Appendix 5: ABI 7500 Fast v2.0 or 2.3 Method (1 of 8)

(A) Define a Run Template Using Software v2.0 or 2.3 on the ABI 7500 Fast Instrument:

Turn on the computer and ABI 7500 FAST Real-Time PCR system. Open the 7500 Software v2.0 or 2.3 and click "New Experiment" (Advanced Setup). Define "Experimental Properties" as shown below with "Experiment Name" \rightarrow "Cyclospora cayetanensis"

Experiment Menu «	Experiment: Cyclospora cayetanensis	Type: Standard Curve	Reagents: TaqMan® Reagents	START RUN (1)	
Setup	Experiment Properties				
Experiment Properties	Enter an experiment name, select the instrument type, select the type of	f experiment to set up, then select materials and methods for the Pr	CR reactions and instrument run.		
Plate Setup	How do you want to identify this experiment?				0
Run Method	* Experiment Name				
Reaction Setup	User Name (Optional)				
Materiais Lest	Comments (Optional):				*
Run	* Which instrument are you using to run the experiment?				
Analysis	7500 (36 Wells) Set up out analyze to experiment using a fast runtion 5-roter 96-well	v 7500 Fast (96 Wells)			
	What type of experiment do you want to set up?				
	V Quantitation - Standard Curve	Quantitation - Relative Standard	Curve Quantitation - C	imparative Cτ (ΔΔCτ)	
	Multi-Crease Use standards to determine the absolute muselfly of fareed nucleic and se	Genetyping Genetyping	Preser	ice/Absence	_
	Which reagants do you want to use to detect the landet se	quence?			
	V TaqMan® Reagents	SYBRD Green Reagents	met sequence	Other.	
	Which ramp ground do you want to use in the instrument ru				
	Which ramp speed ob you want to use in the instrument re		or way		

Click "Plate Setup" on left. Add targets to the "Define Targets and Samples" tab as shown below. Define the targets Ccay18S as "FAM" and synIAC as "CY5" with quencher set as "None".

Add New Target Add Saved Targe	Save farget Delete farget	Add New Sample Add Saved Sample the Semple Control	L Dampin
Target Hame	Reporter Quencher	Color Sample Name	Color
D Ccar18S	FAM None		
synIAC	CY5 None		
Define Biological Replicate Gr	ups		
Define Biological Replicate Gr	ups group in the reaction plate, click Add Biological Group, then	define the biological group.	
Define Biological Replicate Gr	ups plicate group in the reaction plate, click Add Biological Group, then great charge :	define the biological group.	
Define Biological Replicate Gri Clastructions: For each biological to Mod Biological Group 1 Biological Group Name	ups plicate group in the reaction plate, click Add Biological Group, ther gran Christon Color	define the biological group.	
Define Biological Replicate Grn	ups plicate group in the reaction plate, click Add Biological Group, ther girst Onling Color	define the biological group.	
Define Biological Replicate Gri Mathematics For each biological Mathematics For each biological Reclogical Group Name	tips plicate group in the reaction plate, click Add Biological Group, then grant Charge	define the biological group.	
Define Biological Replicate Gri Instructions: For each biological Idda Biological Group Biological Group Name	ups plicate group in the reaction plate, click Add Biological Group, then gran Onlug : Color	define the biological group.	
Cerine Biological Replicate Gr testructions: For each biological Add Biological Group Name	ups plicate group in the reaction plate, click Add Biological Group, then gran Online: Color	define the biological group.	
Oerine Biological Replicate Gr Instructions: For each biological Mod Biological Croup Terror mod Biological Croup Name	ups plicate group in the reaction plate, click Add Biological Group, then byten Christon Color	define the biological group .	
Define Biological Replicate Gri testructions: For each biological Mod Biological Group Biological Group Name	tups pilote group in the reaction plate, dick Add Biological Group, then gran throup	define the biological group.	

Appendix 5: ABI 7500 Fast v2.0 or 2.3 Method (2 of 8)

Click "Run Method" on left and define cycling parameters as shown below for a 20 μ L reaction. Define the program with an initial step of 95°C for 5 min followed by 45 cycles of [95°C for 30 sec + 67°C for 30 sec]. Data collection should be on during the 67°C hold.



After discontinuation of Qiagen QuantiFast Multiplex PCR Kit (400), Cat No. 204654 by the manufacturer, please use TaqMan[™] Fast Advanced Master Mix, ThermoFisher Scientific Applied Biosystem as: Click "Run Method" on left and define cycling parameters as shown below for a 20 µL reaction. Define the program with an initial step of 95°C for **2 min** followed by 45 cycles of [95°C for 30 sec + 67°C for 30 sec]. Data collection should be on during the 67°C hold.



Detection of Cyclospora cayetanensis in Produce

Choose "Analysis" on the left and click "Analysis Settings" in the upper right corner. Define target Ct settings in the pop up window:

- Select the Ccay18S target: Turn off: Default Settings, Automatic Threshold, and Automatic Baseline. Set the Threshold to 0.02 and choose manual baseline Start Cycle at 6 and End Cycle at 15.
- Select the synIAC target: Turn <u>off:</u> Default Settings, Automatic Threshold, and Automatic Baseline. Set the Threshold to 0.01 and choose manual baseline Start Cycle at 6 and End Cycle at 15.
- 3. Click "Apply Analysis Settings".

Experiment Menu «	Experiment: 20160405 o	cay 18S CDC+IAC	liquid qPCR M	LV lest new stds	Type: Standard	Curve Reagents: TaqMan® Rea	gents Awyon Setting
Setup	Amplification Plot		Electronic Contractor		View F	Plate Layout [View Well Table]	
Run	Plot Sottings	Analysis Settings for	20160405 coay 185 CD	C+LAC liquid oPCR MEV test ne	w stda	See St.	
	Plot Tide &Rnvs Crde . Gra	Cr Settings	Elag Settings	Advanced Settings	1		85
Analysis Analy		Reserved Department Desart C setter Desart C setter Desart C setter Travestid AUT Case153 wmAC	Long Antiparties (1999) the task of the task of task	Be at Working Section 2014 (1997) In Control (1997	E ne stithe distuit setterou p. Then datage the settings in setting. To exit the defau units (settings, To exit the defau units) (settings, To exit the defau distuits and the defau distuits and the defau distuits and the defau distuits and the default of the default distuits and the default of the default distuits and the default of the default distuits and the default of the default of the distuits and the default of the default of the distuits and the default of the default of the distuits and the default of the default of the default of the distuits and the default of the default of the default of the distuits and the default of the default of the default of the distuits and the default of the default of the default of the distuits and the default of the default of the default of the distuits and the default of the default of the default of the distuits and the default of the default of the default of the distuits and distuits and distui	. oci "Lei Dahut Genou". To uso difrent settros 8 In ara distance. Cr Settops for "Lat Orbard Settops" Cr Settops for Carris Cr Settops for Carris dubrial: Threshold massion: Gaze dubrial: Threshold massion: Gazetto Baseline Baseline Baseline Stant Optic.	
	Options Target All . Threshold						
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Appendix 5: ABI 7500 Fast v2.0 or 2.3 Method (3 of 8)

Experiment Menu «	Experiment: Cyclospora cayetaner	isis		Type: Standard Curve		Reagen	ts: Taq	Man® I	Reagen	ts		-	/28	Analysis Sette	nga l	?
Setup	Amplification Plot	-			View F	late Layou	t Vie	w Weil T	able							
Run	/ Post Sattings \				>			Select Web	a With:	Select II	sm- 💌	Select item				
Analysis	Plot Type (SRn vs Cycle) Graph Type Log	Plot Color, V	ved (*)		0 She	ow in Wells T	Vier	v Legend						E	一四	8
Amplification Plot			P	PAREE	1	2	3	4	5	6	7	8	9	10	11	12
Standard Curve		Save As Templa	te Li templates			aen										
Nutlic orponent Plot			2 Cyclospora cayetar	tensis				11								
		Recent Borns														
Raw Cela Pios																
QC Summary		Desktop														
Multiple Plots View	5	I														
	com	My Documents														
														_		
		Computer														
			File name: Dicion	cora cayetasensis ed			Şavo	ī –								
	1. A.	Network	Files of type: Esperin	nent Document Template files (* edt)		•	Cancel									
		Ш н														
	- Andrewson and -				0											
	Target All Threshold Take															
	Thow Threaten - Bassine Start Wes	Tarpet de Bane	one fact west Tarte	540 -	н											
	and the second statement of the second															

Click "File" \rightarrow "Save as template..." \rightarrow "Save"

(B) Run Method Using Software v2.0 or 2.3 on the ABI 7500 Fast Instrument

Turn on the computer and ABI 7500 FAST Real-Time PCR system. Open the 7500 Software v2.0 or 2.3 and click "File" \rightarrow "New Experiment" \rightarrow "From Template". Choose the "Cyclospora cayetanensis.edt" template file created according to Appendix 2 instructions above. Under "Setup" on the left click "Plate Setup" and define all unknown samples or DNA extraction controls on the plate on the "Define Targets and Samples" tab by clicking "Add New Sample" until all samples are defined.

and the second s	Define Targets			Define	Samples	
inte Solup	Add New Target Add Saved Target	Save Target Delete Target	1	1	ew Sample Add Saved Sample Save Sample Delete Sample	
m Method	TargetName	Reporter	Quencher	Color Sampl	e Name	Color
	Ccay10S	FAM	· None	· Sampl	e1	
	synIAC	CY5	+ None	· Sampl	02	
aterials List				Sampl	e3	
luo	1			Sampl	e 1 diluted	
Kun				Sampl	e 2 diluted	
Analysis				Samp	e 3 diuted	
	1					
	Define Biological Replicate Gro	ups				
	Instructions: For each biological re	plicate group in the reaction plat	e, click Add Biological Gro	up, then define the biological group.		
	Add Biological Group	рса Слосе				
	Pistopicst Oroug Name		Color		Comments	

Appendix 5: ABI 7500 Fast v2.0 or 2.3 Method (4 of 8)

Click the "Assign Targets and Samples" tab to define well assignments. Define the NTC wells by selecting three wells and checking the box next to the Ccay18S target choosing "N" as task.

Experiment Menu «	Experiment: Untitled	Type: Standard Curve	Reagents: TaqMan® Reagents	START RUN IN
Setup	Define Targets and Samples Assign Targ	ets and Samples		
Experiment Properties	Instructions: To set up standards: Chck: "Define and Set U To set up unknowns: Select wells, assign ta To set up negative controls: Select wells, as	Ip Standards." rge(s), select "U" (Unknown) as the task for each target assignment, the sign target(s), then select "N" (Negative Control) as the task for each targ	n assign a sample et assignment	
Plate Setup	Assign target(s) to the selected wells.	View Plate Layout View Well Table		
Run Method	Auron Target Task Qua	> mty	Select Wells With: - Select item Select item	10 TH 11
Reaction Setup	Ccay185	Show in Wells View Legend		
🐔 Materials List	SURVE UNIT AL	1 2 3 4	5 6 7 8 9	10 11 12
Run	Mixed 🕕 Unknown 🔂 Standard 🗔 Negative Co	A Coay183 Coay183 Coay183		0.00
Analysis	They Define and Set Up Standards	0		
	Assign sample(s) to the selected wells.			
	Assign Sample	C		
	Sample 1			
	Sample 2			
	Sample 1 diluted			
	Assign sample(s) of selected well(s) to biologica	ni gr		
	Assign Biological Group	F		
		6		a a a
	Select the due to use as the passive reference			_
		н		
	Rox at	Wells: 10 0 Unknown S 0 Standard S 3 Negative Control		93 Empty

Define all unknown samples or DNA extraction controls one at a time by selecting three wells for each and checking the box next to the sample name in the "Assign sample(s) to the selected wells" panel.

Experiment Menu «	Experiment:	Untitled		т	ype: Stand	lard Curve		R	leagents:	TaqMan® I	Reagents			START RUN	0
Setup	Define Targ	ets and Samples Assign Target	s and	Samples											
Experiment Properties	Instructions:	To set up standards. Click "Define and Set Up 5 To set up unknowns: Select wells, assign targe To set up negative controls: Select wells, assign	Standard N(s), sele n target(r	s." d."U" (Unknow 1), then select 1	n) as the task f N° (Negative Co	or each target as: ontrol) as the task	lignment, ther for each targe	i assign a samp t assignment	ie.						
Plate Setup	Assign targe	t(s) to the selected wells.	<	liew Plate	Layout	view Well Tab	le	20172122200220							
Run Method	Assign	Target Task Quantity	2					Select Wells V	/ith: - Select It	em - 💌 - Sele	ct Item - 💌				
Reaction Setup	1	Ccay18S		Show in W	ella Y	View Legend								III III	
		synIAC		1	2	3	4	5	6	7	8	9	10	11	12
S Milleriano Lisa			A	Coay185	Coay185	Casy185									
Run	Mixe	d 🛄 Unknown 🔝 Standard 🔝 Negative Contro						_				_	_		
Analysis	The Define a	nd Set Up Standards	8	Sample 1	Sample 1	Sample 1									
	Assign samp	le(s) to the selected wells.													
	Assign	Sample	c	Sample 2	Sample 2	Sample 2									
	E	Sample 1													
	E1 -	Sample 2	_ 0	Sample 3	Sample 3	Sample 3									
	<u> </u>	Sample 3													
		Sample 1 diluted	- 6	Sample 1 diul.	Semple 1 ditut	Sample 1 dout									
	12	Sample 3 diluted	-	Samela 7 dilut	Samale 2 dika	Semale 2 dilut									
	Assign samp	ie(s) of selected well(s) to biological o					-								
	Assign	Biological Group	0	Sample 3 dilut	Sample 3 dilut	Sample 3 dilut		- 185							
	Select the dy	e to use as the passive reference.	н		-		100								
	ROX .	1													
- T	10		e w	ells: 🔟 0 Unk	nown 🔝 🛛 Sta	indard 🔝 3 Neg	ative Control								93 Empty

Appendix 5: ABI 7500 Fast v2.0 or 2.3 Method (5 of 8)

Then select all unknown samples or DNA extraction controls and check the box next to Ccay18S target choosing "U" as task.

Experiment Menu «	Experiment: Untitled	Type: Standard Curve	Reagents: TaqMan® Reagents	START RUN ()
Setup	Define Targets and Samples Assign Tar	gets and Samples		
Experiment Properties	Instructions: To set up standards: Click 'Define and Se To set up unknowns' Select wells, assign To set up negative controls. Select wells, a	t Up Standards." target(s), select "U" (Unknown) as the task for each target assignment, thei assign target(s), then select "N" (Negative Control) as the task for each target	n assign a sample et assignment	
Page Search	Assign target(s) to the selected wells.	View Plate Layout View Well Table		
Fun Method	Target Task Ok	santity	Select Wells With: - Select Item - 💌	Travers I cannot a second
Reaction Solup	Ccay185	O Show in Wells View Legend		
🛒 Materials List	torration (torration)	1 2 3 4	5 6 7 9 9	10 11 12
Run	Mixed 🔃 Unknown 🔂 Standard 🔝 Negative C	ontrol A Court25 Court25		
Analysis	Define and Set Up Standards	B Carcia 1 Sampla 1 Science 1		
	Assign sample(s) to the selected wells.	Sample 2 Sample 2 Sample 2		
	Assign Sample	Centras Centras		
	Sample 1	Sample 3		100 IS 100
	E Sample 3			
	Sample 1 diuted	E Conviss Convis		
	Sample 3 diluted	Dampie 2 diul. barrate 2 diul. barrate 2 diul.		100 100 100
	Assign sample(s) of selected well(s) to biologic	al gro		
	Assign Biological Group	C Convis		
	Select the dye to use as the passive reference			100 (St. 100)
	ROX			
	(m)	Wells: UI 18 Unknown 🔝 0 Standard 🔝 3 Negative Control	1	75 Empty

Define the Positive control (Standard) wells by selecting three wells and checking the box next to the Ccay18S target choosing "S" as task and "1000" as quantity.

Experiment Menu «	Experiment: 1	Untitled		Туре:	Standard	Curve			Rea	igents: Ta	aqMan®	Rea	gents			STAR	RUN (C	0
Setup	Define Targe	ets and Samples Assign Targets an	d Sar	nples											1996			
Experiment Properties	Instructions:	To set up standards: Click 'Define and Set Up Standa To set up unknowns: Select wells, assign target(s), s To set up negative controls: Select wells, assign target	rds " Hect "U t(s), the	" (Unknown) as t In select "N" (Ne	he task for eac gative Control)	h target assignm as the task for ea	ent, then assi ich target ass	on a s Ionme	ample.									
Plate Setup	Assign target	t(s) to the selected wells.	<	View Plate	Layout	View Well Ta	ble]											
Run Method	Asses T	arget Task Quantity	ľ.				1	Select	Wells W	m: - Select	item - 💌	Select	Nem - 💌					
Reaction Setup	1 C	Cay185		O Show in W	INIS V	View Legend											15	10
Wateriste List	<u>D</u> _0		- [1	2	3	4		5	6	7		8	9	10		11	12
				A Coay185	Coay188	Ceay185												
Run	Mixed	d 🕕 Unknown 🔄 Standard 🔝 Negative Control		LISENSU!	2623807	255835.0				_		-				-		
Analysis	The Define an	od Set Up Standarda	_	B Ccay185	Ccay183	Ccay183												
	Assign sampl	ie(s) to the selected wells.		Sample 2	Sample 2	Sample 2												
	Assign	Sample		C Coay105	U Ccay105	U Ccay105												
		Sample 1	1	D Sample 3	Sample 3	Sample 3												
	10	Sample 3		Contras	Ceating	Ceatings												
	10	Sample 1 diluted	1	E Semple 1 dilu.	Sample 1 dile	Sample 1 dile												
	1	Sample 2 diluted		famela 7 dilu	famale 7 dilu	Samela 7 dilu										t	_	
		Sample 3 diuted	-	F Ccay185	Ccay183	Ccay183												
	Assign sampl	le(s) of selected well(s) to biological group		Sample 3 dilu	Sample 3 dilu	Sample 3 dilu.												
	Assign	Biological Group		Ceay185	Ceay185	Ceay185												_
	Select the dy	e to use as the passive reference.	1	H Ceay185	Coay185	Ceay185												
	ROX -																	
	4	m	7	Wells: 🔟 18 Ur	innown 📴 3 S	atandard 🔝 3 N	egative Contro	N								_		72 Empty

Appendix 5: ABI 7500 Fast v2.0 or 2.3 Method (6 of 8)

Assign the internal amplification control by selecting all reaction wells and checking the box next to the synIAC target choosing "U" as task. Assure that ROX is selected as a passive reference dye. "File" \rightarrow "Save as" \rightarrow Experiment Document Single file (*.eds) with a unique name. Insert plate or tube strips and start the run.

Experiment Menu «	Experiment: 1	Untitled		Type: Stan	lard Curve		Rea	gents: Tao	qMan® Re:	agents			TART RUN	
Setup	Define Targ	ets and Samples Assign T	argets and Samp	les										
Experiment Properties	Instructions:	To set up standards: Click 'Define and To set up unknowns: Select wells, assi To set up negative controls: Select welk	Set Up Standards," gn target(s), select "U" (U s, assign target(s), then s	nknown) as the task relect "N" (Negative C	for each target assignm ontrol) as the task for e	ent then assi ich target ass	ign a sample. Ignment							
Tate Senip	Assign target	t(s) to the selected wells.	< 1	ew Plate Layou	t View Well Ta	ble	2250000000							
Run Method	Assian	farnet Task	Osantity				Select Wells W	th: - Select ite	m - 💌 - Sele	ct Item - 💌				
	2 0	Ccay188	1,000	Show in Wells ¥	View Legend									
Keaction Setup	12 s	synlAC		- E	2 3	4	5	6	7	8	. 9	10	11	12
🛒 Materials List					artas Courtas									
Rup	Mixed	d 🕕 Unknown 🔄 Standard 🖸 Negative	Control	U synLAC U syn	anc 🔟 +11 MC									
Analysis	The Define an	nd Set Up Standards		Sample 1 Sam	cle 1 Samole ay188 II Ceay188									
	Assign sampl	le(s) to the selected wells.		Semple 2 Sem	ule 2 Samule 2									
	Assign	Sample	c	Coaytas Co syntAC SyntAC	ey105 Coey105									
	8	Sample 1		Sample 3 Sam	cle 3 Sample 3									
	8	Sample 2	D	U Craytas U Cr	ay185 Caay185									
	13	Sample 3		Bample 1 dilu . Bample	1 dilu . Sample 1 dilu .									
	8	Sample 1 diluted	E	Cuay185 Co	ay185 Ceay185									
	8	Sample 2 diluted		Ramala 2 dilu	2 dily Barrale 2 dily									
	B	Sample 3 diluted	- 5	Coaytas Co	ay185 Ceay185									
	Assign sampl	le(s) of selected well(s) to biolo	gical group.	Samole 3 dilu Samole	3 dilu									
	Assign	Biological Group	0	Coay125 Co	ay185 U Ceay185									
	Select the dy	e to use as the passive referen	ce.	Coay185 Co 1E3	ny185 Coay185									
	ROX		we	lis: 🚺 👥 Unknown	3 Stand 1 3 N	egative Contri	ol							72 Empty

(C) Analysis Using Software v2.0 or 2.3 on the ABI 7500 Fast Instrument

When the run is complete select "Amplification Plot" under "Analysis" on the left. Ensure that all wells are selected on the "View Plate Layout" tab to the right of the amplification curves. In the options panel below the amplifications curves, select the "synIAC" target and check that the show threshold and baseline boxes are both checked. Verify threshold and baseline settings are accurate as defined in the run template above.



Appendix 5: ABI 7500 Fast v2.0 or 2.3 Method (7 of 8)

Next, select the "Ccay18S" target in the options panel and check that the show threshold and baseline boxes are checked. Verify threshold and baseline settings are accurate as defined in the run template above.



Review the amplification plots and Ct's for each target. Verify that all criteria for a valid experimental run are met as defined in the "Interpretation of Results" section of the protocol.

Then, assure that all reaction wells on the plate are selected by clicking the upper left corner of the Plate Layout. Click "Export" to open the Export Tool window.



Appendix 5: ABI 7500 Fast v2.0 or 2.3 Method (8 of 8)

On the "Export Properties" tab, select the following:

- 1. Select "Results" only.
- 2. Choose "One File"
- 3. Name: *use experiment name*. Location: *define a location of your choice*. File type: *choose ".xls"*

Click the "Customize Export" tab and select the following results content: Well, Sample Name, Target Name, Task, Reporter, Ct, Ct Mean, Ct SD.

Click on the "Target Name" column header to sort the table by target name. Click "Start Export". Close the export tool.

Export Properties	Customiz	e Export										-
Customize: Results 💌						File Na	ame: 20160-	406 ccay 185 (DC+IAC liqui	d gPCR MLV te	st 20160122 stds_data Fil	e Type: 💐
Organize Data		Results Ex	port									
Down Rows	sas Columns	Weti	Sample I Targe	tName *1 ask	Re	porter	Quencher	Ст	Cr Mean	CT SD		
	Concession of the	A1	-	UNKNO	WN CY	and the second second	None	27.683271	27.565454	0.16096953		
Select Results Content		A2	syniAd	C UNKNO	WN CYS		None	27.63105	27.565454	0.16096953		
All Results Fields		43	syn/A/	UNKNG	WN CY		INOUG	27.382046	27.565454	0.16096953		
100		81	syntAc	S UNKW	NUTA CYS		rvone	27.427088	25.957669	0.24948272		1
(K) Well		06	symmetry sumit/	2 UNION	WH CVE		None	27,091705	26.967660	0.24940272		
Sample Name		R4	auntal	10000	INTE CY		None	26.800162	26.067660	0 24048272		
E completione		85	evolat	LINKN	WAN CY	-	None	26.904732	26.967669	0.24948272		
Target Name	- E	86	synlad	LINKN	INN CY		None	26.939571	26.957669	0.24948272		
100	11	87	syniad	UNKNO	WN CY		None	26.841892	26 967669	0 24948272		
V Task		BS	synlad	UNKNO	WN CY		None	26.55771	26.967669	0.24948272		
Reporter		C1	syniAd	LINKNO	WN CYS		None	27.127333	26.758644	0.2983527		
(E) contraction		C2	synlAd	UNKNO	WN CY	5	None	26.705702	26.758644	0.2983527		
2 Quencher		C3	syntac	C UNKNO	WIN CYS	5	None	26.722055	26.758644	0.2983627		
100 00		C4	syniAd	C UNKNO	WN CYS		None	26:399403	26.758644	0.2983627		
V CT		D1	syniAd	C UNKNO	WN CYS	5	None	27.02777	26.701876	0.28476223		
CT Mean		D2	syntAd	C UNKNO	WN CY	6	None	26.57681	26.701876	0.28476223		
(e) or mean		03	synlåd	D UNKNO	WN CYS	i	None	26,501049	26 70 1876	0.28476223		
Cr SD		E1	syniA(2 UNKNO	WIN CYS	5	None	26.806679	26.548342	0.23110305		
		E2	syntad	C UNKNO	WN CYS	5.	None	26.477097	26.548342	0.23110306		
Cuantity		E3	synlad	C UNKNO	WN CY	8	None	26,36125	26.548342	0.23110306		
Cuantity Mean	.+	F1	synlAl	C UNKNO	WN CYE	5	None	26.92971	26.654741	0.23936835		
Field Concerns (Dellarity)		F2	syntAc	C UNKING	WN CY:	5/2	None	26.492933	26.654741	0.23936835		
view separator (Destinate)		F3	synlad	UNKNO	WN CY		NORE	26.541584	26.654741	0.23935835		
C Tabs Commas		61	syntAc	CINKNG	ANTA CYS	0	None	27.957405	27.856936	0.3314823		
		GZ	SyriAd	C UNKNK	WN CY:	2	None	28.126562	27.856936	0.3314623		
Open file(5) when expor	t is complete											