	http://theoncologist.alphamedpress.org/content/13/4/382.full
Survivin (BIRC5)	Tyner, J. W., Jemal, A. M., Thayer, M., Druker, B. J., & Chang, B. H. (2012). Targeting survivin and p53 in pediatric acute lymphoblastic leukemia. <i>Leukemia</i> , 26(4), 623–632. http://doi.org/10.1038/leu.2011.249	https://www.ncbi.nlm.nih.gov/pmc/articles/PMC3364442/
SUZ12	D'Angelo, V., Iannotta, A., Ramaglia, M., Lombardi, A., Zarone, M. R., Desiderio, V., ... Caraglia, M. (2015). EZH2 is increased in paediatric T-cell acute lymphoblastic leukemia and is a suitable molecular target in combination treatment approaches. <i>Journal of Experimental & Clinical Cancer Research</i> CR, 34(1), 83. http://doi.org/10.1186/s13046-015-0191-0	https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4535295/
SWI/SNF	St. Pierre, R., & Kadoch, C. (2017). Mammalian SWI/SNF Complexes in Cancer: Emerging Therapeutic Opportunities. <i>Current Opinion in Genetics & Development</i> , 42, 56–67. http://doi.org/10.1016/j.gde.2017.02.004	https://www.ncbi.nlm.nih.gov/pmc/articles/PMC5777332/
TET2	Zhao, Z., Chen, L., Dawlaty, M. M., Pan, F., Weeks, O., Zhou, Y., ... Xu, M. (2015). Combined loss of Tet1 and Tet2 promotes B-cell, but not myeloid malignancies in mice. <i>Cell Reports</i> , 13(8), 1692–1704. http://doi.org/10.1016/j.celrep.2015.10.037	https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4764044/
TGF-beta	Hahm, K. (1999). Correction: Repression of the gene encoding the TGF-β type II receptor is a major target of the EWS-FLI1 oncprotein. <i>Nature Genetics</i> , 23(4), 481–481. doi 10.1038/70611	https://www.ncbi.nlm.nih.gov/pubmed/10508522
Thymidylate synthase	Rocha, J. C. C., Cheng, C., Liu, W., Kishi, S., Das, S., Cook, E. H., ... Relling, M. V. (2005). Pharmacogenetics of outcome in children with acute lymphoblastic leukemia. <i>Blood</i> , 105(12), 4752–4758. http://doi.org/10.1182/blood-2004-11-4544	https://www.ncbi.nlm.nih.gov/pmc/articles/PMC1895006/
Topoisomerase I/II	Bredel, C., Lassmann, S., Pollack, I., Knoth, R., Hamilton, R., Volk, B., ... Bredel, M. (2005). DNA topoisomerase IIa and Her-2/neu gene dosages in pediatric malignant gliomas. <i>International Journal of Oncology</i> . doi 10.3892/ijo.26.5.1187	https://www.ncbi.nlm.nih.gov/pubmed/15809708
TRAIL	Kopp, L. M., & Katsanis, E. (2015). Targeted immunotherapy for pediatric solid tumors. <i>OncImmunology</i> , 5(3). doi 10.1080/2162402x.2015.1087637	https://www.tandfonline.com/doi/full/10.1080/2162402X.2015.1087637
Tubulin	Stanton, R. A., Gernert, K. M., Nettles, J. H., & Aneja, R. (2011). Drugs That Target Dynamic Microtubules: A New Molecular Perspective. <i>Medicinal Research Reviews</i> , 31(3), 443–481. http://doi.org/10.1002/med.20242	https://www.ncbi.nlm.nih.gov/pmc/articles/PMC3155728/
XPO1 (Exportin)	Echin, J., Berezovskaya, A., Conway, A. S., Galinsky, I. A., Stone, R. M., Baloglu, E., ... Look, A. T. (2017). KPT-8602, a second-generation inhibitor of XPO1-mediated nuclear export, is well tolerated and highly active against AML blasts and leukemia-initiating cells. <i>Leukemia</i> , 31(1), 143–150. http://doi.org/10.1038/leu.2016.145	https://www.ncbi.nlm.nih.gov/pmc/articles/PMC5220128/
WDR5	Sun, Y., Bell, J. L., Carter, D., Gherardi, S., Poulos, R. C., Milazzo, G., ... Liu, T. (2015). WDR5 Supports an N-Myc Transcriptional Complex That Drives a Protumorigenic Gene Expression Signature in Neuroblastoma. <i>Cancer Research</i> , 75(23), 5143–5154. doi 10.1158/0008-5472.can-15-0423	https://www.ncbi.nlm.nih.gov/pubmed/26471359
WEE1	Mueller, S., Hashizume, R., Yang, X., Kolkowitz, I., Olow, A. K., Phillips, J., ... Haas-Kogan, D. A. (2014). Targeting Wee1 for the treatment of pediatric high-grade gliomas. <i>Neuro-Oncology</i> , 16(3), 352–360. http://doi.org/10.1093/neuonc/not220	https://www.ncbi.nlm.nih.gov/pmc/articles/PMC3922515/

Target Symbol (Non-Relevant)	Citation(1)	Link(1)	Citation(2)	Link(2)	Comment
AR	Sun, J., Wang, D., Guo, L., Fang, S., Wang, Y., & Xing, R. (2017). Androgen Receptor Regulates the Growth of Neuroblastoma Cells in vitro and in vivo. <i>Frontiers in Neuroscience</i> , 11, 116. http://doi.org/10.3389/fnins.2017.00116	https://www.ncbi.nlm.nih.gov/pmc/articles/PMC5339338/			
ESR1	Lovén, J., Zinin, N., Wahlström, T., Müller, I., Brodin, P., Fredlund, E., ... Henriksson, M. (2010). MYCN-regulated microRNAs repress estrogen receptor- α (ESR1) expression and neuronal differentiation in human neuroblastoma. <i>Proceedings of the National Academy of Sciences of the United States of America</i> , 107(4), 1553–1558. http://doi.org/10.1073/pnas.0913517107	https://www.ncbi.nlm.nih.gov/pmc/articles/PMC2824410/			
ESR2	Scott, R. (2009). Estrogen receptor polymorphisms and the risk of endometrial cancer. <i>BJOG: An International Journal of Obstetrics & Gynaecology</i> , 116(8), 1053–1061. http://doi.org/10.1111/j.1471-0528.2009.02185.x	https://obgyn.onlinelibrary.wiley.com/doi/full/10.1111/j.1471-0528.2009.02185.x			
GnRHR	Cheng, C. K., Chow, B. K., & Leung, P. C. (2003). An Activator Protein 1-Like Motif Mediates 17 β -Estradiol Repression of Gonadotropin-Releasing Hormone Receptor Promoter via an Estrogen Receptor α -Dependent Mechanism in Ovarian and Breast Cancer Cells. <i>Molecular Endocrinology</i> , 17(12), 2613–2629. http://doi.org/10.1210/me.2003-0217	https://academic.oup.com/mend/article-pdf/17/12/2613/10716982/mend2613.pdf			
PSA	Matera, L. (2010). The choice of the antigen in the dendritic cell-based vaccine therapy for prostate cancer. <i>Cancer Treatment Reviews</i> , 36(2), 131-141. doi:10.1016/j.ctrv.2009.11.002	https://www.ncbi.nlm.nih.gov/pubmed/19954892	Cho H, Cockle P, Binder J, Resini W, White P, & Jooss K. Vaccine based immunotherapy regimen (VBIR) for the treatment of prostate cancer. <i>Cancer Res</i> . 76 (14 Supplement) LB-093-LB-093 (2016)		https://www.cancer.org/cancer/prostate-cancer/about/key-statistics.html
PSCA	Matera, L. (2010). The choice of the antigen in the dendritic cell-based vaccine therapy for prostate cancer. <i>Cancer Treatment Reviews</i> , 36(2), 131-141. doi:10.1016/j.ctrv.2009.11.002	https://www.ncbi.nlm.nih.gov/pubmed/19954892	Cho H, Cockle P, Binder J, Resini W, White P, & Jooss K. Vaccine based immunotherapy regimen (VBIR) for the treatment of prostate cancer. <i>Cancer Res</i> . 76 (14 Supplement) LB-093-LB-093 (2016)		https://www.cancer.org/cancer/prostate-cancer/about/key-statistics.html
PSMA	Matera, L. (2010). The choice of the antigen in the dendritic cell-based vaccine therapy for prostate cancer. <i>Cancer Treatment Reviews</i> , 36(2), 131-141. doi:10.1016/j.ctrv.2009.11.002	https://www.ncbi.nlm.nih.gov/pubmed/19954892	Cho H, Cockle P, Binder J, Resini W, White P, & Jooss K. Vaccine based immunotherapy regimen (VBIR) for the treatment of prostate cancer. <i>Cancer Res</i> . 76 (14 Supplement) LB-093-LB-093 (2016)		https://www.cancer.org/cancer/prostate-cancer/about/key-statistics.html
VEGF	Glade Bender, J., Yamashiro, D. J., & Fox, E. (2011). Clinical Development of VEGF Signaling Pathway Inhibitors in Childhood Solid Tumors. <i>The Oncologist</i> , 16(11), 1614–1625. http://doi.org/10.1634/theoncologist.2011-0148	https://www.ncbi.nlm.nih.gov/pmc/articles/PMC3233297/			
VEGFR	Kieran, M. W., Kalluri, R., & Cho, Y.-J. (2012). The VEGF Pathway in Cancer and Disease: Responses, Resistance, and the Path Forward. <i>Cold Spring Harbor Perspectives in Medicine</i> , 2(12), a006593. http://doi.org/10.1101/cshperspect.a006593	https://www.ncbi.nlm.nih.gov/pmc/articles/PMC3543071/			