CERTIFICATE OF ANALYSIS

Chain of Custody: 307491

Client: US Food & Drug Adminitration Address: Office of Cosmetics & Colors

4300 River Road

College Park, MD 20740

Attention: John Gasper

Job Name: Task 3 - Analysis of Official Samples

Job Location: 2nd Group - 10 Samples Job Number: CLIN 1 - Task 3 (10 Samples) PO Number: HHSF223201810337P Date Submitted: 5/23/019

Date Analyzed: 6/20/2019-6/26/2019

Report Date: 7/3/2019

Date Sampled: Not Provided

Person Submitting: Kepal Dewan/Steve Wolfgang

Revised: 8/30/2019, 3rd Revision

SUMMARY OF ANALYSIS

AMA	Client	TEM LOD	TEM LOQ	% Tremolite by TEM	% Chrysotile by TEM	% Total Tremolite & Chrysotile by TEM	%	%	% Acid	%	Community
Sample ID	Sample ID	Using ASTM D5756 Mass	Asbestos bv PLM	Organics	Soluable	Other	Comments				
		Calculation	Calculation	Calculation	Calculation	Calculation	Dy F LIVI				
307491-11	D-51	0 00000144%	0.00000575%	0 00025%	0.00190%	0.00215%	ND	17.8%	14.5%	67.7%	
307491-11A	D-51	0 00000133%	0.0000534%	0 00020%	0.00164%	0.00184%	ND	17.9%	19 2%	62.9%	
307491-11B	D-51	0 00000151%	0.00000602%	ND	0.00010%	0.00010%	ND	18.0%	13.9%	68.0%	

LOQ = Limit of Detection LOQ = Limit of Quantification ND = Not Detected PLM = Polarized Light Microscopy TEM = Transmission Electron Microscopy

Analytical Method(s): PLM by Modified NY ELAP 198.6

TEM by Modified NY ELAP 198.4/ASTM D5756

Analyst(s): PLM



Technical Director: Andreas Saldivar

All results are to be considered preliminary and subject to change unless signed by the Technical Director or Deputy

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Sample Preparation

Samples were prepared for PLM and TEM bulk analysis by Chon Simpha on May 22, 2019 through June 24, 2019. Sample preparation consisted of the following steps:

- 1) Label and weigh two 8mL glass vials for each sample in the set one vial for the PLM preparation and one vial for the TEM preparation.
- 2) Weigh out 0.1 to 0.8 grams of material and place in corresponding 8mL glass vial. Record weight.
- 3) Burn samples at 480° C for at least 12 hours.
- 4) Record Post-Ash Weight.
- 5) Treat ashed sample with concentrated hydrochloric acid.
- 6) Filter acid reduced material onto a pre-weighed 47mm 0.4um PolyCarbonate filter.
- 7) Place filter into drying oven for 30 minutes and then record Post-Acid Reduced weight.
- 8) Make four PLM slide preparations from the PLM residual ash for each sample in 1.550 dispersion oil. Make additional preparations in 1.605, 1.625, 1.680 and 1.700 dispersion oil as necessary for particle identification.
- 9) Weigh a portion of the residue from the TEM residual ash and place it into the corresponding pre-weighed 100ml jar.
- 10) Fill the 100ml jar with deionized water
- 11) Sonicate the jars for approximate 5-minutes.

- 12) Filter 0.2ml to 1ml of the solution onto a 47mm 0.22um MCE filter.
- 13) Dry the filter for 10 minutes then collapse, carbon coat, and place on a 3 TEM grids.

PLM Analysis

Analysis was performed in accordance with NY ELAP 198.6 protocols. The analysis was conducted using an Olympus BH-2 polarized light microscope (PLM) equipped with a dispersion staining objective. All four slide preparations for each aliquot were examined. 400-point count was performed for those samples on which asbestos or a regulated amphibole was observed. If no asbestos was detected on any of the slides, the percentage of fibrous components was determined by visual estimation. The results of this analysis are detailed below in the *Discussion and Interpretation of Analytical Findings* section for each individual sample.

TEM Analysis

Analysis was performed in accordance with modified NY ELAP Method 198.4 protocols. The analysis was performed using a JEOL JEM-100CX II transmission electron microscope (TEM), equipped with a Thermo Fisher Quest Energy Dispersive X-Ray Analyzer (EDXA), at magnifications of 19,000x. Two grids for each aliquot were examined. Twenty (20) grid openings per sample were examined.

Modifications to the NY ELAP 198.4 Method were:

- 1) The residue was not placed in alcohol and prepared using the quick drop method. To obtain a more uniform preparation, the residue was placed in a jar and filled with 100ml of deionized water. The jar was sonicated, and a portion of the solution was filtered onto a 47mm 0.22um MCE filter.
- 2) The tremolite and chrysotile were not visually estimated. The length and width of the observed particles were measured and the mass of each particle was calculated using the ASTM D5756 method. All particles identified as tremolite were included with the counts/concentrations, regardless of size and aspect ratio.

The results of this analysis are detailed below in the *Discussion and Interpretation of Analytical Findings* section for each individual sample.

Calculations

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ASTM D5756 Mass M = \pi/4 L * W^2 * D * 10^{-12} M = mass L = length W = width D = density Percent Calculation EFA(mm^2) * 100ml * MA(g) * RW(g) VF(ml) * IW(g) * AA(mm^2) * RJ(g) The calculated value is then multiplied by 100 to convert it to percent.
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EFA – Effective filter area
MA – Mass of asbestos
RW – Weight of residue
VF – Volume filtered
IW – Initial weight of the sample
AA – Area analyzed
RJ – Weight of residue placed into the jar
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Limit of Detection and Quantification

We used the mass of a 0.5 x 0.04-micron chrysotile fiber as the basis for our calculations. Limit of detection was defined as 1 fiber and limit of quantification was defined as 4 fibers.

Discussion and Interpretation of Analytical Findings:

PLM

All three aliquots of sample D-51 were analyzed by Peerawut Chaikeenee on June 26, 2019. No asbestos or non-asbestos amphibole variants were detected the samples. The results were calculated using the equations detailed in the calculations section.

307491-11	NAD
307491-11A	NAD
307491-11B	NAD

TEM

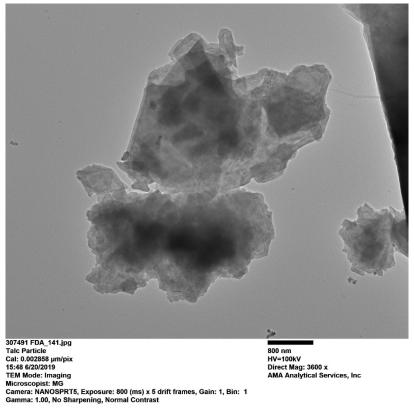
Michael Greenberg analyzed sample 11 on June 20, 2019, 11A on June 24, 2019 and 11B on June 25, 2019. The sample consisted of talc particles and mica particles. Chrysotile was observed on all three aliquots. One tremolite particle of was observed on aliquot 11 and aliquot 11A. No tremolite was observed on aliquot 11B. The results were calculated using the equations detailed in the calculations section.

307491-11	0.00215%
307491-11A	0.00184%
307491-11B	0.00010%

The original preparation of 307491-11B did not have an even particulate distribution on the filter. A new preparation of 11B was made. Out of precaution, additional preparations of 11 and 11A were also made. Analysis was performed on the original preparations of 11 and 11A. For 11B, the second preparation was analyzed.

Below are pictures, diffraction patterns, and chemistry from some of the observed particles. The unidentified peaks in chemistry spectra are copper, zinc, and carbon. Those peaks are from the TEM specimen holder and specimen grid.

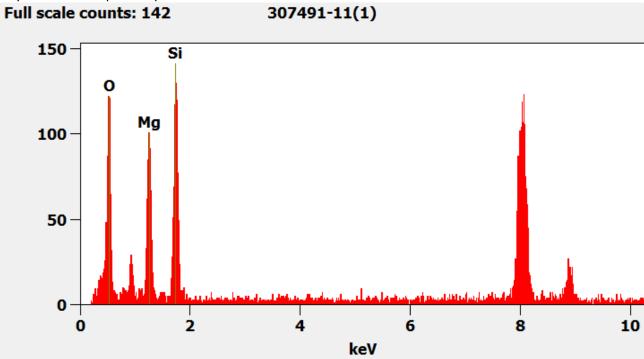
Talc particle from sample 307491-11



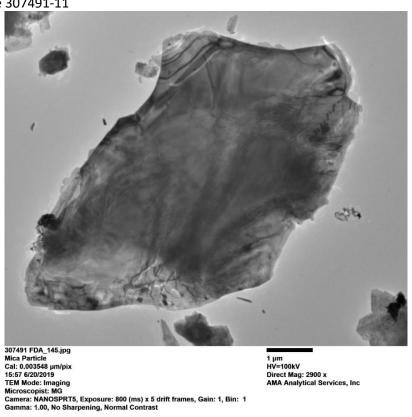
Diffraction pattern for the talc particle pictured above.



Chemistry from the talc particle pictured above.



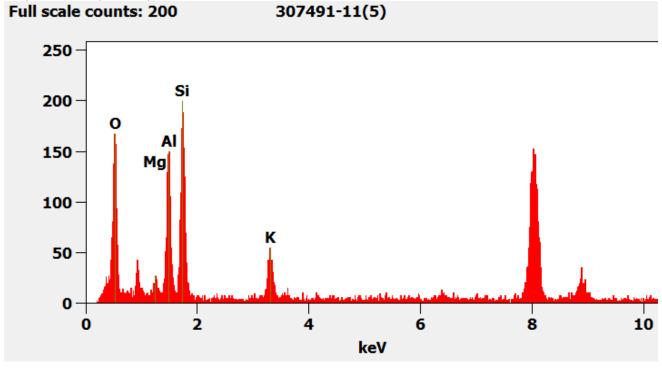
Mica particle from sample 307491-11



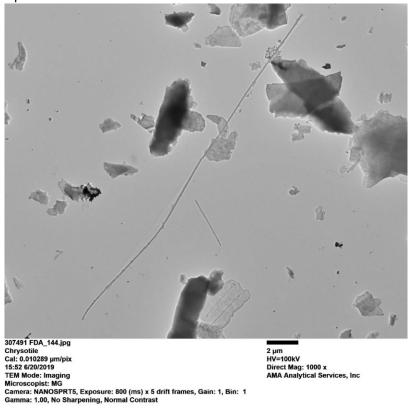
Diffraction pattern from mica particle pictured above.



Chemistry from mica particle pictured above.



Chrysotile particle from sample 11



Diffraction pattern from chrysotile pictured above.

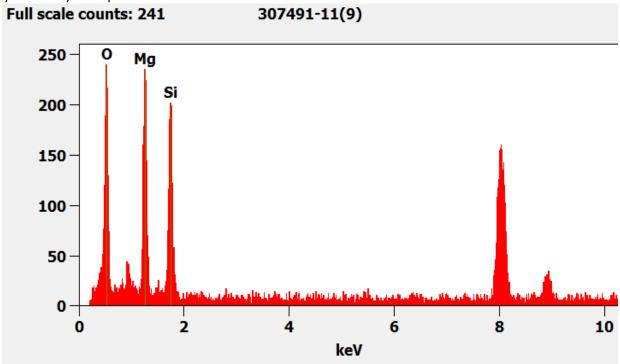


307491 FDA_143.jpg Chrysotile Diffraction 15:51 6/20/2019

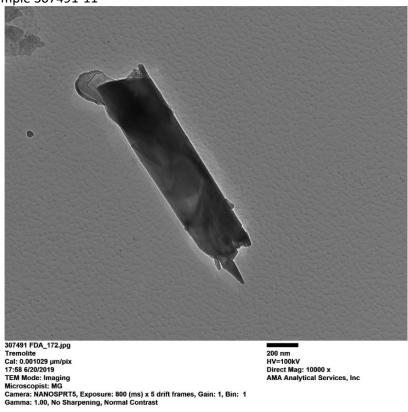
15:51 6/20/2019
TEM Mode: Diffraction
Microscopist: MG
Camera: NANOSPRT5, Exposure: 800 (ms) x 5 drift frames, Gain: 1, Bin: 1
Gamma: 1.00, No Sharpening, Normal Contrast

100 (1/Å) HV=100kV Cam Len: 0.2200 m AMA Analytical Services, Inc

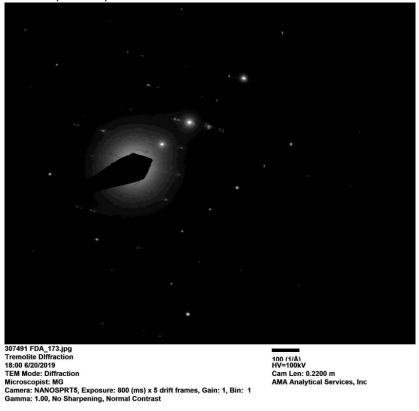
Chemistry from chrysotile pictured above.



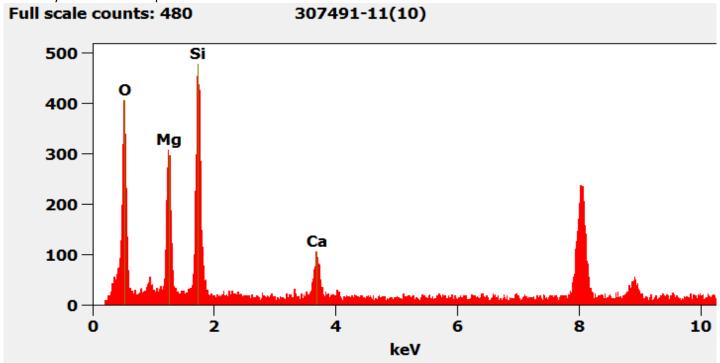
Tremolite particle from sample 307491-11



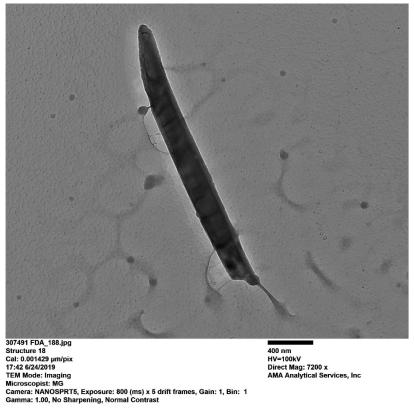
Diffraction pattern from tremolite particle pictured above.



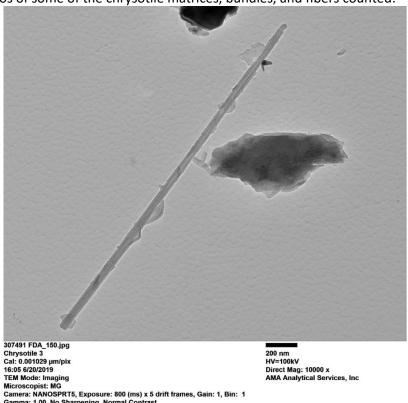
Chemistry from tremolite pictured above.

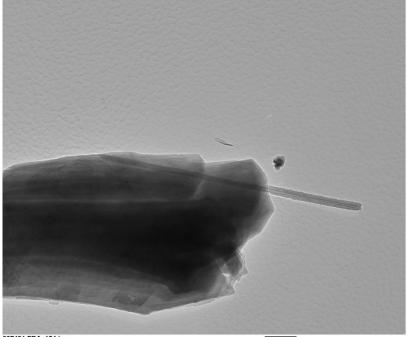


Tremolite particle from 307491-11A



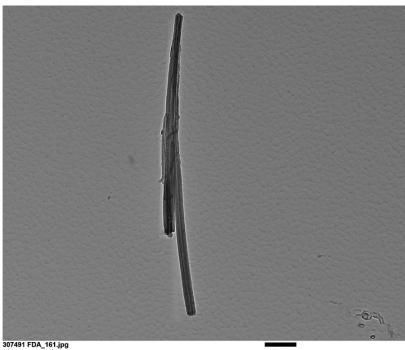
Below are additional photos of some of the chrysotile matrices, bundles, and fibers counted:





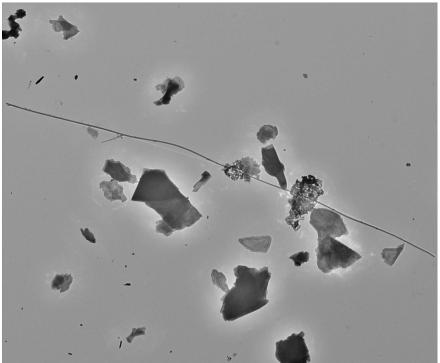
307491 FDA_154.jpg
Chrysotile 4
Cal: 0.001029 µm/pix
16:16 6/20/2019
TEM Mode: Imaging
Microscopist: MG
Camera: NANOSPRT5, Exposure: 800 (ms) x 5 drift frames, Gain: 1, Bin: 1
Gamma: 1.00, No Sharpening, Normal Contrast

200 nm HV=100kV Direct Mag: 10000 x AMA Analytical Services, Inc



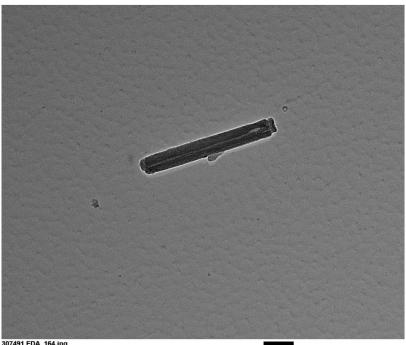
307491 FDA_161.jpg
Chry 9
Cal: 0.001029 µm/pix
16:32 6/20/2019
TEM Mode: Imaging
Microscopist: MG
Camera: NANOSPRT5, Exposure: 800 (ms) x 5 drift frames, Gain: 1, Bin: 1
Gamma: 1.00, No Sharpening, Normal Contrast

200 nm HV=100kV Direct Mag: 10000 x AMA Analytical Services, Inc



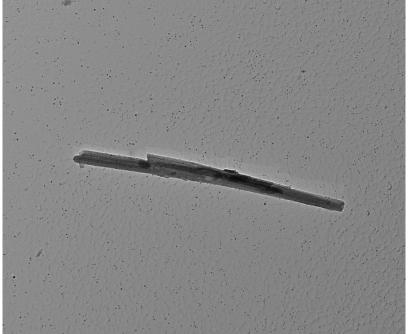
Direct Mag: 1400 x AMA Analytical Services, Inc

307491 FDA_162.jpg
Chry 10
Cal: 0.007349 µm/pix
16:33 6/20/2019
TEM Mode: Imaging
Microscopist: MG
Camera: NANOSPRT5, Exposure: 800 (ms) x 5 drift frames, Gain: 1, Bin: 1
Gamma: 1.00, No Sharpening, Normal Contrast



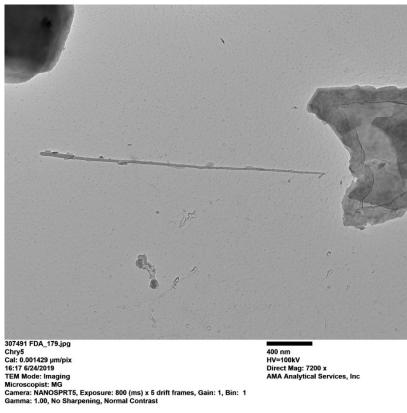
307491 FDA_164.jpg
Chry 18
Cal: 0.541520 nm/pix
16:43 6/20/2019
TEM Mode: Imaging
Microscopist: MG
Microscopist: MG
Camera: NANOSPRT5, Exposure: 800 (ms) x 5 drift frames, Gain: 1, Bin: 1
Gamma: 1.00, No Sharpening, Normal Contrast

100 nm HV=100kV Direct Mag: 19000 x AMA Analytical Services, Inc

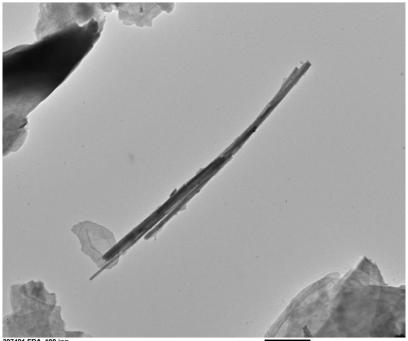


307491 FDA_178.jpg
Chry2
Cal: 0.001029 µm/pix
16:02 6/24/2019
TEM Mode: Imaging
Microscopist: MG
Camera: NANOSPRT5, Exposure: 800 (ms) x 5 std. frames, Gain: 1, Bin: 1
Gamma: 1.00, No Sharpening, Normal Contrast

200 nm HV=100kV Direct Mag: 10000 x AMA Analytical Services, Inc

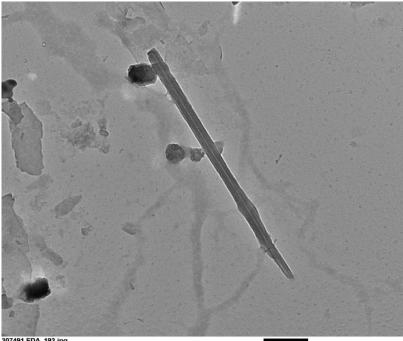


400 nm HV=100kV Direct Mag: 7200 x AMA Analytical Services, Inc



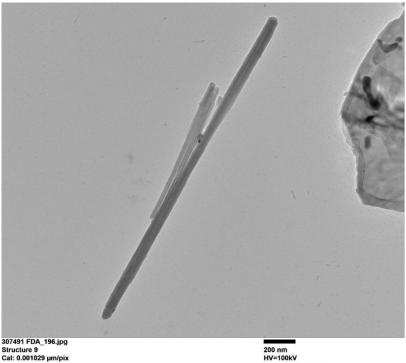
307491 FDA_190.jpg
Structure 19
Cal: 0.002858 µm/pix
17:52 6I24/2019
TEM Mode: Imaging
Microscopist: MG
Camera: NANOSPRT5, Exposure: 800 (ms) x 5 drift frames, Gain: 1, Bin: 1
Gamma: 1.00, No Sharpening, Normal Contrast

800 nm HV=100kV Direct Mag: 3600 x AMA Analytical Services, Inc



307491 FDA_193.jpg
Structure 7
Cal: 0.734921 nm/pix
10:18 6/25/2019
TEM Mode: Imaging
Microscopist: MG
Microscopist: MG
Gamera: NANOSPRT5, Exposure: 800 (ms) x 5 std. frames, Gain: 1, Bin: 1
Gamma: 1.00, No Sharpening, Normal Contrast

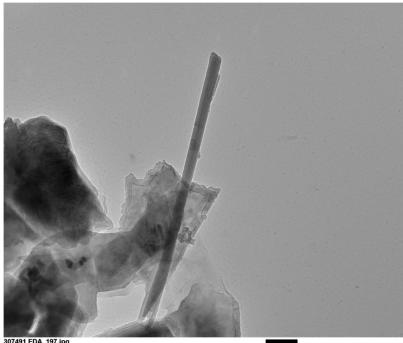
200 nm HV=100kV Direct Mag: 14000 x AMA Analytical Services, Inc



Structure 9 Cal: 0.001029 µm/pix 10:36 6/25/2019

10:36 6/Z5/Z019
TEM Mode: Imaging
Microscopist: MG
Camera: NANOSPRT5, Exposure: 800 (ms) x 5 drift frames, Gain: 1, Bin: 1
Gamma: 1.00, No Sharpening, Normal Contrast

200 nm HV=100kV Direct Mag: 10000 x AMA Analytical Services, Inc



307491 FDA_197.jpg Structure 15 Cal: 0.001029 μm/pix 11:01 6/25/2019

11:01 ot.2012/13 TEM Mode: Imaging Microscopist: MG Camera: NANOSPRT5, Exposure: 800 (ms) x 5 drift frames, Gain: 1, Bin: 1 Gamma: 1.00, No Sharpening, Normal Contrast

200 nm 200 nm HV=100kV Direct Mag: 10000 x AMA Analytical Services, Inc

QC Discussion:

During preparation, one blank control sample and one reference control sample were prepared. These samples were prepared alongside the customer samples. The blank sample was prepared using Sigma-Aldrich Talc Powder, <10



micron. No asbestos was detected on the blank sample. The reference sample was made from the same Sigma-Aldrich talc powder spiked with 1% Chrysotile. The reference sample was analyzed and found to be within acceptable limits.

Our LIMS randomly selects samples for additional replicate and duplicate QC. 307491-11, 11A, and 11B/D-51 were not selected for any additional QC analysis.

Attachments:

The following items are attached to this case narrative for your reference:

- 1) Sample Log-In Sheet
- 2) Daily PLM Scope Calibration Log
- 3) Refractive Index Oil Calibration Log
- 4) Daily TEM Scope Calibration Log
- 5) QC Results Summary
- 6) Raw Data Sheets
 - a. Gravimetric Data
 - b. Filtration Worksheets
 - c. PLM Analysis
 - d. TEM Analysis
 - e. QC Samples

I certify that all information contained in this report pertaining to laboratory events, procedures, and protocols is true and accurately describes the handling of this project by AMA Analytical Services, Inc. and its personnel.

7/9/2019

Andreas Saldivar Laboratory Director Date