Equivalence of Complex Products Cyclosporine Ophthalmic Emulsion

Robert A. Bellantone, Ph.D.

President, Physical Pharmaceutica LLC

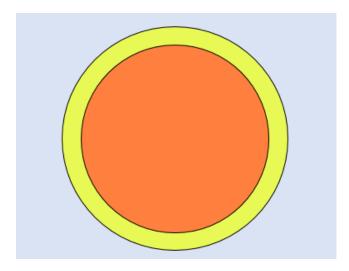


Ophthalmic emulsions as complex dosage forms

- Two marketed products (cyclosporine 0.05% and difluprednate 0.05%)
- Ophthalmic emulsions are complex materials
 - Drug is distributed in several phases
 - Complex set of conditions governing release
- Ophthalmic emulsions are subject to a complex route of delivery
 - The formulation and target region can affect each other
 - Special considerations for ocular delivery
- Two special considerations must be taken into account
 - Short residence time in the ocular region
 - Administration leaves a thin film of formulation on the ocular surfaces (~50 micron)
 - Thin film does not act as a drug depot- % depletion per time is large
 - Formulation temperature goes to ~35 °C (ocular surface temp) in about 1 second
 - The film thickness is a critical factor affecting in vitro release testing
- Cyclosporine property: as formulation temperature increases from storage temp to 35 °C, cyclosporine solubility decreases in water but increases in globules

Cyclosporine ophthalmic emulsions

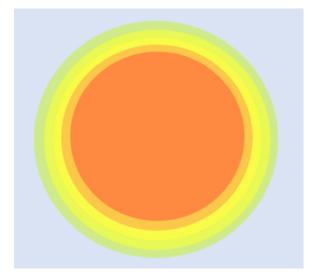
- Microemulsion
 - Globule size ~ 100-200 nm, globules occupy ~2% of the formulation volume
 - Surface to surface separation ~250-500 nm
 - In 0.1 mL, 5-40 x 10^{11} globules with total surface area $^{\circ}600-1200$ cm²
 - In a 50 micron film, estimate about 1% of globules are within 500 nm of ocular surfaces
- Structure likely affected by geometry and miscibility of Tween 80 and castor oil



If pure Tween-80, surfactant layer thickness

would be 10-20 nm (~10-20 molecules)

"Surfactant layer" may be more like a transition layer from oil to water due to miscibility



Comparing ophthalmic emulsions

- If two ophthalmic emulsion formulations are "equivalent", they will perform in the same way when administered in vivo
- One approach: two formulations will perform equivalently in vivo if they
 - Start out the same (same during storage- static measurements)
 - Respond in the same way to in vivo perturbations (kinetic processes)
- Starting state reflects storage conditions, static parameter measurements
- Response- process(es) induced by perturbations encountered in vivo
 - Rapid temperature change, redistribution and drug loss by absorption
 - Other possible factors (tearing related, for instance)
 - These perturbations are <u>large</u> and <u>occur rapidly</u> (thin film effects)

Factors affecting drug availability vs. time

- Contact time in the ocular region
 - Globule size and surface area
 - Formulation viscosity
 - Surface interactions
 - Tearing (pH, osmolality)

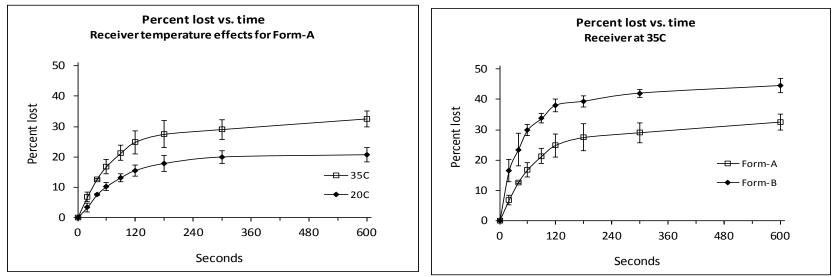
- Drug availability to tissue vs. time (transfer)
 - Initial distribution
 - Release kinetics from globule phases
 - Tearing and dilution
- Parameters to measure (static, initial conditions)
 - Globule size (contact area, surfactant distribution)
 - Viscosity, zeta potential, surface tension
 - Tearing (pH, osmolality)
 - Distribution of the drug in the formulation
- Processes that follow a change in environment (kinetic response)
 - IVRT (in vitro release test)
 - Measure release of drug in the presence of a sudden temperature change
- Data supports that all of the above are necessary
 – cannot theoretically relate the
 variables to reduce the measurement set

Release of cyclosporine from ophthalmic emulsions

- Two Q1/Q2 formulations (Form-A and Form-B) produced by different processes
- Looked at effect of temperature change, and effect of processing method
- Release measured using pulsatile microdialysis (PMD)
- See biphasic patterns. We think that
 - Drug in aqueous phase is immediately available to ocular tissues
 - Drug in globules takes longer to partition into ocular tissues
 - In vitro release data shows biphasic release patterns







Note: 100% release corresponds to ~2.85 μ g/cm² for all plots

Comments on comparative in vitro release tests

An ideal in vitro release test accounts for factors relevant to the in vivo conditions

- The ocular residence time is short
 - Release test should obtain data in a timeframe similar to the ocular residence time
 - Should avoid extrapolation of data from long times to short times
- Test should expose the formulation to perturbations from the stored state that are similar in magnitude and timescale to in vivo perturbations
 - Formulation increases temperature from 20 to 35 °C (nominally) nearly instantly
 - In the ocular region, large fraction of drug lost per time– affects diffusion and redistribution

Observation: Typical in vitro release rate tests (example, Franz cells) are far from ideal

- Release data are typically obtained over hours and require extrapolation to early times
 - Data typically obtained from 30 minutes to hours, so must extrapolate close to time = 0
 - Extrapolation requires a model with intercept = 0 (M vs. t, M vs. t^{0.5}, or ???)
 - If uncertainties in the intercept are not small compared to the differences in formulations, extrapolation cannot discriminate at the early (relevant) times
- Release experiment reflects a much more gentle and slow perturbation than occurs in vivo
 - Cannot raise temperature instantly, so perform constant temperature experiment
 - Fraction released per time is slow because of depot effect (formulation layer >> 50 microns)

Summary

- Ophthalmic emulsions are complex
 - Complex form of matter
 - Complex interactions with the ocular environment when administered in vivo
 - Cyclosporine is particularly difficult due to solubility properties
- The complexity makes it difficult (if possible at all) to model drug delivery
- We like the "same starting state" and "same response" approach
- Starting state: Static parameters to measure before administering the drug
- Response: release kinetics induced by changes reflective of those incurred in vivo
- All of the above are candidates for further research
 - Mechanistic studies of what affects release are feasible
 - Mechanistic studies of how formulation process affects the final product are more difficult

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