

FOOD AND DRUG ADMINISTRATION
CENTER FOR DRUG EVALUATION AND RESEARCH

Oncologic Drugs Advisory Committee

Tuesday, December 16, 2008

8:00 a.m. to 3:30 p.m.

Hilton Washington, D.C. North Gaithersburg
620 Perry Parkway
Gaithersburg, Maryland

**Oncologic Drugs Advisory Committee
December 16, 2008**

Hilton Washington DC North/Gaithersburg
The Ballrooms, 620 Perry Parkway, Gaithersburg, MD
Meeting Roster

ONCOLOGIC DRUGS ADVISORY COMMITTEE MEMBERS (Voting)

Jean Grem, M.D.

Professor of Medicine
Department of Internal Medicine
Section of Hematology/Oncology
University of Nebraska Medical Center
987680 Nebraska Medical Center
Omaha, NE 68198

David Harrington, Ph.D.

Department of Biostatistics &
Computational Biology
Dana-Farber Cancer Institute
44 Binney Street
Boston, MA 02115

Michael Link, M.D.

The Lydia J. Lee Professor of Pediatrics
Chief, Division of Hematology/Oncology
Stanford University School of Medicine
1000 Welch Road, Suite 300
Palo Alto, CA 94304

Gary Lyman, M.D., M.P.H.

Director, Health Services and Outcomes
Research Program- Oncology
Duke University Medical Center
Box 3645
Durham, NC 27710

Virginia Mason, R.N. (Consumer Representative)

Executive Director
Inflammatory Breast Cancer Research Foundation
P.O. Box 786
Citronelle, AL 36522

Ronald Richardson, M.D.

Consultant, Department of Medical Oncology
Mayo Clinic
200 First Street, SW, Gonda 10
Rochester, MN 55905

**Oncologic Drugs Advisory Committee
December 16, 2008**

Hilton Washington DC North/Gaithersburg
The Ballrooms, 620 Perry Parkway, Gaithersburg, MD
Meeting Roster

**ONCOLOGIC DRUGS ADVISORY COMMITTEE MEMBERS
(Voting - Continued)**

Wyndham Wilson, M.D.
Chief, Lymphoma Therapeutics Section
Metabolism Branch
Center for Cancer Research
National Cancer Institute
9000 Rockville Pike, Bldg. 10, Room 4N-115
Rockville, MD 20892

INDUSTRY REPRESENTATIVE (Non-Voting)

Gregory Curt, M.D.
U.S. Medical Science Lead, Emerging Products
AstraZeneca Oncology
P.O. Box 223
Garrett Park, MD 20896

TEMPORARY VOTING MEMBERS

Ralph D'Agostino, Ph.D.
Chair, Mathematics and Statistics Department
Boston University
111 Cummington Street
Boston, MA 02215

Jo-Ellen De Luca (Patient Representative)
Spartanburg, SC 29302

Janice Dutcher, M.D. (Acting Chair)
Associate Director for Clinical Affairs
Cancer Center
North Division, Montefiore Medical Center
Bronx, NY 10466

William Funkhouser, M.D., Ph.D.
Associate Professor
Director, Anatomic and Surgical Pathology
Department of Pathology and Lab Medicine
University of North Carolina Hospitals
CB#7525, Brinkhous-Bullitt Building
University of North Carolina
Chapel Hill, NC 27599

**Oncologic Drugs Advisory Committee
December 16, 2008**

Hilton Washington DC North/Gaithersburg
The Ballrooms, 620 Perry Parkway, Gaithersburg, MD
Meeting Roster

TEMPORARY VOTING MEMBERS (Continued)

Joanne Mortimer, M.D.
Vice Chair Medical Oncology
City of Hope Comprehensive Cancer Center
1500 East Duarte Road
Duarte, CA 91010

George Netto, M.D.
Associate Professor of Pathology, Urology and Oncology
Johns Hopkins Medical Institutions
Division of Surgical Pathology
401 N. Broadway, Weinberg Bldg/Room 2242
Baltimore, MD 21231

Ronald Przygodzki, M.D.
Acting Director, Biomedical Laboratory
Research and Development
Associate Director, Genomic Medicine
Office of Research and Development (121E)
Department of Veterans Affairs
810 Vermont Ave, NW
Washington, DC 20420

Derek Raghavan M.D., Ph.D.
M. Frank & Margaret Domiter Rudy
Distinguished Chair & Director
Taussig Cancer Institute
Cleveland Clinic
9500 Euclid Ave, R35
Cleveland, OH, 44195

Richard Simon, D.Sc.
Chief, Biometric Research Branch
National Cancer Institute
9000 Rockville Pike
MSC 7434
Bethesda MD 20892

**Oncologic Drugs Advisory Committee
December 16, 2008**

Hilton Washington DC North/Gaithersburg
The Ballrooms, 620 Perry Parkway, Gaithersburg, MD
Meeting Roster

TEMPORARY VOTING MEMBERS (Continued)

Xiao-Hua Andrew Zhou, Ph.D.
Professor, Department of Biostatistics
School of Public Health and
Community Medicine
University of Washington
Office H655E, HSB Box #35723
Seattle, WA 98198

FDA (Non-Voting)

Richard Pazdur, M.D.
Director, Office of Oncology Drug Products (OODP)
Office of New Drugs (OND), CDER, FDA

Patricia Keegan, M.D.
Director, Division of Biologic Oncology Products (DBOP)
OODP, OND, CDER, FDA

Robert O'Neill, Ph.D.
Director, Office of Biostatistics (OB)
Office of Translational Science (OTS), CDER, FDA

Ruthann Giusti, M.D.
Medical Officer, Division of Biologic Oncology
Products (DBOP), OODP, OND, CDER, FDA

Robert Becker, M.D., Ph.D.
Chief Medical Officer, Office of In-Vitro
Diagnostics (OIVD)
Center for Devices and Radiological Health (CDRH), FDA

**FOOD AND DRUG ADMINISTRATION
Center for Drug Evaluation and Research**

Oncologic Drugs Advisory Committee

AGENDA

December 16, 2008

- | | | |
|--|---|--|
| 8:00 a.m. | Call to Order
Introduction of Committee | Janice Dutcher, M.D.
Acting Chair, ODAC |
| | Conflict of Interest Statement | Nicole Vesely, Pharm.D.
Designated Federal Official, ODAC |
| <i>The committee will discuss biologic license application (BLA) 125084, trade name ERBITUX (cetuximab), ImClone Systems, Incorporated, and BLA 125147, trade name VECTIBIX (panitumumab), Amgen, Incorporated, in the context of K-ras as a predictive and/or prognostic biomarker in oncology drug development. <u>The discussion at this meeting will focus on general considerations for clinical trial designs involving the use of diagnostic tests and conducting retrospective analyses.</u></i> | | |
| 8:10 a.m. | Opening Remarks | Richard Pazdur, M.D.
Director, Office of Oncology Drug Products (OODP), Office of New Drugs (OND), CDER, FDA |
| 8:15 a.m. | <u>FDA Presentation</u>
Regulatory History | Ruthann Giusti, M.D.
Medical Officer, Division of Biologic Oncology Products, OODP, OND, CDER, FDA |
| 8:30 a.m. | <u>Sponsor Presentation</u>
Role of K-ras Mutation Status In Optimizing Selection of Colorectal Cancer Patients for Treatment with Erbitux [®] (Cetuximab) | <u>ImClone Systems Inc.</u>
Hagop Youssofian, M.D.
Senior Vice President, Clinical Research and Development
ImClone Systems, a wholly-owned subsidiary of Eli Lilly and Company |
| 9:00 a.m. | <u>Sponsor Presentation</u>
Introduction and Overview | <u>Amgen, Inc.</u>
Paul Eisenberg, M.D., MPH
Senior Vice President,
Global Regulatory Affairs & Safety
Amgen Inc. |
| | KRAS as a Predictive Biomarker for Vectibix [®] (panitumumab) Monotherapy | David Reese, M.D. , Executive Director
Global Clinical Development
Amgen Inc. |

**FOOD AND DRUG ADMINISTRATION
Center for Drug Evaluation and Research**

Oncologic Drugs Advisory Committee

AGENDA

**December 16, 2008
(continued)**

- 9 :30 a.m. **FDA Presentation**
Prospective vs. Non-Prospective Design in Companion Drug/Diagnostic Studies **Robert Becker, Jr., M.D., Ph.D.**
Chief Medical Officer, Office of In Vitro Diagnostics (OIVD), CDRH
- Some Considerations for Statistical Design, Analysis, and Interpretation for Biomarker Classifier Based Clinical Trials in Establishing Efficacy in Support of Regulatory Marketing and Promotional Claims **Robert O'Neill, Ph.D.**
Director, Office of Biostatistics (OB), Office of Translational Sciences (OTS), CDER, FDA
- 10:30 a.m. *Break*
- 10:45 a.m. Questions to the Presenters
- 11:30 a.m. Open Public Hearing
- 12:30 p.m. *Lunch*
- 1:30 p.m. Questions to ODAC and ODAC Discussion
- 3:00 p.m. *Break*
- 3:15 p.m. Questions to ODAC and ODAC Discussion
- 4:00 p.m. Adjourn

C O N T E N T S

	<u>Page</u>
Call to Order and Introduction of Committee Janice Dutcher, M.D., Acting Chair, ODAC	10
Conflict of Interest Statement Nicole Vesely, Pharm.D. Designated Federal Official, ODAC	12
Opening Remarks Richard Pazdur, M.D.	17
FDA Presentation Ruthann Giusti, M.D.	26
Sponsor Presentation ImClone Systems Inc. Hagop Youssoufian, M.D.	39
Sponsor Presentation Amgen, Inc. Paul Eisenberg, M.D., MPH David Reese, M.D.	60 66
FDA Presentation Robert Becker, Jr., M.D., Ph.D. Robert O'Neill, Ph.D.	82 103
Questions to Presenters	127
Open Public Hearing	172
Questions to ODAC and ODAC Discussion	188
Adjournment	303

P R O C E E D I N G S

1
2 DR. DUTCHER: Good morning. We're going to
3 get started.

4 My name is Janice Dutcher. I'm chairing this
5 special meeting of the ODAC Committee. I need to read
6 some statements at the beginning.

7 For topics such as those being discussed at
8 today's meeting, there are often a variety of opinions,
9 some of which are quite strongly held. Our goal is
10 that today's meeting will be a fair and open forum for
11 discussion of these issues and that individuals can
12 express their views without interruption. Thus, as a
13 gentle reminder, individuals will be allowed to speak
14 into the record only if recognized by the chair. We
15 look forward to a productive meeting.

16 In the spirit of the Federal Advisory
17 Committee Act and the Government in the Sunshine Act,
18 we ask that the Advisory Committee members take care
19 that their conversations about the topic at hand take
20 place in the open forum of the meeting.

21 We are aware that members of the media are
22 anxious to speak with the FDA about these proceedings.

1 However, FDA will refrain from discussing the details
2 of this meeting with the media until its conclusion.
3 Also, the committee is reminded to please refrain from
4 discussing the meeting topic during breaks or lunch.

5 Thank you.

6 We're going to go around the table and ask
7 members of the committee to introduce themselves.

8 As I said, my name is Janice Dutcher. I'm at
9 Montefiore Medical Center in New York, New York Medical
10 College.

11 DR. GREM: I'm Jean Grem. I am from the
12 University of Nebraska Medical Center in Omaha.

13 DR. RICHARDSON: Ron Richardson, Mayo Clinic,
14 Rochester, Minnesota, a medical oncologist.

15 DR. DUTCHER: Yes. Please say your
16 discipline, as well. Thank you.

17 MS. MASON: I'm Jenny Mason, Virginia Mason.
18 I'm the Executive Director of the Inflammatory Breast
19 Cancer Research Foundation and a nurse.

20 MS. DE LUCA: Jo-Ellen De Luca. I'm your
21 patient advocate.

22 DR. D'AGOSTINO: Ralph D'Agostino from Boston

1 University, statistician.

2 DR. FUNKHOUSER: Bill Funkhouser from Chapel
3 Hill, North Carolina, at UNC, in atomic pathology.

4 DR. MORTIMER: Joanne Mortimer, Medical
5 Oncologist, City of Hope.

6 DR. NETTO: George Netto from Johns Hopkins
7 University, pathology, molecular pathology and
8 oncology.

9 DR. RAGHAVAN: Derek Raghavan, Cleveland
10 Clinic, medical oncology.

11 DR. CURT: Gregory Curt, medical oncology,
12 AstraZeneca, industry representative to ODAC.

13 DR. PAZDUR: Richard Pazdur, Office of
14 Oncology Drug Products, FDA.

15 DR. KEEGAN: Patricia Keegan, FDA.

16 DR. BECKER: Robert Becker, Center for
17 Devices and Radiological Health, FDA.

18 DR. GIUSTI: Ruthann Giusti, medical
19 oncology, medical officer, Office of Oncology Drug
20 Products, FDA.

21 DR. ZHOU: Andrew Zhou from University of
22 Washington, biostatistician.

1 DR. SIMON: Richard Simon, National Cancer
2 Institute. I'm chief of the Biometric Research Branch.

3 DR. PRZYGODZKI: I'm Ron Przygodzki. I'm a
4 molecular genetic pathologist and I'm the director in
5 Biomedical Labs and Research Development in the
6 Veterans Affairs.

7 DR. LINK: Michael Link, pediatric
8 oncologist, Stanford.

9 DR. HARRINGTON: David Harrington,
10 statistician, Dana-Farber Cancer Institute.

11 DR. LYMAN: Gary Lyman, medical oncologist
12 and health outcomes researcher from Duke University.

13 DR. WILSON: Wyndham Wilson, intramural
14 program, National Cancer Institute, medical oncologist.

15 MS. VESELY: Nicole Vesely, designated
16 federal official, ODAC.

17 The Food and Drug Administration is convening
18 today's meeting of the Oncologic Drugs Advisory
19 Committee under the authority of the Federal Advisory
20 Committee Act of 1972.

21 With the exception of the industry
22 representative, all members and temporary voting

1 members are special government employees or regular
2 federal employees from other agencies and are subject
3 to federal conflict of interest laws and regulations.

4 The following information on the status of
5 the committee's compliance with federal ethics and
6 conflict of interest laws covered by, but not limited
7 to, those found at 18 USC Section 208 and Section 712
8 of the Federal Food, Drug and Cosmetic Act is being
9 provided to participants in today's meeting and to the
10 public.

11 FDA has determined that members and temporary
12 voting members of this committee are in compliance with
13 federal ethics and conflict of interest laws.

14 Under 18 USC Section 208(3), Congress has
15 authorized FDA to grant waivers to special government
16 employees who have potential financial conflicts when
17 it is determined that the agency's need for a
18 particular individual's services outweighs his or her
19 potential financial conflict of interest.

20 Under Section 712 of the FD&C Act, Congress
21 has authorized FDA to grant waivers to special and
22 regular government employees with potential personal

1 financial conflicts when necessary to afford the
2 committee essential expertise.

3 Related to the discussions of today's
4 meeting, members and temporary voting members of this
5 committee have been screened for potential financial
6 conflicts of interest of their own, as well as those
7 imputed to them, including those of their spouses or
8 minor children and for purposes of 18 USC Section 208,
9 their employers.

10 These interests may include investments,
11 consulting, expert witness testimony, contracts,
12 grants, CRADAs, teaching, speaking, writing, patents
13 and royalties, and primary employment.

14 Today's agenda involves discussion of ImClone
15 Systems' Erbitux and Amgen's Vectibix regarding types
16 of studies and data needed to establish KRAS mutational
17 status as predictive of response to drug therapy or as
18 a prognostic biomarker in colon cancer. This topic is
19 a particular matter involving specific parties.

20 Based on the agenda for today's meeting and
21 all financial interests reported by the committee
22 members and temporary voting members, conflict of

1 interest waivers have been issued in accordance with
2 18 USC Section 208(b)(3) to the following participants:
3 Dr. Jean Grem for imputed interest in three Eastern
4 Oncology Cooperative Group sponsored trials and one
5 Cancer and Leukemia Group B sponsored trial. The
6 magnitude of the funding for each trial is between zero
7 and \$50,000.

8 Dr. David Harrington for an imputed interest
9 in a cooperative group related to a competing product.
10 The magnitude of the subcontract is between \$50,001 and
11 \$100,000.

12 Dr. Derek Raghavan for an imputed interest in
13 the sponsor of one of the products to be discussed.
14 The magnitude of the contract is between \$50,001 and
15 \$100,000.

16 The waivers allow these individuals to
17 participate fully in today's deliberations. FDA's
18 reasons for issuing the waivers are described in the
19 waiver documents which are posted on FDA's Website at
20 www.fda.gov/ohrms/dockets/default.htm.

21 Copies of the waivers may also be obtained by
22 submitting a written request to the agency's Freedom of

1 Information Office, Room 6-30 of the Parklawn Building.

2 A copy of this statement will be available
3 for review at the registration table during this
4 meeting and will be included as part of the official
5 transcript.

6 With respect to FDA's invited industry
7 representative, we would like to disclose that
8 Dr. Gregory Curt is participating in this meeting as a
9 non-voting industry representative, acting on behalf of
10 all regulated industry.

11 Dr. Curt's role at this meeting is to
12 represent industry interests in general and not any
13 particular company. Dr. Curt is employed by
14 AstraZeneca.

15 We would like to remind members and temporary
16 voting members that if the discussions involve any
17 other products or firms not already on the agenda for
18 which an FDA participant has a personal or imputed
19 financial interest, the participants need to exclude
20 themselves from such involvement and their exclusion
21 will be noted for the record.

22 FDA encourages all other participants to

1 advise the committee of any financial relationships
2 that they may have with any firms at issue.

3 We also just wanted to remind everyone to
4 please silence your cell phones if you have not already
5 done so. And I would also like to identify the FDA
6 press contact, Karen Riley.

7 Thank you.

8 And we did also want to make one announcement
9 that there will be a fire alarm test at 11:00 a.m. this
10 morning. It will last approximately 30 seconds.
11 There's no need for us to leave the room, but not to
12 alarm anyone, it will just be a test.

13 DR. DUTCHER: All right. We're going to
14 proceed with the meeting.

15 The first agenda item is the opening remarks
16 by Dr. Pazdur.

17 DR. PAZDUR: Good morning. The selection of
18 a drug based on a biomarker profile is desirable
19 because it may limit drug exposure to patients who will
20 benefit from drug treatment, may avoid drug use in
21 patients who may be harmed by drug treatment, or may
22 enhance safe use by optimizing drug dosing.

1 In the ideal case, the development of the
2 assay methodology for a biomarker should be an integral
3 part of the clinical drug development program. The
4 clinical studies required to establish the drug's
5 efficacy and those needed to establish the prognostic
6 and/or predictive value of the biomarker should occur
7 in tandem.

8 However, there are multiple examples of
9 retrospective or post hoc biomarker assessment. The
10 worst example involves a retrospective reanalysis of a
11 failed clinical trial, that is, a trial that did not
12 meet its primary endpoint. An attempt to salvage the
13 trial is made by examining non-pre-specified subgroups.

14 The FDA discourages such practice and this
15 practice should not be considered during this advisory
16 meeting discussion. However, FDA also recognizes that
17 there may be legitimate reasons for the lack of
18 consideration of biomarkers early in drug development,
19 primarily due to advances in the scientific knowledge
20 of a drug or disease occurring during drug development.

21 In today's meeting, the FDA seeks guidance
22 regarding how to incorporate new scientific information

1 without compromising our mandate to ensure that the
2 marketed drugs show substantial evidence of efficacy
3 and are safe.

4 The FDA and commercial sponsors during this
5 meeting will present a recent example of retrospective
6 biomarker analyses intended to support changes to
7 product labeling.

8 ImClone, the license-holder for Erbitux, and
9 Amgen, the license-holder for Vectibix, will describe
10 the results of retrospective analyses assessing
11 efficacy outcomes determined by KRAS genomic status.
12 We have asked these sponsors to present this data to
13 provide a context for the questions and discussions
14 posed to the committee.

15 The KRAS presentations provide a real world
16 situation faced by the FDA in which considerations of
17 the type and extent of data needed to support labeling
18 claims must be made. The issues posed to the committee
19 during the afternoon discussions deal with general
20 considerations of incorporating retrospectively
21 identified biomarkers in regulatory decisions rather
22 than the specifics of the KRAS example.

1 As previously stated, an ideal scenario is
2 one in which the relationship of the biomarker to the
3 potential action of the drug is recognized early.

4 Indeed, such a relationship might be the motivation for
5 starting the drug's development. In this setting, many
6 milestones for development of the in vitro diagnostic,
7 or IVD, might be reached in an orderly manner.

8 The identity of the biomarker should be
9 established early, along with reliable means for its
10 measurements. If the biomarker has an impact on the
11 natural course of the disease, being prognostic, such a
12 relationship might be elucidated.

13 Through preclinical studies and early
14 clinical trials, support might grow for the
15 applicability of the biomarker as an indicator of drug
16 effect. Formulation of an intended use of the
17 biomarker might emerge and resources are committed to
18 complete the analytical validation of a fully specified
19 in vitro diagnostic.

20 When a definitive efficacy trial for an
21 investigational drug is undertaken, its design might
22 incorporate a test of the IVD so that conclusions can

1 be drawn concerning both the drug's safety and efficacy
2 and the effectiveness of the in vitro diagnostic. With
3 a trial that is successful from all perspectives, the
4 drug will be approved and the test will be clinically
5 validated and approved for prediction of the drug
6 effect.

7 For many reasons, the ideal situation is
8 unusual. When a definitive efficacy trial has been
9 conducted and completed without reference to the
10 biomarker, there may be an intense interest to
11 retrospectively examine the biomarker in available
12 clinical trial specimens.

13 The follow-up of patients accrued to an
14 efficacy trial is already at hand. The patients who
15 accrue to the completed trial include both patients who
16 were, quote, "positive" and patients who were also
17 negative for the biomarker of interest, a likely
18 requirement for gaining insight on a predictive claim
19 for the in vitro diagnostic.

20 There are many issues to be addressed with
21 the strategy of retrospectively examining biomarker
22 data. Some of the points of discussion should include

1 the following.

2 First, the chance of erroneously concluding
3 that there is a real treatment effect, when, in fact,
4 it is not true, or the chance of concluding there is no
5 treatment effect, when, in fact, one actually exists,
6 are two critical concerns for the design and
7 interpretation of study results of any clinical trial.

8 There are many examples of sub-population
9 findings that are spurious. To address this problem,
10 it is necessary to control the chances of making these
11 false conclusions, usually by pre-specifying the
12 hypotheses and the number of subgroups for which a
13 treatment effect in a sub-population is sought as a
14 primary objective of the trial.

15 Secondly, an additional issue to be discussed
16 with the anticipated use of retrospective analysis is
17 the concept of replication, that is, the likelihood for
18 reproducing a treatment effect identified in a
19 subpopulation in a single trial in another independent
20 trial.

21 A third consideration in using retrospective
22 biomarker analysis is that the required sample size for

1 the biomarker negative population should be sufficient
2 to detect a treatment effect, if it exists, considering
3 that the effect may not be of the same magnitude as in
4 the biomarker positive subpopulation.

5 The minimum performance characteristics, for
6 example, sensitivity, specificity, reproducibility, of
7 the assay used to define patient subgroups and the
8 consequences of the performance for correct
9 decision-making and inferences from the study should be
10 understood. In addition, the proportion of patients
11 whose biomarker specimens are available for analysis
12 needs to be considered in any request for a
13 retrospective analysis of a completed trial.

14 Clarity on whether the biomarker is being
15 considered a prognostic and/or predictive marker and
16 the consequences of these definitions on study design
17 planning, sample size and the ability to draw
18 conclusions must be understood.

19 Lastly, there should be an understanding if
20 randomization has been preserved in a retrospective
21 analysis, especially if small sample size
22 subpopulations identified after completion of the

1 clinical trial are utilized.

2 In today's meeting, we will discuss the
3 concept of a, quote, "prospective/retrospective study."
4 A working definition follows.

5 In a completed or post-interim analysis
6 trial, biomarker samples were collected prior to
7 treatment initiation, whether or not full
8 ascertainment. The hypothesis is prospectively
9 specified prior to diagnostic assay testing. However,
10 the clinical outcome data without biomarker information
11 have been collected, un-blinded and analyzed. The
12 biomarker data analysis might arguably be prospectively
13 performed; however, this is a retrospective analysis.

14 In essence, in a prospective/retrospective
15 study, the classification factor or biomarker is not
16 known at the time of study initiation and the study is,
17 at first, not analyzed with that factor as part of the
18 hypothesis. The initial hypothesis and endpoints of
19 the study are not changed, except if pre-specified as
20 part of a planned adaptive study design.

21 The control of false positive conclusions
22 from the study are appropriately dealt with. The

1 randomization is not stratified on a biomarker status
2 as one of the hypotheses to be tested. Biomarker
3 should be ascertained at baseline on all subjects
4 randomized to treatment groups, ideally.

5 The FDA is seeking ODAC's deliberations on
6 issues raised in using biomarkers after trials have
7 been initiated or completed. In particular, the
8 committee should discuss the conditions where a
9 prospective/retrospective clinical study design may
10 provide evidence for treatment effects that are limited
11 to biomarker classified subpopulations, thereby being
12 judged as evidence of a predictive marker.

13 In addition, if a retrospective analysis can
14 be performed to show benefit in a subset and it is
15 considered acceptable that randomization on a biomarker
16 status was not done, what level of evidence should be
17 considered for reproducibility of the finding?

18 As stated previously, our purpose of
19 presenting KRAS data is to provide an illustrative
20 example of the complexities in decision-making
21 regarding retrospective analyses. We view these
22 discussions at today's ODACs to be an educational

1 dialogue, examining the incorporation of new scientific
2 information without compromising our mandate to ensure
3 that marketed drugs show substantial evidence of
4 efficacy and are safe.

5 Thank you.

6 DR. DUTCHER: Thank you, Dr. Pazdur.

7 Dr. Ruthann Giusti will present the FDA's
8 presentation and regulatory history.

9 DR. GIUSTI: Good morning. As Dr. Pazdur has
10 previously alluded to, the selection of a drug based on
11 a genomic biomarker profile has the potential to limit
12 drug exposure to patients who are most likely to
13 benefit from the drug, to avoid or modify drug use in
14 patients who are most at risk of developing
15 drug-related toxicities, and to optimize drug dosing
16 based on pharmacogenomic models of drug activation and
17 elimination.

18 In the ideal case, the development of the
19 genomic biomarker is an integral part of the clinical
20 development. The assay methodology for the assessment
21 of the biomarker is well characterized prior to
22 initiation of the randomized trial. The clinical trial

1 required to establish the efficacy of the drug and
2 those required to establish the prognostic and
3 predictive value of the genomic biomarker then can
4 proceed in tandem.

5 FDA strongly endorses this prospective and
6 scientifically guided approach to drug development as
7 part of the critical path initiative. This approach
8 permits labeling considerations to be data driven based
9 on clinical trial results.

10 However, increasingly, examples can be cited
11 of the reanalysis of clinical trials data to explore
12 efficacy in a subset of patients retrospectively
13 defined by a genomic biomarker, and FDA acknowledges
14 that there may be legitimate reasons for failure to
15 assess a genomic biomarker during the course of drug
16 development, the primary reason being that scientific
17 advances may outpace drug development and require a
18 redefinition of the original study question. However,
19 this is distinct from a post-hoc subgroup analysis in
20 an attempt to salvage a failed clinical trial. This is
21 a practice that is to be discouraged and one that the
22 FDA cannot endorse.

1 To provide a context for today's meeting, FDA
2 presents a real world example. Two anti-EGFR
3 antibodies have been approved for use in the United
4 States for the treatment of patients with metastatic
5 colorectal cancer.

6 These are ImClone's cetuximab and Amgen's
7 panitumumab. Based on retrospective analyses assessing
8 efficacy as a function of KRAS mutation subset, both
9 ImClone and Amgen propose to include information
10 concerning drug use in the subset of patients with
11 metastatic colorectal cancer whose tumors express
12 wild-type KRAS.

13 The basis for these proposals is the apparent
14 lack of efficacy of cetuximab and panitumumab in
15 patients whose tumors express a mutated version of
16 KRAS. This observation has been made in the
17 retrospective analyses of several completed randomized
18 trials.

19 While the example for today's discussion
20 involves KRAS, FDA seeks general guidance from ODAC
21 this morning concerning how to incorporate new and
22 evolving scientific information from retrospective

1 biomarker analyses into product labeling, this without
2 compromising FDA's legal mandate, which is to ensure
3 that marketed drugs show substantial evidence of
4 efficacy and are reasonably safe.

5 FDA hasn't advised ImClone and Amgen that the
6 optimal approach is to conduct a trial prospectively
7 designed to assess efficacy in patients who are
8 randomized based on KRAS genomic status, as determined
9 using a validated assay. However, the results of
10 recently completed retrospective analyses have been
11 widely disseminated and there is a perception within
12 the oncology community that the lack of efficacy of the
13 anti-EGFR therapies in patients whose tumors express a
14 mutated KRAS is well established, and conducting a
15 prospectively designed trial at this point in time may
16 no longer be feasible.

17 So given these practical constraints, FDA has
18 indicated that submission of a retrospective analysis
19 could be considered if the following conditions are
20 met.

21 These conditions are that the trial is
22 adequate, well controlled and well conducted; that the

1 sample size is large enough to approximate random
2 allocation for factors not used as stratification
3 variables for randomization; that an evaluable
4 biomarker result is obtained on a high number of
5 randomized participants to minimize selection bias;
6 that the biomarker assay has acceptable performance
7 characteristics in terms of sensitivity, specificity
8 and precision under the conditions proposed for
9 clinical use; the genomic analysis is performed by
10 individuals who are masked to treatment assignment and
11 to the clinical outcome in the original trial; and,
12 finally, that an acceptable plan for the biomarker
13 analysis has been pre-specified.

14 With these conditions in mind, I will now
15 review the randomized trials that have been
16 retrospectively submitted to the FDA in which efficacy
17 in the KRAS genomic subgroups has been assayed.

18 It must be noted that FDA has not reviewed
19 any of these studies in detail or verified the results
20 of any of the retrospective KRAS analyses that will be
21 presented today.

22 In April 2008, ImClone submitted summary

1 results from four large completed randomized clinical
2 trials of cetuximab in which available tumor tissue was
3 obtained and then retrospectively tested for KRAS
4 genomic status.

5 These trials include two trials of
6 chemotherapy with or without cetuximab in first line
7 metastatic colorectal cancer, the CRYSTAL and the OPUS
8 trials, the EPIC trial of irinotecan with or without
9 cetuximab in second line metastatic colorectal cancer,
10 and the NCIC trial of best supportive care with or
11 without cetuximab in third line therapy.

12 The primary study endpoint was met in only
13 two of these trials, the CRYSTAL trial and the NCIC
14 trial.

15 In this slide, and in subsequent slides,
16 those trials in which the primary study endpoint was
17 met have been highlighted. The NCIC study is the only
18 trial of an anti-EGFR antibody to have shown an overall
19 survival benefit in metastatic colorectal cancer.

20 These four studies have been retrospectively
21 analyzed to assess efficacy by treatment arm and by
22 KRAS genomic status. In these studies, the percent of

1 the originally randomized study population assessed for
2 KRAS genomic status ranged from 23 percent in the EPIC
3 trial to 69 percent in the NCIC and the OPUS trials.
4 Both direct sequencing and PCR-based methodologies were
5 used for retrospective KRAS testing.

6 Amgen has submitted summary results from the
7 retrospective analysis of two large randomized studies
8 in metastatic colorectal cancer, the 408 trial of best
9 supportive care with or without panitumumab in the
10 third line treatment of metastatic colorectal cancer
11 and the PACCE trial of chemotherapy with bevacizumab
12 with or without panitumumab in first line metastatic
13 colorectal cancer. The primary endpoint for both of
14 these trials was progression-free survival.

15 In the 408 trial, the primary endpoint for
16 the analysis was met, which resulted in the accelerated
17 approval of panitumumab. The PACCE trial failed to
18 meet its primary endpoint and the study was terminated
19 when statistically inferior progression-free survival
20 was observed in the panitumumab treated patients.

21 As part of an active biomarker exploration
22 program, Amgen acquired tissue on patients enrolled in

1 the 408, as well as other ongoing trials. In the 408
2 trial, tissue availability was required for study entry
3 and, consequently, KRAS genomic status was available in
4 a large proportion of both of these trials, over 90
5 percent in the 408 trial and 82 percent from patients
6 enrolled in the PACCE trial.

7 Both studies used a PCR-based assay to assess
8 KRAS genomic status. And while the assay used to
9 demonstrate KRAS subgroup was the prototype of the DxS
10 assay, the analytical performance of this assay had not
11 been characterized at the time. And the PACCE trial,
12 which failed to meet its primary endpoint, had the
13 largest number of patients tested for KRAS mutation
14 status of any of the retrospectively analyzed trials.

15 Even with a high rate of KRAS mutation
16 ascertainment, the number of patients within each
17 subgroup can be relatively small. For example, here,
18 despite KRAS mutation ascertainment in 92 percent of
19 the randomized population in the 408 trial, the number
20 of patients, when stratified by KRAS mutation and
21 treatment assignment, is approximately 100 patients in
22 each subgroup.

1 What general observations can be made about
2 the retrospective KRAS studies?

3 Summary data from the retrospective KRAS
4 analyses from six randomized studies in metastatic
5 colorectal cancer have been presented to the FDA.

6 The pre-specified primary endpoint for the
7 studies were met in only three of these trials, the
8 CRYSTAL, the NCIC, and the 408 trial. These studies
9 were heterogeneous with respect to the line of therapy,
10 additional treatments administered, and in the percent
11 of the originally randomized study population assessed
12 for KRAS. None of the studies were assessed using KRAS
13 assay methodology with well characterized analytical
14 performance.

15 Six studies were reported in which a time to
16 event endpoint was the primary endpoint. The direction
17 of interaction of effects favored the wild-type KRAS
18 group in three studies, the CRYSTAL, the NCIC and the
19 408 trial.

20 In these three trials, the originally defined
21 primary study endpoint was met. In two trials, the
22 EPIC and the PACCE trials, the direction of interaction

1 of effects did not favor the wild-type KRAS subgroup.
2 And in the PACCE trial, the direction of interaction of
3 effects favored the KRAS mutated subgroup. These two
4 trials failed to meet their primary originally defined
5 study endpoint.

6 The direction of and the interaction of
7 effects also favored KRAS in the OPUS -- the KRAS
8 wild-type subgroup in the OPUS trial, in which the
9 primary endpoint was response rate. However, this
10 study did not meet its primary endpoint.

11 In five of the six trials in which the
12 retrospective KRAS analysis reported progression-free
13 survival results, the direction of the interaction of
14 effects favored the KRAS wild-type subgroup.

15 In three of these trials, the CRYSTAL, the
16 NCIC, and the 408 trial, the study met the originally
17 defined primary study endpoint. The other three trials
18 failed to meet the originally defined study endpoint.

19 In the retrospective KRAS analysis, the
20 direction of the interaction of effects favored the
21 KRAS wild-type subgroup in both the EPIC and the OPUS
22 trial and, as previously stated, the direction of the

1 interaction of effects favored the KRAS mutated
2 subgroup in the PACCE trial.

3 Looking at the report of overall survival as
4 an endpoint in the retrospective analysis, the
5 direction of interaction of effects favored the
6 wild-type KRAS subgroup in two studies, the CRYSTAL and
7 the NCIC.

8 In the 408 and EPIC trials, the retrospective
9 KRAS analysis for overall survival, the interaction of
10 effects was neutral, and, in the PACCE, favored the
11 KRAS mutated subgroup. Overall survival results were
12 not provided for the OPUS trial.

13 What then is the potential to obtain
14 additional data to address the question of efficacy of
15 EGFR antibodies within KRAS defined genomic subsets?

16 Amgen has completed enrollment of two large
17 randomized trials and plans retrospective testing of
18 treatment effects by KRAS status in these studies.
19 These include a randomized study of FOLFOX with or
20 without panitumumab in first line metastatic colorectal
21 cancer and a study of FOLFIRI with or without
22 panitumumab in second line therapy.

1 The CALGB and NCCTG are currently actively
2 accruing two randomized trials. However, both trials
3 have recently been modified to limit enrollment to
4 patients with KRAS wild-type tumors through completion
5 of the trial.

6 This slide shows the accrual to these studies
7 at the time the decision was made to modify the
8 inclusion criteria. KRAS mutation testing of all four
9 of these trials will be done with the to be
10 commercialized DxS test kit.

11 Representatives from ImClone and Amgen will
12 present data concerning their retrospective analyses
13 and following these presentations, FDA will have two
14 additional presentations.

15 Dr. Robert Becker will discuss the optimal
16 approach to drug biomarker co-development, which
17 includes the prospective development of assay method
18 qualification and clinical validation.

19 Dr. Robert O'Neill will then discuss the
20 limitations associated with the use of retrospective
21 subset biomarker data.

22 We note again that the purpose of today's

1 discussion is not to render an opinion on the action
2 FDA should take on any given application. This can
3 only be completed upon a full review of the study data.
4 Rather, today, FDA asks ODAC to consider the following
5 four general questions.

6 First, when is it appropriate to limit the
7 use of a drug to a subset based on a retrospective
8 analysis of one or more studies that were not initially
9 designed for this purpose?

10 Secondly, when is a prospective study
11 designed explicitly for the purpose of examining
12 treatment effects in a predefined subgroup needed to
13 establish treatment effects in this group?

14 Third, what properties of clinical studies
15 originally designed for non-selected populations make
16 such studies unsuitable for demonstrating efficacy in a
17 biomarker subgroup?

18 And when is it acceptable to limit future
19 enrollment to a biomarker selected subgroup of an
20 actively accruing clinical trial based on data external
21 to that trial?

22 And, finally, we ask ODAC to consider the

1 importance of timing and rigor in determining the
2 analytical performance of the companion diagnostic
3 test.

4 Thank you.

5 DR. DUTCHER: Thank you very much.

6 We're going to move on to the sponsors'
7 presentations. The first presentation will be by
8 ImClone.

9 Dr. Youssoufian?

10 DR. YOUSSEOUFIAN: Good morning, ladies and
11 gentlemen, and thank you, Dr. Pazdur and Dr. Giusti,
12 for outlining these critical issues so well.

13 My name is Hagop Youssoufian from ImClone
14 Systems and, as of several weeks ago, a wholly-owned
15 subsidiary of Eli Lilly and Company.

16 I have the pleasure of representing our
17 extended family today, as well as our development
18 partner, Bristol-Myers Squibb, in this discussion.
19 We've been working in collaboration with the scientific
20 community and the FDA to refine the use of Erbitux in
21 patients with colorectal cancer. KRAS testing gives us
22 this opportunity. And as we describe our data today,

1 it is with the belief that the level of evidence
2 justifies its rapid implementation to identify the most
3 appropriate candidates for Erbitux therapy.

4 In this presentation, I will review the large
5 and consistent body of evidence on the predictive value
6 of KRAS for Erbitux therapy in colorectal cancer.

7 Secondly, I will touch on the impact of the
8 new KRAS data, which has been nothing less than
9 transformational in both academia and the medical
10 community.

11 Patients with KRAS mutated colorectal cancer,
12 as Dr. Giusti outlined, are no longer eligible for
13 clinical trials utilizing Erbitux. This development
14 has profound implications for the conduct of
15 prospective conformational studies.

16 Thirdly, we believe that the current labeling
17 of Erbitux should be revisited, and I will reflect on
18 ongoing discussions with the FDA to arrive at
19 appropriate next steps.

20 Briefly, Erbitux is a recombinant chimeric
21 monoclonal antibody that was first approved by the FDA
22 in 2004. Currently, Erbitux is indicated in EGFR

1 expressing colon cancer as monotherapy or in
2 combination with irinotecan, but without KRAS
3 selection, and in head and neck cancer.

4 The development of Erbitux has always
5 included a search for meaningful biomarkers. Although
6 I mentioned that EGFR expression by
7 immunohistochemistry is specified in our label for
8 colon cancer, most experts would agree that this is not
9 fully reliable. KRAS, by contrast, is a great example
10 of a biomarker that represents the convergence of a
11 molecular concept and clinical data.

12 From the work of Bert Vogelstein, it's been
13 known for over two decades that mutations in KRAS
14 represent early events in the pathogenesis of colon
15 cancer and are, thus, relevant to both early and
16 advanced disease. But more directly to the point, KRAS
17 plays a pivotal role in the antitumor effect of
18 anti-EGFR monoclonal antibodies.

19 This is illustrated on this slide. In the
20 wild-type state, the binding of cetuximab to EGFR can
21 shut down receptor-mediated signaling. However,
22 mutations in KRAS place the system in a perpetually

1 active state that is no longer responsive to cetuximab.
2 Hence, it follows from the biology that cetuximab
3 should work primarily in KRAS wild-type tumors.

4 In over the past two years, there's been a
5 profusion of reports demonstrating correlations between
6 KRAS status and efficacy. This slide summarizes
7 several such reports that come from a variety of
8 sources and geographies. This is not an exhaustive
9 list.

10 It is apparent that the response rates of
11 patients with KRAS wild-type tumors are superior to
12 those with KRAS mutated tumors. Collectively, these
13 large differences between wild type and mutant tumors
14 and the consistency among different studies began to
15 suggest that KRAS is a strong predictor of Erbitux
16 efficacy. It's also remarkable just how rapidly the
17 new KRAS data in colon cancer permeated the medical
18 community.

19 In the case of Erbitux, several milestones
20 are worth noting.

21 In April of 2008, we submitted
22 pharmacogenomic data to the FDA through the mechanism

1 of voluntary genomic data submission, or VGDS.

2 In May and June, there were a number of
3 presentations of novel data in both front line and
4 refractory settings.

5 As a consequence of these developments, NCI
6 CTEP issued an action letter to cooperative groups
7 requiring exclusion of patients with KRAS mutant colon
8 cancer from clinical trials of Erbitux-containing
9 regimens.

10 And within this timeframe, the European
11 indication of Erbitux in colon cancer was also revised
12 to target KRAS wild-type tumors only. This was
13 followed by a publication in the *New England Journal*
14 and by the NCCN issuing revised guidelines for
15 KRAS-based treatment with Erbitux.

16 Again, as Dr. Giusti outlined, the VGDS
17 included pharmacogenomic data from four
18 company-sponsored randomized trials, shown here.
19 Although KRAS analysis was done retrospectively,
20 archive tissue had to be available prospectively for
21 EGFR testing, which was then retrieved for KRAS
22 testing, and this was possible in 23 to 69 percent of

1 the cases. A common reason for our inability to obtain
2 tissue was the lack of informed consent.

3 The analysis was also planned prospectively,
4 with a specific marker, KRAS, and pre-specified cut
5 points. We also communicated data and performance
6 characteristics of two assay methodologies, indicated
7 here, that we used and compared those methods against a
8 test being developed by DxS.

9 The DxS test uses allele-specific PCR and
10 this test is currently undergoing review by the FDA.
11 The correlation in a limited number of samples that we
12 tested was high.

13 So I will now review these studies in greater
14 detail.

15 The NCIC CO.17 study forms by far the most
16 important foundation for how we think about the
17 clinical activity and validity of KRAS in Erbitux.

18 In this study, 572 patients with
19 EGFR-positive refractory colorectal cancer were
20 randomized to Erbitux with best supportive care or best
21 supportive care alone. The primary endpoint was
22 overall survival. Patients were not selected by KRAS

1 status. Nevertheless, in this KRAS unselected
2 population and for the first time, there was a
3 significant improvement in survival, with a median of
4 6.1 months and 4.6 months in favor of Erbitux,
5 translating to a 23 percent reduction in the risk of
6 death.

7 The NCIC made a concerted effort to collect
8 archived tissue for KRAS analysis. These samples,
9 again, were obtained at the time of diagnosis and
10 eventually retrieved from 69 percent of the randomized
11 patients, stored in a central repository maintained by
12 the NCIC, and then analyzed for mutations by
13 Bristol-Myers Squibb. The mutation frequency was 42
14 percent, which is consistent with the data and the
15 literature.

16 Now, in an effort to make the analysis as
17 rigorous as possible, it's important to take note of
18 the following.

19 First, KRAS analysis was performed in a
20 blinded fashion by a different party, BMS, and in
21 accordance with a statistical analysis plan written
22 before the mutation data was analyzed.

1 Second, every randomly assigned patient for
2 whom KRAS data was available was included in the
3 analysis.

4 And third, the focus of the analysis was
5 KRAS. This was not an unspecified, broad exploratory
6 program.

7 Baseline demographic features were comparable
8 between the overall and the KRAS evaluable populations
9 and within the KRAS subsets. Therefore, the data from
10 the KRAS subset should be a faithful reflection of that
11 of the total study population.

12 This slide shows the survival curves in the
13 KRAS evaluable population. Aside from the demographic
14 comparability, the KRAS evaluable and the ITT
15 populations were also comparable with regard to
16 outcome, in this case, the median survival times and
17 hazard ratios.

18 Now, for the key efficacy data within the
19 KRAS subset. This was analyzed by treatment arm.

20 In the KRAS wild-type group, the median
21 survival was 9.5 months with Erbitux versus 4.8 months
22 without Erbitux and the hazard ratio was 0.55 and the

1 P value, less than 0.0001. By contrast, there was
2 little, if any, effect attributable to Erbitux in the
3 mutant population. A formal interaction test between
4 KRAS status and Erbitux treatment yielded a P value of
5 0.01, indicating that KRAS status is a significant
6 predictor of survival with Erbitux treatment.

7 Now, given the overall survival advantage
8 already attributable to Erbitux in the unselected
9 population, the near doubling of survival with the
10 inclusion of KRAS information demonstrates a striking
11 enhancement of Erbitux activity that is clearly
12 attributable to Erbitux -- I'm sorry -- to KRAS.

13 We then asked whether a KRAS evaluable
14 dataset that represents 69 percent of the primary
15 analysis population is sufficiently robust to support
16 this conclusion. To do so, we made several extreme
17 assumptions about the missing data. I will give two
18 examples.

19 If we assume that all of the missing data
20 were from the KRAS mutant population, the interaction
21 test remains significant. Conversely, if the KRAS
22 wild-type population was the culprit for all missing

1 data, the interaction is also strong. These results
2 essentially imply that the KRAS data from the KRAS
3 evaluable population are so robust that even with
4 extreme assumptions, an evaluable sample size that is
5 large and otherwise satisfies the criteria that were
6 outlined previously is very likely sufficient to
7 justify our conclusions.

8 Besides survival, KRAS status also
9 consistently affected other efficacy endpoints. For
10 PFS, in the KRAS wild-type background, the effect size
11 increased from a median of 1.9 months in the control
12 group to 3.8 months with Erbitux, amounting to a
13 statistically significant 60 percent reduction in the
14 progression risk. No such effect was noted in the KRAS
15 mutant population. This interaction test was highly
16 significant.

17 Although not shown here, there were no
18 responders in the best supportive care group and only a
19 single responder in the KRAS mutant group treated with
20 Erbitux. All other responders were in the KRAS
21 wild-type group treated with Erbitux.

22 These data demonstrate robust internally

1 consistent effects with Erbitux that are essentially
2 restricted to patients harboring wild-type tumors.

3 While I won't discuss the safety data, by and
4 large, safety and tolerability were comparable between
5 the ITT and the KRAS populations, indicating that KRAS
6 selection enhances the already favorable risk-benefit
7 profile of Erbitux.

8 Importantly, the role of KRAS as a prognostic
9 marker can also be assessed cleanly in this setting
10 without a potential confounding effect by chemotherapy.
11 And as shown here, there were no differences in
12 survival between the wild-type and mutant populations,
13 indicating that KRAS is largely, if not purely, a
14 predictive marker of efficacy.

15 In summary, the NCIC data demonstrate
16 convincing, robust effects on the KRAS-dependent
17 efficacy of Erbitux in colon cancer. By virtue of the
18 fact that this was a monotherapy study, these data, we
19 believe, are the cleanest possible demonstration of the
20 impact of KRAS on Erbitux therapy.

21 Next, I will go on to describe additional
22 combination studies in the first and second line

1 settings that were also outlined by Dr. Giusti that
2 confirm and extend these data.

3 CRYSTAL was a first line study of
4 approximately 1,200 patients that compared in a
5 randomized fashion the benefit of adding Erbitux to
6 FOLFIRI in chemotherapy naive EGFR-positive metastatic
7 colon cancer. This was a positive trial in that the
8 primary endpoint of PFS, as assessed by an independent
9 review committee, was met. Survival was a secondary
10 endpoint. Since the submission of VGDS, our
11 development partner, Merck Serono, has provided
12 survival information and updated the KRAS data.

13 In the KRAS unselected population, there was
14 a statistically significant improvement in PFS, with a
15 median of 8.9 months versus eight months in favor of
16 Erbitux. Note the hazard ratio of 0.85 to which I will
17 return shortly.

18 Here, again, tissue for KRAS analysis was
19 obtained from archived material and the analysis
20 performed in a blinded fashion and under carefully
21 defined standard operating procedures. The tissue
22 collection rate was 45 percent and reasons for not

1 having a higher yield were very similar to those of the
2 NCIC study. The 36 percent frequency of KRAS mutation
3 was also similar. Baseline demographic characteristics
4 between the ITT and the KRAS population, as well as
5 within the KRAS evaluable subsets, were comparable.

6 First, and not shown on this slide, beside
7 the demographic comparability, in the KRAS evaluable
8 population, the hazard for PFS was 0.82 in favor of
9 Erbitux and you'll recall the hazard of 0.85 in the ITT
10 population, again, indicating comparability between
11 these populations and suggesting that selection bias is
12 limited.

13 As shown here, within the KRAS evaluable
14 subsets, the benefit of adding Erbitux to FOLFIRI was
15 limited to the KRAS wild-type population. The hazard
16 in this case was 0.68 compared to 0.85 in the ITT
17 population. Again, no benefit was apparent in the KRAS
18 mutant population with Erbitux above chemotherapy.
19 There was a strong association between KRAS status and
20 Erbitux treatment with a P value of 0.07.

21 Parenthetically, we've also performed sensitivity
22 analyses of missing data scenarios similar to the NCIC

1 study and the outcome, again, supports the predictive
2 value of KRAS in this setting.

3 With regard to secondary endpoints, the
4 median survival times in the ITT population were
5 19.9 months with Erbitux versus 18.6 months without
6 Erbitux, and this was not significant.

7 In the KRAS subtype, shown here, the median
8 survival times in the wild-type subset were 24.9 months
9 and 21.1 months with and without Erbitux, respectively,
10 which remained non-significant. But even here, it
11 appears that any potential impact of Erbitux on
12 survival might occur in the wild-type setting.

13 The overall response rate in the KRAS
14 wild-type setting was improved to an extent greater
15 than that in the ITT population, 59 percent versus
16 43 percent, without a similar effect in the mutant
17 population. The interaction test was significant.

18 Therefore, as in the NCIC monotherapy study,
19 the impact of KRAS is notable consistently across most,
20 if not all, efficacy endpoints and, in this case, the
21 KRAS dependency of Erbitux appears to be relevant to
22 front line therapy in combination with FOLFIRI.

1 The notion that KRAS is a predictive marker
2 of Erbitux efficacy, even in the setting of combination
3 therapy, is supported by OPUS, a Phase II study, in
4 which 337 unselected patients were randomized to first
5 line treatment with FOLFOX alone or in combination with
6 Erbitux. The overall response rate, the primary
7 endpoint was not met.

8 The strategy of tissue collection and KRAS
9 analysis were quite similar to those of other studies.
10 The tissue collection yield in this case was 69 percent
11 with a breakdown for wild-type and mutant populations
12 consistent with the other studies. The demographic
13 variables were comparable across all populations.

14 With the benefit of KRAS selection, the
15 response rate increased to 61 percent in the Erbitux
16 arm compared to 37 percent in the control arm, which
17 was significant. By contrast, patients with KRAS
18 mutations derived no such benefit.

19 Similarly, for PFS, which was identical for
20 both arms in the KRAS unselected population, an
21 improvement could be demonstrated only in patients with
22 wild-type tumors, with a 43 percent risk reduction in

1 progression and a P value of 0.016. The interaction
2 between KRAS status and each of these endpoints was
3 significant. Although this is a much smaller study,
4 the results are consistent with both the NCIC study and
5 CRYSTAL in the sense that the efficacy of Erbitux is
6 enhanced in the KRAS wild-type milieu.

7 This slide summarizes the main efficacy
8 endpoints from these three studies. Two met their
9 primary endpoints in the ITT population, while OPUS did
10 not. In all three studies, the ITT and KRAS
11 populations were comparable, both by baseline and key
12 efficacy characteristics.

13 KRAS selection led to notable enhancements of
14 the primary analysis outcomes, which were validated by
15 interaction tests, as well as the enhancements in
16 secondary efficacy endpoints in many cases, indicating
17 internally consistent results.

18 This last study that I wish to discuss is
19 EPIC, a second line Phase III study included in the
20 VGDS that randomized about 1,300 patients with
21 EGFR-positive colorectal cancer to Erbitux plus
22 irinotecan or irinotecan alone. It failed to meet its

1 primary endpoint of overall survival, but it is
2 instructive to look at this study mostly to point out
3 concerns that we all share about the adequacy of some
4 retrospective data.

5 Areas of concern, which were highlighted
6 before, included the following. Per protocol, tissue
7 for KRAS testing could only be obtained from U.S.
8 patients, about 23 percent of the overall population,
9 which differed from the overall population on certain
10 baseline characteristics.

11 Moreover, with regard to the primary efficacy
12 endpoint, the survival outcomes, shown here, in the ITT
13 and the KRAS evaluable populations were inconsistent
14 and, in fact, trended in different directions. This
15 may have been impacted by differences in crossover
16 rates.

17 Although the other outcomes may be
18 directionally consistent in the two populations, this
19 study serves to illustrate, perhaps too obviously, that
20 not all retrospective data can be given equal weight
21 and that distinctions must be made between studies with
22 larger sample retrievals and internally consistent

1 results, such as the randomized studies I have
2 presented earlier and others that lack those features.

3 I will now describe the impact of these data
4 on current research and clinical practice.

5 Following the 2008 ASCO meeting, NCI CTEP
6 issued an action letter to their investigators
7 requiring temporary suspension and amendments to
8 Erbitux-containing colon cancer trials. Three major
9 trials, shown here, all combination studies, were
10 affected.

11 Our companies endorsed this action for
12 sponsored trials and, at the same time, Merck Serono
13 and the European cooperative groups amended the
14 PETACC-8 adjuvant trial and a first line trial, COIN.

15 All of this was done rapidly in a highly
16 collaborative fashion. I'd like to use the intergroup
17 N-0147 adjuvant trial to illustrate this collaborative
18 dynamic.

19 The NCCTG proposed the amendment to limit
20 eligibility to KRAS wild-type tumors, changed the
21 primary analysis population to patients with wild-type
22 KRAS tumors, increased the sample size with power

1 calculations sufficient to detect a treatment effect in
2 the KRAS wild-type population, and incorporate KRAS
3 testing using the DxS kit.

4 At the same time that protocol changes were
5 being discussed with sponsors in CBER, CDRH was
6 evaluating the technical aspects of the assay itself
7 and agreed with DxS on a set of criteria that defined
8 technical success. So thanks to these efforts, accrual
9 on the amended study resumed quickly.

10 The status of the other studies is as shown.
11 COIN had already completed accrual, but the analysis
12 plan was revised to focus on the KRAS wild-type subset.

13 In time, these studies could provide
14 additional data relevant to today's discussion, but
15 their incremental value is unlikely to alter the
16 substantial body of knowledge that we believe is
17 already established.

18 The impact of KRAS testing is also
19 increasingly apparent at the community level. Only
20 last month, the NCCN issued new guidelines for
21 colorectal cancer that reflect these new realities, to
22 not only recommend KRAS testing in the workup of all

1 patients with metastatic cancer, colon cancer, but also
2 recommend Erbitux and Vectibix as single agents and
3 Erbitux in combination with irinotecan for patients
4 with wild-type KRAS tumors.

5 So in summary, although the current labeling
6 of Erbitux makes no mention of KRAS, we believe that
7 these data provide strong support to consider a
8 labeling change. The data are reproducible and
9 consistent.

10 With regard to the attributes of the
11 sponsored studies, we used all the mechanisms available
12 to us to carry out the tissue collection and the
13 analysis in a highly rigorous fashion.

14 I would again emphasize the following. The
15 cooperative groups and the sponsors spared no effort to
16 collect tissue. The NCIC and the CRYSTAL studies were
17 well conducted and well controlled, that were positive
18 studies.

19 The sample sizes of the KRAS evaluable
20 populations are sufficiently large to ensure
21 comparability of the KRAS evaluable and the ITT
22 populations both with respect to baseline

1 characteristics and overall outcome.

2 The analyses were performed in a blinded
3 fashion. There is consistency across different assays
4 with regard to their performance characteristics and
5 the additional validation work for DxS assay is in
6 progress.

7 Given these developments, prospective
8 comparisons of Erbitux activity in patients with
9 wild-type versus mutant tumors and stratified at
10 randomization by KRAS status are no longer possible.

11 So for all of these reasons, we believe the
12 timing is right to discuss a labeling change with the
13 FDA and, at a minimum, incorporate the NCIC KRAS data
14 in the labeling, perhaps in the clinical study section
15 of that label.

16 In closing, we strongly believe that these
17 data are clinically relevant and compelling and the
18 recent change in the NCCN guidelines is but one
19 affirmation. It's especially gratifying that all of
20 this is anchored firmly in science. And so I'd like to
21 end with an essential point.

22 While these data open a new era in the

1 treatment of colorectal cancer, we as sponsors cannot
2 effectively communicate them to practitioners without a
3 labeling change. A labeling change would certainly
4 make it congruent with state-of-the-art medical
5 practice.

6 We have submitted a meeting request to the
7 FDA and very much look forward to a continued dialogue
8 in the near future. In due course, our hope is that we
9 will be able to convey this information to physicians
10 in the most compliant and responsible way to enable
11 them to make the best risk-benefit assessments for
12 their patients.

13 Thank you very much.

14 DR. DUTCHER: Thank you very much.

15 We're going to go on to the second sponsor
16 presentation and another FDA presentation, and then
17 we'll have questions of the presenters.

18 Dr. Eisenberg is going to be presenting for
19 Amgen.

20 DR. EISENBERG: Dr. Dutcher, members of the
21 committee, thank you.

22 I'll be making some brief comments on behalf

1 of Amgen and then Dr. David Reese, a medical oncologist
2 in our Global Development Group, will walk through the
3 data with respect to KRAS and our data with panitumumab
4 monotherapy and combination therapy.

5 First of all, we appreciate the opportunity
6 to present our data today. Our data with respect to
7 the response of patients with colorectal cancer to
8 panitumumab with KRAS mutations, we believe, is
9 compelling and are supportive of the importance of KRAS
10 as a predictive biomarker.

11 We also appreciate the opportunity to have a
12 discussion, as FDA has framed it, around the general
13 technical considerations that one must consider in
14 evaluating the use of biomarkers in the treatment of
15 patients with cancer, particularly in an era where
16 emerging science almost always will be outpacing our
17 ability to test the results in rigorous clinical trials
18 in the manner that we ideally would consider
19 appropriate.

20 I think it's also important, and it was
21 implicit in the comments, actually explicitly stated by
22 the other sponsor, that the sponsors and FDA are,

1 obviously, interested in communicating benefit-risk to
2 patients and, clearly, the information we obtain from
3 biomarker data is an important component of that.

4 One component, as the committee is being
5 asked to address, are the technical considerations to
6 validate a biomarker. Another component, and one I
7 think that is reflected in the comments regarding how
8 clinical practice has already changed, is the fact that
9 medical judgment regarding weight of evidence plays an
10 important role in making decisions ultimately as to how
11 to conservatively communicate potential risks and
12 potential benefits. It's an important topic, one which
13 we take seriously.

14 With regard to the KRAS data, Amgen's
15 position is quite clear. We believe that there is a
16 predictive value to the KRAS mutation status and the
17 treatment of patients with colorectal cancer with
18 panitumumab as monotherapy should be restricted to
19 patients with wild-type KRAS or, that is, KRAS without
20 activating mutations. This is reflected in our product
21 label outside the U.S. We believe it should be
22 reflected in the U.S. label to avoid treatment of

1 patients who are highly unlikely to benefit as a
2 consequence of having activated mutations. Clearly,
3 the committee's deliberations, discussion will guide
4 both FDA and the sponsors in the most appropriate way
5 to communicate this to patients.

6 I'd like to now briefly provide you an
7 overview of what Dr. Reese will cover in greater detail
8 regarding the basis of our studies, which, as noted by
9 Dr. Giusti, were part of a program which allowed us to
10 have a very high ascertainment of KRAS status, because
11 we anticipated that there would be emerging data that
12 would inform the use of panitumumab in patients with
13 colorectal cancer and, therefore, included in our
14 trials, informed consent to allow collection of samples
15 as a criteria for entry and, therefore, a high
16 ascertainment of KRAS on archived samples from the
17 clinical program.

18 With regard to the KRAS analyses, we have
19 data from both Phase II and Phase III studies.
20 However, again, importantly, reflective of the comments
21 FDA has made regarding level of evidence, we'll discuss
22 the 408 study, which has already been alluded to, which

1 was the pivotal registration study, that is, the
2 randomized control study that led to accelerated
3 approval of panitumumab as monotherapy for treatment of
4 patients with metastatic colorectal cancer.

5 There was a pre-specified analysis plan, that is,
6 after initial validation, which Dr. Reese will review
7 for you, of the potential value of KRAS mutations in
8 determining response. A pre-specified analysis plan
9 was prepared prior to a blinded analysis of all of the
10 samples in the hypothesis testing study, which were
11 samples from the 408 study.

12 In addition, there was a fair amount of work
13 done to identify a reliable assay. You've heard
14 mention of the DxS assay. We have worked with DxS and
15 are happy to discuss that, if the panel wishes to
16 identify that assay, and then to use a single assay.
17 And, again, I believe Dr. Giusti appropriately noted
18 that in her presentation, a PCR-based assay to assess
19 KRAS status in the samples that were archived from 408.

20 All of our studies, the hypothesis testing
21 studies, as well as generation studies, have been very
22 consistent with regard to the high negative predictive

1 value, that is, that KRAS activating mutations predict
2 non-response in virtually all patients in which that
3 has been evaluated in the clinical programs with
4 panitumumab in the monotherapy setting.

5 Now, we recognize -- and, clearly, the
6 science outpaced the clinical study. So the predictive
7 value regarding KRAS potential in determining response
8 to panitumumab, those data emerged after the 408 study
9 was fully enrolled and, in fact, had been completed and
10 analyzed.

11 However, the work going on to validate KRAS
12 was occurring in parallel to the completion of that
13 study. And we also recognize the importance that when
14 a recommendation is made regarding benefit-risk to
15 patients, it has to be implementable; that is that
16 there has to be an FDA-approved test to allow a
17 clinician to have a high degree of certainty that if
18 we're making a recommendation to a patient, it's on the
19 basis of high evidence scientifically regarding the
20 test validity.

21 Outside the U.S., that test is available. It
22 is the DxS test, and FDA is currently working with DxS

1 to evaluate what the requirements would be for that to
2 be available in the U.S., though we would note that
3 there are other means through clear labs, pathology
4 labs, for testing for KRAS and, in fact, KRAS testing
5 clinically has been implemented, but recognize the
6 challenges FDA has in assuring that that clinical
7 implementation meets the reliability standards, and I'm
8 sure they'll revisit that in their presentation.

9 But ultimately, we believe the use of KRAS as
10 a predictive marker will improve the benefit-risk
11 profile for panitumumab monotherapy decisions regarding
12 treatment of patients with colorectal cancer. And I'll
13 have Dr. Reese now walk through the data that support
14 our position.

15 DR. REESE: Good morning. My name is Dave
16 Reese. I'm a medical oncologist in the Clinical
17 Development Group at Amgen, and I'm pleased to be here
18 with you today to share some of our data regarding KRAS
19 as a predictive biomarker for panitumumab.

20 This is a diagram that many of you will be
21 familiar with, illustrating EGFR signaling pathways.
22 When ligand binds to the EGFR, the receptor homo or

1 heterodimerizes. This causes intracellular tyrosine
2 phosphorylation and subsequent activation of a number
3 of downstream signaling pathways.

4 The RAS-MAP kinase pathway is one of the most
5 important effectors of EGFR signaling in colorectal
6 cancer. RAS was identified as a human oncogene more
7 than 25 years ago, when it was recognized that point
8 mutations in the molecule render it constitutively
9 active. Our understanding of this biology subsequently
10 gave rise to the hypothesis that upstream inhibitors of
11 EGFR, such as EGFR antibodies, would be ineffective in
12 tumors in the presence of RAS mutations.

13 With the availability of panitumumab and
14 cetuximab, a number of investigators began examining
15 KRAS as a potential predictive biomarker. Shown here
16 are representative studies. In these trials, patients
17 had received panitumumab or cetuximab alone or in
18 combination with chemotherapy.

19 As you can see in the middle of the slide,
20 approximately 30 to 50 percent of tumors bore KRAS
21 mutations, and, on the right of the slide, you can see
22 that the large majority of responses clustered in

1 patients with KRAS wild-type tumors. Some of the
2 responses reported in these studies in patients with
3 KRAS mutant tumors may have been due to concomitant
4 chemotherapy that some of these patients were
5 receiving. Well, these data, as they emerged, really
6 prompted us to focus on KRAS as a predictive biomarker
7 in the panitumumab development program.

8 This slide overlays key milestones from our
9 biomarker program, as well as the pivotal 408 study
10 that led to accelerated approval for panitumumab.

11 Shown at the bottom of the slide are key
12 events in the panitumumab development and regulatory
13 pathway. The 408 pivotal monotherapy trial completed
14 enrollment early in 2005 and ultimately formed the
15 basis for accelerated approval for panitumumab at the
16 end of 2006.

17 Beginning in 2004, as shown on the top of the
18 slide, we began exploring multiple biomarkers, multiple
19 targets across tumor indications within the panitumumab
20 program. With the emergence of Phase II data regarding
21 KRAS, we elected to focus on this potential predictive
22 biomarker and, first, performed an analysis of our

1 Phase II studies.

2 These analyses, on which we've provided
3 additional information in the briefing book, comprised
4 about ten percent of all patients enrolled in our
5 Phase II trials and indicated that responses were
6 reported only in patients with KRAS wild-type tumors.
7 Those data, in concert with data from the literature,
8 then prompted us to perform a comprehensive analysis of
9 the pivotal 408 trial for KRAS as a predictive
10 biomarker.

11 As you can see by the overlay of these
12 timelines, and as Dr. Pazdur has pointed out, this is a
13 real world example of how knowledge of a biomarker may
14 emerge in parallel with the development program of a
15 molecule.

16 Prior to conducting the analysis on our
17 Phase III study, we needed to select a KRAS assay. Our
18 goal here was really to identify an assay that could be
19 used in a high throughput fashion on routinely
20 available clinical specimens, specifically, formal and
21 fixed paraffin-embedded tumor samples that would be
22 normally obtainable from patients.

1 When thinking about KRAS testing, there are a
2 few important points to bear in mind. First, more than
3 95 percent of activating KRAS mutations affect Exon 2,
4 specifically codons 12 and 13.

5 Prior to analyzing our Phase III data, we
6 performed a comparison study using 40 commercially
7 available tumor samples that were sent to a variety of
8 independent laboratories to compare their assays
9 against bidirectional DNA sequencing performed at
10 Amgen. Bidirectional DNA sequencing is often regarded
11 as the gold standard in these sorts of cases.

12 Prior to the analysis of specimens in our
13 Phase III trial, the performance characteristics of the
14 specific assay that we selected, the DxS mutation test
15 kit, were determined by the manufacturer and, also,
16 independently validated in the central independent
17 laboratory performing testing on our specimens. As you
18 can see in the table at the bottom of this slide, the
19 DxS assay compared to bidirectional sequencing produced
20 concordant results in 39 out of 40 tumors.

21 I'd now like to turn to a description of our
22 Phase III monotherapy data.

1 This is the study schema, illustrating the
2 design for the 408 trial that led to accelerated
3 approval for panitumumab in 2006.

4 In this trial, patients with advanced
5 chemotherapy refractory colorectal cancer were
6 randomized to panitumumab or best supportive care. An
7 important design feature of this trial was an optional
8 crossover to panitumumab among patients initially
9 assigned to best supportive care at the time of disease
10 progression. In fact, more than three-quarters of
11 patients crossed over to panitumumab at a median time
12 to crossover of between seven and eight weeks. The
13 primary endpoint of this trial was progression-free
14 survival.

15 These are the PFS curves from the 408 study
16 that led to accelerated approval in 2006. As you can
17 see, panitumumab significantly improved
18 progression-free survival compared to best supportive
19 care with a hazard ratio of 0.54.

20 Well, with the emergence of the KRAS
21 hypothesis, we believed that the 408 study provided an
22 opportune setting to assess KRAS as a potential

1 predictive biomarker. The protocol had required tumor
2 samples for EGFR assessment. These were archived with
3 the specific intent of performing correlative biomarker
4 analyses in the future. In addition, based on an
5 anticipated high KRAS ascertainment rate, we believed
6 that the sample size would be sufficient to provide
7 balance between treatment arms. Importantly, KRAS was
8 the only biomarker tested in this particular exercise;
9 in other words, we were not examining a panel of
10 biomarkers, but, rather, KRAS alone.

11 Finally, statistical calculations had
12 indicated that we should have high power to test
13 whether KRAS was a predictive biomarker for
14 progression-free survival.

15 A statistical analysis plan was developed and
16 finalized prior to knowledge of any patient's KRAS
17 status. The objectives of this plan were, first, to
18 test whether the relative improvement in PFS was
19 greater in patients with KRAS wild-type tumors as
20 opposed to those with KRAS mutant tumors. Then we
21 would test specifically the treatment effect on PFS,
22 response and overall survival in the KRAS wild-type

1 stratum. An important feature of this statistical
2 analysis plan is that the analysis was designed to
3 control the overall Type I error rate for the set of
4 planned comparisons within the KRAS analysis.

5 What did we find? KRAS was ascertained in an
6 independent laboratory, blinded to treatment assignment
7 and clinical outcome. We obtained tumor samples from
8 96 percent of all patients enrolled in the trial. KRAS
9 testing was successful in a very high proportion of
10 those. Ultimately, KRAS results were available for
11 92 percent of all patients enrolled in the initial
12 study.

13 As you can see at the bottom of the slide,
14 the distribution of wild-type to mutant KRAS status is
15 as one would expect from the prior literature;
16 57 percent of patients had KRAS wild-type tumors.

17 Baseline demographic and tumor
18 characteristics were well balanced between treatment
19 arms when broken out by tumor KRAS status and treatment
20 assignment. Specifically, there were no imbalances for
21 known prognostic factors, such as age or performance
22 status.

1 This is probably the most important slide
2 that I'll share with you today. This shows
3 progression-free survival curves broken out by tumor
4 KRAS status and treatment assignment. On the left, in
5 patients with KRAS wild-type tumors, panitumumab
6 significantly improved progression-free survival with a
7 hazard ratio of 0.45. In contrast, in patients with
8 KRAS mutant tumors, there was no difference in
9 progression-free survival with a hazard ratio of 0.99.
10 As you can see, the PFS curves here largely overlap. A
11 quantitative interaction test evaluating a treatment by
12 KRAS interaction was highly significant, with a P value
13 of less than 0.0001.

14 Consistent with the progression-free survival
15 data were data from tumor response. Shown on this
16 slide are waterfall plots that graph the maximum
17 percent change in individual tumors from individual
18 patients. A green bar below the line indicates a
19 decline in tumor size. A blue bar above the zero line
20 indicates an increase in tumor size.

21 As you can see in the upper left corner,
22 61 percent of patients with KRAS wild-type tumors

1 receiving panitumumab experienced some degree of tumor
2 shrinkage. Declines in tumor size were quite unusual
3 in all three other groups. The partial response rate
4 was 17 percent among patients with KRAS wild-type
5 tumors receiving panitumumab. It was zero percent in
6 all other groups.

7 Well, we've shared with you efficacy data.
8 What about safety data, an important consideration when
9 we're evaluating overall risk-benefit.

10 This shows selected adverse events of
11 interest to EGFR inhibitors broken down by tumor, KRAS
12 status, and treatment assignment.

13 The first thing to note when assessing these
14 data is that patients with KRAS wild-type tumors
15 receiving panitumumab received, on average, double the
16 number of infusions as those with KRAS mutant tumors
17 receiving panitumumab.

18 There were some increases in the KRAS
19 wild-type group in specific events, such as skin rash,
20 diarrhea, or hypomagnesaemia. These are well described
21 side effects of EGFR inhibitors.

22 When we attempted to control for drug

1 exposure and analyze safety, for example, by looking
2 across the first eight weeks of therapy, as you can
3 see, the numerical differences between these groups
4 become much smaller.

5 We do not believe that there are any
6 intrinsic differences in the safety profile of
7 panitumumab between patients with KRAS wild-type tumors
8 and those with KRAS mutant tumors. There is nothing
9 about the biology of the disease that would lend us to
10 that speculation.

11 Well, once we had completed our Phase III
12 analysis, we wished to extend our understanding of KRAS
13 as a potential predictive biomarker. We performed a
14 pooled analysis of four monotherapy studies that had
15 been performed in the panitumumab development program.
16 The four trials comprising this analysis had very
17 similar eligibility criteria and all were conducted in
18 patients with advanced chemotherapy refractory disease.
19 KRAS was tested with the same methodology, the DxS kit,
20 in the same independent central laboratory, blinded to
21 clinical outcome, in each of these studies. A high
22 rate of KRAS ascertainment was achieved within each

1 study and overall, as I'll show you in a moment.

2 Finally, each study had consistent outcomes by KRAS
3 status.

4 These are the results from the pooled
5 analysis. The four studies included in this analysis
6 included the 408 pivotal trial, this is the study I've
7 just shown to you; the 194 trial, this was the
8 crossover trial available to patients in the pivotal
9 trial initially assigned to best supportive care; and,
10 two large Phase II monotherapy studies, the 167 and 250
11 trials. These trials were performed in patients whose
12 tumors expressed high or low or negative EGFR,
13 respectively. A total of 715 patients were ultimately
14 included in these analyses, representing a KRAS
15 ascertainment rate of 90 percent across all of these
16 studies.

17 The objective response rate was 14 percent in
18 patients with KRAS wild-type tumors and zero percent in
19 patients with KRAS mutant tumors. In fact, among 320
20 patients in the panitumumab development program, for
21 whom we have data and who have KRAS mutant tumors, no
22 objective responses have been recorded.

1 Finally, I'd like to conclude with a glimpse
2 of some of our plans for the future. These have been
3 mentioned by Dr. Giusti earlier, and I'd like to take
4 you through a few of the details regarding these
5 trials. They specifically focus on panitumumab in the
6 combination chemotherapy setting and in evaluation of
7 KRAS as a predictive biomarker.

8 This table illustrates the currently ongoing
9 or completed trials that are controlled studies that
10 will evaluate panitumumab in combination regimens and
11 have incorporated KRAS analysis into the study designs.

12 As Dr. Giusti has shown you, the 181 and 203
13 studies are fully enrolled, Phase III trials in the
14 second and first line settings, respectively. In these
15 trials, patients are randomized to a chemotherapy
16 regimen with or without panitumumab. We expect data
17 from these studies in 2009.

18 The 141 or SPIRIT trial is a randomized
19 Phase II study in which patients with second line
20 colorectal cancer receive FOLFIRI chemotherapy with
21 either panitumumab or bevacizumab. This trial is
22 currently enrolling. With the availability of some of

1 the data you heard earlier in the day, this trial has
2 been amended to enroll patients only with KRAS
3 wild-type tumors.

4 Finally, you've heard brief mention of the
5 249 or PACCE study. This was a trial that examined
6 chemotherapy with bevacizumab, with or without
7 panitumumab. This trial was terminated prematurely
8 when outcomes were reported to be inferior on an
9 interim analysis in the panitumumab arms. We've
10 provided additional data on this trial in the briefing
11 book and would be happy to take you through some of
12 that during the question-and-answer session, if you so
13 desire.

14 I'd like to turn now and describe the ongoing
15 pivotal trials.

16 As I noted, the 181 and 203 studies are fully
17 enrolled and we believe these will provide relatively
18 comprehensive information about KRAS as a predictive
19 biomarker for panitumumab in the combination
20 chemotherapy setting.

21 The 181 trial is being conducted in patients
22 with second line colorectal cancer. They are

1 randomized to receive FOLFIRI chemotherapy with or
2 without panitumumab. This trial completed enrollment
3 in the early part of this year, with just under 1,200
4 patients participating. We anticipate that data from
5 this trial will be available in 2009.

6 The statistical analysis plan for this study,
7 after consultation with the FDA and European regulatory
8 authorities, has been amended to focus the analysis on
9 an outcomes by KRAS analysis. Likewise, the 203 study,
10 which is being conducted in the first line setting,
11 randomizes patients to FOLFOX chemotherapy with or
12 without panitumumab.

13 This trial was also initiated and largely
14 enrolled as our Phase III data became available and
15 prior to any of the cetuximab combination therapy data
16 being available. We have also amended the statistical
17 analysis plan for this trial to focus on an outcome by
18 KRAS analysis. Together, these two trials enroll
19 nearly 2,400 patients and we believe will provide a
20 relatively definitive answer to the role of KRAS as a
21 predictive biomarker in the combination chemotherapy
22 setting for panitumumab.

1 In conclusion, I'd like to note that we all
2 share a number of goals in being here today. First, we
3 wish to avoid toxicity of drugs in patients who are
4 unlikely to benefit from those drugs. We believe that
5 in the monotherapy setting, our data are compelling and
6 consistent and that patients with KRAS mutant tumors
7 are very unlikely to derive benefit from therapy. We
8 are committed to enhancing benefit from therapy and
9 trying to get the right drug to the right patient at
10 the right time.

11 We have an obligation to ensure that KRAS
12 testing is available. We are working closely with the
13 FDA, our diagnostic partner, and our collaborators to
14 ensure that this happens as soon as possible. We
15 believe we have an obligation to provide reliable
16 information to physicians and patients, specifically,
17 through the label.

18 Based on the totality of the evidence that
19 we've presented, we believe that in the monotherapy
20 setting, the use of KRAS as a predictive biomarker will
21 improve the risk-benefit profile for panitumumab in
22 patients with advanced colorectal cancer.

1 It's been a privilege to be here with you
2 today. We hope these data are of some value to the
3 agency and the committee as we debate the optimal
4 development of biomarkers in oncology.

5 Thank you.

6 DR. DUTCHER: Thank you very much.

7 We now have two presentations by FDA
8 regarding design issues and study issues.

9 Dr. Becker will begin, followed by
10 Dr. O'Neill.

11 DR. BECKER: So this is an unusual example of
12 inter-center participation in an advisory council
13 meeting, and I think it reflects a welcome and growing
14 collaboration on important questions such as those that
15 you'll be considering today.

16 Good morning. I will present some issues
17 concerning drug device development from the perspective
18 of medical device regulation. I will touch on the
19 risk-based approach that CDRH takes to regulation of
20 medical devices as this applies to companion
21 diagnostics.

22 I will discuss some aspects of medical

1 devices for predictive claims for companion
2 diagnostics; that is, their use to distinguish patients
3 who will benefit by treatment with a drug from patients
4 who will not.

5 I will discuss best approaches to the
6 analytical validation of the companion diagnostic.
7 These topics will connect up with topics in
8 retrospective testing of clinical trial specimens in
9 order to clinically validate a drug-device pair.

10 Authority for FDA's regulation of medical
11 devices in the United States stems from the 1976
12 medical device amendments to the Federal Food, Drug and
13 Cosmetic Act. The standard for legal marketing is that
14 such devices are safe and effective, meaning there is
15 reasonable assurance that probable benefits outweigh
16 any probable risks, and that use of the device will
17 provide clinically significant results. These
18 assurances involve assessments of risk.

19 One element of risk for in vitro diagnostic
20 devices, or IVDs, is whether use of a device explicitly
21 drives a clinical decision. Important new devices are
22 now emerging in which the result from a single

1 laboratory test can rule in or rule out the use of a
2 specific potentially effective therapy for a
3 life-threatening disease. For companion diagnostics,
4 the risk for the test is equivalent to the risk for the
5 drug whose use is guided by the test result. Such
6 usage requires availability of an FDA-approved test for
7 the biomarker. Companion diagnostics are at the heart
8 of personalized medicine, with some uses that
9 Dr. Giusti has already described. Taken together,
10 these uses are meant to identify patients for whom drug
11 selection and dose ensure a satisfactory risk-benefit
12 ratio.

13 It is worthwhile to consider how biomarkers
14 can be helpful in this regard. Within broad classes of
15 ailment, we generally narrow the scope in which drugs
16 are studied. In cancer, for example, the focus may be
17 specific to an organ or tissue, such as colon or lung
18 or lymphoid. Further subdivision is commonly done; for
19 example, small cell versus non-small cell cancers in
20 lung.

21 The point is that these narrowings usually
22 follow extensive study and clinical confirmation and

1 serve to distinguish diseases that differ for features
2 like pathophysiologic mechanism, aggressiveness or
3 response to therapy.

4 In an important sense, each addition to our
5 growing portfolio of biomarkers can serve to further
6 sub-classify disease, with a caveat that the time and
7 study needed to understand many biomarkers'
8 significance have not yet been achieved before a drug
9 trial starts. Therefore, care is needed in applying
10 biomarkers for patient management. Incomplete
11 understanding of what the biomarker means can lead to
12 misperceptions about the best population for use of the
13 drug.

14 An example is in the selection of patients
15 for drug clinical trials. A trial designed under the
16 expectation that effectiveness of the drug depends on
17 expression of a biomarker might establish a benefit for
18 the studied subpopulation; that is, a biomarker
19 positive group. However, such a trial does nothing to
20 establish the biomarker's meaning. The presupposition
21 that biomarker positive patients are enriched for
22 responders remains a presupposition.

1 By itself, such a trial leaves the promise of
2 personalized medicine unfulfilled, since we obtain only
3 half an answer to the question of optimal drug use,
4 absent evidence establishing that biomarker negative
5 patients will not significantly benefit from the drug.
6 We really want to know whether biomarker expression
7 distinguishes patients who will benefit from patients
8 who will not benefit from the drug. This can be a
9 complex question to resolve, partly because there are
10 two ways in which the biomarker's expression might have
11 medical significance.

12 In one way, the biomarker predicts which
13 patients will benefit from the drug. For example,
14 patients who express the biomarker will benefit and
15 patients who do not express the biomarker will not
16 benefit. This is the essence of a predictive biomarker
17 claim and it requires comparison of the drug effect in
18 one biomarker defined group with the drug effect in
19 another biomarker defined group.

20 It is also feasible that the biomarker's
21 expression simply denotes the aggressiveness of the
22 disease without regard to how the disease is treated.

1 This is the essence of a prognostic biomarker claim, by
2 which the outcomes among a group of patients are
3 associated with the biomarker status. Prognostic
4 claims are not framed in terms of drug effect since
5 different treatments are not part of the outcome
6 comparisons.

7 For illustrative purposes, via a set of
8 sketches, let us presume that a new targeted drug has
9 beneficial effect compared to standard therapy for some
10 population. Perhaps a beneficial effect can be
11 detected even in an unselected population, as depicted
12 in this panel.

13 What follows are illustrations of three of
14 the many ways in which predictive and prognostic
15 effects might occur.

16 In the first example, a biomarker is
17 predictive for benefit in that marker positive patients
18 benefit from the targeted drug, while marker negative
19 patients do not.

20 As another illustration, it is also feasible
21 that a biomarker is simply prognostic, with no
22 predictive value. It is prognostic in that marker

1 positive patients do better than marker negative
2 patients. However, the drug effect does not differ
3 between the marker positive and marker negative
4 patients. Therefore, the biomarker has no predictive
5 effect and may be ineffective in assessing who should
6 or should not receive the drug.

7 It is also feasible that a marker is neither
8 predictive nor prognostic if the benefit from the
9 targeted drug does not vary between marker positive and
10 marker negative patients and there is no difference in
11 outcome between marker positive and marker negative
12 patients within treatment groups.

13 Examples of both success and challenges in
14 development of oncology drugs and companion diagnostic
15 tests are seen in our experience with receptor targeted
16 monoclonal antibodies and the IVDs for related
17 biomarkers.

18 Accrual of HER2 positive patients to the
19 pivotal Herceptin trials is widely credited with having
20 drastically shortened the time and expense needed to
21 demonstrate drug effect in marker positive patients.
22 Since marker negative patients were not studied, we do

1 not know well Herceptin's effect or lack of effect in
2 such patients. The HER2 biomarker test requires expert
3 visual interpretation of images and there are concerns
4 in recent years about the consistency with which those
5 interpretations are made. Further complications arise
6 with availability of multiple assays spanning multiple
7 technologies for presence of the HER2 protein or
8 amplification of the gene for it.

9 Lastly, the demonstration of safety and
10 efficacy for Herceptin in settings beyond the original
11 advanced stage cancer trials has relied on the same
12 HER2 tests and decision points as before. There is
13 little knowledge, and none reviewed by FDA, concerning
14 different performance of the HER2 assays for the
15 various drug indications.

16 Erbitux and Vectibix both came to market in
17 the United States on the basis of trials that accrued
18 marker positive patients; that is, patients whose
19 tumors expressed epidermal growth factor receptor. As
20 with HER2 testing, the biomarker assay requires visual
21 interpretation of complex images. The readout for EGFR
22 is perhaps even more challenging than for HER2 because

1 the cut point was set at expression of the marker by
2 just one percent of the tumor cells.

3 Over the last two years or so, there are
4 reports of responses to cetuximab by patients who did
5 not meet marker positive criteria, and these have
6 raised uncertainty about whether the biomarker is
7 helpful in discriminating patients who will or will not
8 respond to the drug.

9 The lessons to be drawn from these precedents
10 have been discussed and debated. The benefit is that
11 important new drugs are on the market. From the
12 medical device perspective, it is beneficial to have
13 biomarker assays marketed subject to quality system
14 practices, including post-market monitoring. However,
15 the claims for the devices are quite limited. In
16 particular, predictive claims are not established for
17 distinguishing patients who will benefit or not benefit
18 from the drugs.

19 Experience so far provides imperfect, but
20 important precedence. Looking forward, questions
21 remain concerning the extent of knowledge that is
22 needed for adequate analytical and clinical validation

1 of paired drugs and diagnostic devices.

2 Studies done ahead of a clinical drug trial
3 can establish a biomarker's role in the pathophysiology
4 of disease, perhaps including insights on disease
5 prognosis. This information might give ample incentive
6 for developing the drug to target the biomarker.
7 However, it cannot directly address whether a
8 particular biomarker test set for a particular cutoff
9 to distinguish marker positive from marker negative
10 disease distinguishes patients who will or will not
11 benefit from the drug.

12 This is the setting for our questions today
13 about the use of data from retrospective analyses of
14 clinical trials. In peeling off patient subsets from
15 clinical trials, developers of drugs and IVDs will aim
16 to establish claims for the biomarkers to guide the use
17 of the drugs. Insight and consensus are needed about
18 the extent of knowledge that is sufficient for
19 analytical and clinical validation of such uses.

20 I'll now discuss how and when, relative to
21 pivotal clinical trials, knowledge about satisfactory
22 performance characteristics for the biomarker test

1 should be obtained. In a broad sense, this knowledge
2 concerns the analytical validation of the medical
3 device.

4 There are hurdles that must be cleared in
5 order to interpret data from pivotal clinical trials
6 competently, regardless of whether the trials accrue
7 patient subpopulations or all comers. One hurdle is
8 that practical problems in obtaining the right kind of
9 specimen might occur. Pre-analytical variables, though
10 often neglected, have a large effect on results from
11 many tests. Clearly, alteration or degradation of the
12 biomarker during specimen acquisition, storage or
13 processing can raise implications about the meaning of
14 test results.

15 Also important are the effects of
16 sub-sampling the specimen for analysis and variation in
17 biomarker presence or state with different kinds of
18 samples or prior treatment of the patient.

19 Another hurdle is that the analytical
20 performance characteristics of the biomarker
21 measurement process might be insufficient to
22 distinguish benefitting from non-benefitting patients.

1 It is essential to understand and control the
2 analytical sensitivity, specificity, accuracy and
3 precision of the test.

4 For IVDs operating at very low levels of the
5 analyte, a stable limit of detection is critically
6 important. Reproducibility of analytical results
7 across different laboratories is a requirement for
8 medical devices that are fielded widely. In essence,
9 we need confidence that patient classifications
10 according to a biomarker cut point are consistent and
11 correct.

12 Other hurdles concern the biology of the
13 marker itself and the drug's interaction with the
14 biomarker or alternatives to interaction with it.
15 Presence of the biomarker analyte beyond the cells of
16 therapeutic interest may cloud the meaning of biomarker
17 measurements. Some biomarkers might vary widely in
18 their expression without proportional implications for
19 the effect of the drug. Finally, the biomarker might
20 relate to only one of several modes by which the drug
21 can act.

22 All of these considerations should be

1 resolved before the biomarker is brought into a pivotal
2 clinical trial to establish use of the biomarker in
3 guiding drug therapy.

4 All that I've said so far readily applies
5 when a new drug and a new biomarker test travel
6 together through a co-development effort. However,
7 there are various ways in which product development can
8 diverge from the idealized concurrent path.

9 One way is when there is late recognition of
10 the biomarker or its putative relevance to drug effect
11 so that a new test bears on the use of a previously
12 studied drug. This is the case with the anti-EGFR
13 monoclonal antibody drugs and KRAS testing discussed
14 today.

15 Another situation is when a biomarker that
16 has been previously well characterized and reduced to a
17 test approved for clinical use seems relevant to
18 development of a new drug. FDA has seen this situation
19 in evaluating next in class drugs targeted at a
20 previously studied biomarker. An example was the
21 review coordinated between CDER and CDRH for the new
22 drug application for Vectibix and the pre-market

1 approval supplement of the Dako pharmDx EGFR
2 immunohistochemistry assay. Vectibix carried
3 indications very similar to Erbitux and the Dako test
4 was unchanged from the version previously studied with
5 Erbitux. The PMA supplement review for the unchanged
6 Dako test was uncomplicated and rapid.

7 It is also feasible that a previously studied
8 drug will be reevaluated in the context of a previously
9 studied IVD test; for example, when the drug and the
10 test must be reviewed for a new indication. Changes in
11 indication have occurred already; for example, when
12 Herceptin was evaluated for use in adjuvant therapy of
13 breast cancer.

14 It is an open question whether the companion
15 diagnostic gives equivalent performance in selecting
16 different kinds of patients for Herceptin treatment.
17 To date, re-review of the companion diagnostic has not
18 been undertaken for such a situation.

19 These variant co-development situations may
20 ultimately prove to be more common than classical
21 concurrent development of new drugs and devices. It is
22 important to recognize and accommodate these

1 challenges, the challenges that they raise.

2 In cases like today's example with anti-EGFR
3 monoclonals and KRAS, analytical validation of the new
4 IVD test may come very late in the development process
5 and it is tempting for clinical validation plans to
6 rely on the analysis of specimens that were retained
7 from ongoing or already completed clinical trials.

8 I'll speak more about the analytical
9 validation timing and then briefly about some
10 retrospective trial analysis issues that will be
11 treated in more detail by Dr. O'Neill.

12 FDA strongly recommends that sponsors
13 complete the analytical validation of their IVD test
14 product before applying the test to specimens in the
15 clinical trials that would be submitted for review of
16 the drug and the test.

17 This is not to say that the device must be
18 locked down for specifications and performance very
19 early in the development program. Test performance
20 depends partly on recognizing what analytical
21 performance characteristics are needed and designing
22 the test to meet these needs. Substantial and often

1 iterative effort is spent during test development to
2 ensure that the test design yields analytical
3 performance characteristics that support the test's
4 clinical performance. Interim data may prompt changes
5 in test design, followed by another round of analytical
6 validation studies. The expense of these efforts
7 motivates sponsors to time them carefully.

8 It is essential to recognize that changes in
9 the design of the test and its analytical performance
10 characteristics become very suspect if they are
11 prompted by results from testing pivotal trial
12 specimens. Such changes are likely to raise the need
13 for additional clinical trials to establish the
14 performance characteristics of the test for use with
15 the drug.

16 Some test developers consider taking a
17 non-final version of the test into the clinical trial,
18 intending to reduce the test to a final version later,
19 in order to demonstrate clinical validity using pivotal
20 trial specimens.

21 Even assuming that final configuration of the
22 test is not contaminated by insights from the pivotal

1 trial material, this strategy carries significant
2 risks. One is that the final configuration of the test
3 might simply perform unacceptably worse than did the
4 earlier test version. Another risk is that the
5 specimens from the pivotal trial will be unavailable or
6 unsuitable for testing with the final device, so that
7 performance cannot be directly assessed. Bridge
8 studies using specimens somehow like those from the
9 clinical trial are a markedly less desirable approach
10 to validation.

11 There are special issues if a non-final
12 version of the IVD test is used as a gate in accruing
13 patients to the pivotal trial, as in, for example, a
14 trial confined to marker positive patients. Here is
15 why.

16 In accruing only patients who are marker
17 positive, according to a prototype IVD assay, patients
18 in blocks B and D go off study. When the final version
19 of the device is applied, patients in block C are also
20 removed from the analysis. This is not a problem if
21 the two device versions are perfectly concordant, since
22 then no patient will be in block B or block C.

1 To the degree that the two versions are not
2 concordant, either or both of these blocks will be
3 populated. Patients in block A will be the only ones
4 carried through the statistical analysis in a marker
5 positive study. Such patients might not be
6 representative of the patients in blocks A and B, who
7 will receive the drug if the drug and the final device
8 are approved.

9 An alternative would be to consider patients
10 who test positive with either version of the device as
11 positive for the biomarker. However, this does not
12 match the manner in which patients will be tested
13 post-approval since only the final IVD will be approved
14 and performance characteristics of the test will be
15 skewed by including block C patients in the analysis.

16 Summarizing my points on analytical
17 validation. Regardless of the co-development path
18 needed for a drug-device pair, it is highly desirable
19 to complete the specification and analytical validation
20 of the IVD before using the device to test clinical
21 trial samples. Strategies for very late analytical
22 validation, depending on like performance between the

1 final test and an earlier version, might raise
2 significant review issues.

3 As discussed already, trials that assess
4 efficacy of the drug in both marker positive and marker
5 negative patients are most informative. If the trial,
6 nevertheless, is designed to accrue only a subset of,
7 say, marker positive patients, then it is especially
8 desirable to avoid accruing patients based on one
9 version of the device with the intent of completing the
10 clinical validation using a second version of the
11 device.

12 You've previously seen this graphic depicting
13 an idealized pathway for developing the companion
14 diagnostic test along with the drug. Such a
15 development path provides for early identification of
16 the biomarker. It provides for exploration of the
17 biomarker's relevance to the disease and for relevant
18 measurement of the biomarker itself. It provides for
19 early assessment concerning relevance of the biomarker
20 to use of the drug and it provides for final assessment
21 of the drug and the analytically validated device in
22 pivotal trials.