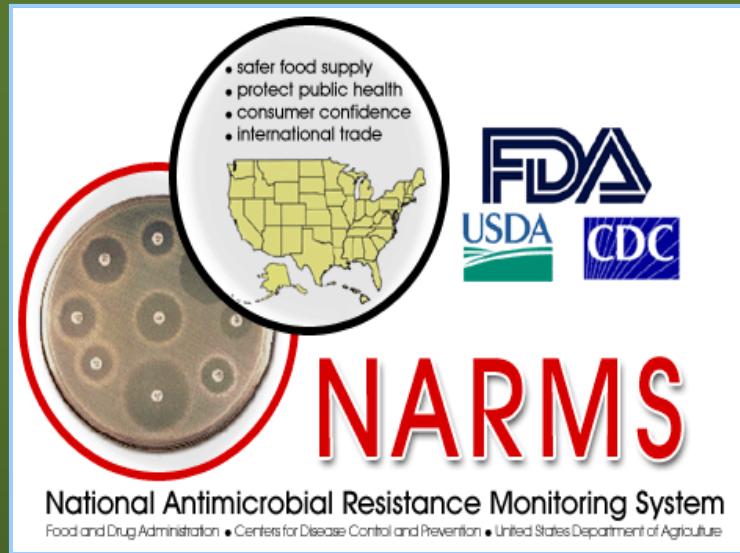


NARMS Retail Meat Annual Report, 2004



Enable informed decision making

Prolong the efficacy and useful life of antimicrobials

Guide prescription practices

Encourage standardization of laboratory techniques

Identify areas for more detailed investigation

Promote collaboration



ABBREVIATIONS USED IN THE REPORT, 2004

AR	Antimicrobial Resistance
BAP	Blood Agar Plate
CCA	Campy-Cefex Agar Plate
CDC	Centers for Disease Control and Prevention
CVM	Center for Veterinary Medicine
EAP	Enterococcose Agar Plate
EIP	Emerging Infections Program
EMB	Eosin Methylene Blue
FDA	Food and Drug Administration
FoodNet	Foodborne Disease Active Surveillance Network
MIC	Minimum Inhibitory Concentration
NARMS	National Antimicrobial Resistance Monitoring System
CLSI	Clinical and Laboratory Standards Institute
PCR	Polymerase Chain Reaction
PFGE	Pulsed Field Gel Electrophoresis
PulseNet	The National Molecular Subtyping Network for Foodborne Disease Surveillance
QC	Quality Control
RVR10	Rappaport-Vassiliadis
USDA	United States Department of Agriculture
XLD	Xylose Lysine Deoxycholate

Antimicrobial Abbreviations:

AMC	Amoxicillin/Clavulanic Acid	FOX	Cefoxitin
AMI	Amikacin	GEN	Gentamicin
AMP	Ampicillin	KAN	Kanamycin
AXO	Ceftriaxone	LIN	Lincomycin
AZI	Azithromycin	LZD	Linezolid
BAC	Bacitracin	NAL	Nalidixic Acid
CHL	Chloramphenicol	NIT	Nitrofurantoin
CIP	Ciprofloxacin	PEN	Penicillin
CLI	Clindamycin	QDA	Quinupristin/Dalfopristin
COT	Trimethoprim/Sulfamethoxazole	STR	Streptomycin
DAP	Daptomycin	TEL	Telithromycin
ERY	Erythromycin	TET	Tetracycline
FFN	Florfenicol	TYL	Tylosin
FIS	Sulfisoxazole	TIO	Ceftiofur
FLA	Flavomycin	VAN	Vancomycin

Meat Types

CB	Chicken Breast	GT	Ground Turkey
GB	Ground Beef	PC	Pork Chop

State Abbreviations:

CA	California	MN	Minnesota
CO	Colorado	NM	New Mexico
CT	Connecticut	NY	New York
GA	Georgia	OR	Oregon
MD	Maryland	TN	Tennessee

NARMS Retail Meat Annual Report 2004

Background:

Food animal products destined for human consumption are known to harbor enteric bacteria, including zoonotic foodborne pathogens. Antimicrobial resistance (AR) among these organisms may be associated with the use of antimicrobial agents in food animals. Retail meats represent a point of exposure close to the consumer and, when combined with data from slaughter plants and on-farm studies, provides insight into the prevalence of AR in foodborne pathogens originating from food animals. To gain a better understanding of AR among enteric bacteria in the food supply, the NARMS monitors antimicrobial susceptibility/resistance phenotypes in bacteria isolated from retail meats.

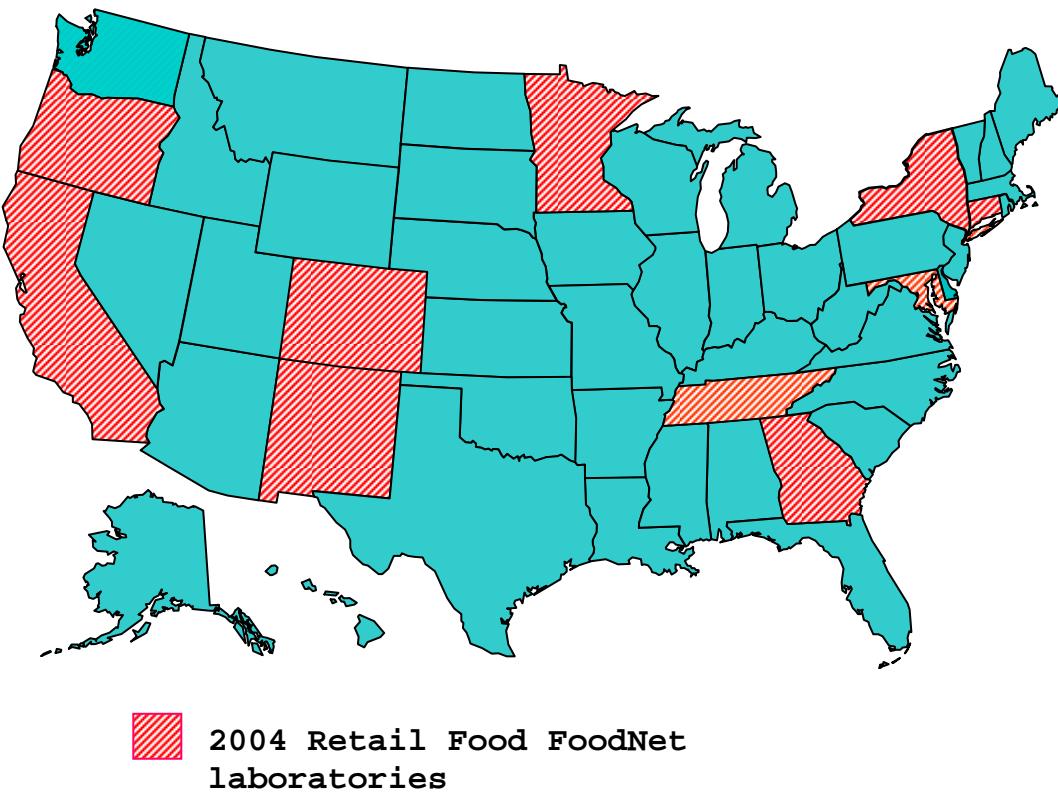
The primary purpose of the NARMS retail meat surveillance program is to monitor the prevalence of antimicrobial resistance among foodborne pathogenic and commensal organisms, in particular, *Salmonella*, *Campylobacter*, *Enterococcus* and *E. coli*. The results generated by the NARMS retail meat program will establish a reference point for analyzing trends of antimicrobial resistance among these foodborne bacteria. NARMS retail meat surveillance is an ongoing collaboration between the U.S. Food and Drug Administration (Center for Veterinary Medicine), the Centers for Disease Control and Prevention, and in 2004, all 10 of the current FoodNet laboratories: California, Colorado, Connecticut, Georgia, Maryland, Minnesota, New Mexico, New York, Oregon, and Tennessee. Retail meats are collected at these FoodNet sites and cultured for the presence of the selected organisms. Bacterial isolates are sent to FDA/CVM for confirmation of species, antimicrobial susceptibility testing, and genetic analysis.

NARMS Retail Meat Annual Report 2004

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FoodNet is the principal foodborne disease component of CDC's Emerging Infections Program (<http://www.cdc.gov/foodnet/>). It is a collaborative project of the CDC, ten EIP sites (California, Colorado, Connecticut, Georgia, New York, Maryland, Minnesota, Oregon, Tennessee, and New Mexico), the [U.S. Department of Agriculture \(USDA\)](#), and the [Food and Drug Administration \(FDA\)](#). The project consists of active surveillance for foodborne diseases and related epidemiologic studies designed to help public health officials better understand the epidemiology of foodborne diseases in the United States. The NARMS/FoodNet Retail Food Study was developed to monitor the presence of AR among *E. coli*, *Salmonella*, *Campylobacter*, and *Enterococcus* from convenience samples of fresh meat and poultry purchased monthly from grocery stores in the participating States. These isolates are then subjected to standardized antimicrobial susceptibility testing methods in order to determine the prevalence of resistance.

Retail meat sampling:

For calendar year 2004, retail meat sampling started in January among the 10

participating FoodNet laboratories. In each of the FoodNet sites monthly sampling, an attempt was made to go to as many different stores as possible. The object was to purchase as many different brands of fresh (not frozen) meat and poultry as possible. A total of 40 food samples were purchased per month comprised of 10 samples each of chicken breast, ground turkey, ground beef, and pork chops. For each meat and poultry sample, the FoodNet sites recorded the store name, brand name, lot number (if available), sell-by date, purchase date and lab processing date on log sheets ([A-9](#)). Additional information with regard to whether or not the meat or poultry was ground or cut in-store was also collected, if possible. Samples were kept cold during transport from the grocery store(s) to the laboratory.

Microbiological analysis:

In the laboratory, samples were refrigerated at 4°C and were processed no later than 96 hours after purchase. After microbiological examination, the sites recorded on the log sheets whether or not the meat and poultry samples were presumptively positive for *Salmonella*, *Campylobacter*, *E. coli*, and *Enterococcus*. Each laboratory used essentially the same procedure for sample collection. Retail meat and poultry packages were kept intact until they were aseptically opened in the laboratory at the start of examination. For chicken and pork samples, one piece of meat was examined. For ground beef and ground turkey samples, 25 g of ground product was analyzed. The analytical portions from each sample were placed in separate sterile plastic bags, 250 mL of buffered peptone water was added to each bag, and the bags were vigorously shaken. Fifty mL of the rinsate from each sample was transferred to separate sterile containers for isolation and identification of *Salmonella*, *Campylobacter*, *E. coli*, or *Enterococcus* using standard microbiological procedures. Once isolated and identified, bacterial isolates were sent to FDA's CVM Office of Research for further characterization including species confirmation, antimicrobial susceptibility testing and PFGE analysis (*Salmonella* and

Campylobacter only).

All ten FoodNet sites cultured the meats and poultry rinsates for the presence of *Salmonella* and *Campylobacter*. Four of the ten FoodNet laboratories (Georgia, Maryland, Oregon, and Tennessee) also cultured meat and poultry rinsates for the presence of *E. coli* and *Enterococcus*.

Changes in 2004

Several notable updates in the NARMS Retail Meat program occurred in 2004. A total of 4699 meats samples were collected, up from 3533 in 2003. This was due to the addition of FoodNet laboratories in Colorado and New Mexico, increasing the number of test sites from 8 to 10.

In 2004, we adopted a broth microdilution antimicrobial susceptibility testing method for *Campylobacter*, which also increased the number of agents tested from 5 to 9. The 9 antimicrobials tested in 2004 were: Azithromycin, Ciprofloxacin*, Clindamycin, Erythromycin*, Florfenicol, Gentamicin*, Nalidixic Acid, Telithromycin, and Tetracycline (* indicates agents also tested in 2003). Meropenem and Doxycycline were dropped from the list of *Campylobacter* agents tested.

The interpretive criteria used for *Campylobacter* antimicrobials is shown in Table 1. Based on the upcoming CLSI M45-P document (*Methods for Antimicrobial Dilution and Disk Susceptibility Testing of Infrequently-Isolated or Fastidious Bacteria; Proposed Guideline*, CLSI June 2006), the Erythromycin resistance breakpoint was changed from 8 µg/ml to 32 µg/ml. Based on the MIC distribution published in this report, along with other *Campylobacter* data generated using broth microdilution testing, several other breakpoints have been modified from those used in previous NARMS reports. For resistance breakpoints, these revised values include: Azithromycin (changed from 2 µg/mL to 8 µg/ml); Clindamycin (changed from 4

$\mu\text{g/mL}$ to $8\mu\text{g/ml}$); Gentamicin (changed from $16\mu\text{g/mL}$ to $8\mu\text{g/ml}$); and Nalidixic acid (changed from $32\mu\text{g/mL}$ to $64\mu\text{g/mL}$).

Two content changes were made in the panel formats. Cephalothin was omitted from the *E. coli/Salmonella* testing panel and Sulfamethoxazole was replaced with Sulfisoxazole. Daptomycin was used to replace Salinomycin on the *Enterococcus* panel.

NARMS retail meat working group, 2004

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A-3a	California
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A-3d	Georgia
A-3e	Maryland
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A-3h	New York
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A-4f	<i>Salmonella</i> Kentucky
A-4g	<i>Salmonella</i> Mbandaka
A-4h	<i>Salmonella</i> Montevideo
A-4i	<i>Salmonella</i> Muenster
A-4j	<i>Salmonella</i> Newport
A-4k	<i>Salmonella</i> Reading
A-4l	<i>Salmonella</i> Saintpaul
A-4m	<i>Salmonella</i> Schwarzengrund
A-4n	<i>Salmonella</i> Typhimurium
A-4o	<i>Salmonella</i> IIIa 18:z4,z32:-
A-4p	<i>Salmonella</i> 4, 12:i:-

A-4q
A-4r

Campylobacter coli
Campylobacter jejuni

ANTIMICROBIAL RESISTANCE AMONG

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Salmonella

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Campylobacter
Campylobacter jejuni
Campylobacter coli

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Enterococcus
Enterococcus faecium
Enterococcus faecalis

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Escherichia coli

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Log Sheet Example

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Experimental Design & Procedures

Table 1. Antimicrobial Susceptibility Test Methods and Interpretive Criteria: NARMS Retail Meat, 2004

Genus: *Campylobacter*

Susceptibility Testing Method: Broth microdilution

Sensititre Plate: CAMPY

QC Organism: *Campylobacter jejuni* ATCC 33560

Drug	Susceptible ($\mu\text{g}/\text{ml}$)	Intermediate ($\mu\text{g}/\text{ml}$)	Resistant ($\mu\text{g}/\text{ml}$)
Azithromycin*	≤ 2	4	≥ 8
Ciprofloxacin	≤ 1	2	≥ 4
Clindamycin*	≤ 2	4	≥ 8
Erythromycin	≤ 8	16	≥ 32
Florfenicol* [^]	≤ 4		
Gentamicin*	≤ 2	4	≥ 8
Nalidixic Acid*	≤ 16	32	≥ 64
Telithromycin*	≤ 4	8	≥ 16
Tetracycline	≤ 4	8	≥ 16

Genus: *Enterococcus*

Susceptibility Testing Method: Broth microdilution

Sensititre Plate: CMV1AGPF

QC Organisms: *Enterococcus faecalis* ATCC 29212 and *Enterococcus faecalis* ATCC 51299

Drug	Susceptible ($\mu\text{g}/\text{ml}$)	Intermediate ($\mu\text{g}/\text{ml}$)	Resistant ($\mu\text{g}/\text{ml}$)
Bacitracin*	≤ 32	64	≥ 128
Chloramphenicol	≤ 8	16	≥ 32
Ciprofloxacin	≤ 1	2	≥ 4
Daptomycin*	≤ 4		
Erythromycin	≤ 0.5	1,2,4	≥ 8
Flavomycin*	≤ 8	16	≥ 32
Gentamicin	< 500		≥ 500
Kanamycin*	≤ 128	256	≥ 512
Lincomycin*	≤ 8	16	≥ 32
Linezolid	≤ 2	4	≥ 8
Nitrofurantoin	≤ 32	64	≥ 128
Penicillin	≤ 8		≥ 16
Streptomycin*	< 1000		≥ 1000
Quinupristin/Dalfopristin	≤ 1	2	≥ 4
Tetracycline	≤ 4	8	≥ 16
Tylosin*	≤ 8	16	≥ 32
Vancomycin	≤ 4	8,16	≥ 32

* No CLSI interpretative criteria for this bacterium / antimicrobial combination currently available.

[^]Absence of resistant strains precludes defining any results category other than "susceptible."

Genus: *Escherichia coli* and *Salmonella*

Susceptibility Testing Method: Broth microdilution

Sensititre Plate: CMV1AGNF

QC Organisms: *Escherichia coli* ATCC 25922, *Staphylococcus aureus* ATCC 29213,

Pseudomonas aeruginosa ATCC 27853, and *Enterococcus faecalis* ATCC 29212

Drug	Susceptible ($\mu\text{g}/\text{ml}$)	Intermediate ($\mu\text{g}/\text{ml}$)	Resistant ($\mu\text{g}/\text{ml}$)
Amikacin	≤ 16	32	≥ 64
Amoxicillin/Clavulanic acid	$\leq 8/4$	16/8	$\geq 32/16$
Ampicillin	≤ 8	16	≥ 32
Cefoxitin	≤ 8	16	≥ 32
Ceftiofur	≤ 2	4	≥ 8
Ceftriaxone	≤ 8	16,32	≥ 64
Chloramphenicol	≤ 8	16	≥ 32
Ciprofloxacin	≤ 1	2	≥ 4
Gentamicin	≤ 4	8	≥ 16
Kanamycin	≤ 16	32	≥ 64
Nalidixic acid	≤ 16		≥ 32
Streptomycin*	≤ 32		≥ 64
Sulfisoxazole	≤ 256		≥ 512
Tetracycline	≤ 4	8	≥ 16
Trimethoprim/sulfamethoxazole	$\leq 2/38$		$\geq 4/76$

* No CLSI interpretative criteria for this bacterium / antimicrobial combination currently available.

Table 2. Number of Retail Meat Samples Tested by Site and Meat Type, 2004

Site	Chicken Breast	Ground Turkey	Ground Beef	Pork Chop	Total
CA	120	120	120	120	480
CO	97	101	106	99	403
CT	120	120	120	120	480
GA	120	120	120	120	480
MD	120	120	120	120	480
MN	120	120	120	120	480
NM	119	118	120	119	476
NY	120	120	120	120	480
OR	120	120	120	120	480
TN	116	106	120	118	460
Total	1172	1165	1186	1176	4699

Table 3. Percent Positive Samples by Bacterium and Meat Type, 2004

Bacterium	Chicken Breast		Ground Turkey		Ground Beef		Pork Chop	
	N	(%)	N	(%)	N	(%)	N	(%)
<i>Campylobacter</i>	706	(60.2)	12	(1.0)	0	(0.0)	3	(0.3)
<i>Salmonella</i>	157	(13.4)	142	(12.2)	14	(1.2)	11	(0.9)
<i>Enterococcus</i>	466	(97.9)	437	(93.8)	448	(93.3)	404	(84.5)
<i>Escherichia coli</i>	400	(84.0)	376	(80.7)	338	(70.4)	232	(48.5)

4699 = Total number of retail meats tested for *Salmonella* and *Campylobacter*

1172 = Total Chicken Breast tested

1165 = Total Ground Turkey tested

1186 = Total Ground Beef tested

1176 = Total Pork Chop tested

1900 = Total number of retail meats tested for *Enterococcus* and *Escherichia coli*

476 = Total Chicken Breast tested

466 = Total Ground Turkey tested

480 = Total Ground Beef tested

478 = Total Pork Chop tested

Table 4. Number of Isolates by Site, Bacterium, and Meat Type, 2004

	Chicken Breast	Ground Turkey	Ground Beef	Pork Chops
Site: CA				
<i>Campylobacter</i>	96	0	0	1
<i>Salmonella</i>	17	1	9	1
Site: CO				
<i>Campylobacter</i>	21	0	0	0
<i>Salmonella</i>	1	0	8	0
Site: CT				
<i>Campylobacter</i>	86	0	2	1
<i>Salmonella</i>	30	5	26	5
Site: GA				
<i>Campylobacter</i>	61	0	1	0
<i>Salmonella</i>	6	1	38	64
<i>Enterococcus</i>	120	117	120	116
<i>Escherichia coli</i>	115	91	119	68
Site: MD				
<i>Campylobacter</i>	76	0	2	0
<i>Salmonella</i>	24	1	13	0
<i>Enterococcus</i>	114	100	106	62
<i>Escherichia coli</i>	110	83	109	77
Site: MN				
<i>Campylobacter</i>	73	0	6	0
<i>Salmonella</i>	20	0	14	0
Site: NM				
<i>Campylobacter</i>	53	0	6	0
<i>Salmonella</i>	3	0	14	0
Site: NY				
<i>Campylobacter</i>	96	0	0	0
<i>Salmonella</i>	16	0	11	3
Site: OR				
<i>Campylobacter</i>	73	0	0	0
<i>Salmonella</i>	25	6	6	2
<i>Enterococcus</i>	118	115	105	108
<i>Escherichia coli</i>	73	99	53	51
Site: TN				
<i>Campylobacter</i>	71	0	1	0
<i>Salmonella</i>	15	0	8	0
<i>Enterococcus</i>	114	116	106	103
<i>Escherichia coli</i>	102	65	96	55

Figure 1a. Percent Positive Samples for *Campylobacter* & *Salmonella* by Meat Type and Site, 2004

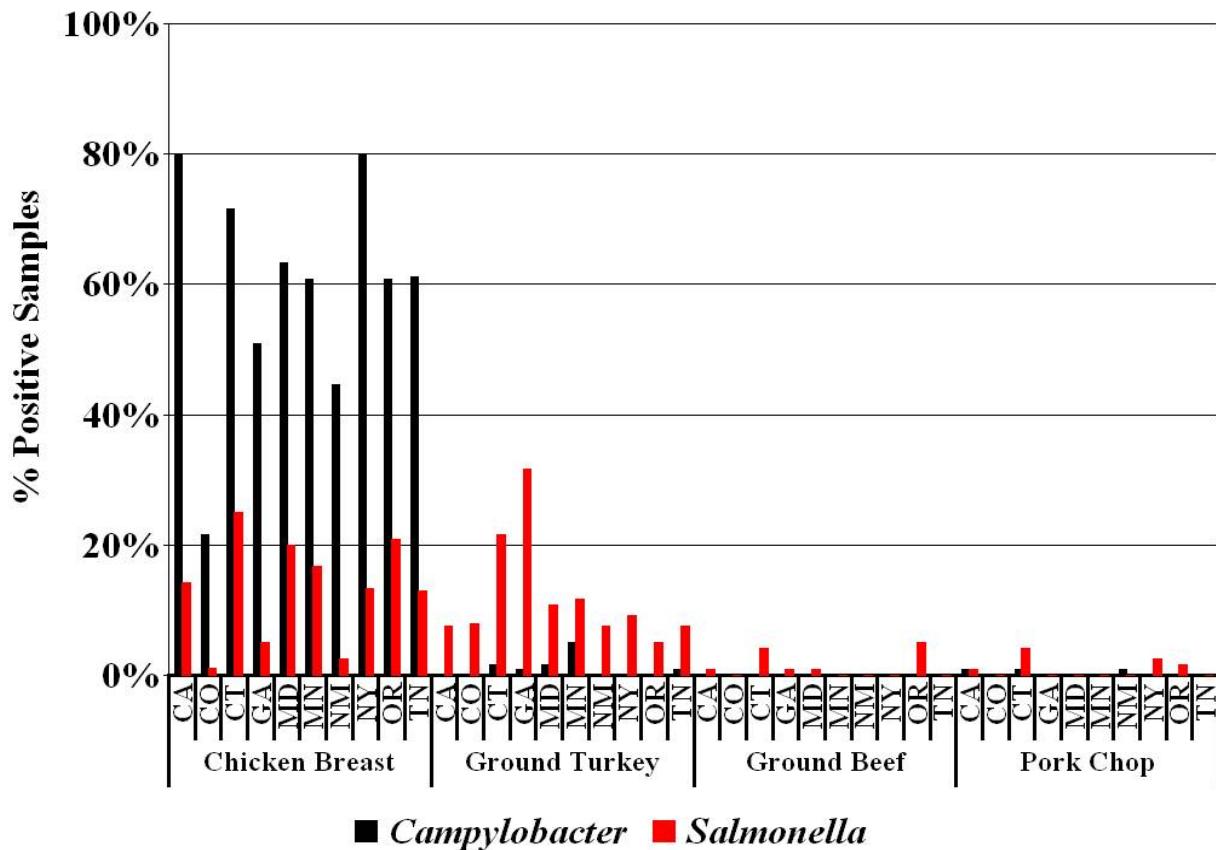


Figure 1b. Percent Positive Samples for *Enterococcus* & *E. coli* by Meat Type and Site, 2004

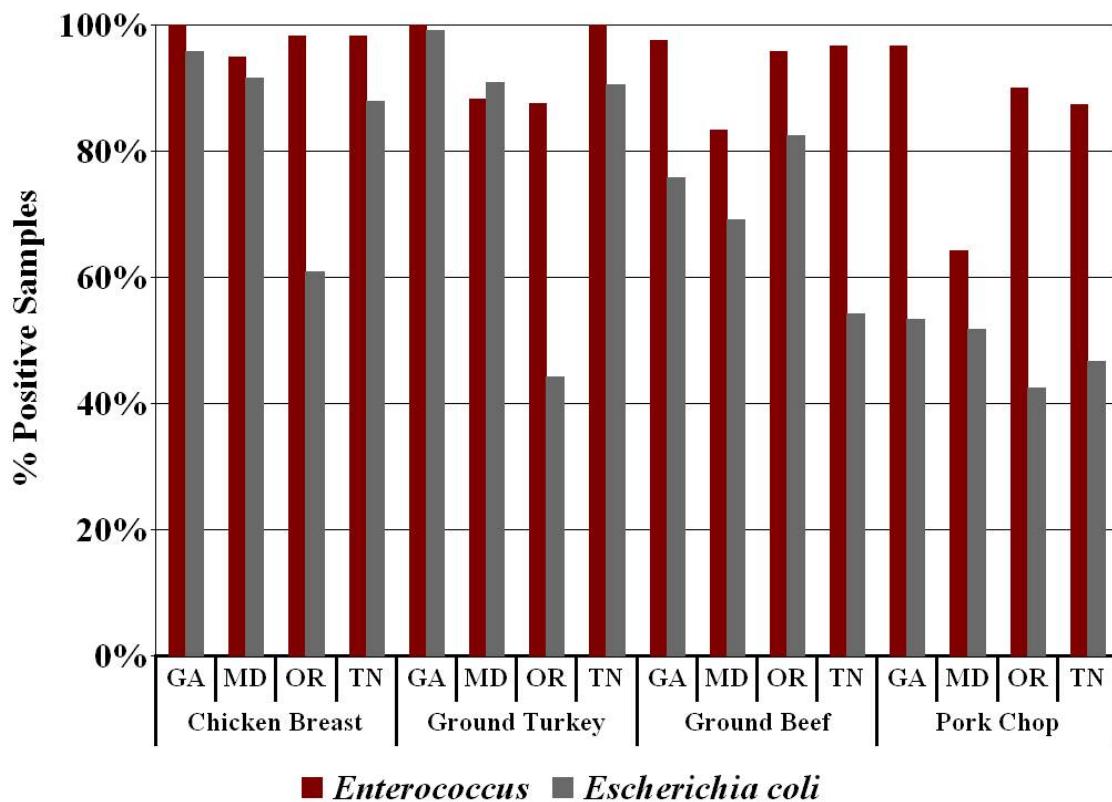


Figure 2a. Percent Positive Samples for *Campylobacter* & *Salmonella* by Meat Type for All Sites, 2004

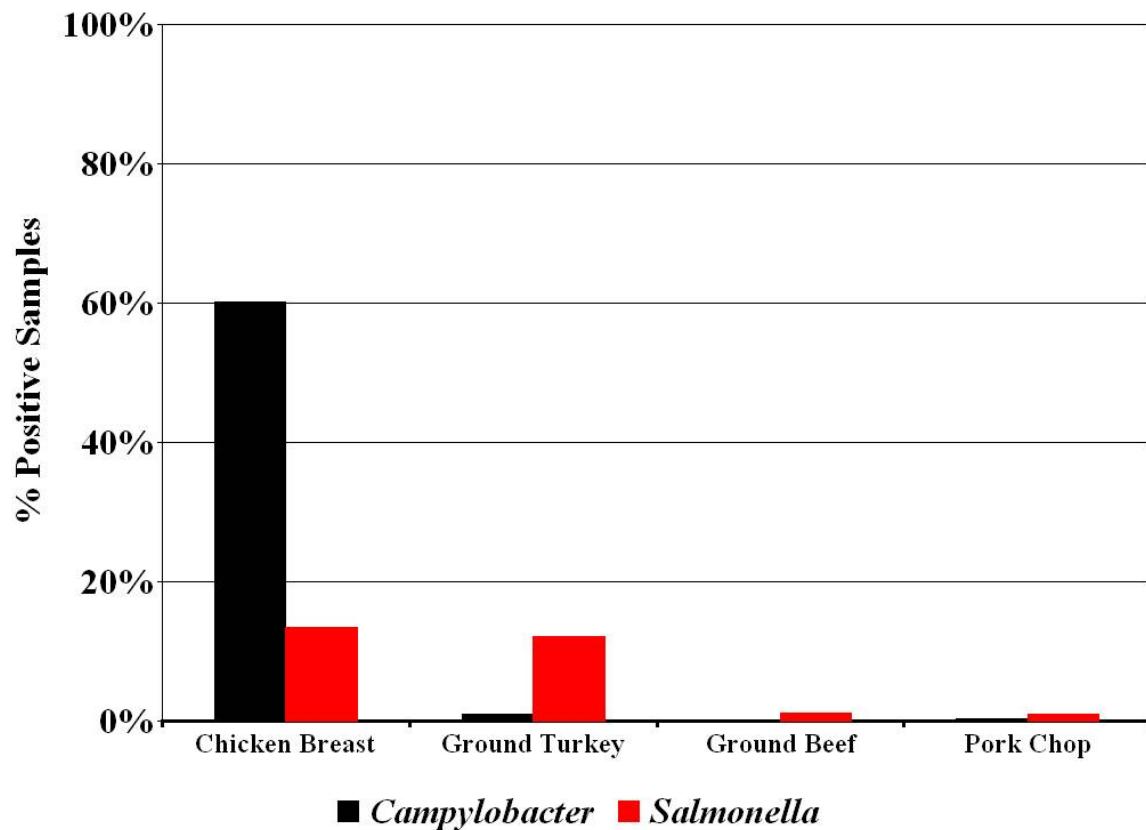


Figure 2b. Percent Positive Samples for *Enterococcus* & *E. coli* by Meat Type for All Sites, 2004

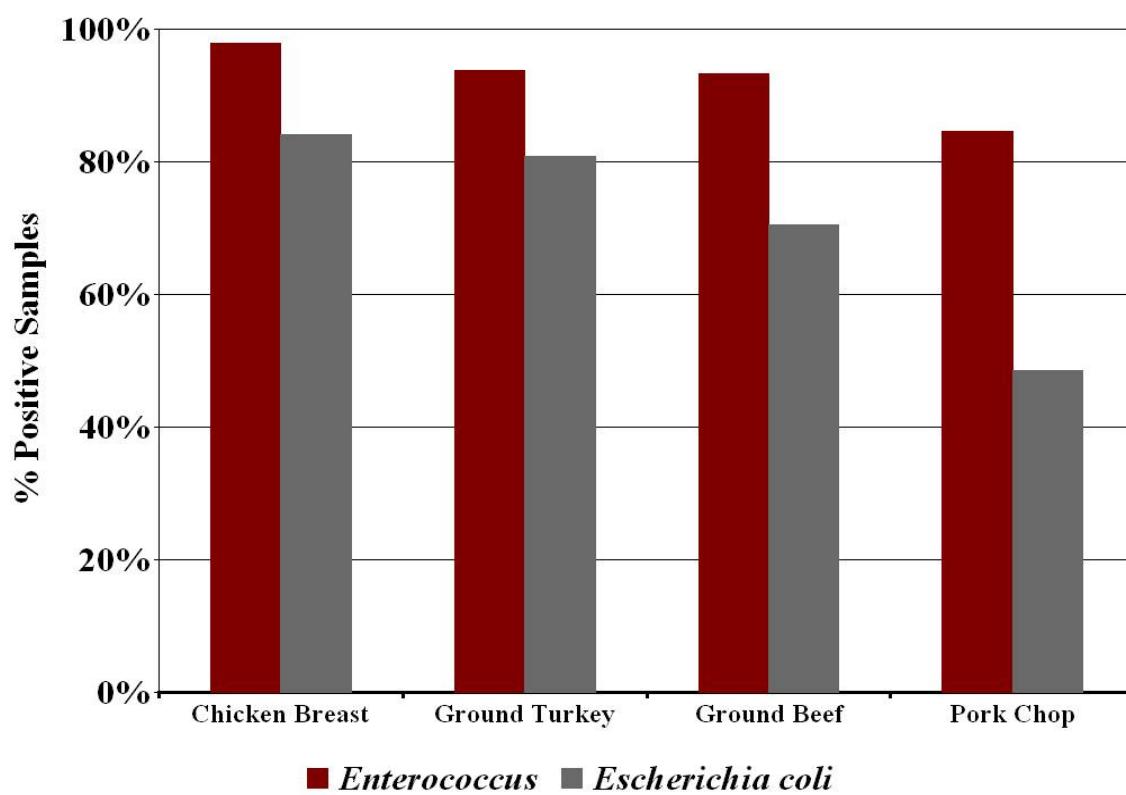


Figure 3a. Percent Positive Samples for *Campylobacter* & *Salmonella* and *Enterococcus* & *E. coli* by Month and Meat Type for All Sites, 2004

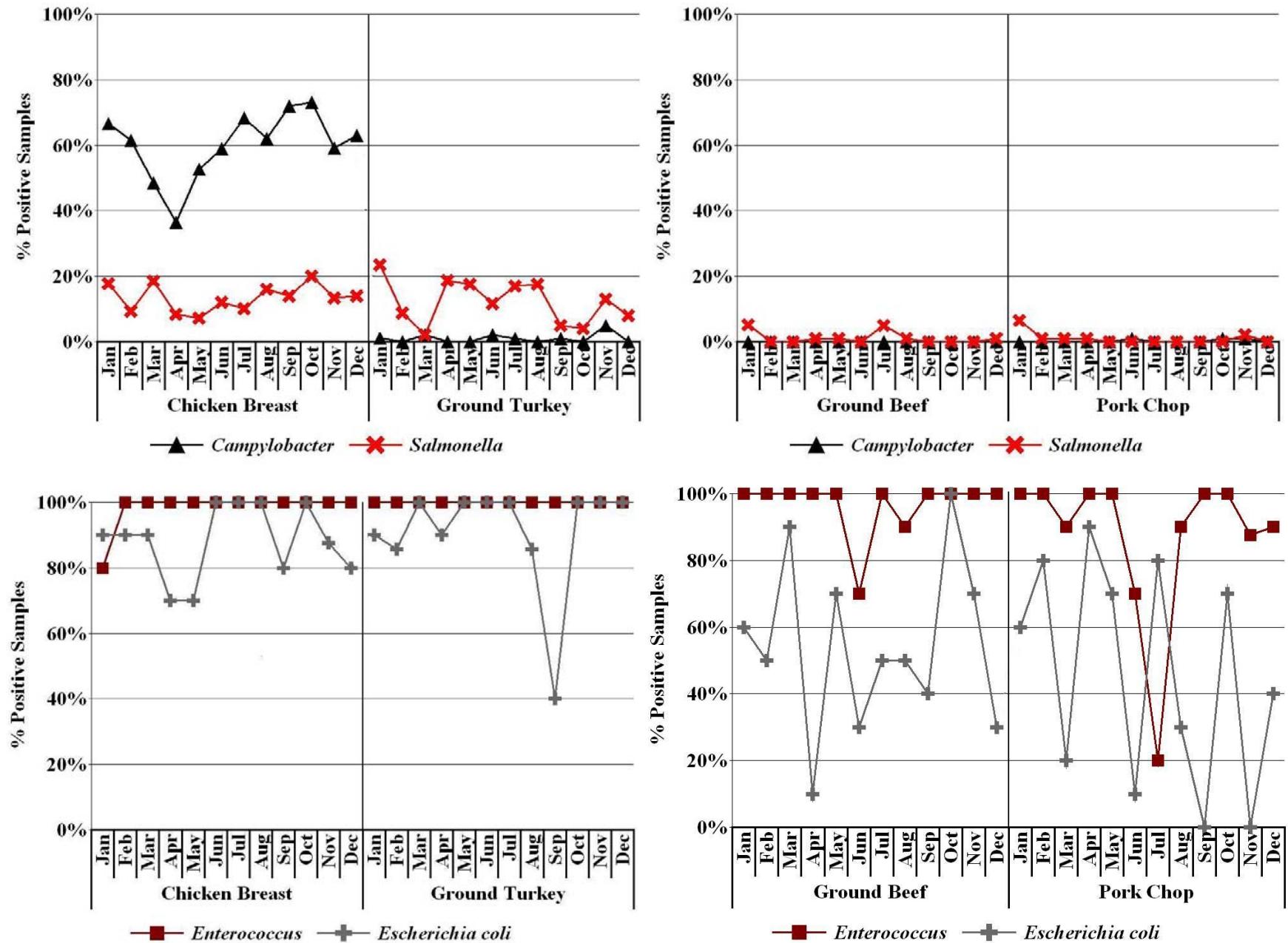


Figure 3b. Percent Positive Samples for *Campylobacter* & *Salmonella* by Month and Meat Type in California, 2004

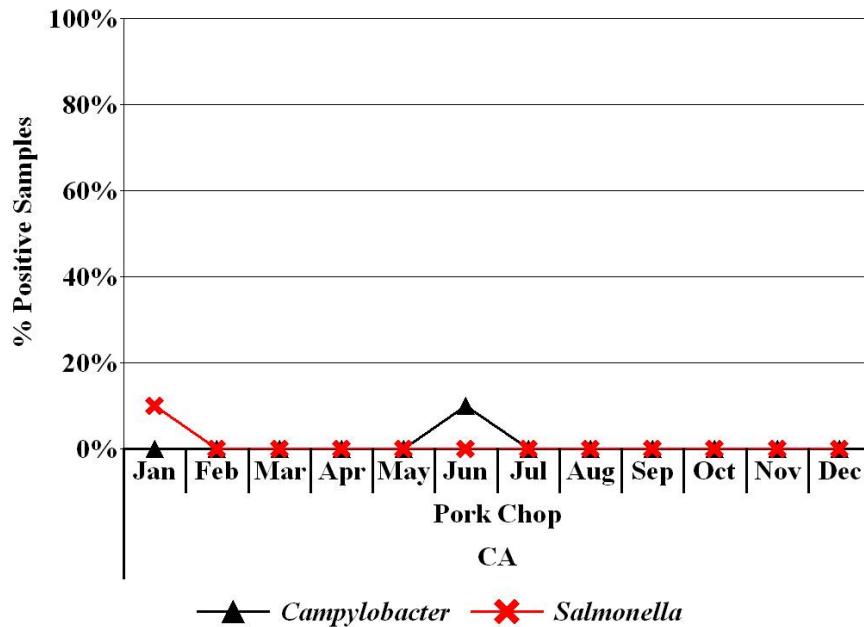
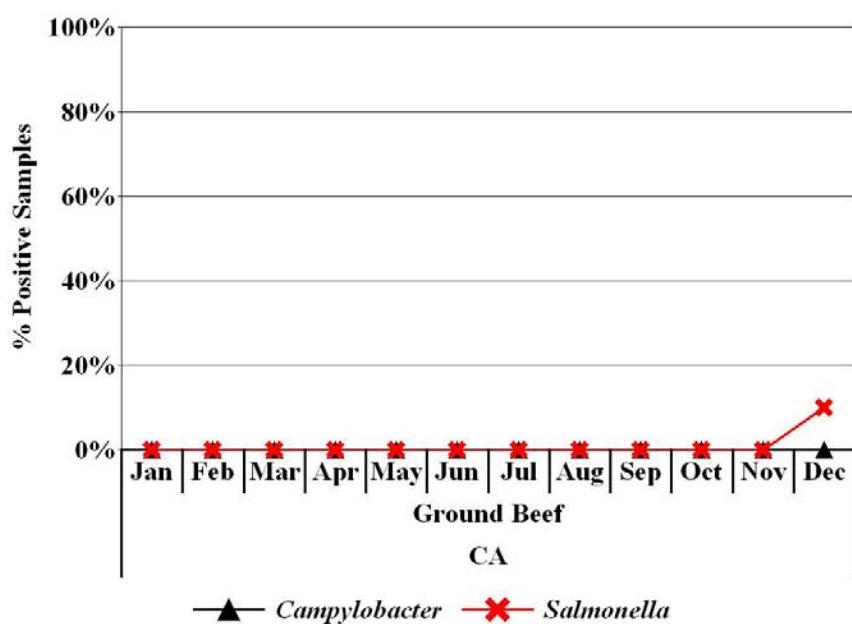
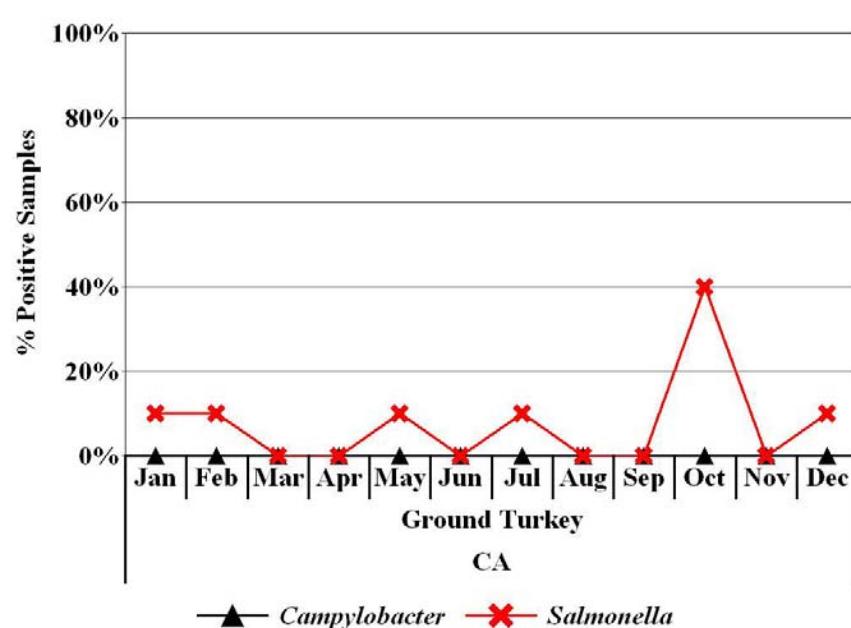
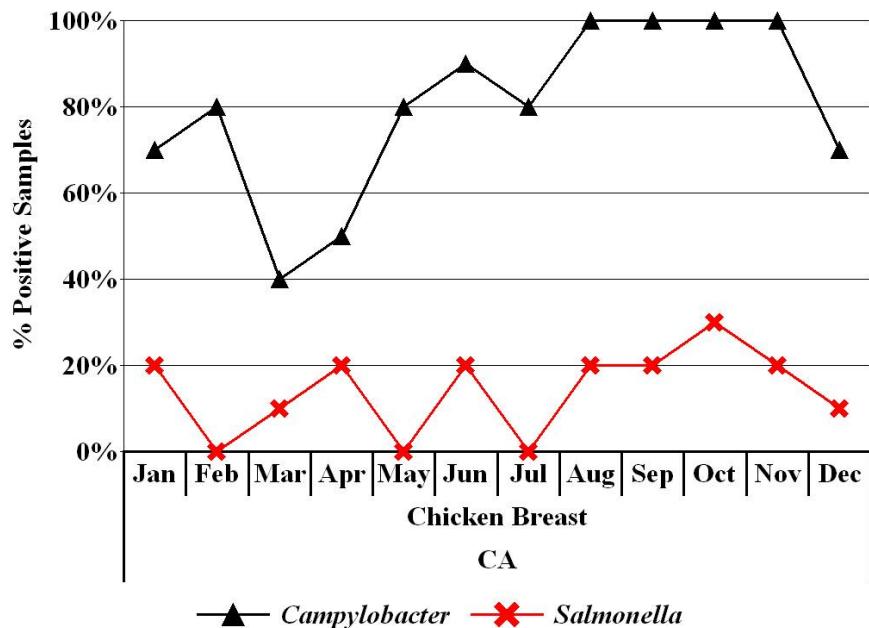


Figure 3c. Percent Positive Samples for *Campylobacter* & *Salmonella* by Month and Meat Type in Colorado, 2004

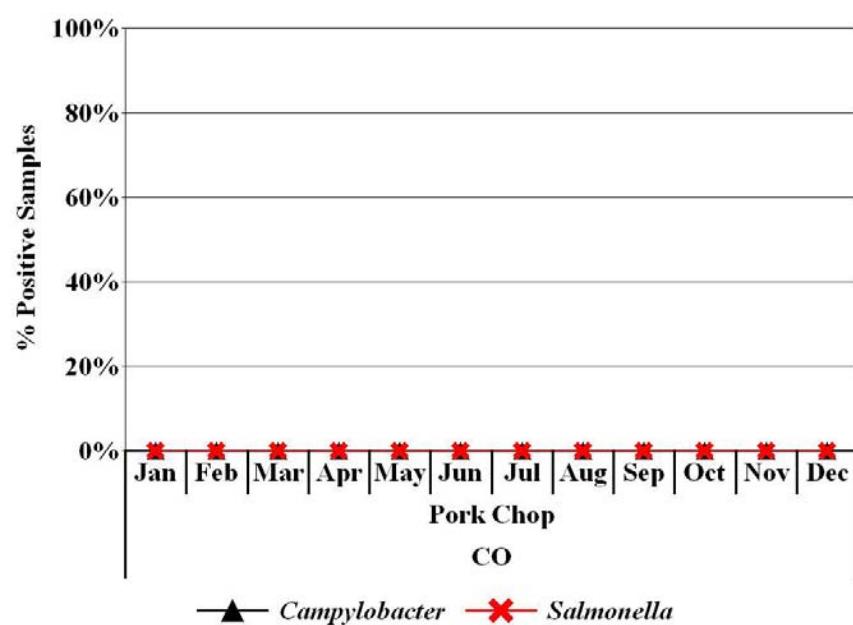
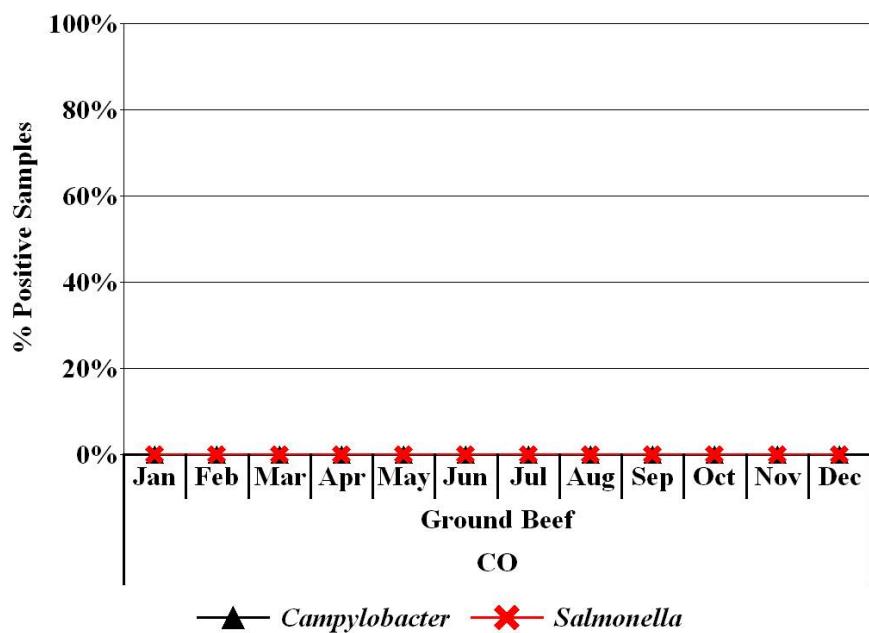
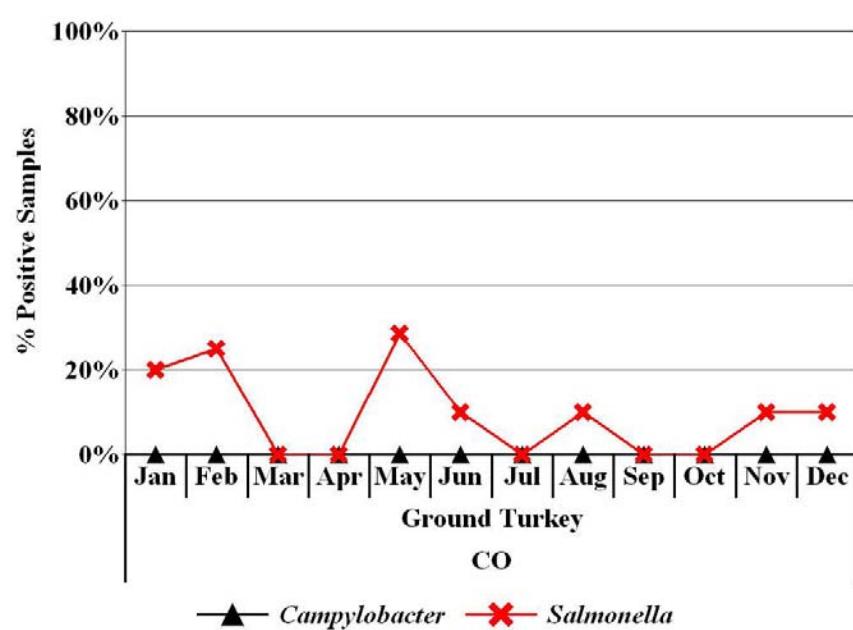
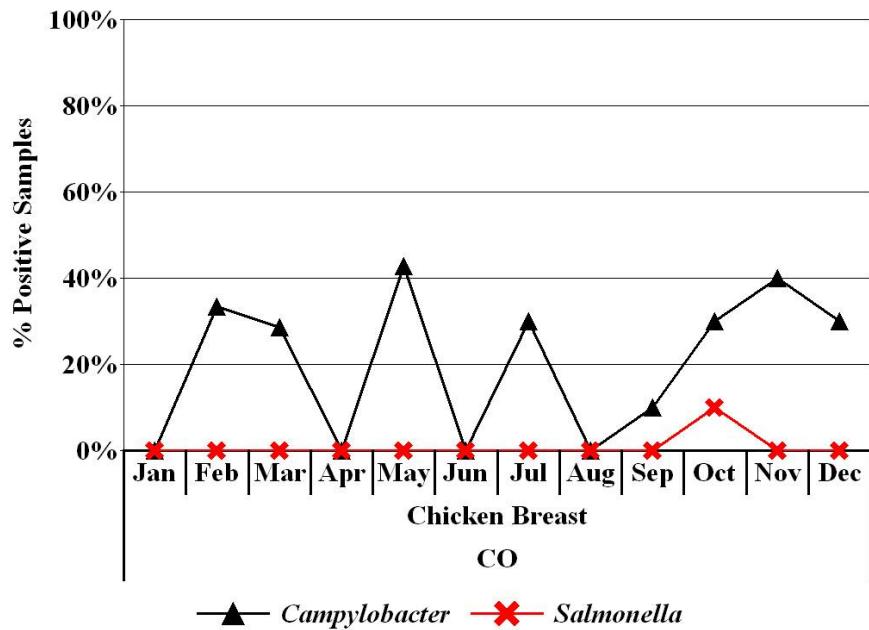


Figure 3d. Percent Positive Samples for *Campylobacter* & *Salmonella* by Month and Meat Type in Connecticut, 2004

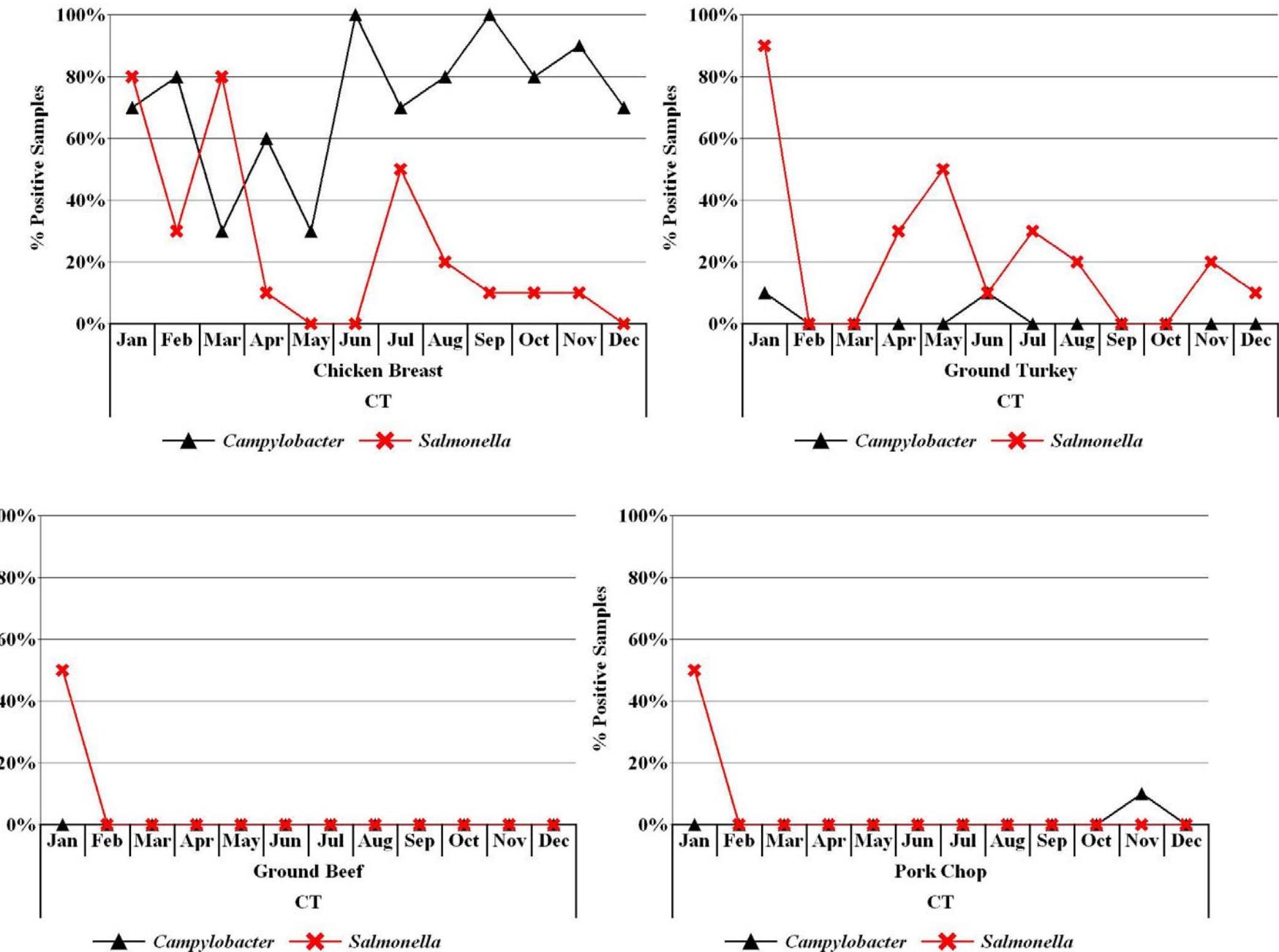


Figure 3e. Percent Positive Samples for *Campylobacter* & *Salmonella* by Month and Meat Type in Georgia, 2004

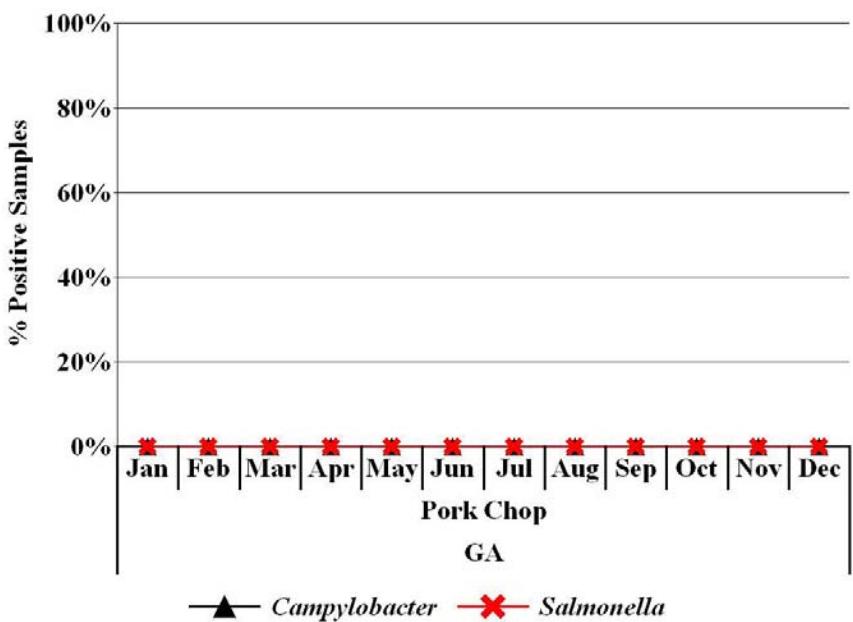
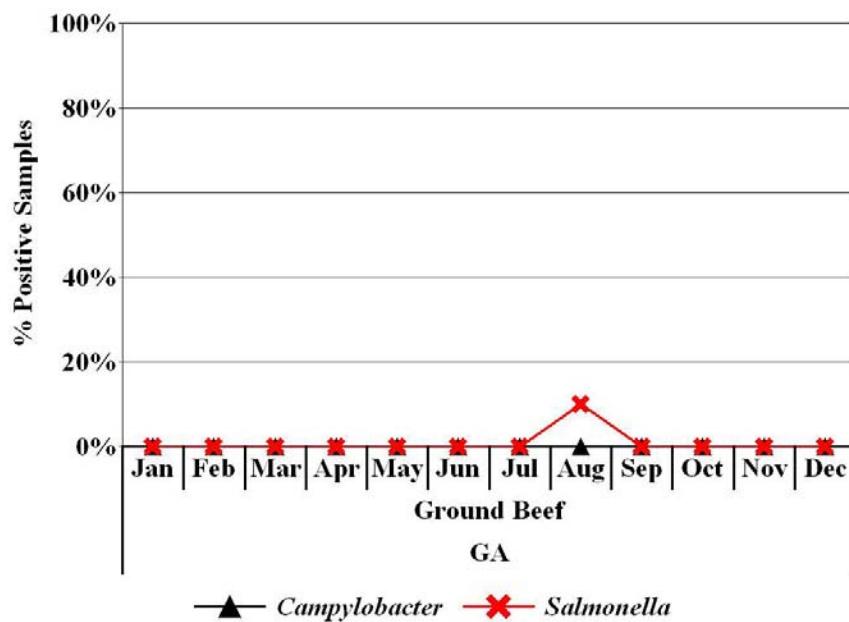
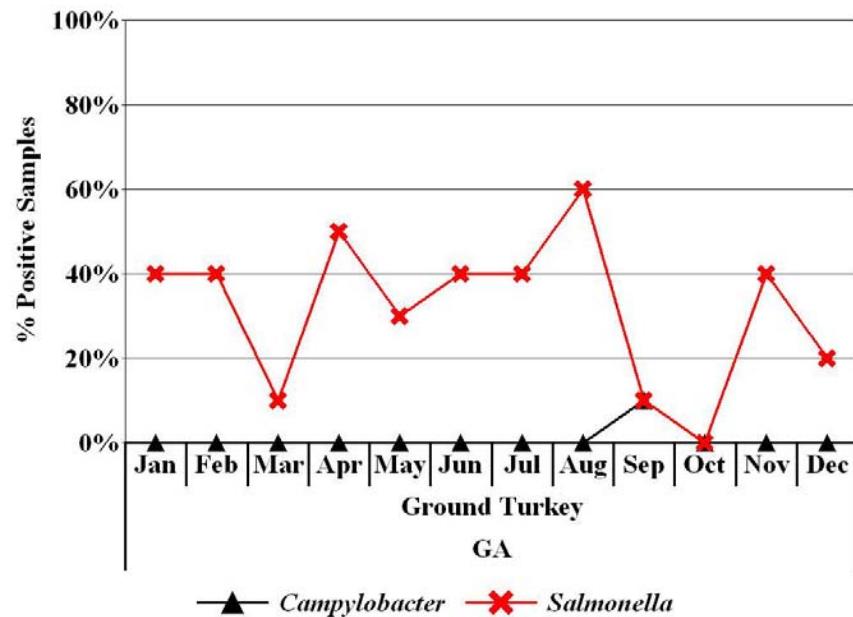
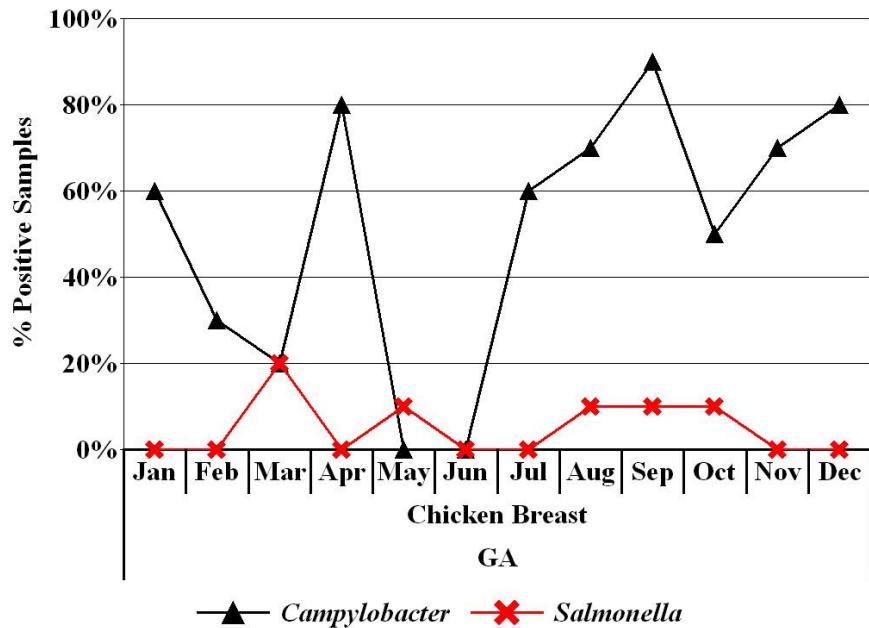


Figure 3f. Percent Positive Samples for *Enterococcus* & *E. coli* by Month and Meat Type in Georgia, 2004

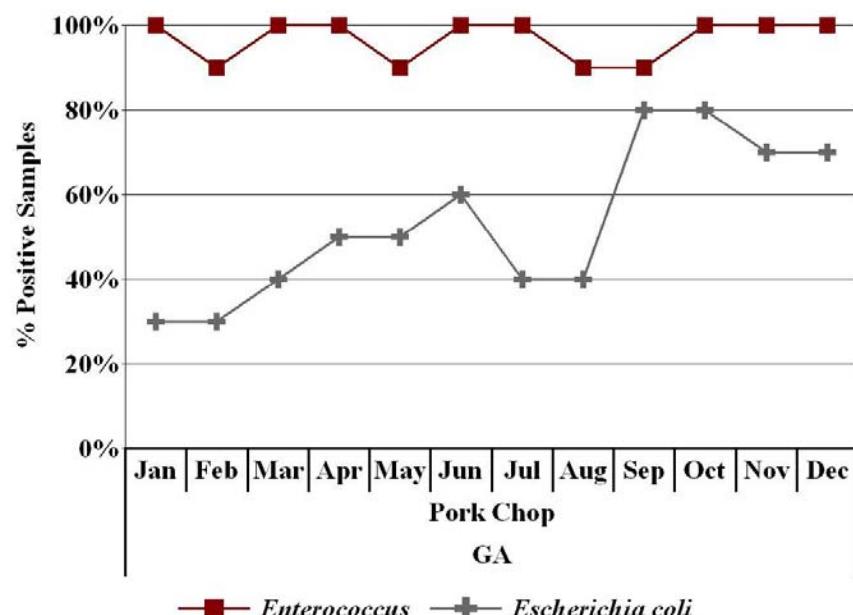
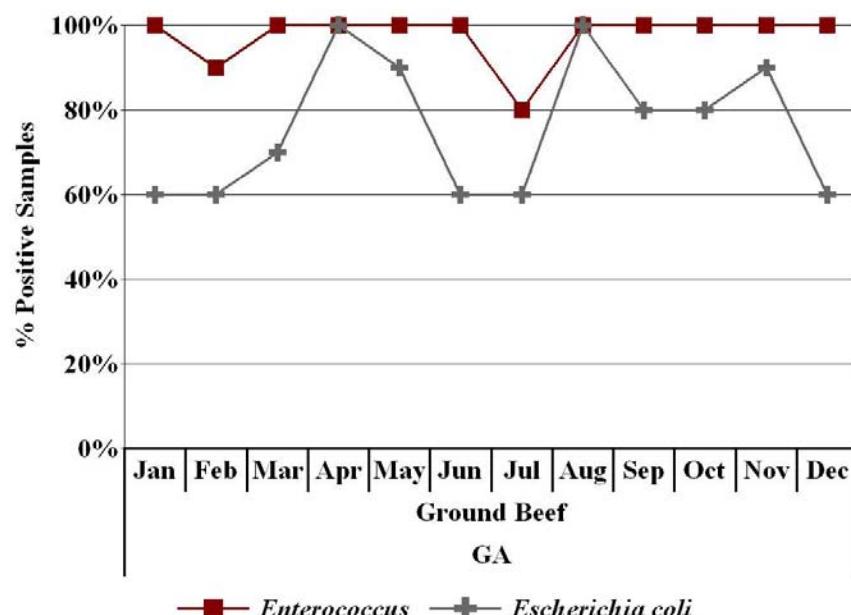
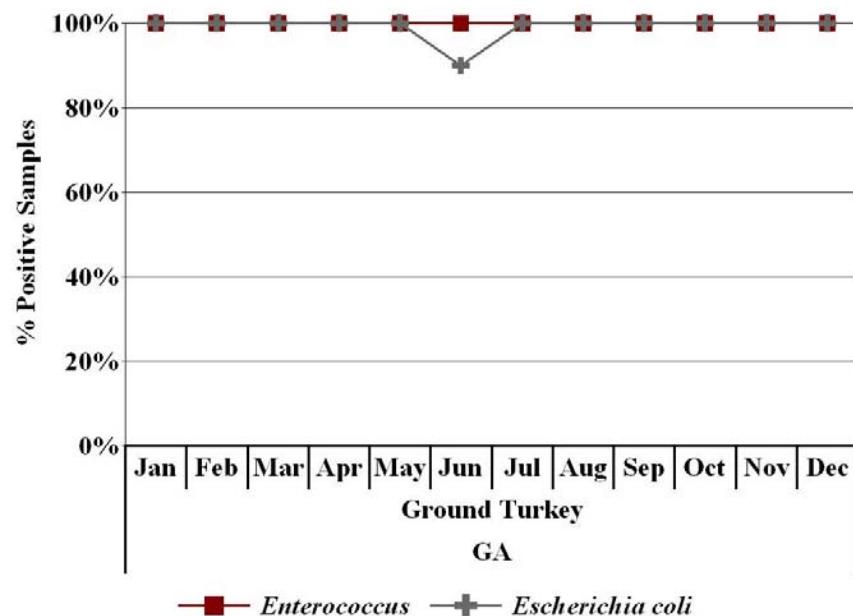
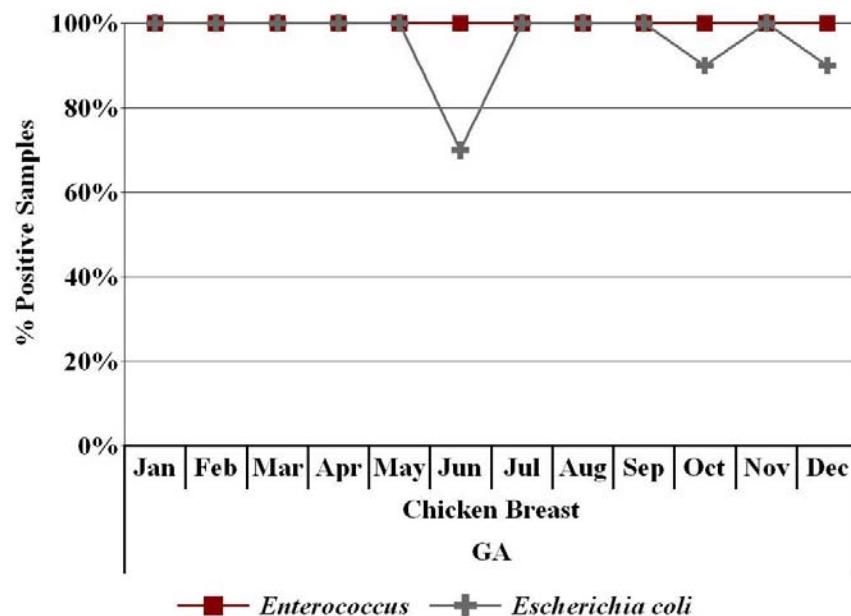


Figure 3g. Percent Positive Samples for *Campylobacter* & *Salmonella* by Month and Meat Type in Maryland, 2004

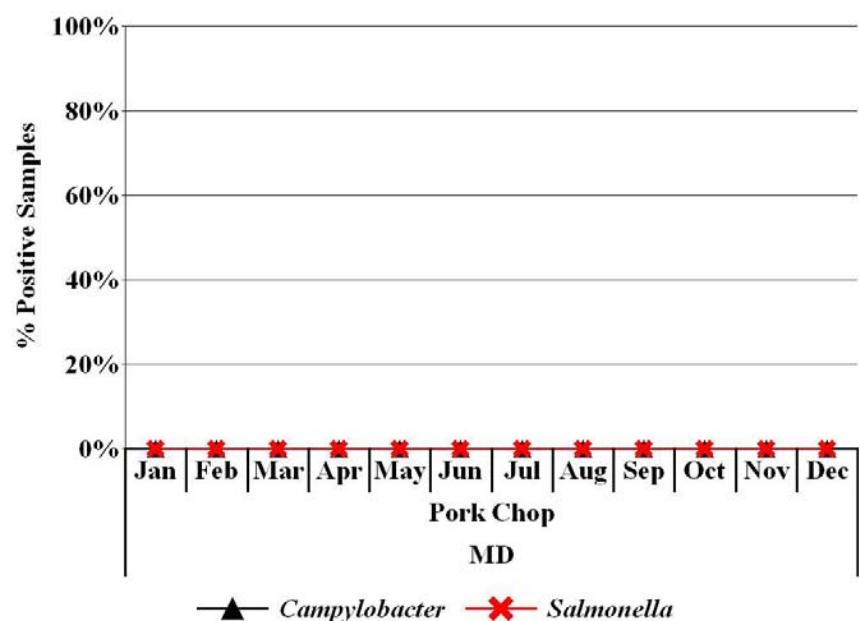
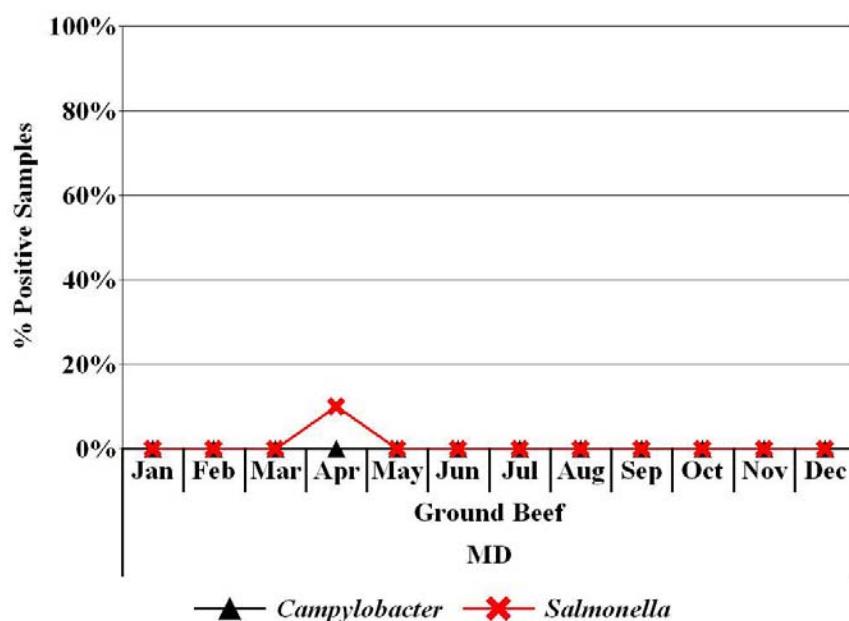
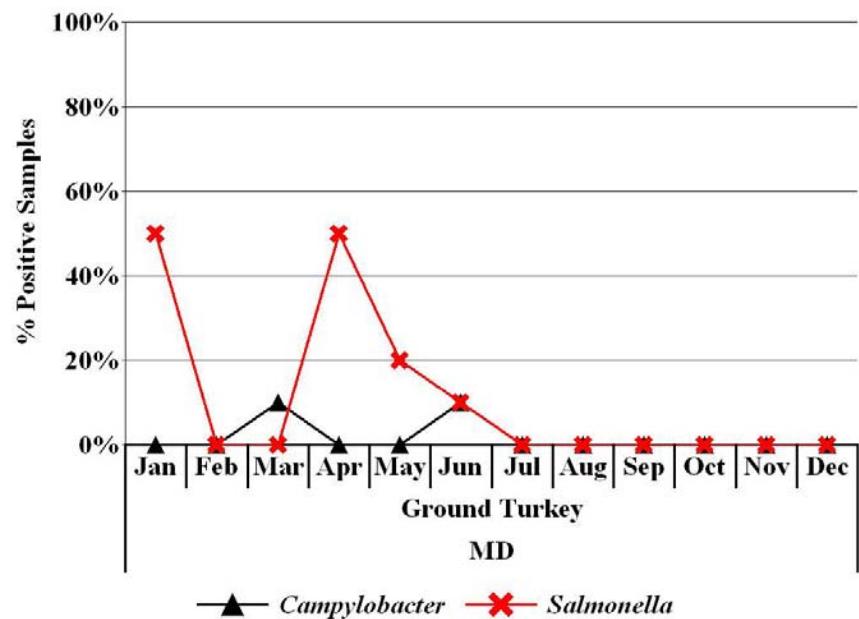
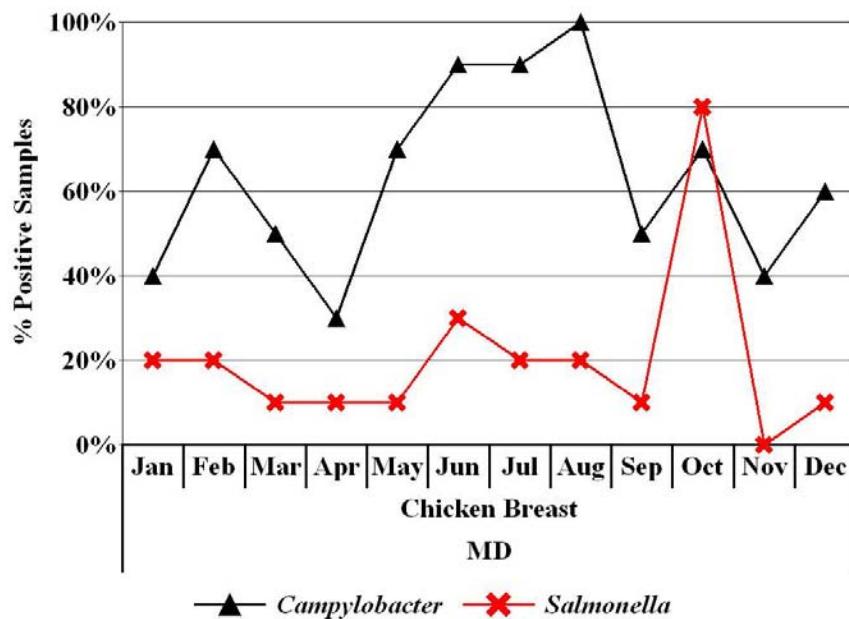


Figure 3h. Percent Positive Samples for *Enterococcus* & *E. coli* by Month and Meat Type in Maryland, 2004

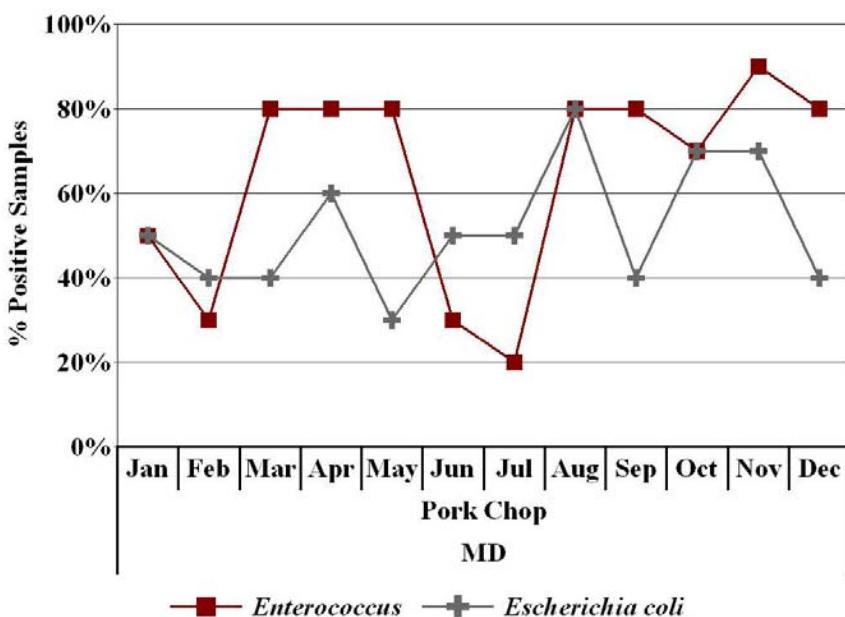
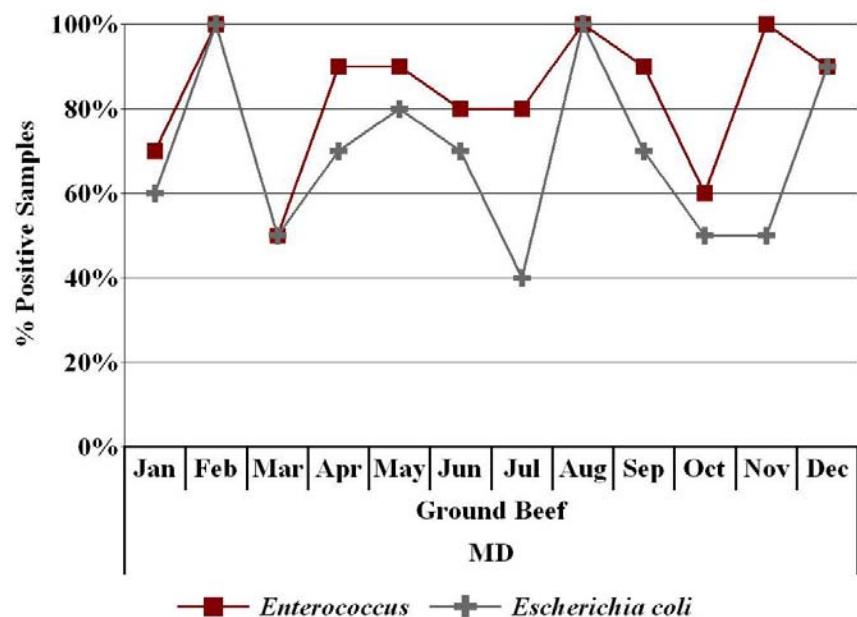
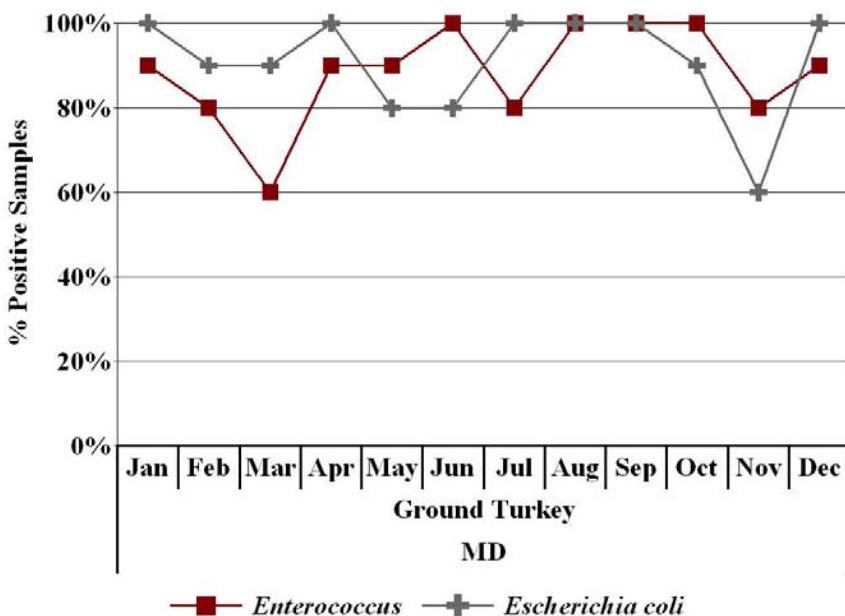
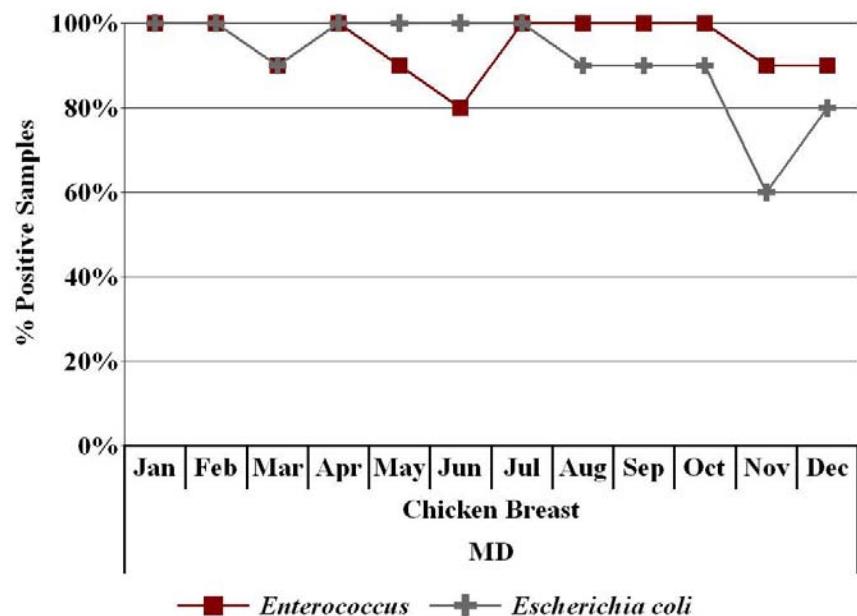


Figure 3i. Percent Positive Samples for *Campylobacter* & *Salmonella* by Month and Meat Type in Minnesota, 2004

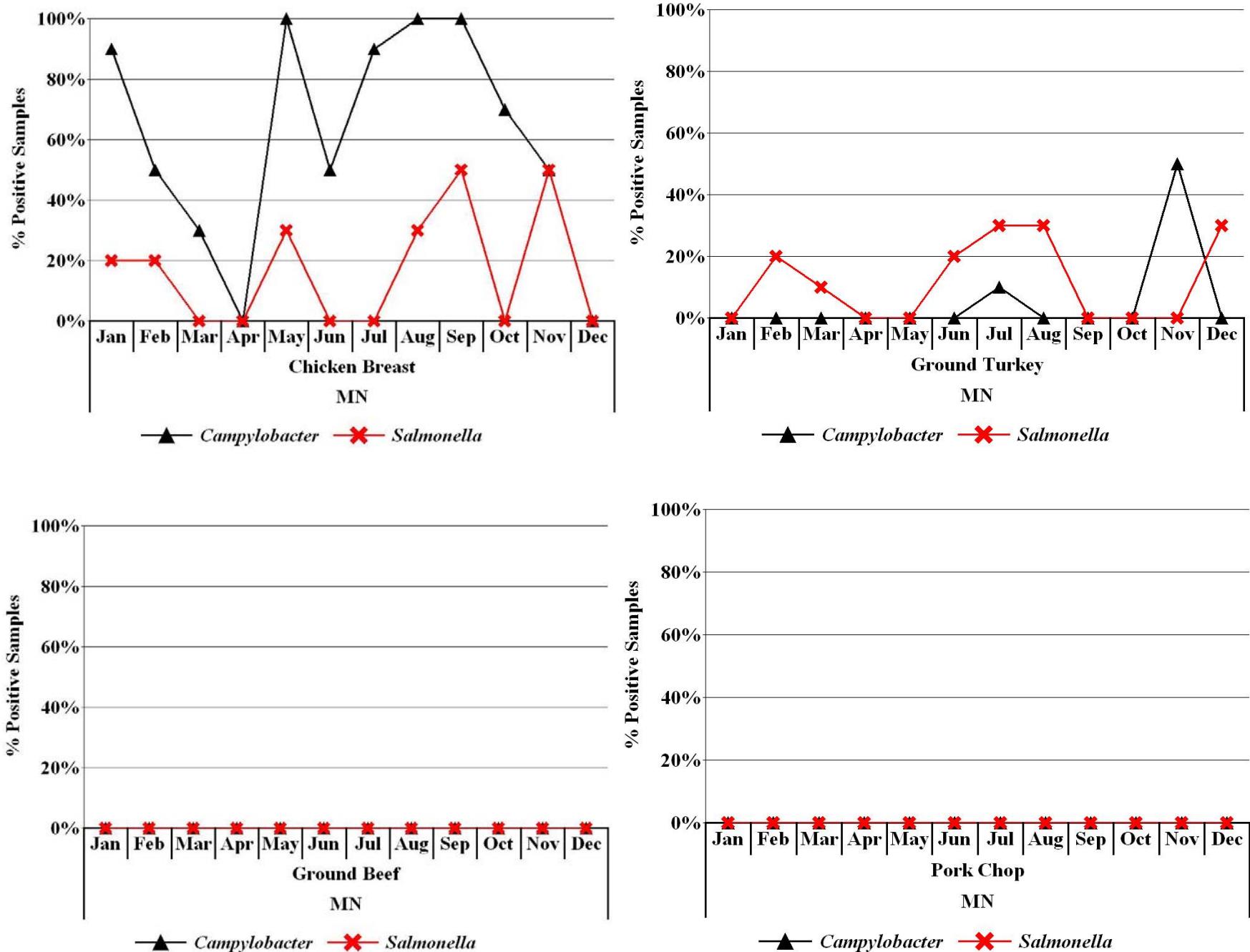


Figure 3j. Percent Positive Samples for *Campylobacter* & *Salmonella* by Month and Meat Type in New Mexico, 2004

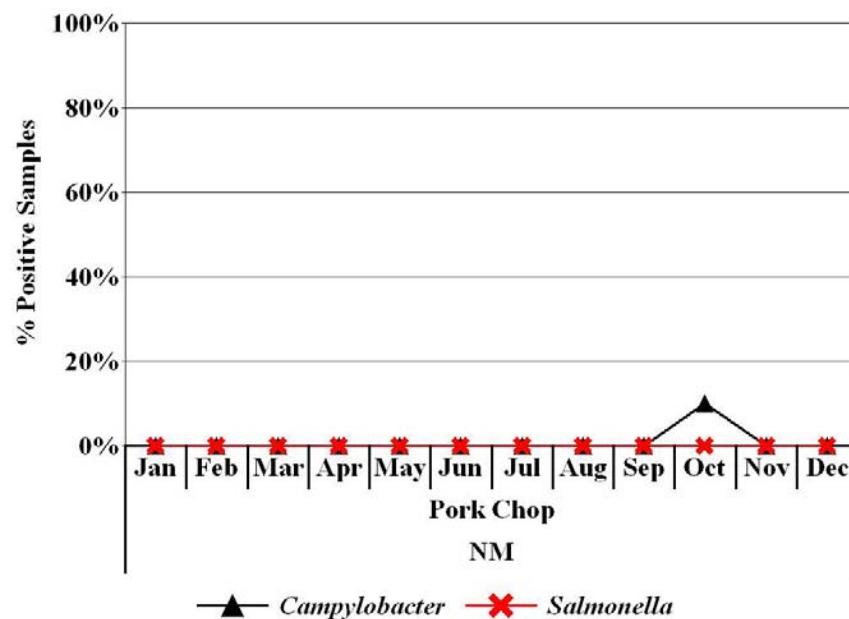
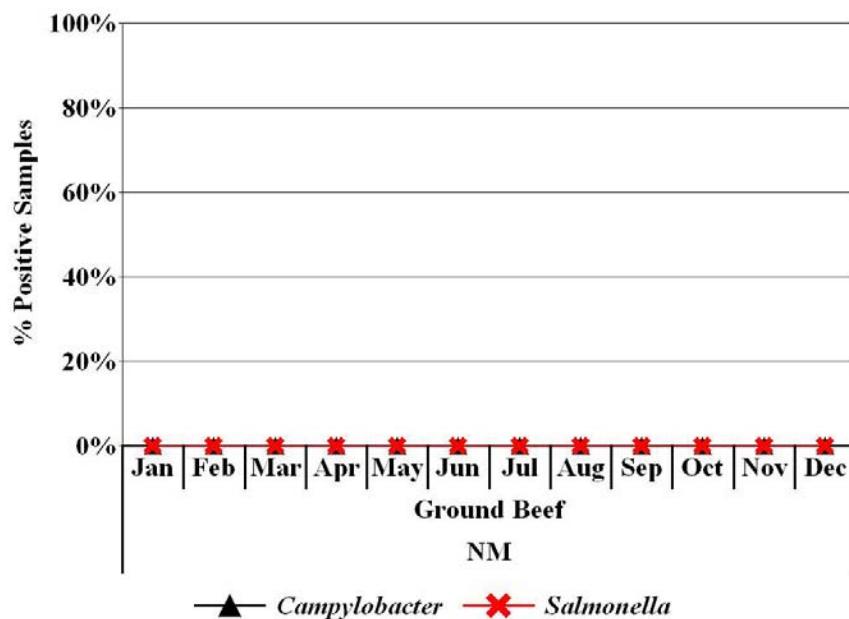
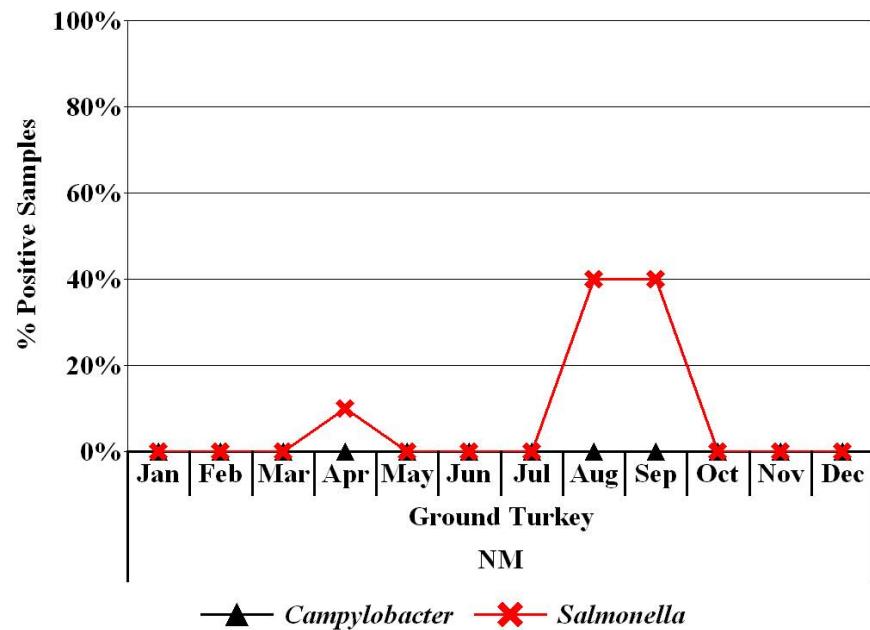
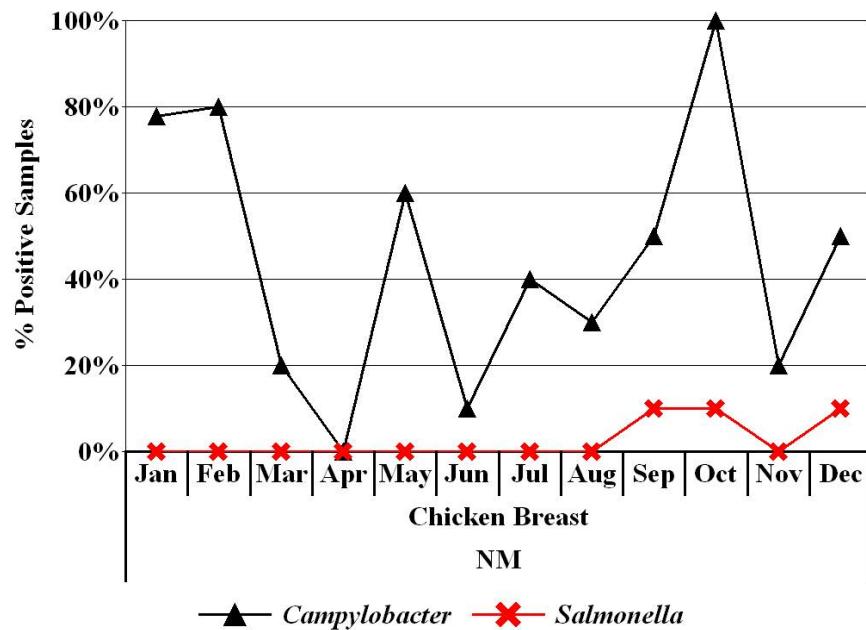


Figure 3k. Percent Positive Samples for *Campylobacter* & *Salmonella* by Month and Meat Type in New York, 2004

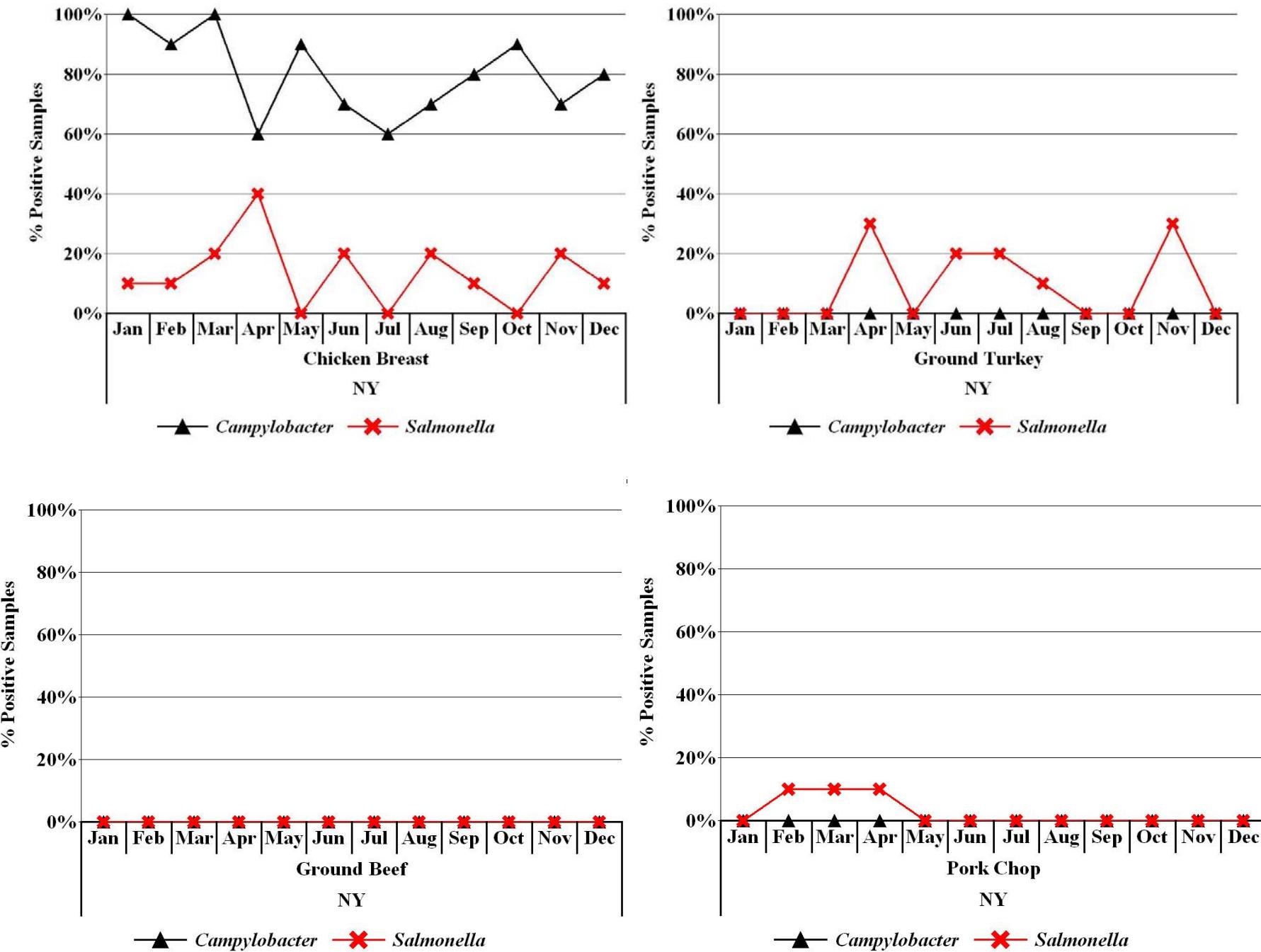


Figure 3l. Percent Positive Samples for *Campylobacter* & *Salmonella* by Month and Meat Type in Oregon, 2004

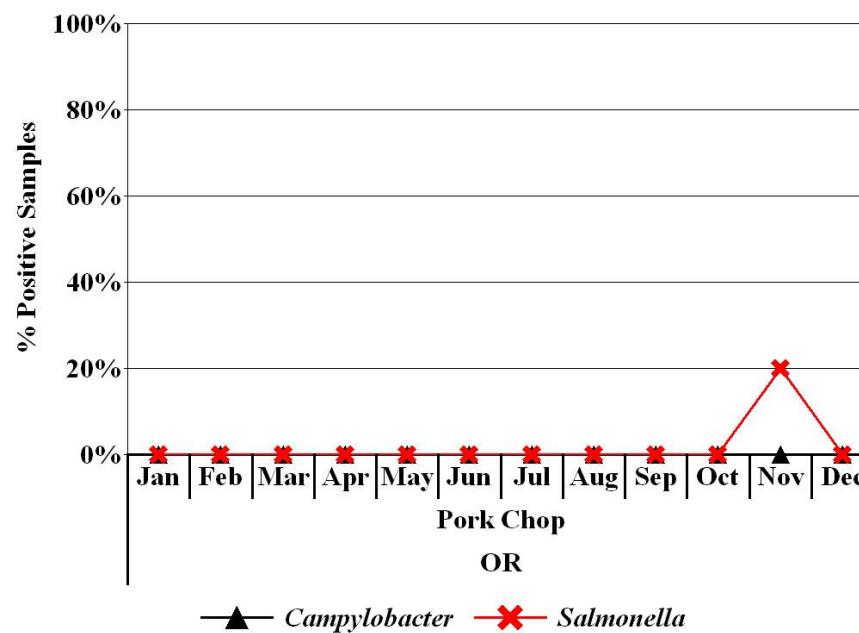
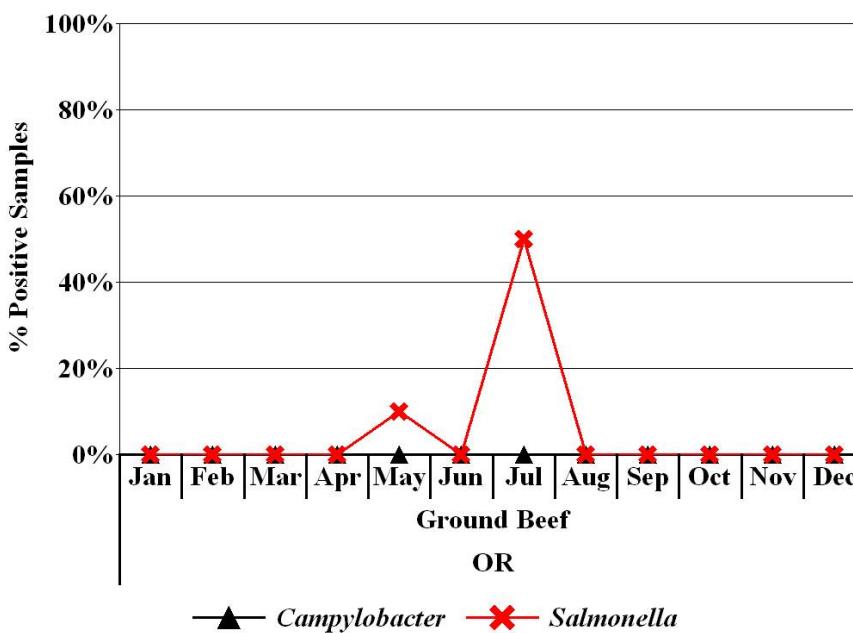
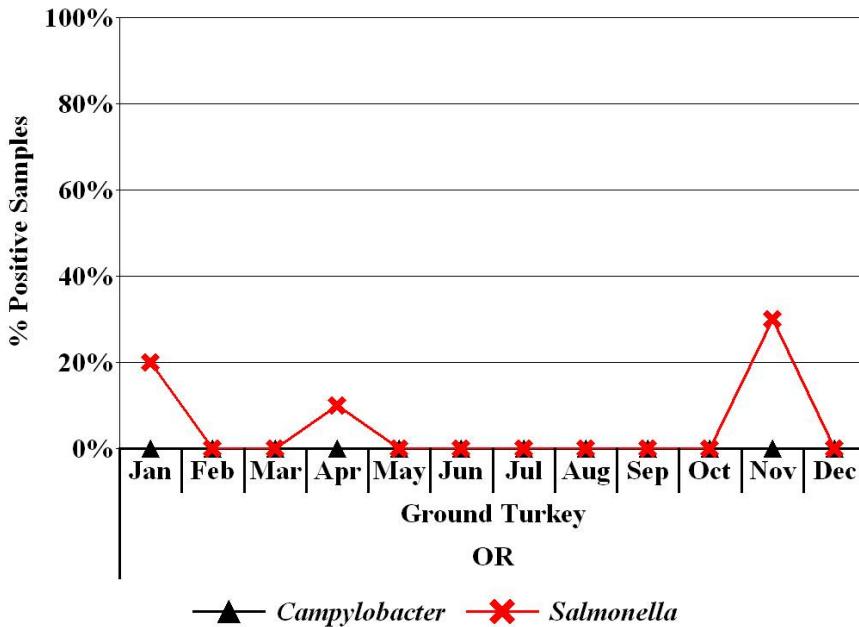
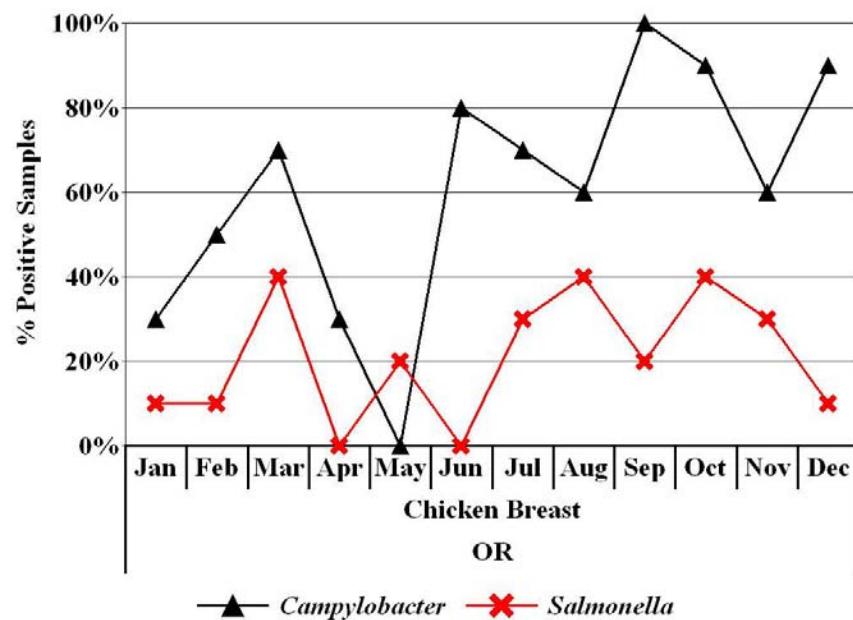


Figure 3m. Percent Positive Samples for *Enterococcus* & *E. coli* by Month and Meat Type in Oregon, 2004

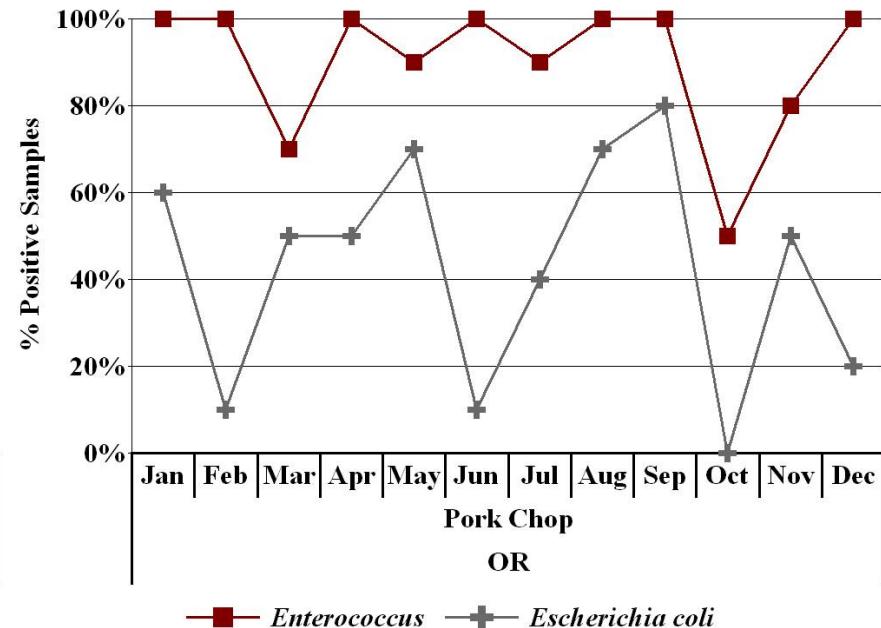
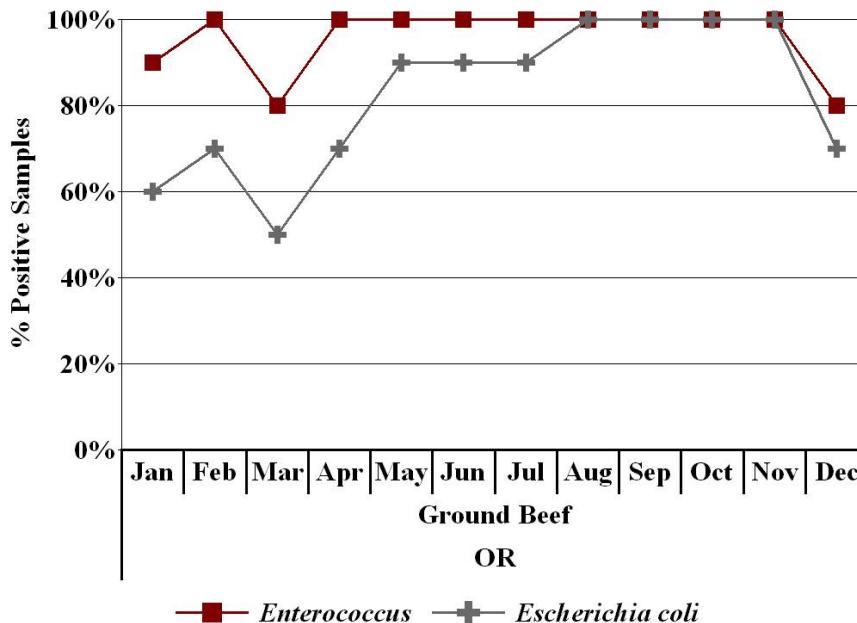
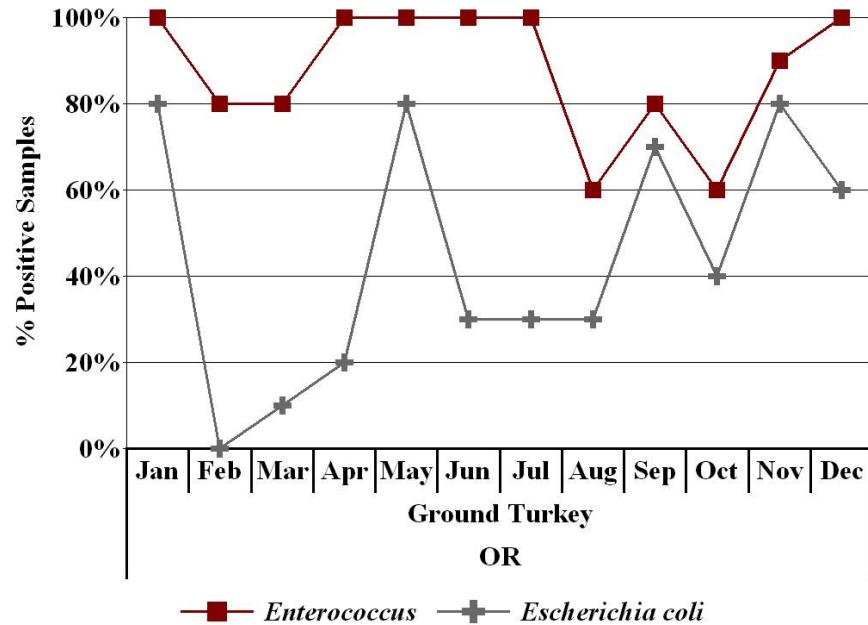
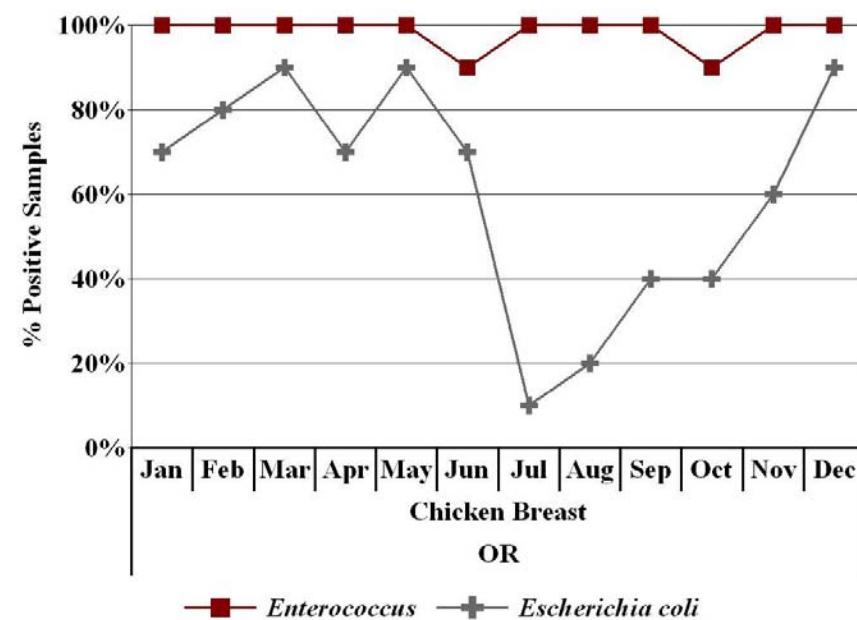


Figure 3n. Percent Positive Samples for *Campylobacter* & *Salmonella* by Month and Meat Type in Tennessee, 2004

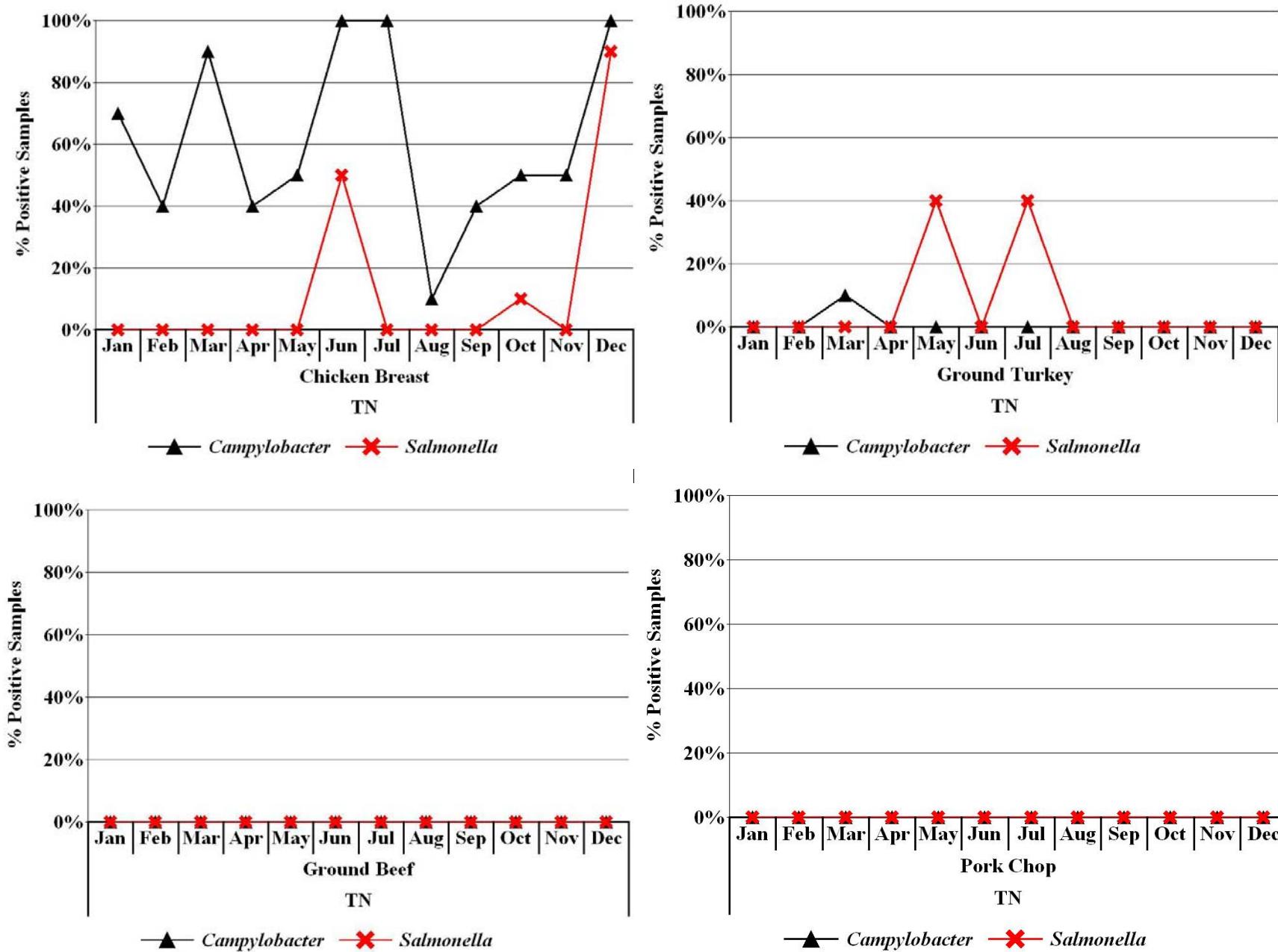


Figure 3o. Percent Positive Samples for *Enterococcus* & *E. coli* by Month and Meat Type in Tennessee, 2004

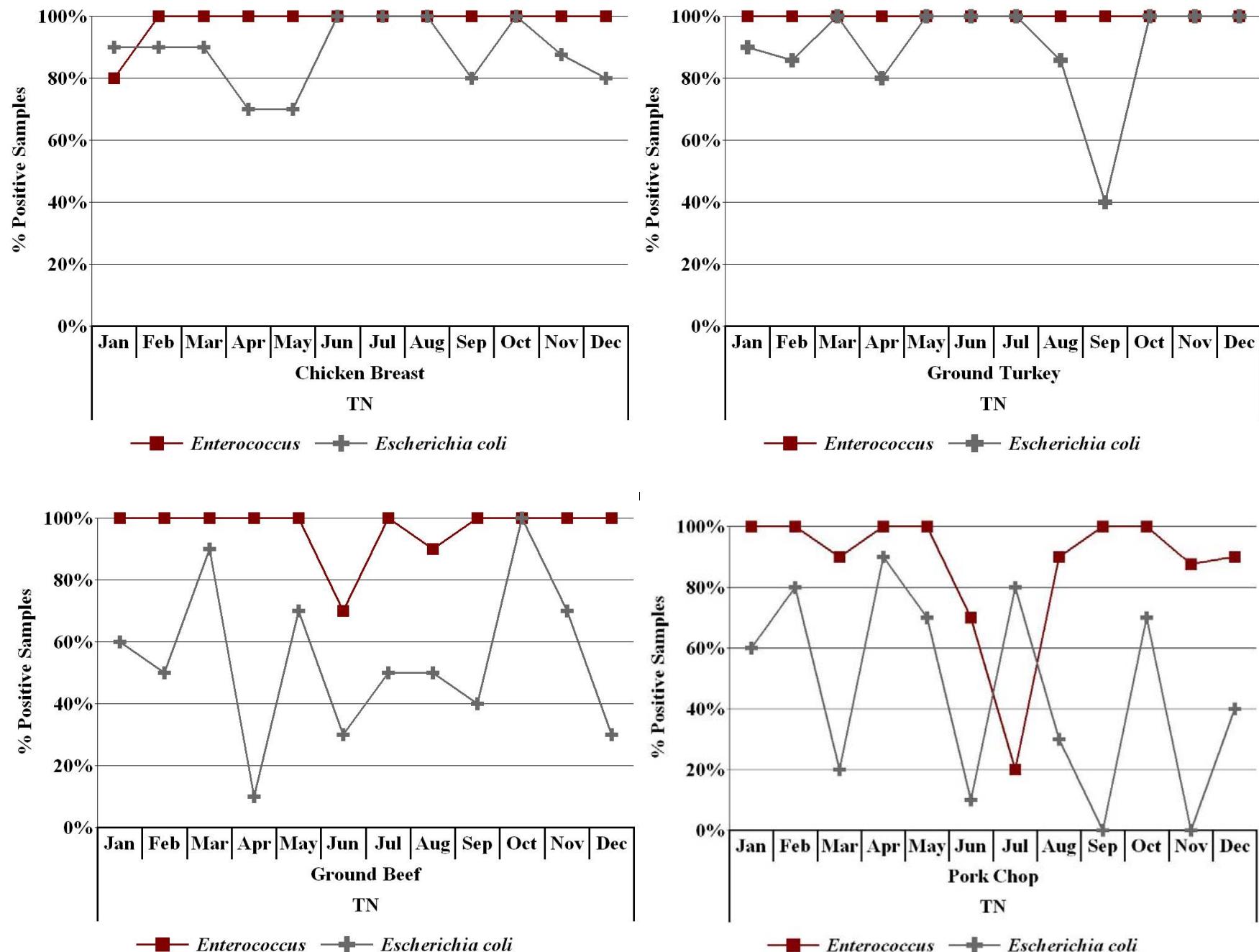


Table 5. Overall *Salmonella* Serotypes Identified, 2004

Serotype	n
1. <i>S. Heidelberg</i>	71
2. <i>S. Typhimurium</i>	53
3. <i>S. Kentucky</i>	43
4. <i>S. Saintpaul</i>	24
5. <i>S. Schwarzengrund</i>	21
6. <i>S. Hadar</i>	19
7. <i>S. Reading</i>	16
8. <i>S. Braenderup</i>	11
9. <i>S. Muenster</i>	10
10. <i>S. Agona</i>	9
11. <i>S. III 18a: z4, z32:-</i>	6
12. <i>S. Berta</i>	5
13. <i>S. Montevideo</i>	5
14. <i>S. Mbandaka</i>	4
15. <i>S. Newport</i>	4
16. <i>S. I 4, 12 : I :-</i>	4
17. <i>S. Derby</i>	3
18. <i>S. Enteritidis</i>	3
19. <i>S. IIIa 18: z4, z23: -</i>	2
20. <i>S. I 4, 12 : r :-</i>	2
21. <i>S. Senftenberg</i>	2
22. <i>S. Bredeney</i>	1
23. <i>S. Dublin</i>	1
24. <i>S. Livingstone</i>	1
25. <i>S. Minnesota</i>	1
26. <i>S. Muenchen</i>	1
27. <i>S. I 4, 12 : d :-</i>	1
28. <i>S. Urbana</i>	1
Total	324

Table 6. *Salmonella* by Serotype and Meat Type, 2004

Serotype	Chicken Breast		Ground Turkey		Ground Beef		Pork Chop	
	n	%*	n	%	n	%	n	%
<i>S.</i> Heidelberg (n=71)	31	43.7%	37	52.1%	0	-†	3	4.2%
<i>S.</i> Typhimurium‡ (n=53)	49	92.5%	2	3.8%	0	-	2	3.8%
<i>S.</i> Kentucky (n=43)	42	97.7%	1	2.3%	0	-	0	-
<i>S.</i> Saintpaul (n=24)	0	-	24	100.0%	0	-	0	-
<i>S.</i> Schwarzengrund (n=21)	5	23.8%	16	76.2%	0	-	0	-
<i>S.</i> Hadar (n=19)	8	42.1%	11	57.9%	0	-	0	-
<i>S.</i> Reading (n=16)	0	-	16	100.0%	0	-	0	-
<i>S.</i> Braenderup (n=11)	1	9.1%	0	-	5	45.5%	5	45.5%
<i>S.</i> Muenster (n=10)	1	10.0%	4	40.0%	5	50.0%	0	-
<i>S.</i> Agona (n=9)	2	22.2%	6	66.7%	0	-	1	11.1%
<i>S.</i> III 18a: z4, z32: - (n=6)	0	-	6	100.0%	0	-	0	-
<i>S.</i> Berta (n=5)	2	40.0%	2	40.0%	1	20.0%	0	-
<i>S.</i> Montevideo (n=5)	3	60.0%	2	40.0%	0	-	0	-
<i>S.</i> Mbandaka (n=4)	4	100.0%	0	-	0	-	0	-
<i>S.</i> Newport (n=4)	0	-	2	50.0%	2	50.0%	0	-
<i>S.</i> I 4, 12 : i : - (n=4)	4	100.0%	0	-	0	-	0	-
<i>S.</i> Derby (n=3)	0	-	3	100.0%	0	-	0	-
<i>S.</i> Enteritidis (n=3)	3	100.0%	0	-	0	-	0	-
<i>S.</i> III 18a: z4, z23: - (n=2)	0	-	2	100.0%	0	-	0	-
<i>S.</i> I 4, 12 : r : - (n=2)	0	-	2	100.0%	0	-	0	-
<i>S.</i> Senftenberg (n=2)	0	-	2	100.0%	0	-	0	-
<i>S.</i> Bredeney (n=1)	0	-	1	100.0%	0	-	0	-
<i>S.</i> Dublin (n=1)	0	-	0	-	1	100.0%	0	-
<i>S.</i> Livingstone (n=1)	1	100.0%	0	-	0	-	0	-
<i>S.</i> Minnesota (n=1)	0	-	1	100.0%	0	-	0	-
<i>S.</i> Muenchen (n=1)	1	100.0%	0	-	0	-	0	-
<i>S.</i> I 4, 12 : d : - (n=1)	0	-	1	100.0%	0	-	0	-
<i>S.</i> Urbana (n=1)	0	-	1	100.0%	0	-	0	-
Total 324	157	48.5%	142	43.8%	14	4.3%	11	3.4%

* Where % = (# isolates per serotype per meat) / (total # isolates per serotype).

† Dashes indicate that no isolates from serotype were isolated from meat type.

‡ Includes Typhimurium var. 5-.

Table 7. *Salmonella* Serotype by Site and Meat Type, 2004

Site	Serotype	Chicken Breast		Ground Turkey		Ground Beef		Pork Chop	
		n	% *	n	%	n	%	n	%
CA	<i>S. Heidelberg</i> (n=8)	4	50.0%	4	50.0%	0	- †	0	-
	<i>S. Kentucky</i> (n=5)	5	100.0%	0	-	0	-	0	-
	<i>S. Typhimurium</i> ‡ (n=3)	1	33.3%	1	33.3%	0	-	1	33.3%
	<i>S. III 18a: z4, z32: -</i> (n=2)	0	-	2	100.0%	0	-	0	-
	<i>S. Montevideo</i> (n=2)	2	100.0%	0	-	0	-	0	-
	<i>S. Hadar</i> (n=2)	2	100.0%	0	-	0	-	0	-
	<i>S. Agona</i> (n=1)	1	100.0%	0	-	0	-	0	-
	<i>S. Braenderup</i> (n=1)	1	100.0%	0	-	0	-	0	-
	<i>S. Saintpaul</i> (n=1)	0	-	1	100.0%	0	-	0	-
	<i>S. Reading</i> (n=1)	0	-	1	100.0%	0	-	0	-
	<i>S. Dublin</i> (n=1)	0	-	0	-	1	100.0%	0	-
	<i>S. Livingstone</i> (n=1)	1	100.0%	0	-	0	-	0	-
	Total (n=28)	17	60.7%	9	32.1%	1	3.6%	1	3.6%
CO	<i>S. Heidelberg</i> (n=4)	0	-	4	100.0%	0	-	0	-
	<i>S. Saintpaul</i> (n=2)	0	-	2	100.0%	0	-	0	-
	<i>S. Reading</i> (n=1)	0	-	1	100.0%	0	-	0	-
	<i>S. Agona</i> (n=1)	1	100.0%	0	-	0	-	0	-
	<i>S. Minnesota</i> (n=1)	0	-	1	100.0%	0	-	0	-
	Total (n=9)	1	11.1%	8	88.9%	0	-	0	-
CT	<i>S. Typhimurium</i> (n=20)	19	95.0%	1	5.0%	0	-	0	-
	<i>S. Heidelberg</i> (n=10)	2	20.0%	8	80.0%	0	-	0	-
	<i>S. Braenderup</i> (n=10)	0	-	0	-	5	50.0%	5	50.0%
	<i>S. Schwarzengrund</i> (n=9)	0	-	9	100.0%	0	-	0	-
	<i>S. Kentucky</i> (n=8)	8	100.0%	0	-	0	-	0	-
	<i>S. Saintpaul</i> (n=3)	0	-	3	100.0%	0	-	0	-
	<i>S. III 18a: z4, z32: -</i> (n=2)	0	-	2	100.0%	0	-	0	-
	<i>S. I 4, 12 : r : -</i> (n=2)	0	-	2	100.0%	0	-	0	-
	<i>S. Enteritidis</i> (n=1)	1	100.0%	0	-	0	-	0	-
	<i>S. Bredeney</i> (n=1)	0	-	1	100.0%	0	-	0	-
	Total (n=66)	30	45.5%	26	39.4%	5	7.6%	5	7.6%
	<i>S. Heidelberg</i> (n=12)	1	8.3%	11	91.7%	0	-	0	-
GA	<i>S. Saintpaul</i> (n=6)	0	-	6	100.0%	0	-	0	-
	<i>S. Reading</i> (n=5)	0	-	5	100.0%	0	-	0	-
	<i>S. Agona</i> (n=4)	0	-	4	100.0%	0	-	0	-
	<i>S. III 18a: z4, z32: -</i> (n=3)	0	-	3	100.0%	0	-	0	-
	<i>S. Newport</i> (n=3)	0	-	2	66.7%	1	33.3%	0	-
	<i>S. Kentucky</i> (n=2)	2	100.0%	0	-	0	-	0	-
	<i>S. Schwarzengrund</i> (n=2)	0	-	2	100.0%	0	-	0	-
	<i>S. Montevideo</i> (n=2)	0	-	2	100.0%	0	-	0	-
	<i>S. Mbandaka</i> (n=2)	2	100.0%	0	-	0	-	0	-
	<i>S. Hadar</i> (n=1)	0	-	1	100.0%	0	-	0	-
	<i>S. Derby</i> (n=1)	0	-	1	100.0%	0	-	0	-
	<i>S. Enteritidis</i> (n=1)	1	100.0%	0	-	0	-	0	-
	<i>S. Senftenberg</i> (n=1)	0	-	1	100.0%	0	-	0	-
	Total (n=45)	6	13.3%	38	84.4%	1	2.2%	0	-

* Where % = (# isolates per serotype per meat type per site)/(total # isolates per serotype per site).

† Dashes indicate that no isolates from serotype were isolated from meat type.

‡ Includes Typhimurium var. 5-.

Table 7_(cont'd). *Salmonella* Serotype by Site and Meat Type, 2004

Site	Serotype	Chicken Breast		Ground Turkey		Ground Beef		Pork Chop	
		n	%	n	%	n	%	n	%
MD	<i>S. Typhimurium</i> (n=14)	14	100.0%	0	-	0	-	0	-
	<i>S. Muenster</i> (n=5)	1	20.0%	4	80.0%	0	-	0	-
	<i>S. Schwarzengrund</i> (n=4)	2	50.0%	2	50.0%	0	-	0	-
	<i>S. Berta</i> (n=3)	0	-	2	66.7%	1	33.3%	0	-
	<i>S. Kentucky</i> (n=3)	3	100.0%	0	-	0	-	0	-
	<i>S. Derby</i> (n=2)	0	-	2	100.0%	0	-	0	-
	<i>S. Enteritidis</i> (n=1)	1	100.0%	0	-	0	-	0	-
	<i>S. Heidelberg</i> (n=1)	1	100.0%	0	-	0	-	0	-
	<i>S. Hadar</i> (n=1)	1	100.0%	0	-	0	-	0	-
	<i>S. Montevideo</i> (n=1)	1	100.0%	0	-	0	-	0	-
	<i>S. Senftenberg</i> (n=1)	0	-	1	100.0%	0	-	0	-
	<i>S. I 4, 12 : d :-</i> (n=1)	0	-	1	100.0%	0	-	0	-
	<i>S. Urbana</i> (n=1)	0	-	1	100.0%	0	-	0	-
	Total (n=38)	24	63.2%	13	34.2%	1	2.6%	0	-
MN	<i>S. Heidelberg</i> (n=10)	7	70.0%	3	30.0%	0	-	0	-
	<i>S. Kentucky</i> (n=10)	9	90.0%	1	10.0%	0	-	0	-
	<i>S. Hadar</i> (n=5)	0	-	5	100.0%	0	-	0	-
	<i>S. Reading</i> (n=4)	0	-	4	100.0%	0	-	0	-
	<i>S. Mbandaka</i> (n=2)	2	100.0%	0	-	0	-	0	-
	<i>S. Berta</i> (n=2)	2	100.0%	0	-	0	-	0	-
	<i>S. Saintpaul</i> (n=1)	0	-	1	100.0%	0	-	0	-
	Total (n=34)	20	58.8%	14	41.2%	0	-	0	-
NM	<i>S. Reading</i> (n=5)	0	-	5	100.0%	0	-	0	-
	<i>S. Heidelberg</i> (n=3)	1	33.3%	2	66.7%	0	-	0	-
	<i>S. Saintpaul</i> (n=2)	0	-	2	100.0%	0	-	0	-
	<i>S. Schwarzengrund</i> (n=1)	1	100.0%	0	-	0	-	0	-
	<i>S. Kentucky</i> (n=1)	1	100.0%	0	-	0	-	0	-
	Total (n=12)	3	25.0%	9	75.0%	0	-	0	-
NY	<i>S. Kentucky</i> (n=10)	10	100.0%	0	-	0	-	0	-
	<i>S. Typhimurium</i> (n=7)	6	85.7%	0	-	0	-	1	14.3%
	<i>S. Saintpaul</i> (n=3)	0	-	3	100.0%	0	-	0	-
	<i>S. Schwarzengrund</i> (n=3)	0	-	3	100.0%	0	-	0	-
	<i>S. Agona</i> (n=3)	0	-	2	66.7%	0	-	1	33.3%
	<i>S. Heidelberg</i> (n=2)	0	-	1	50.0%	0	-	1	50.0%
	<i>S. Hadar</i> (n=1)	0	-	1	100.0%	0	-	0	-
	<i>S. III 18a: z4, z32: -</i> (n=1)	0	-	1	100.0%	0	-	0	-
	Total (n=30)	16	53.3%	11	36.7%	0	-	3	10.0%
OR	<i>S. Heidelberg</i> (n=21)	15	71.4%	4	19.0%	0	-	2	9.5%
	<i>S. Hadar</i> (n=5)	5	100.0%	0	-	0	-	0	-
	<i>S. Muenster</i> (n=5)	0	-	0	-	5	100.0%	0	-
	<i>S. Kentucky</i> (n=3)	3	100.0%	0	-	0	-	0	-
	<i>S. Saintpaul</i> (n=2)	0	-	2	100.0%	0	-	0	-
	<i>S. Schwarzengrund</i> (n=2)	2	100.0%	0	-	0	-	0	-
	<i>S. Newport</i> (n=1)	0	-	0	-	1	100.0%	0	-
	Total (n=39)	25	64.1%	6	15.4%	6	15.4%	2	5.1%

Table 7_(cont'd). *Salmonella* Serotype by Site and Meat Type, 2004

Site	Serotype	Chicken Breast		Ground Turkey		Ground Beef		Pork Chop	
		n	%	n	%	n	%	n	%
TN	<i>S. Typhimurium</i> (n=9)	9	100.0%	0	-	0	-	0	-
	<i>S. Hadar</i> (n=4)	0	-	4	100.0%	0	-	0	-
	<i>S. Saintpaul</i> (n=4)	0	-	4	100.0%	0	-	0	-
	<i>S. I 4, 12 : i : -</i> (n=4)	4	100.0%	0	-	0	-	0	-
	<i>S. Kentucky</i> (n=1)	1	100.0%	0	-	0	-	0	-
	<i>S. Muenchen</i> (n=1)	1	100.0%	0	-	0	-	0	-
	Total (n=23)	15	65.2%	8	34.8%	0	-	0	-
Grand Total (N=324)		157	48.5%	142	43.8%	14	4.3%	11	3.4%

Table 8. *Salmonella* Isolates by Month for All Sites, 2004

Month	n	%*
January	49	15.1%
February	18	5.6%
March	21	6.5%
April	28	8.6%
May	25	7.7%
June	23	7.1%
July	32	9.9%
August	34	10.5%
September	19	5.9%
October	24	7.4%
November	28	8.6%
December	23	7.1%
Total (N)	324	100.0%

* Where % = (n/N).

Table 9. Antimicrobial Resistance among *Salmonella* Isolates (N=324), 2004

Antimicrobial Agent	n	%R*
Tetracycline	161	49.7%
Streptomycin	98	30.2%
Sulfisoxazole	89	27.5%
Ampicillin	81	25.0%
Amoxicillin/Clavulanic Acid	52	16.0%
Cefoxitin	48	14.8%
Ceftiofur	48	14.8%
Kanamycin	45	13.9%
Gentamicin	35	10.8%
Chloramphenicol	11	3.4%
Ceftriaxone	1	0.3%
Trimethoprim/Sulfamethoxazole	1	0.3%
Amikacin	0	0.0%
Ciprofloxacin	0	0.0%
Nalidixic Acid	0	0.0%

* Where %R = (n/N).

Figure 4. Antimicrobial Resistance among *Salmonella* isolates (n =324), 2004

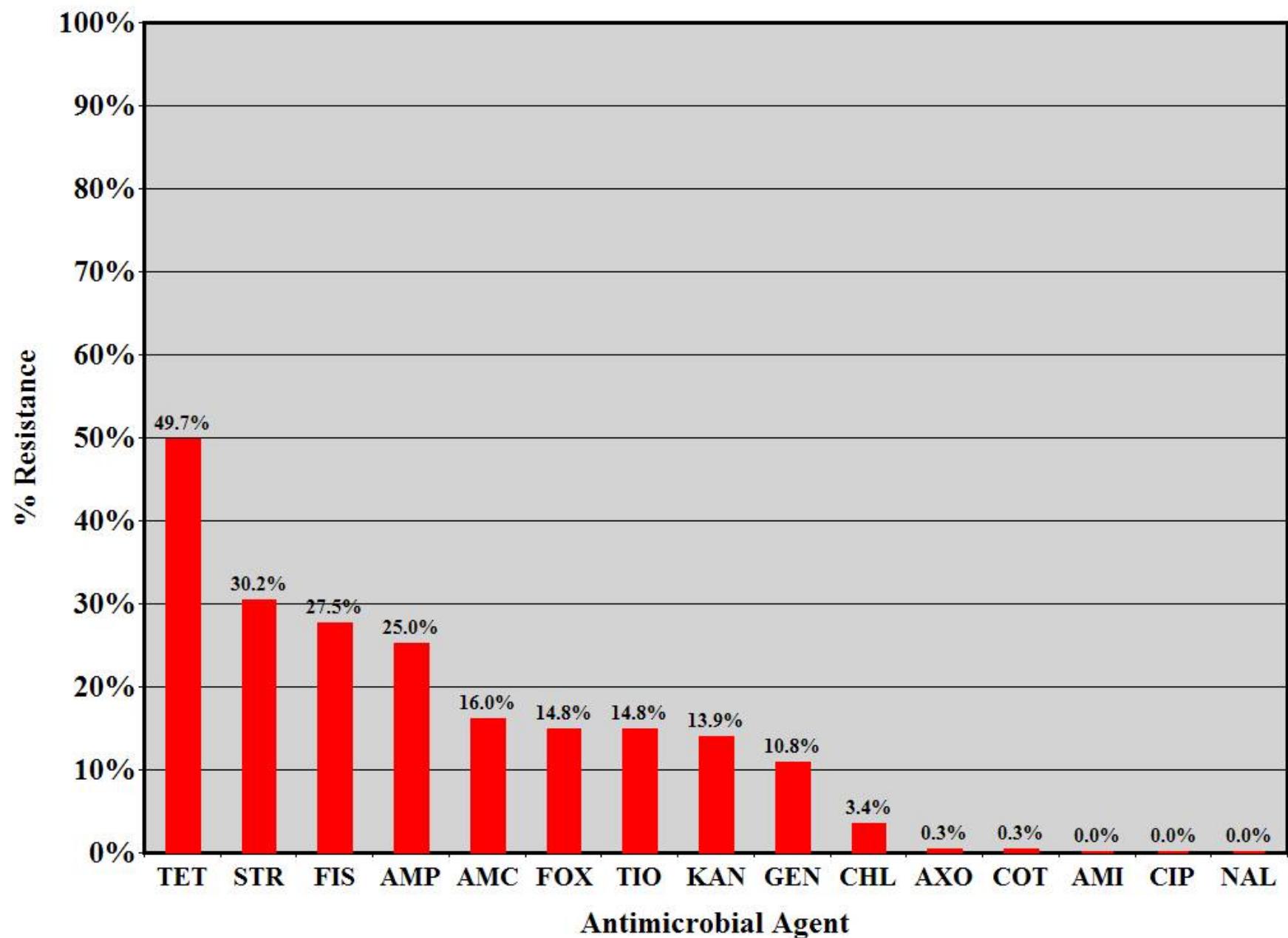


Figure 5. MIC Distribution among all Antimicrobial Agents, 2004

<i>Salmonella</i> from All Meats (N=324)		Distribution (%) of MICs (in µg/ml)															
Antimicrobial Agent	%R [†]	0.015	0.03	0.06	0.12	0.25	0.5	1	2	4	8	16	32	64	128	256	>256
Ampicillin	25.0%							63.6	10.5	0.9					25.0		
Amoxicillin/Clavulanic Acid	16.0%							67.0	8.0	4.0	4.9	1.2	14.8				
Cefoxitin	14.8%							1.9	59.0	20.7	3.1	0.6	3.1	11.7			
Ceftiofur	14.8%					0.3	46.3	36.7	1.9				14.8				
Ceftriaxone	0.3%						84.9			0.9	9.9	4.0	0.3				
Nalidixic Acid	0.0%							8.0	84.9	6.8	0.3						
Ciprofloxacin	0.0%	95.4	4.0	0.6								8.3	15.4	48.1	0.6		
Sulfisoxazole	27.5%											8.3	15.4	48.1	0.6	27.5	
Trimethoprim/Sulfamethoxazole	0.3%		93.5	4.3	1.9							0.3					
Amikacin	0.0%					4.6	49.4	41.0	4.9								
Gentamicin	10.8%					42.0	41.4	4.0	0.3	1.5	4.9	5.9					
Kanamycin	13.9%									82.7	2.5	0.9	3.1	10.8			
Streptomycin*	30.2%									69.8	17.3	13.0					
Chloramphenicol	3.4%							1.2	13.0	80.2	2.2			3.4			
Tetracycline	49.7%								46.6	3.7	1.9	0.9	46.9				

*Currently no CLSI breakpoints have been established for this organism/antimicrobial combination. Indicated breakpoints were established by NARMS.

[†]Discrepancies between %R and sums of distribution %s are due to rounding.

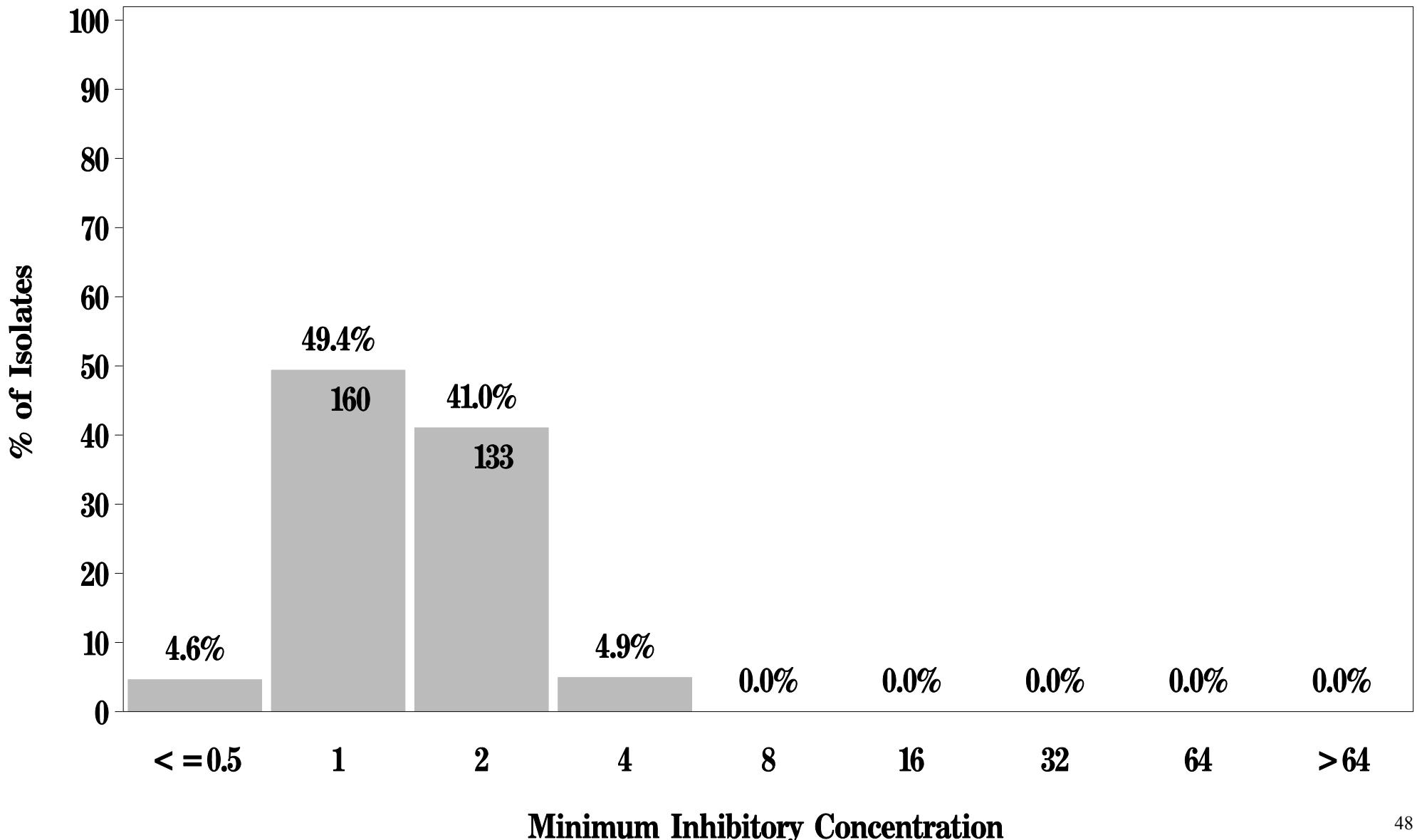
Vertical bars show the CLSI Susceptible/Resistant breakpoints for each drug.

Unshaded areas indicate the dilution ranges of the Sensititre plate used to test the 2004 isolates.

NARMS

**Figure 5a: Minimum Inhibitory Concentration of Amikacin
for *Salmonella* (N=324 Isolates)**

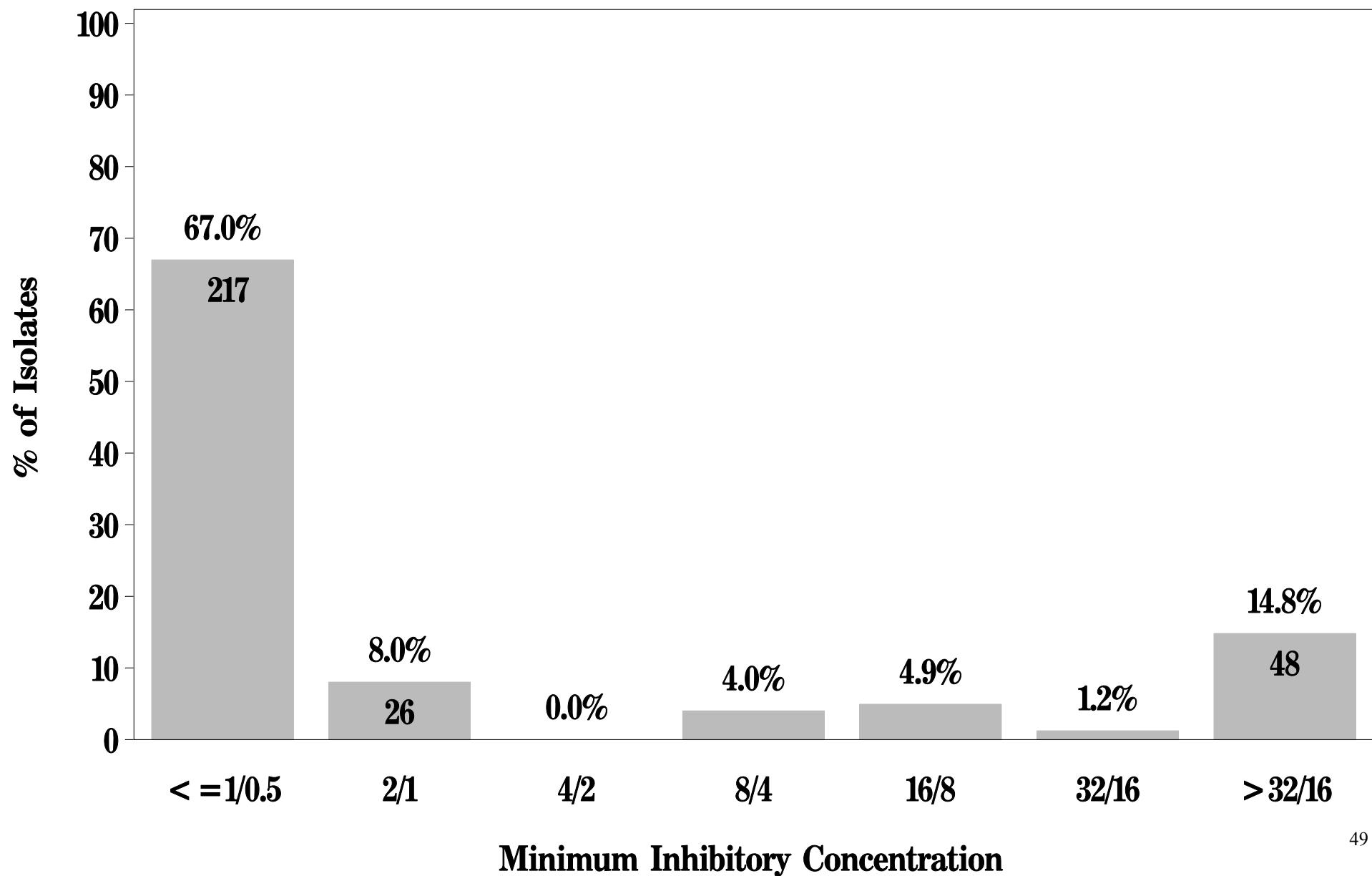
Breakpoints: Susceptible $\leq 16 \text{ } \mu\text{g/mL}$ Resistant $\geq 64 \text{ } \mu\text{g/mL}$



NARMS

**Figure 5b: Minimum Inhibitory Concentration of Amoxicillin/Clavulanic acid
for *Salmonella* (N=324 Isolates)**

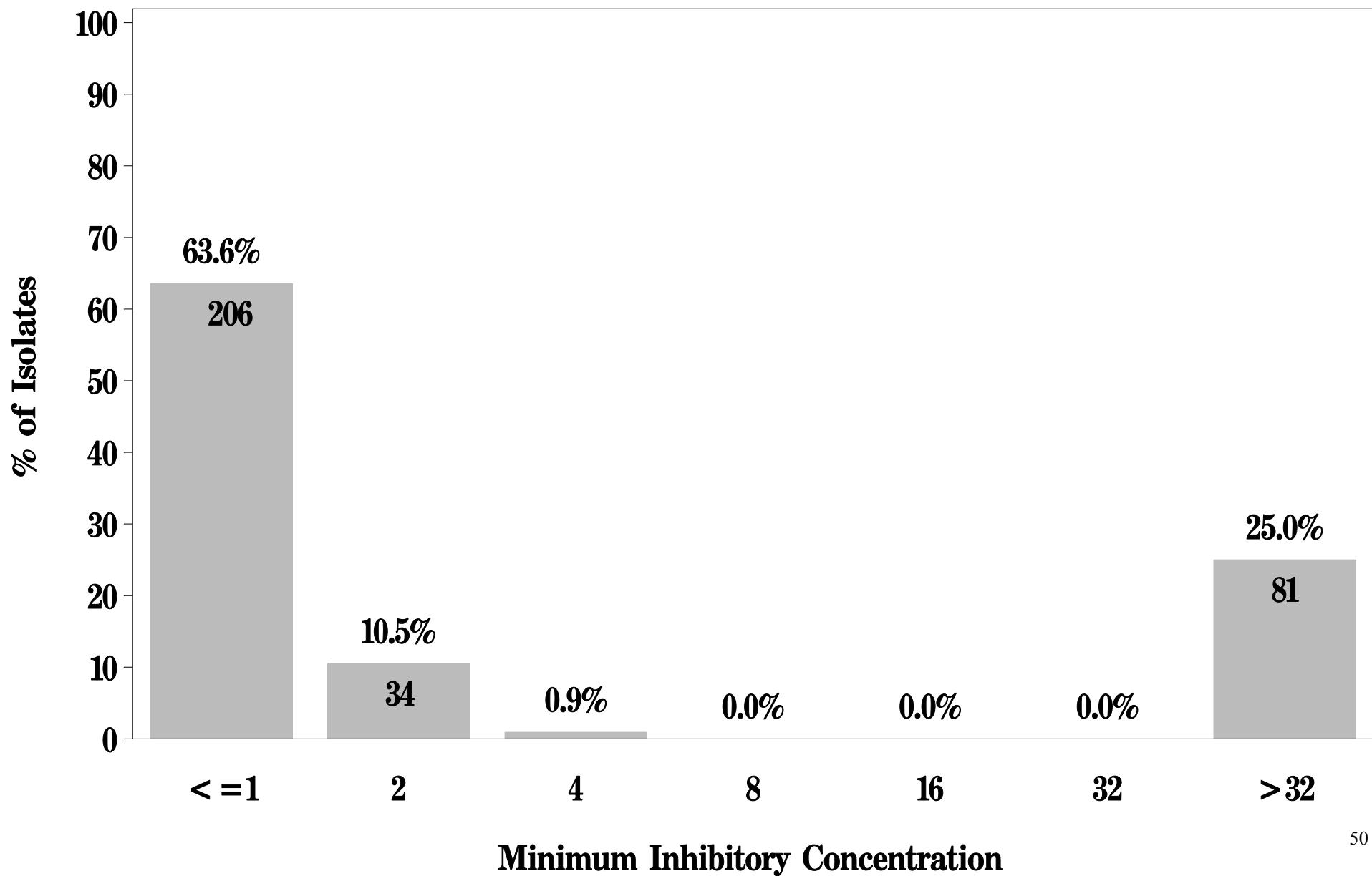
Breakpoints: Susceptible $\leq 8 \text{ } \mu\text{g/mL}$ Resistant $\geq 32 \text{ } \mu\text{g/mL}$



NARMS

**Figure 5c: Minimum Inhibitory Concentration of Ampicillin
for *Salmonella* (N=324 Isolates)**

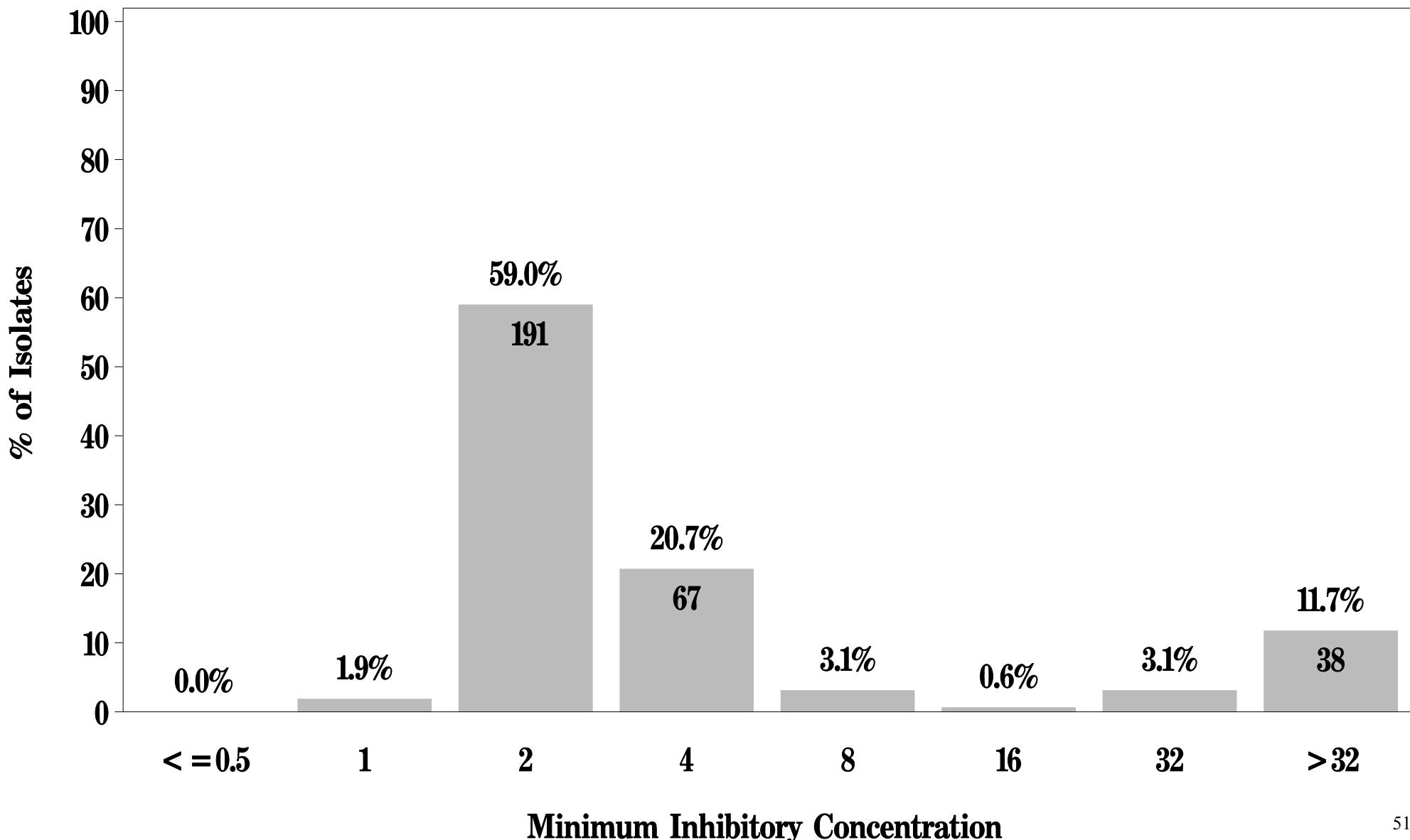
Breakpoints: Susceptible $\leq 8 \text{ } \mu\text{g/mL}$ Resistant $\geq 32 \text{ } \mu\text{g/mL}$



NARMS

**Figure 5d: Minimum Inhibitory Concentration of Cefoxitin
for *Salmonella* (N=324 Isolates)**

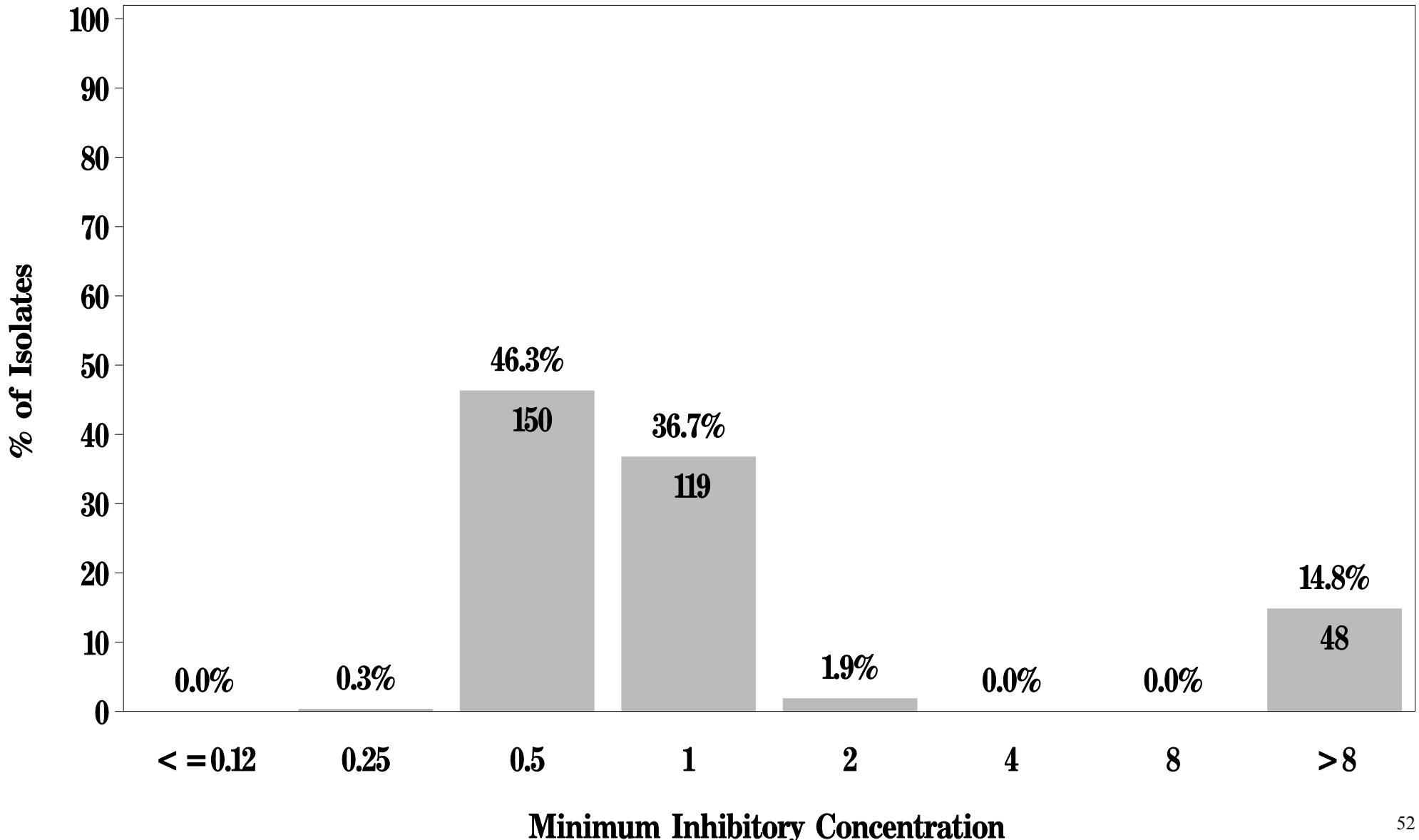
Breakpoints: Susceptible $\leq 8 \text{ } \mu\text{g/mL}$ Resistant $\geq 32 \text{ } \mu\text{g/mL}$



NARMS

**Figure 5e: Minimum Inhibitory Concentration of Ceftiofur
for *Salmonella* (N=324 Isolates)**

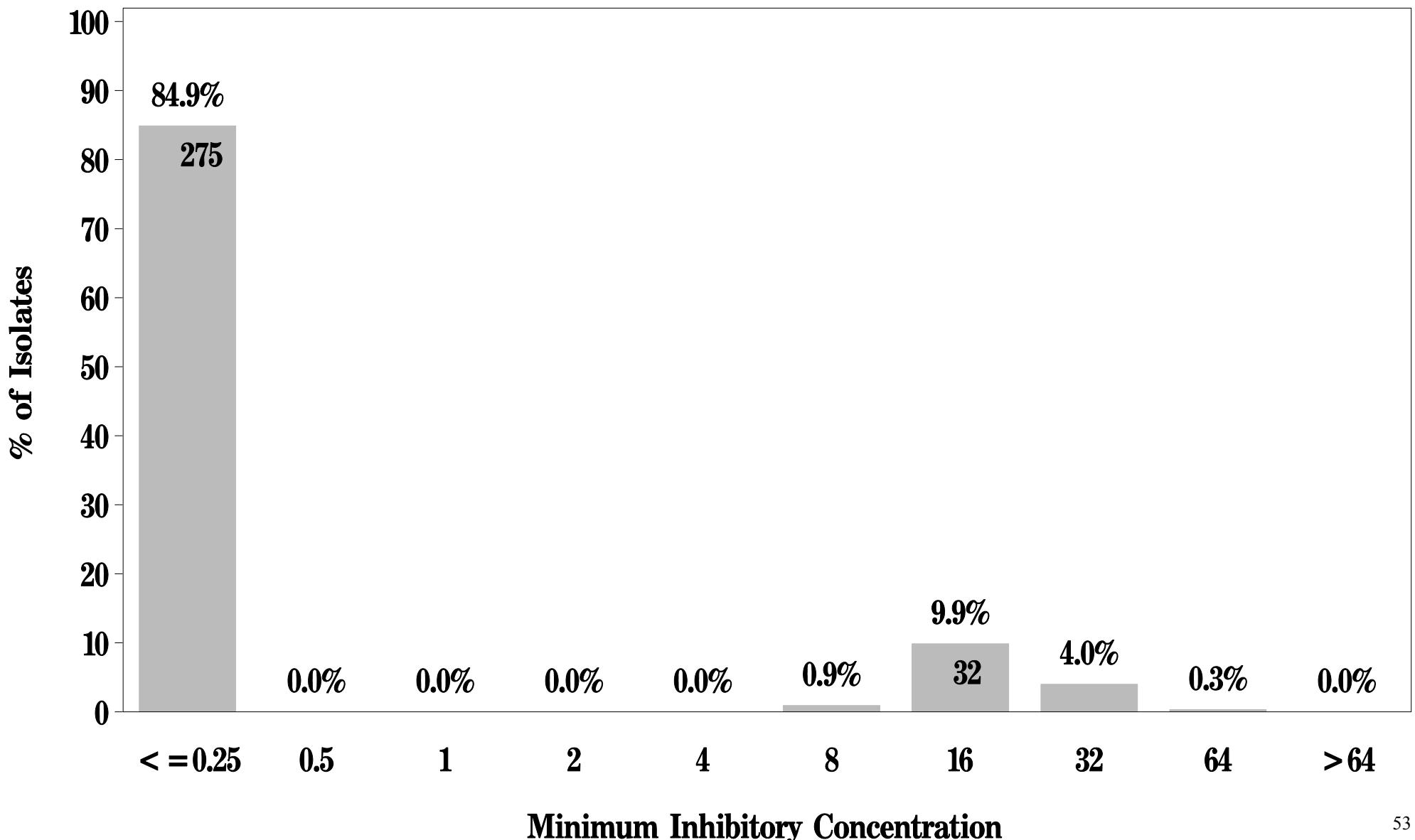
Breakpoints: Susceptible $\leq 2 \text{ } \mu\text{g/mL}$ Resistant $\geq 8 \text{ } \mu\text{g/mL}$



NARMS

**Figure 5f: Minimum Inhibitory Concentration of Ceftriaxone
for *Salmonella* (N=324 Isolates)**

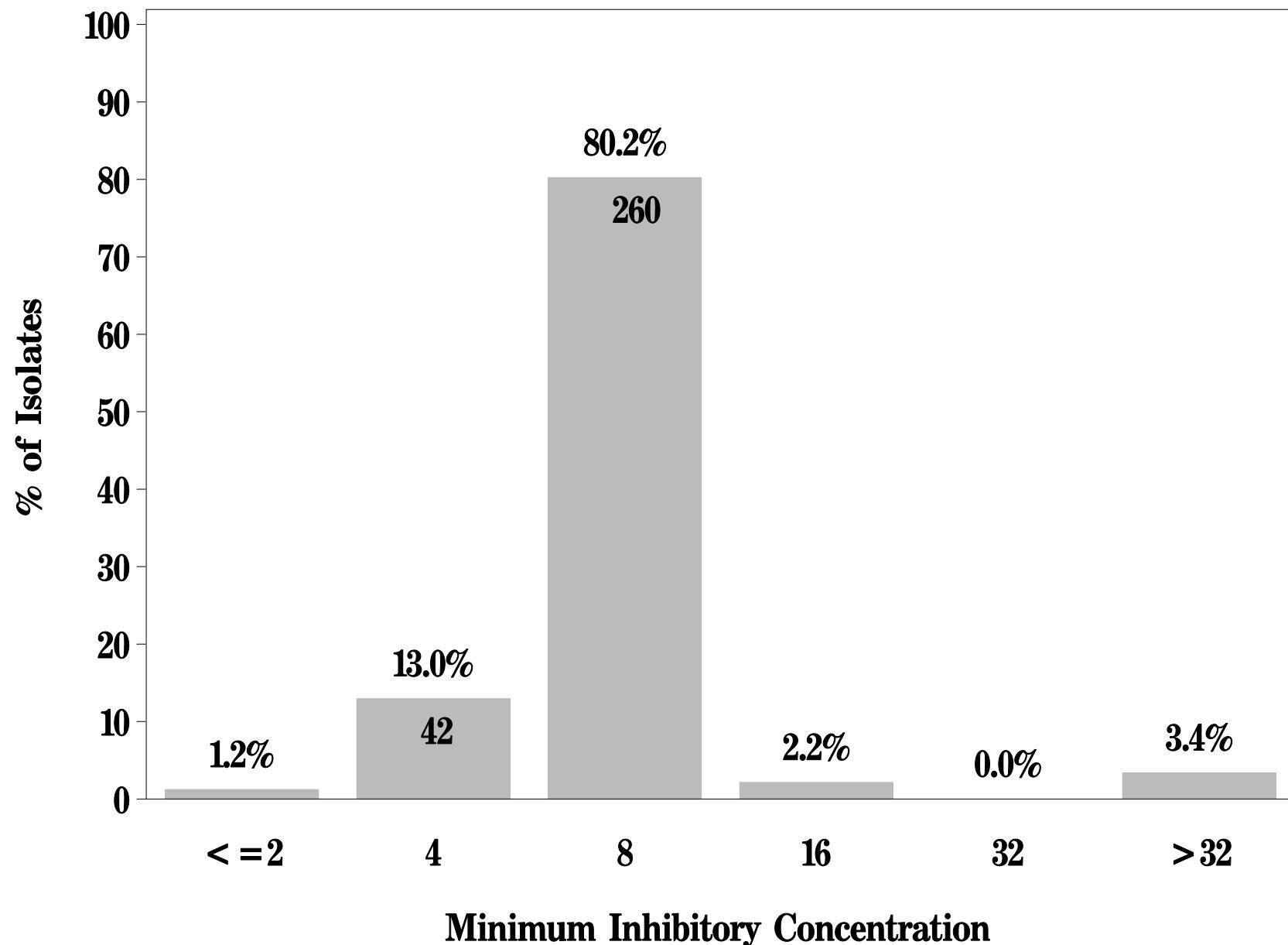
Breakpoints: Susceptible $\leq 8 \text{ } \mu\text{g/mL}$ Resistant $\geq 16 \text{ } \mu\text{g/mL}$



NARMS

**Figure 5g: Minimum Inhibitory Concentration of Chloramphenicol
for *Salmonella* (N=324 Isolates)**

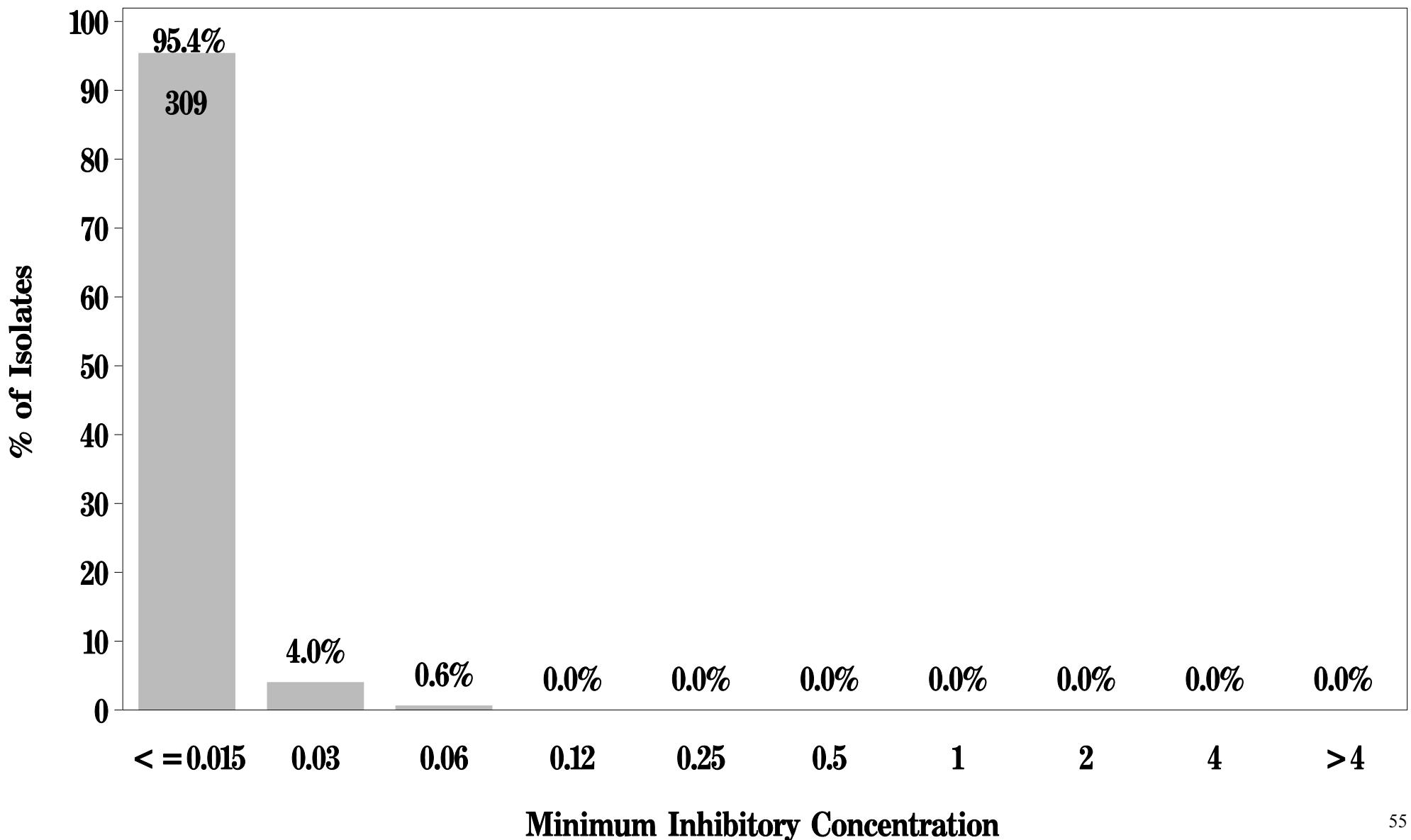
Breakpoints: Susceptible $\leq 8 \text{ } \mu\text{g/mL}$ Resistant $\geq 32 \text{ } \mu\text{g/mL}$



NARMS

**Figure 5h: Minimum Inhibitory Concentration of Ciprofloxacin
for *Salmonella* (N=324 Isolates)**

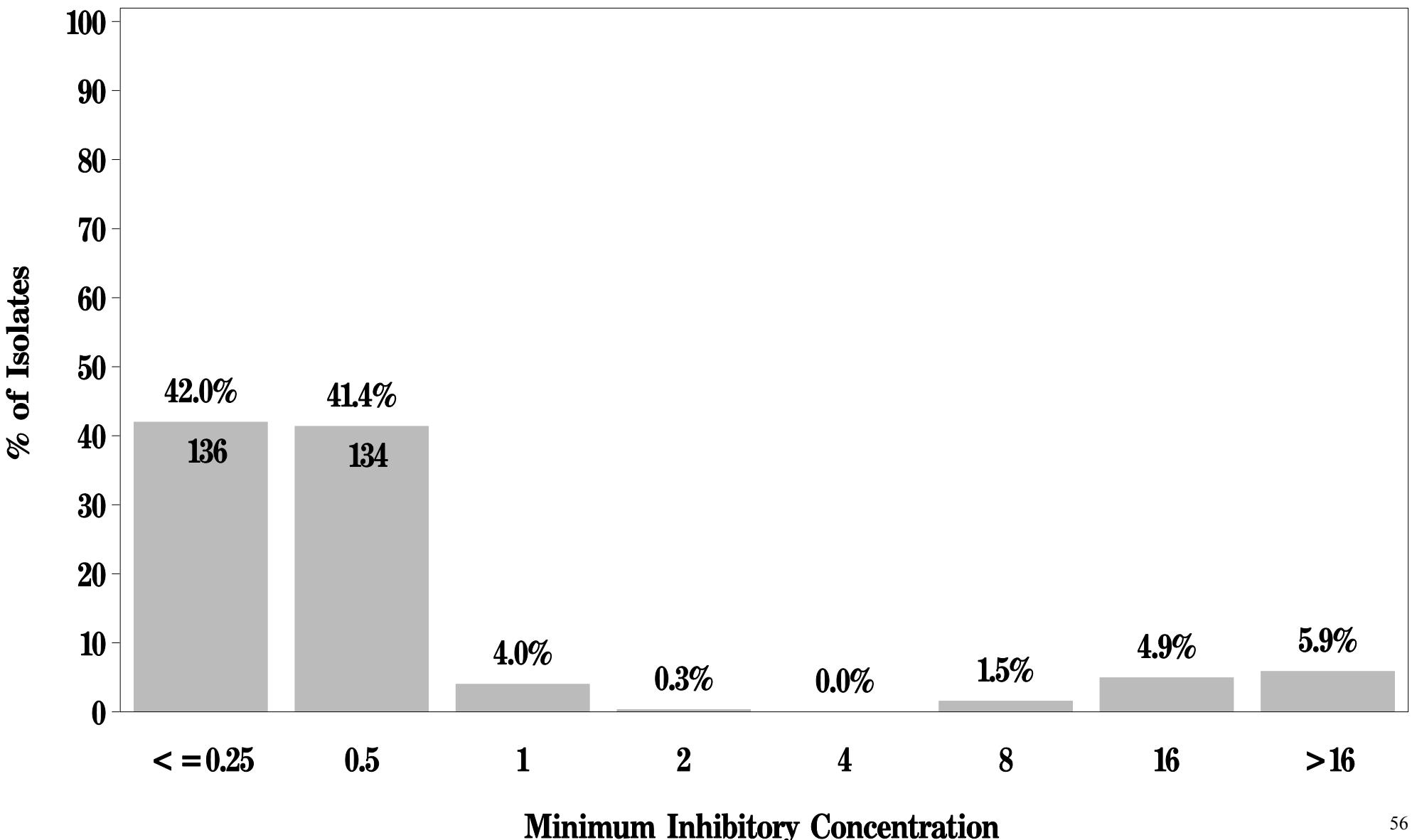
Breakpoints: Susceptible $\leq 1 \mu\text{g/mL}$ Resistant $\geq 4 \mu\text{g/mL}$



NARMS

**Figure 5i: Minimum Inhibitory Concentration of Gentamicin
for *Salmonella* (N=324 Isolates)**

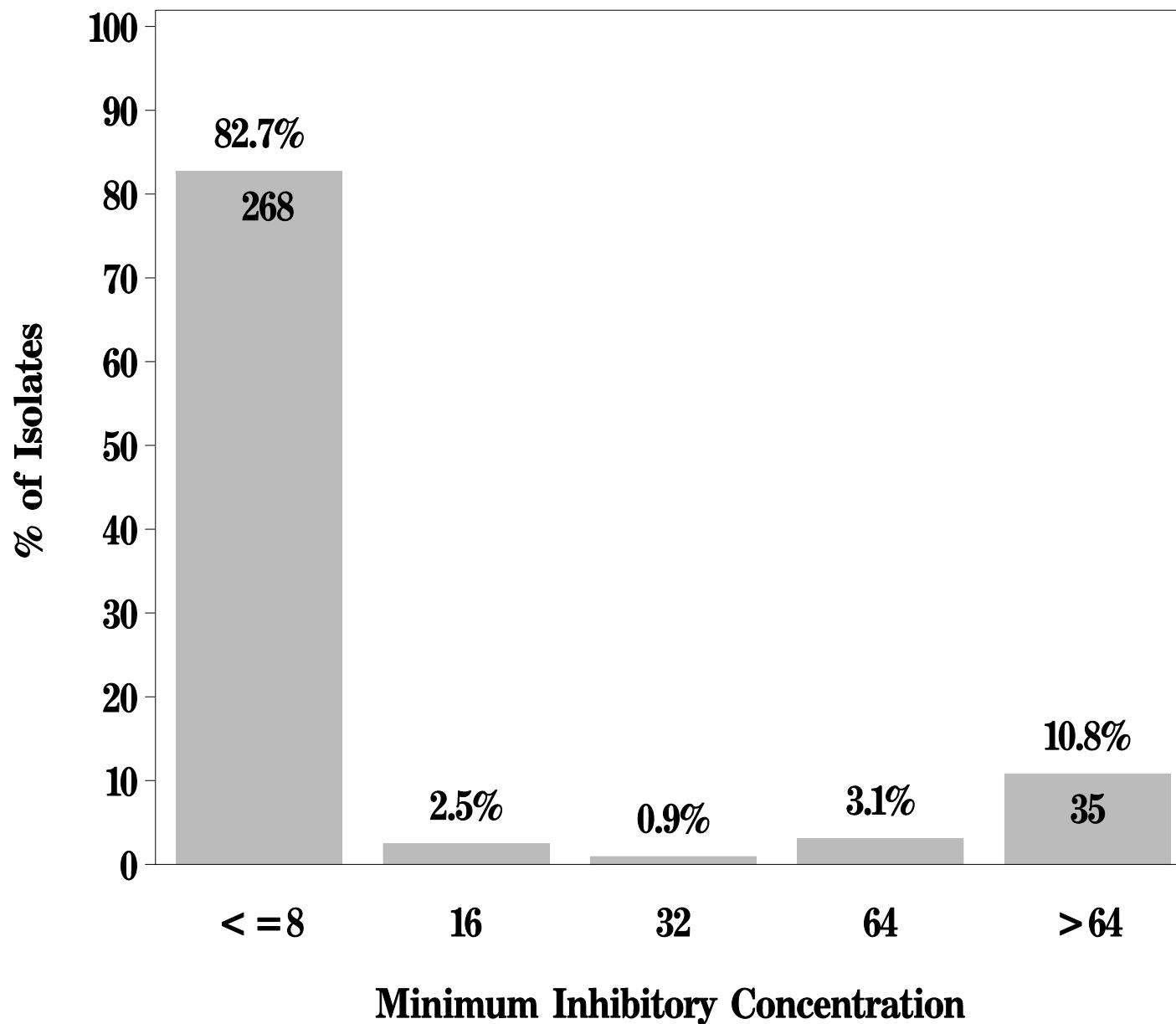
Breakpoints: Susceptible $\leq 4 \text{ } \mu\text{g/mL}$ Resistant $\geq 16 \text{ } \mu\text{g/mL}$



NARMS

**Figure 5j: Minimum Inhibitory Concentration of Kanamycin
for *Salmonella* (N=324 Isolates)**

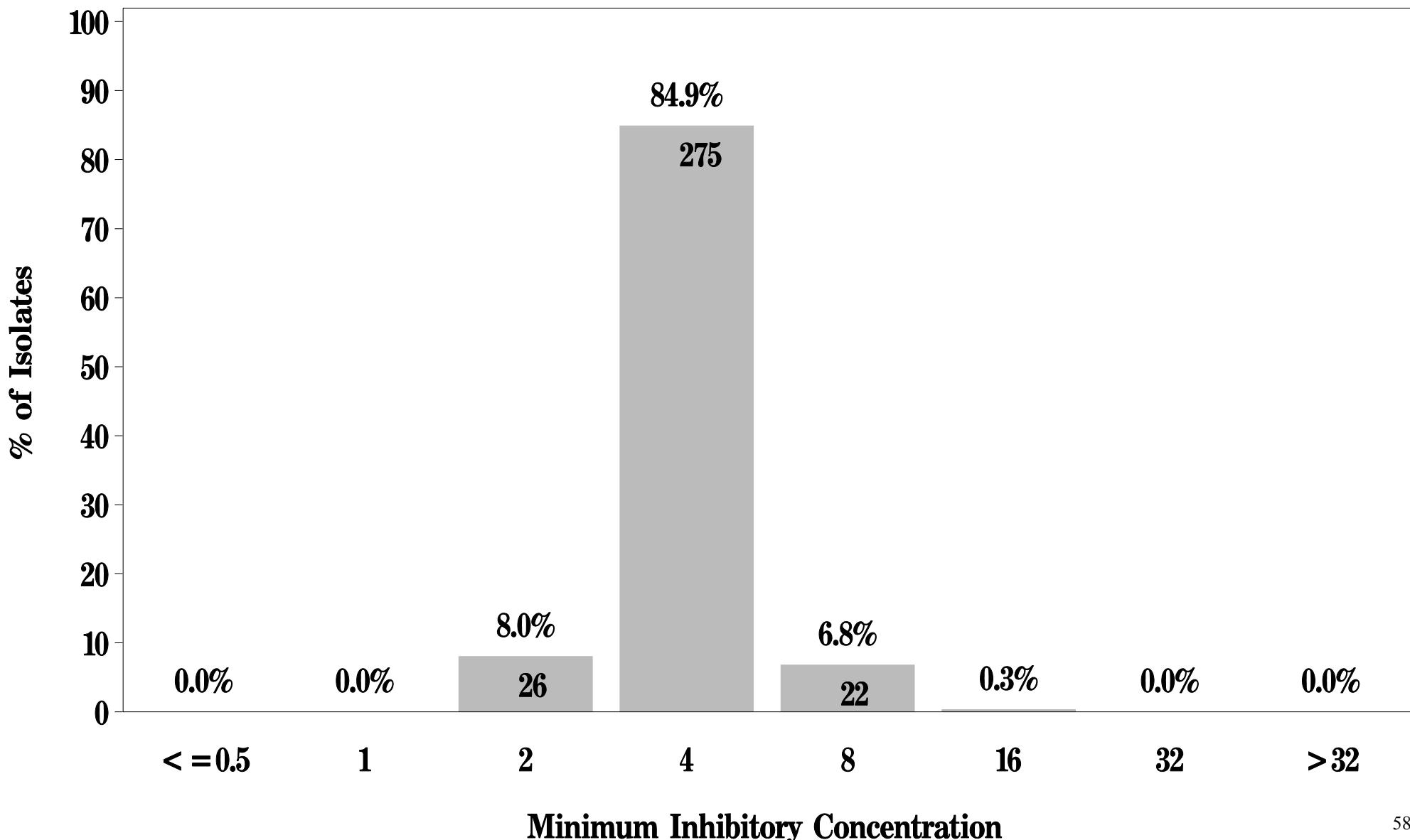
Breakpoints: Susceptible $\leq 16 \text{ } \mu\text{g/mL}$ Resistant $\geq 64 \text{ } \mu\text{g/mL}$



NARMS

**Figure 5k: Minimum Inhibitory Concentration of Nalidixic acid
for *Salmonella* (N=324 Isolates)**

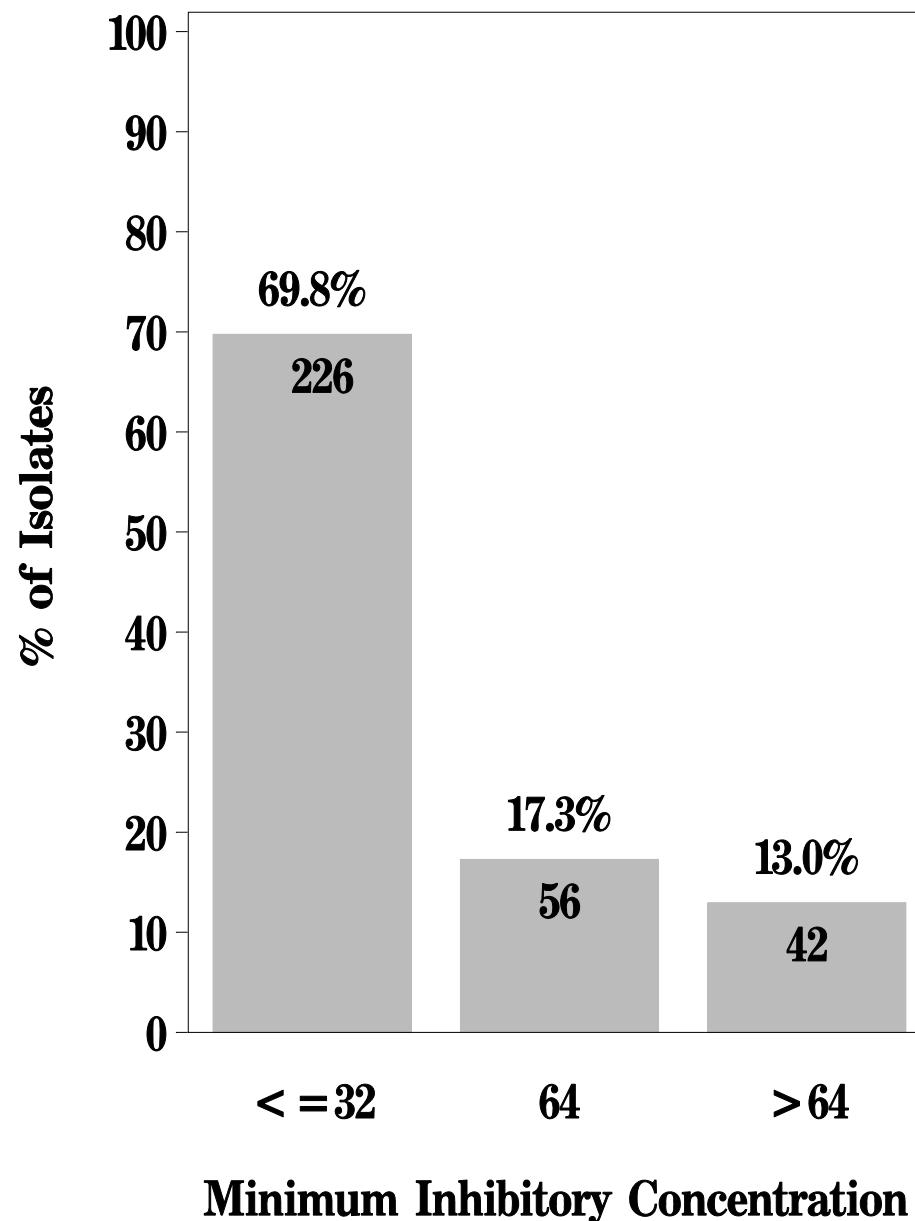
Breakpoints: Susceptible $\leq 16 \mu\text{g/mL}$ Resistant $\geq 32 \mu\text{g/mL}$



NARMS

**Figure 5l: Minimum Inhibitory Concentration of Streptomycin
for *Salmonella* (N = 324 Isolates)**

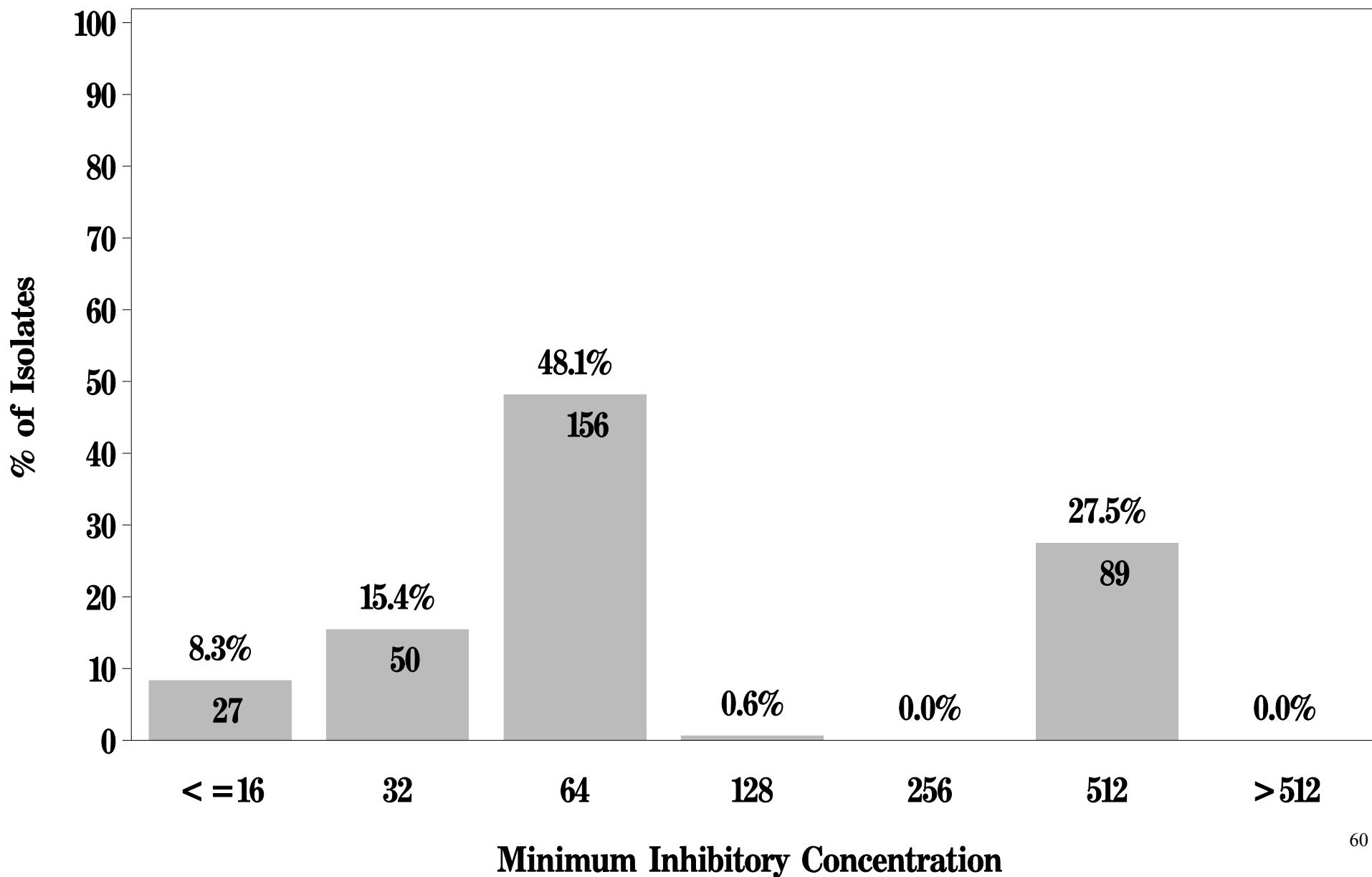
Breakpoints: Susceptible $\leq 32 \text{ } \mu\text{g/mL}$ Resistant $> 64 \text{ } \mu\text{g/mL}$



NARMS

**Figure 5m: Minimum Inhibitory Concentration of Sulfisoxazole
for *Salmonella* (N=324 Isolates)**

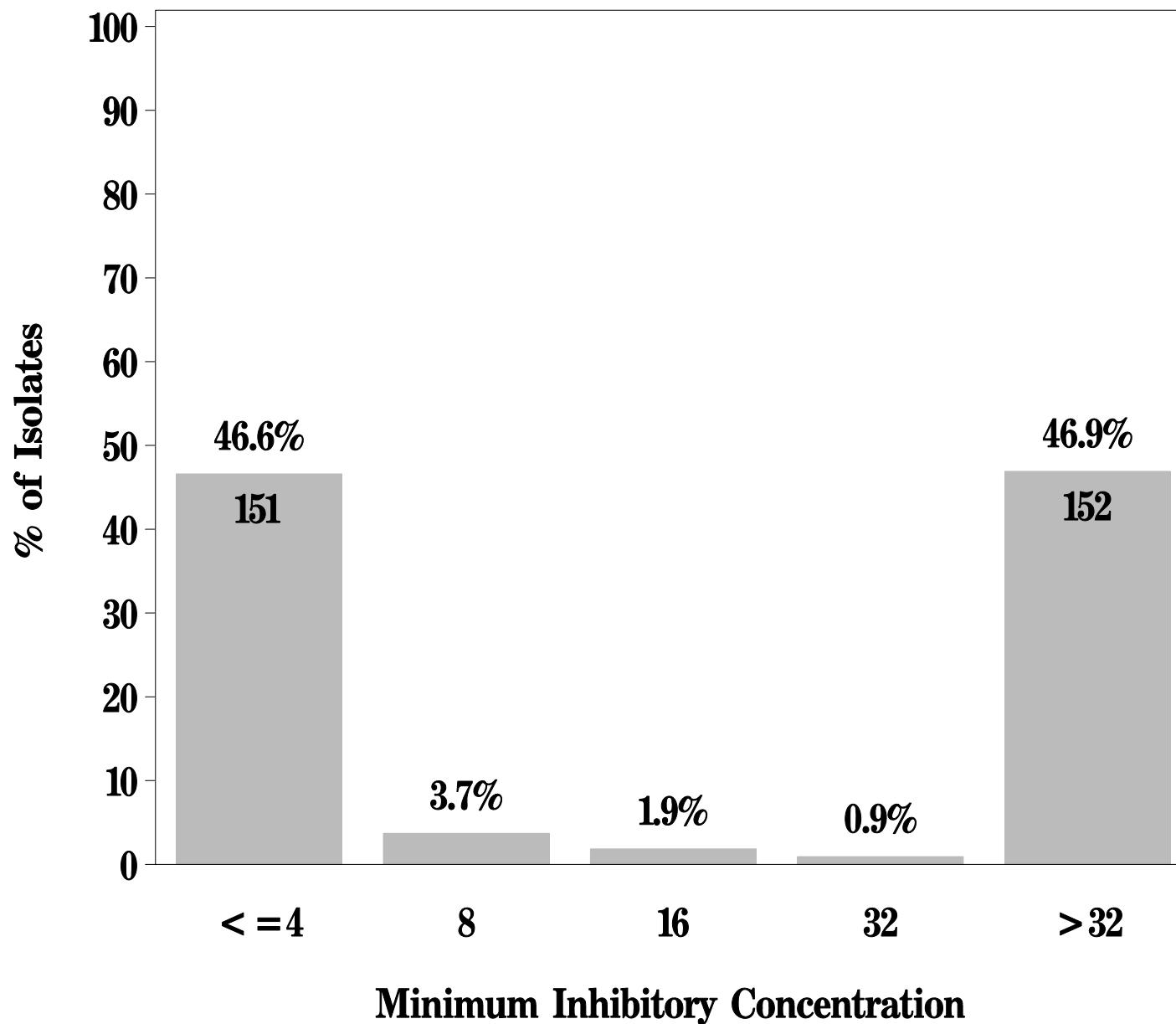
Breakpoints: Susceptible $\leq 256 \text{ } \mu\text{g/mL}$ Resistant $> 512 \text{ } \mu\text{g/mL}$



NARMS

**Figure 5n: Minimum Inhibitory Concentration of Tetracycline
for *Salmonella* (N=324 Isolates)**

Breakpoints: Susceptible $\leq 4 \text{ } \mu\text{g/mL}$ Resistant $\geq 16 \text{ } \mu\text{g/mL}$



NARMS

**Figure 5o: Minimum Inhibitory Concentration of Trimethoprim/sulfamethoxazole
for *Salmonella* (N=324 Isolates)**

Breakpoints: Susceptible $\leq 2 \text{ } \mu\text{g/mL}$ Resistant $\geq 4 \text{ } \mu\text{g/mL}$

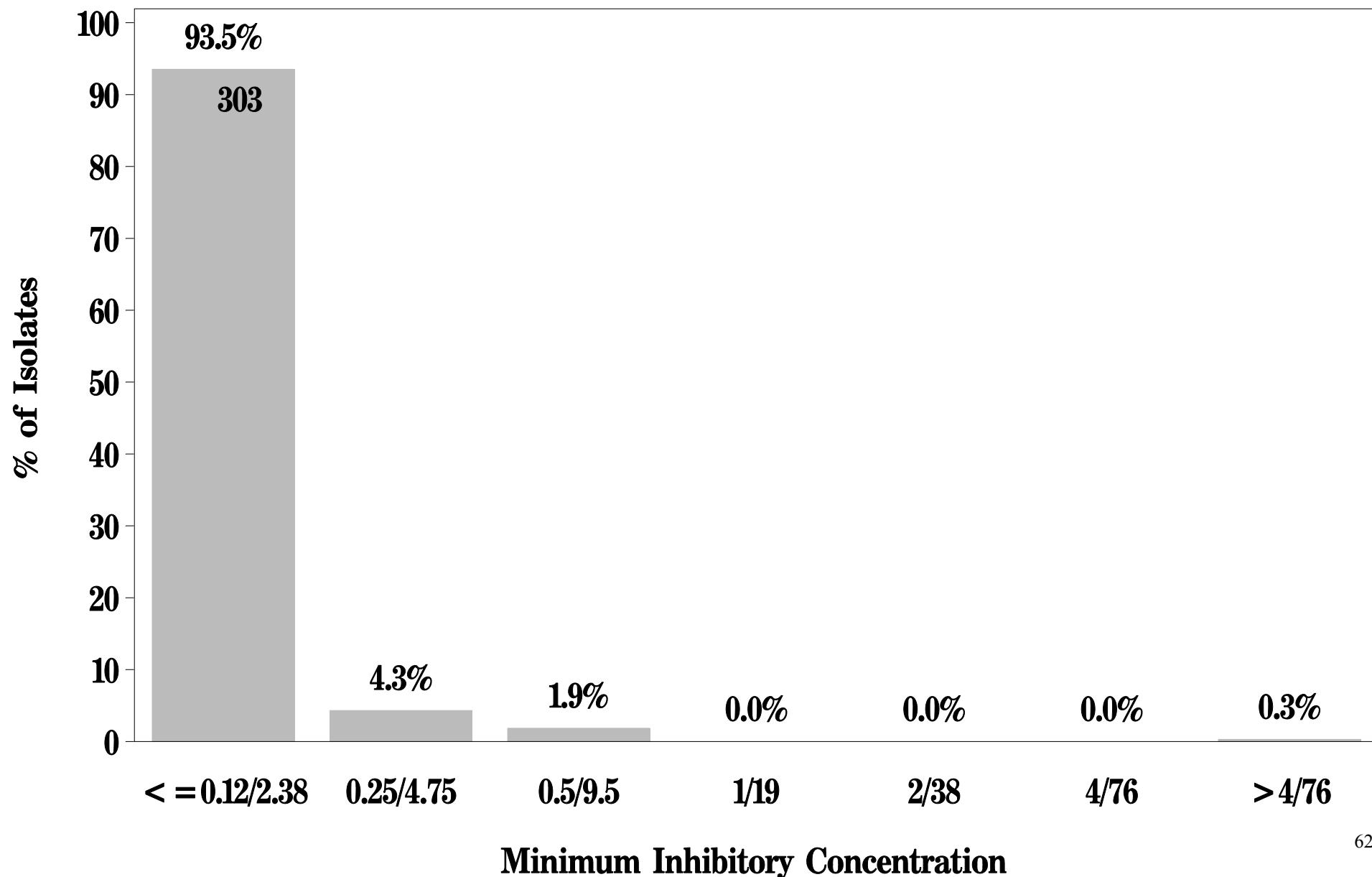


Figure 6a. MIC Distribution among *Salmonella* from Chicken Breast, 2004

<i>Salmonella</i> from Chicken Breast (N=157)		Distribution (%) of MICs (in µg/ml)															
Antimicrobial Agent	%R [†]	0.015	0.03	0.06	0.12	0.25	0.5	1	2	4	8	16	32	64	128	256	>256
Ampicillin	30.6%							60.5	8.9						30.6		
Amoxicillin/Clavulanic Acid	24.8%							61.8	7.6	4.5	1.3				24.8		
Cefoxitin	24.8%							2.5	56.7	14.6	1.3				5.7	19.1	
Ceftiofur	24.8%					0.6	47.1	27.4							24.8		
Ceftriaxone	0.0%						75.2				1.9	18.5	4.5				
Nalidixic Acid	0.0%							12.1	82.8	5.1							
Ciprofloxacin	0.0%	96.2	3.8														
Sulfisoxazole	28.7%											12.1	14.6	43.3	1.3		28.7
Trimethoprim/Sulfamethoxazole	0.0%			96.8	3.2												
Amikacin	0.0%					7.6	46.5	40.1	5.7								
Gentamicin	3.8%					46.5	45.2	3.8		0.6	1.9	1.9					
Kanamycin	11.5%									84.7	3.2	0.6				11.5	
Streptomycin*	28.0%										72.0	16.6	11.5				
Chloramphenicol	1.9%							2.5	14.6	80.3	0.6				1.9		
Tetracycline	46.5%								52.9	0.6					46.5		

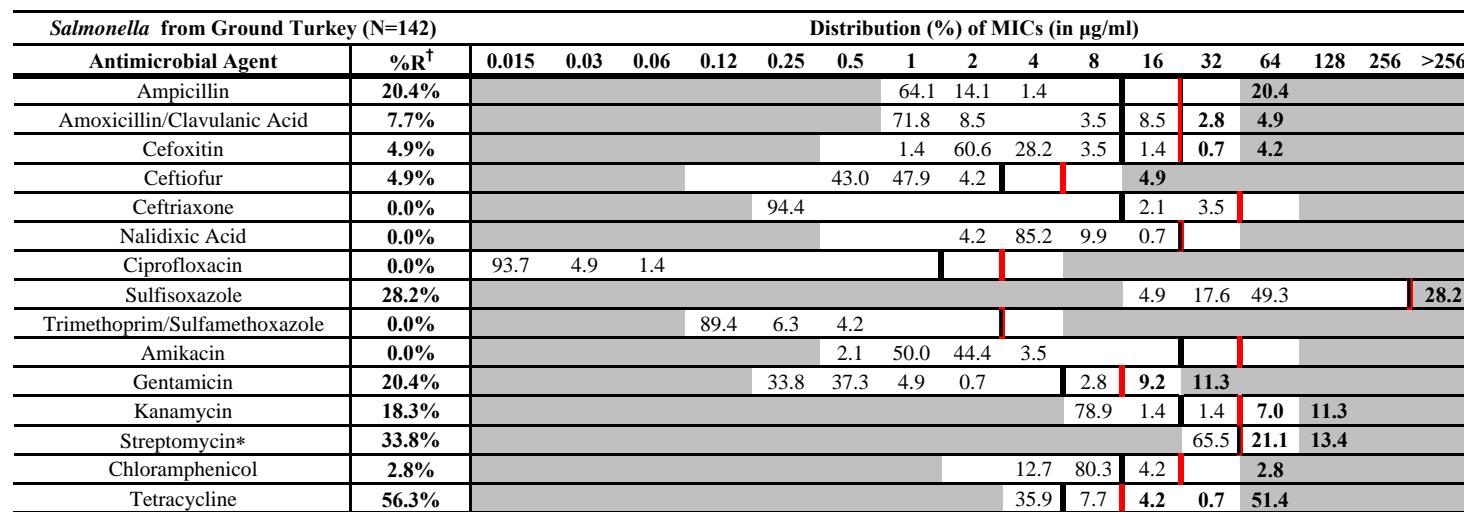
*Currently no CLSI breakpoints have been established for this organism/antimicrobial combination. Indicated breakpoints were established by NARMS.

[†]Discrepancies between %R and sums of distribution %s are due to rounding.

Vertical bars show the CLSI Susceptible/Resistant breakpoints for each drug.

Unshaded areas indicate the dilution ranges of the Sensititre plate used to test the 2004 isolates.

Figure 6b. MIC Distribution among *Salmonella* from Ground Turkey, 2004



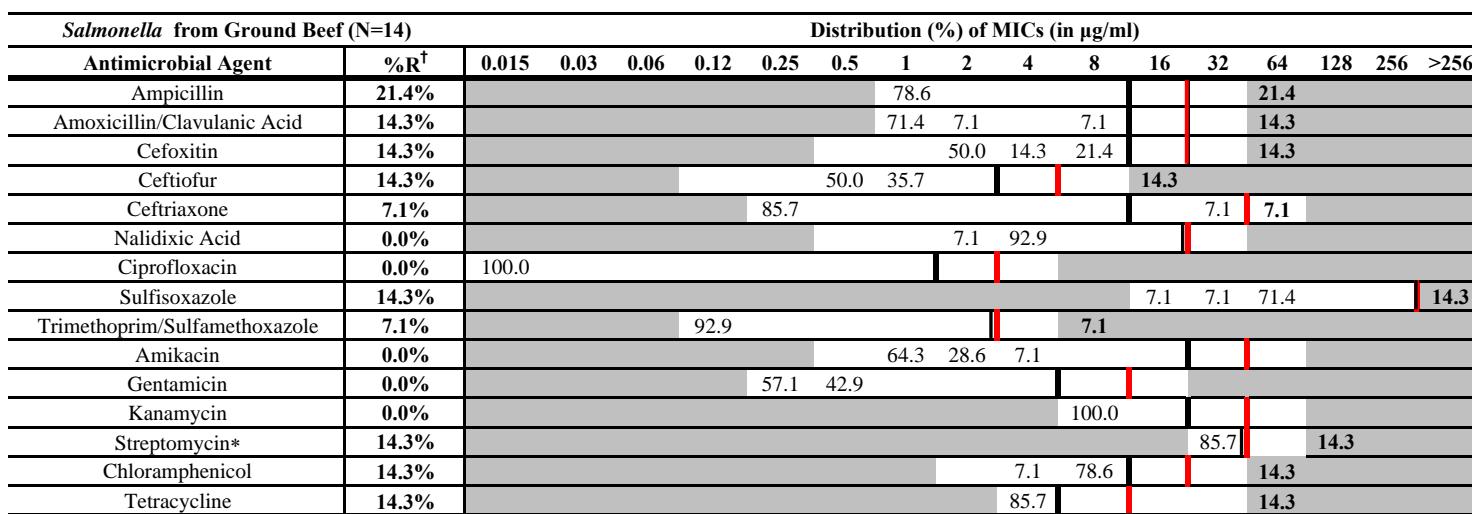
*Currently no CLSI breakpoints have been established for this organism/antimicrobial combination. Indicated breakpoints were established by NARMS.

[†]Discrepancies between %R and sums of distribution %s are due to rounding.

Vertical bars show the CLSI Susceptible/Resistant breakpoints for each drug.

Unshaded areas indicate the dilution ranges of the Sensititre plate used to test the 2004 isolates.

Figure 6c. MIC Distribution among *Salmonella* from Ground Beef, 2004



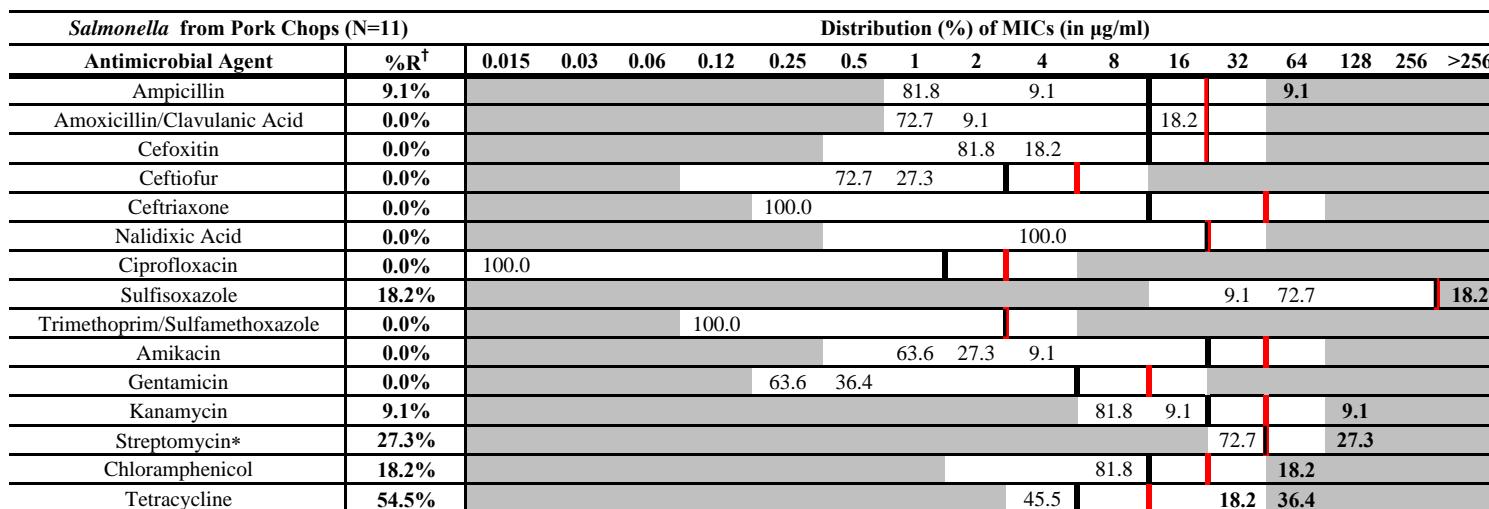
*Currently no CLSI breakpoints have been established for this organism/antimicrobial combination. Indicated breakpoints were established by NARMS.

[†]Discrepancies between %R and sums of distribution %s are due to rounding.

Vertical bars show the CLSI Susceptible/Resistant breakpoints for each drug. Indicated breakpoints were established by NARMS.

Unshaded areas indicate the dilution ranges of the Sensititre plate used to test the 2004 isolates.

Figure 6d. MIC Distribution among *Salmonella* from Pork Chops, 2004



*Currently no CLSI breakpoints have been established for this organism/antimicrobial combination. Indicated breakpoints were established by NARMS.

[†]Discrepancies between %R and sums of distribution %s are due to rounding.

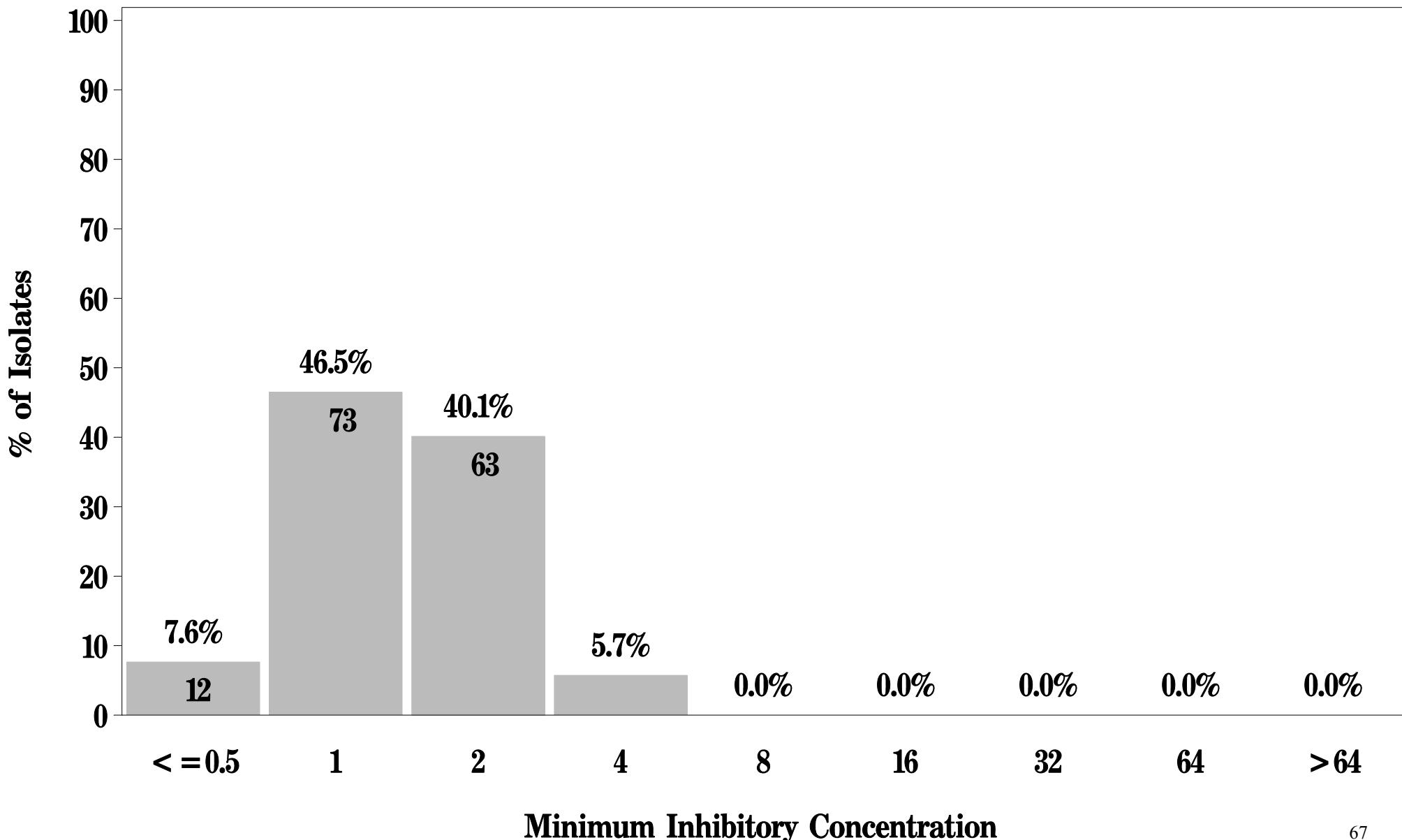
Vertical bars show the CLSI Susceptible/Resistant breakpoints for each drug.

Unshaded areas indicate the dilution ranges of the Sensititre plate used to test the 2004 isolates.

NARMS

**Figure 7a: Minimum Inhibitory Concentration of Amikacin
for *Salmonella* in Chicken Breast (N=157 Isolates)**

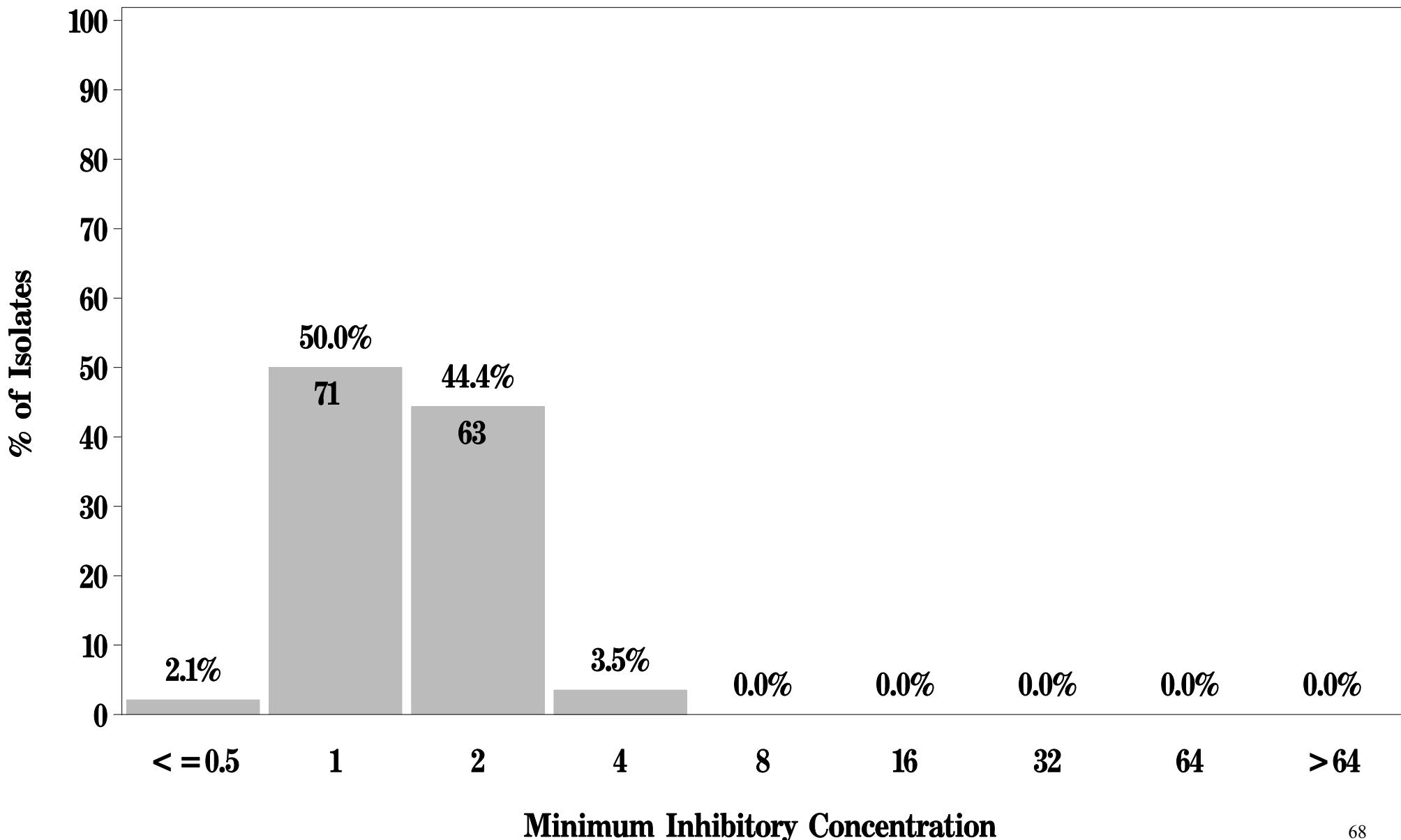
Breakpoints: Susceptible $\leq 16 \mu\text{g/mL}$ Resistant $\geq 64 \mu\text{g/mL}$



NARMS

**Figure 7a: Minimum Inhibitory Concentration of Amikacin
for *Salmonella* in Ground Turkey (N=142 Isolates)**

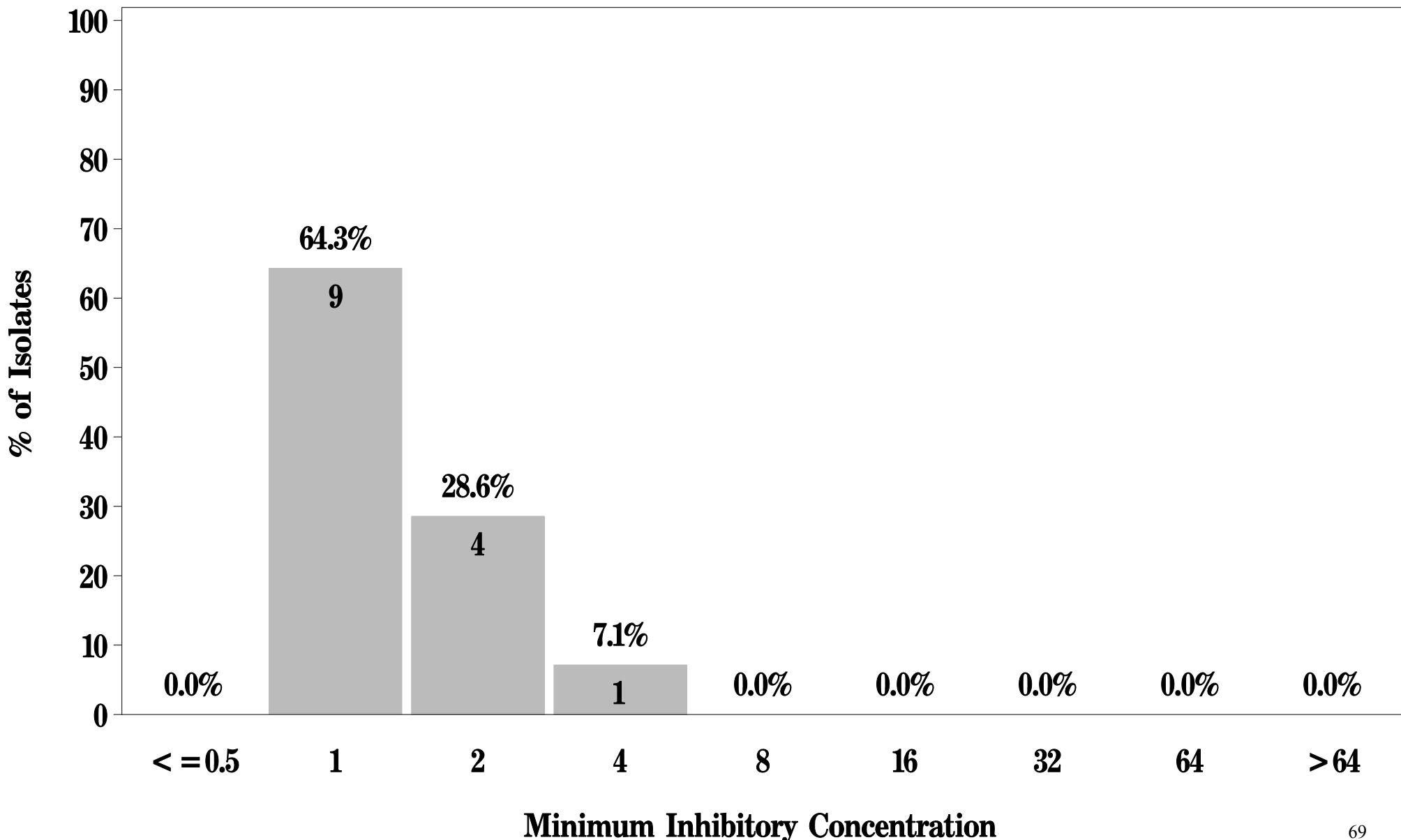
Breakpoints: Susceptible $\leq 16 \mu\text{g/mL}$ Resistant $\geq 64 \mu\text{g/mL}$



NARMS

**Figure 7a: Minimum Inhibitory Concentration of Amikacin
for *Salmonella* in Ground Beef (N=14 Isolates)**

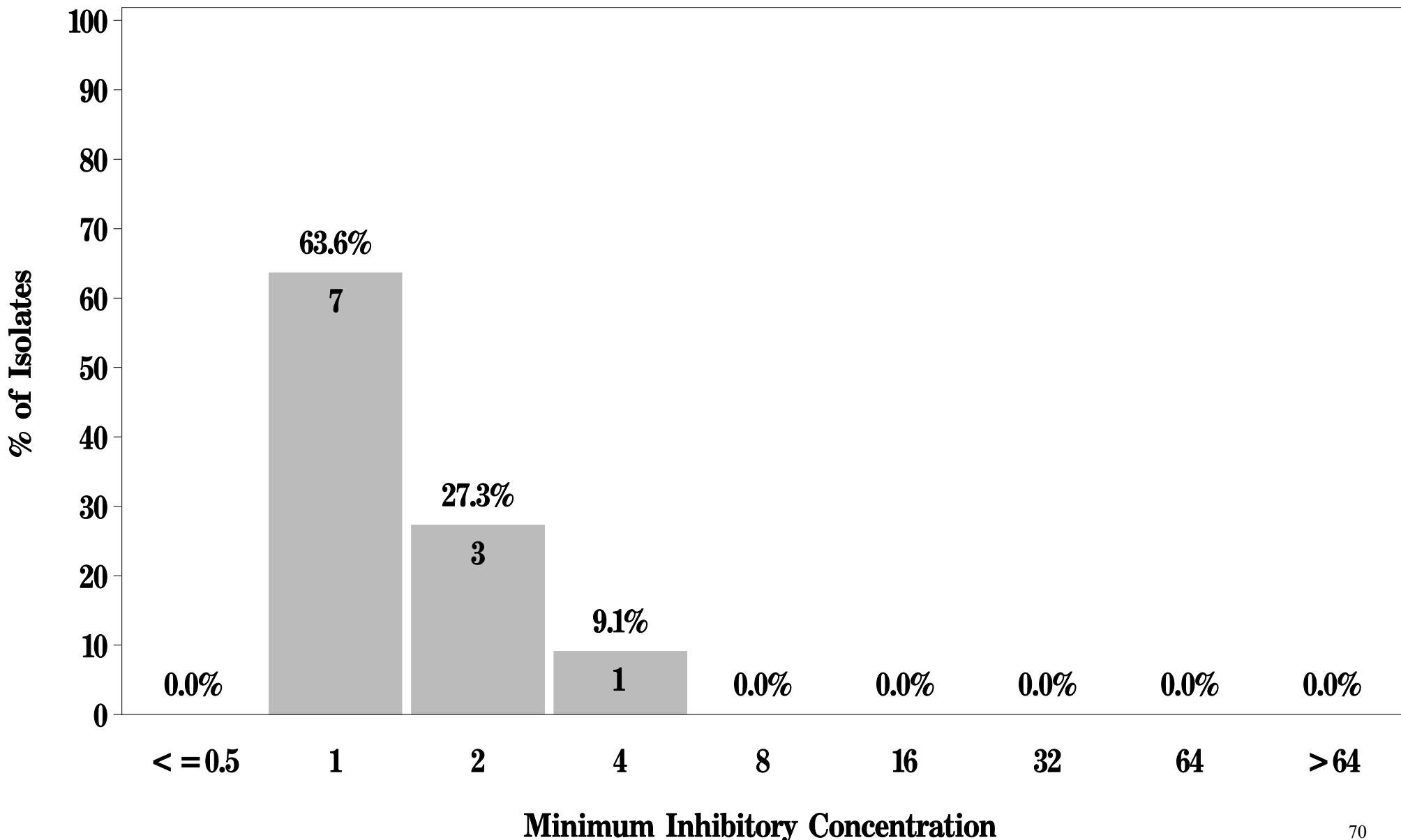
Breakpoints: Susceptible $\leq 16 \mu\text{g/mL}$ Resistant $\geq 64 \mu\text{g/mL}$



NARMS

**Figure 7a: Minimum Inhibitory Concentration of Amikacin
for *Salmonella* in Pork Chop (N=11 Isolates)**

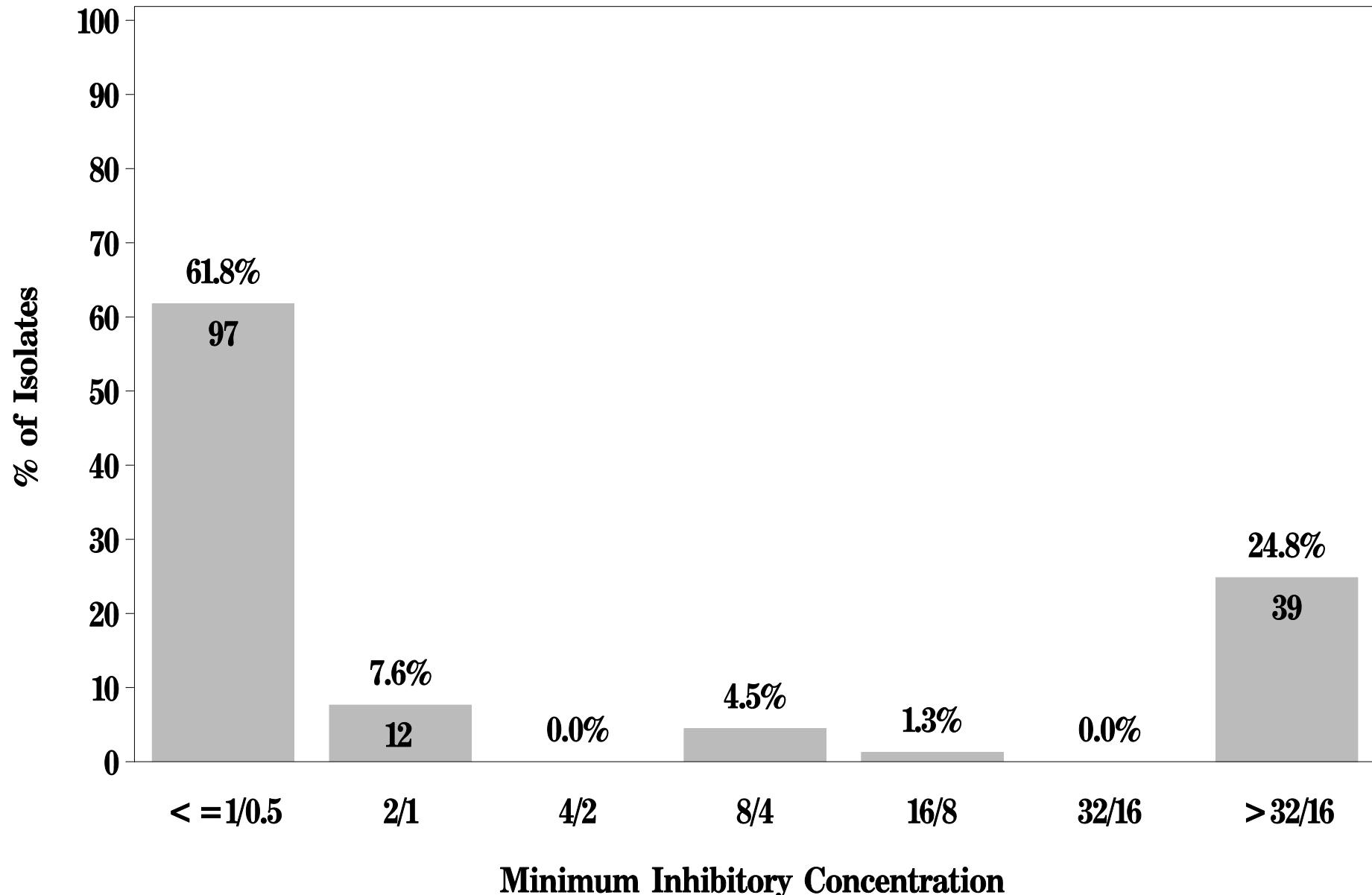
Breakpoints: Susceptible $\leq 16 \mu\text{g/mL}$ Resistant $\geq 64 \mu\text{g/mL}$



NARMS

**Figure 7b: Minimum Inhibitory Concentration of Amoxicillin/Clavulanic acid
for *Salmonella* in Chicken Breast (N=157 Isolates)**

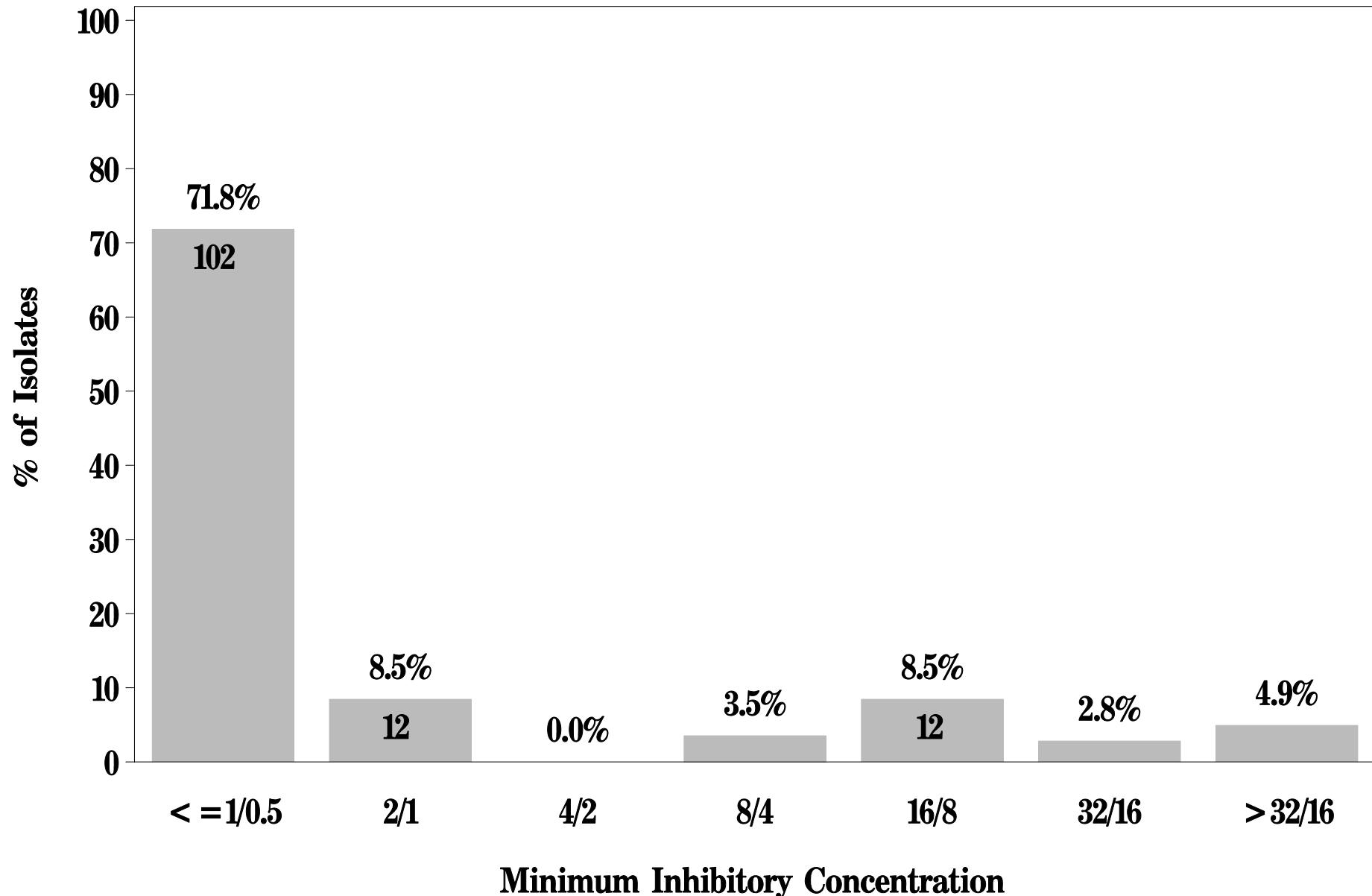
Breakpoints: Susceptible $\leq 8 \mu\text{g/mL}$ Resistant $\geq 32 \mu\text{g/mL}$



NARMS

**Figure 7b: Minimum Inhibitory Concentration of Amoxicillin/Clavulanic acid
for *Salmonella* in Ground Turkey (N=142 Isolates)**

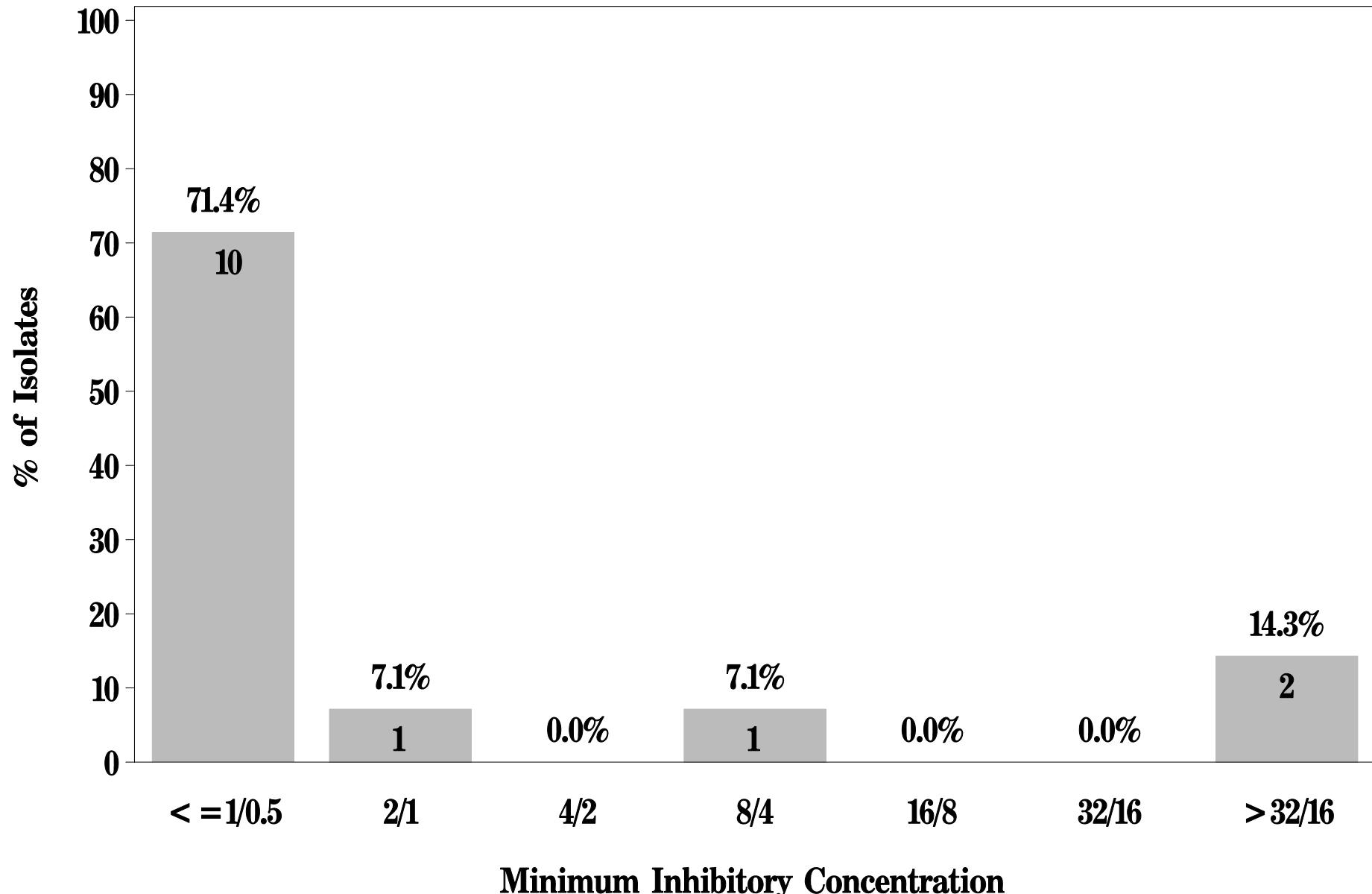
Breakpoints: Susceptible $\leq 8 \mu\text{g/mL}$ Resistant $\geq 32 \mu\text{g/mL}$



NARMS

**Figure 7b: Minimum Inhibitory Concentration of Amoxicillin/Clavulanic acid
for *Salmonella* in Ground Beef (N=14 Isolates)**

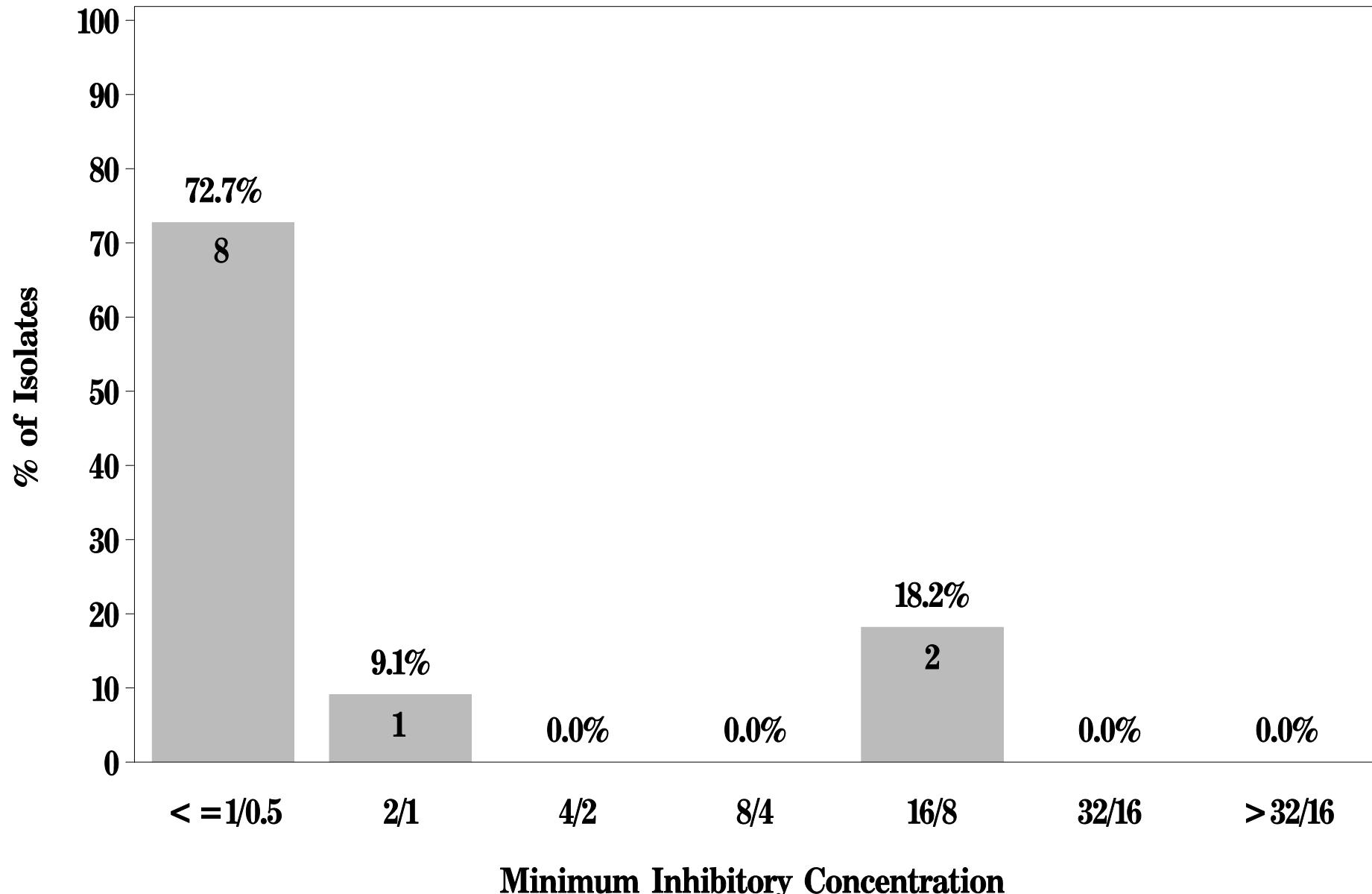
Breakpoints: Susceptible $\leq 8 \mu\text{g/mL}$ Resistant $\geq 32 \mu\text{g/mL}$



NARMS

**Figure 7b: Minimum Inhibitory Concentration of Amoxicillin/Clavulanic acid
for *Salmonella* in Pork Chop (N=11 Isolates)**

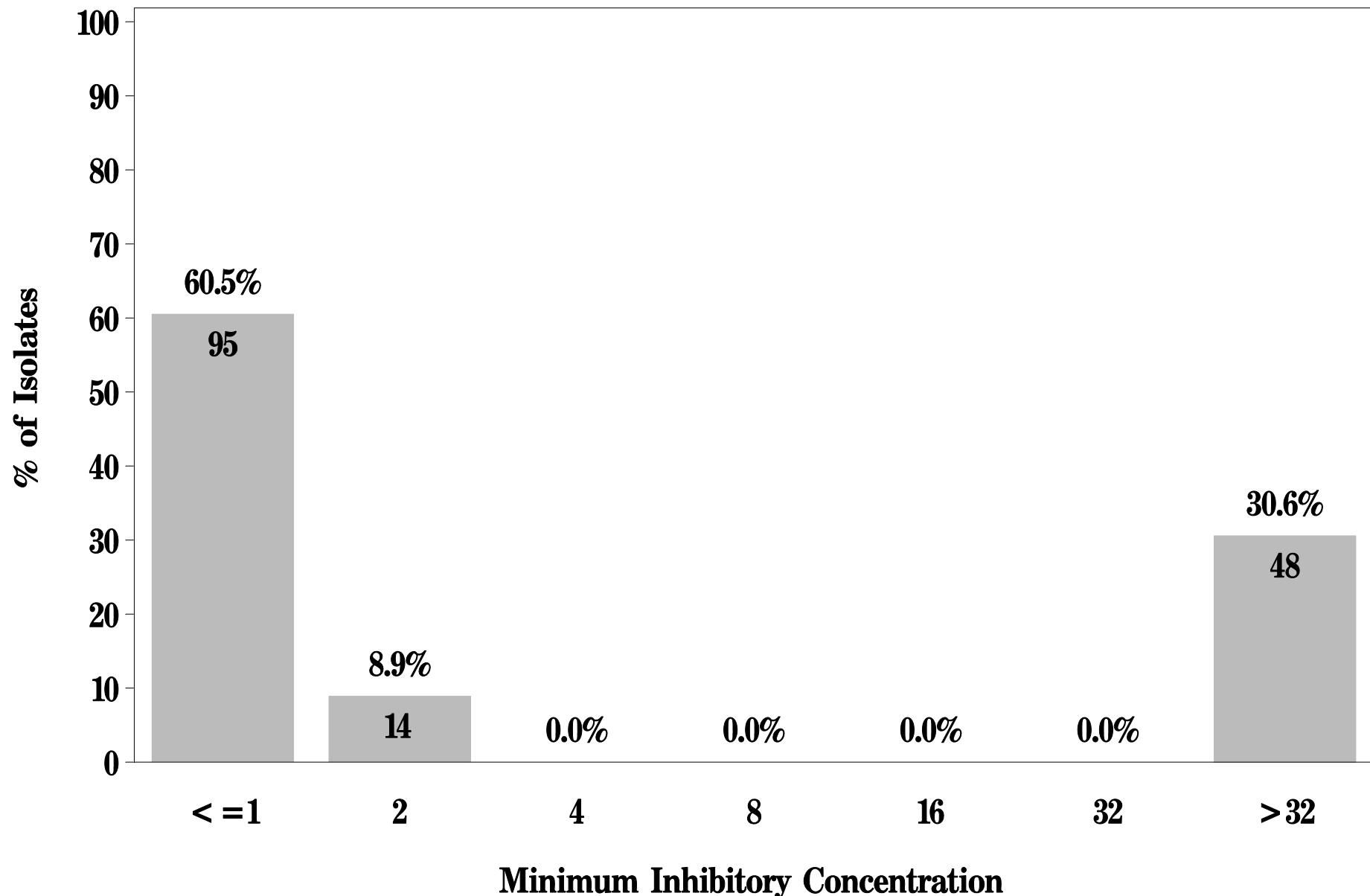
Breakpoints: Susceptible $\leq 8 \mu\text{g/mL}$ Resistant $\geq 32 \mu\text{g/mL}$



NARMS

**Figure 7c: Minimum Inhibitory Concentration of Ampicillin
for *Salmonella* in Chicken Breast (N=157 Isolates)**

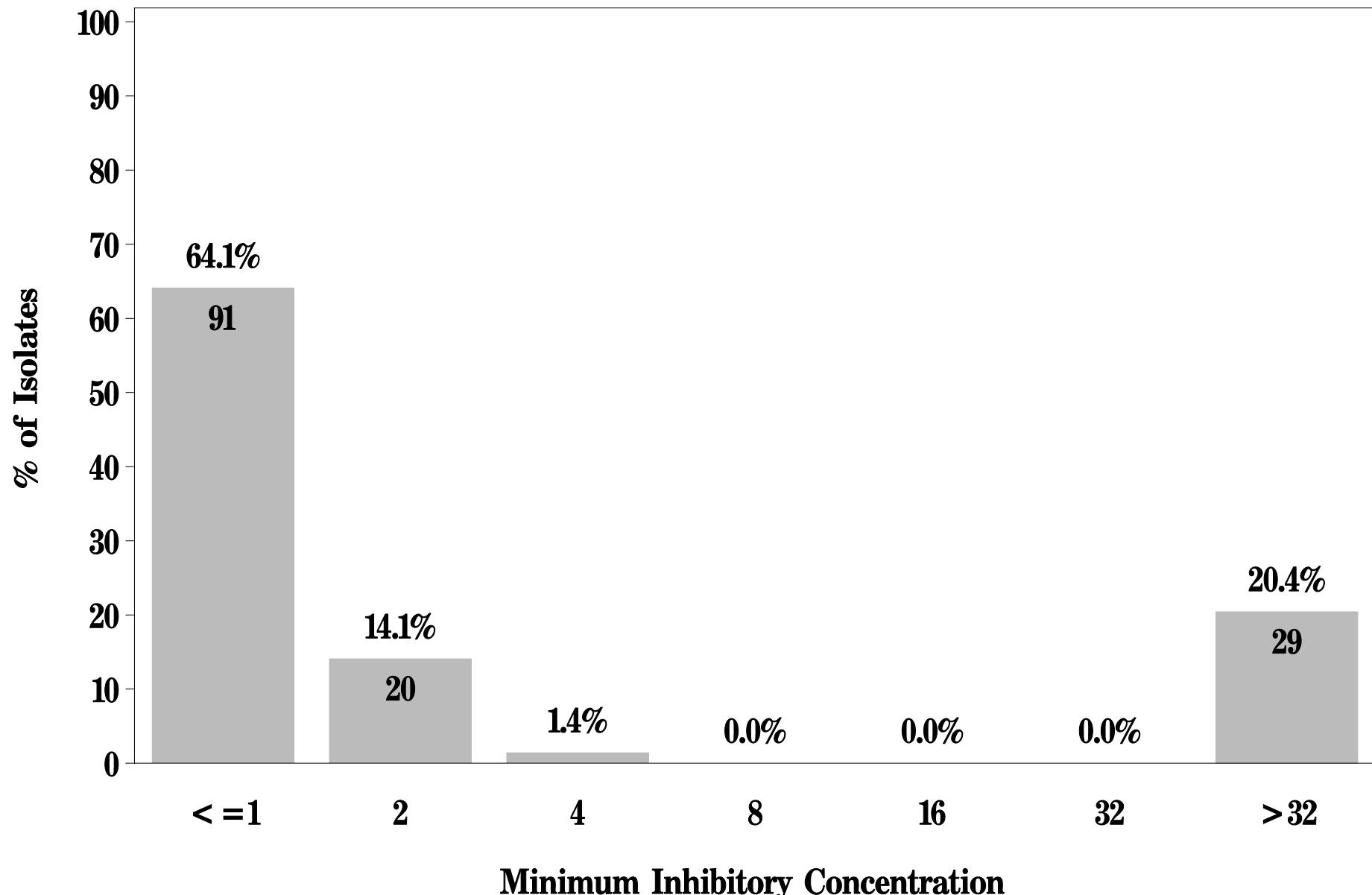
Breakpoints: Susceptible $\leq 8 \mu\text{g/mL}$ Resistant $\geq 32 \mu\text{g/mL}$



NARMS

**Figure 7c: Minimum Inhibitory Concentration of Ampicillin
for *Salmonella* in Ground Turkey (N=142 Isolates)**

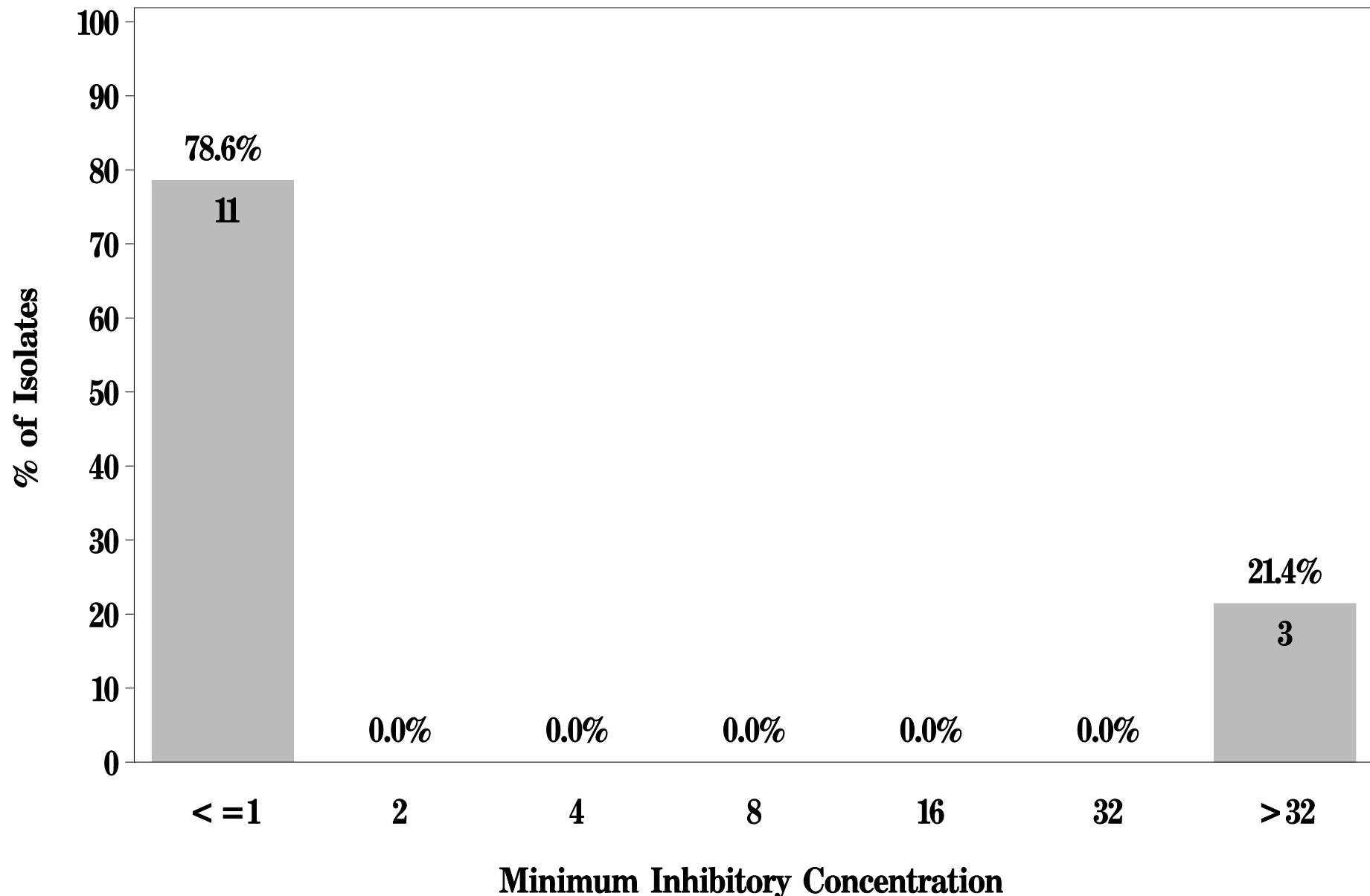
Breakpoints: Susceptible $\leq 8 \mu\text{g/mL}$ Resistant $\geq 32 \mu\text{g/mL}$



NARMS

**Figure 7c: Minimum Inhibitory Concentration of Ampicillin
for *Salmonella* in Ground Beef (N=14 Isolates)**

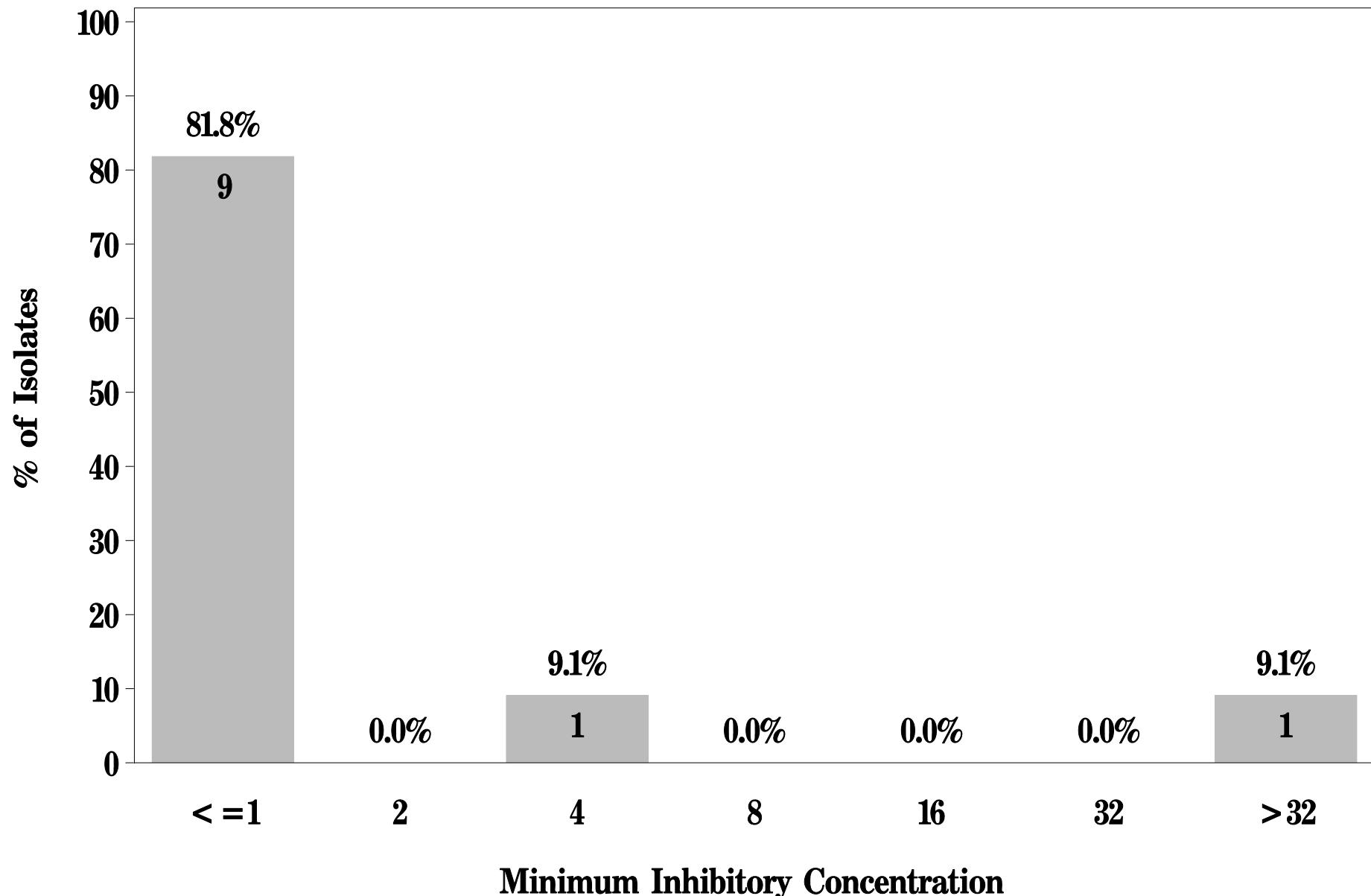
Breakpoints: Susceptible $\leq 8 \mu\text{g/mL}$ Resistant $\geq 32 \mu\text{g/mL}$



NARMS

**Figure 7c: Minimum Inhibitory Concentration of Ampicillin
for *Salmonella* in Pork Chop (N=11 Isolates)**

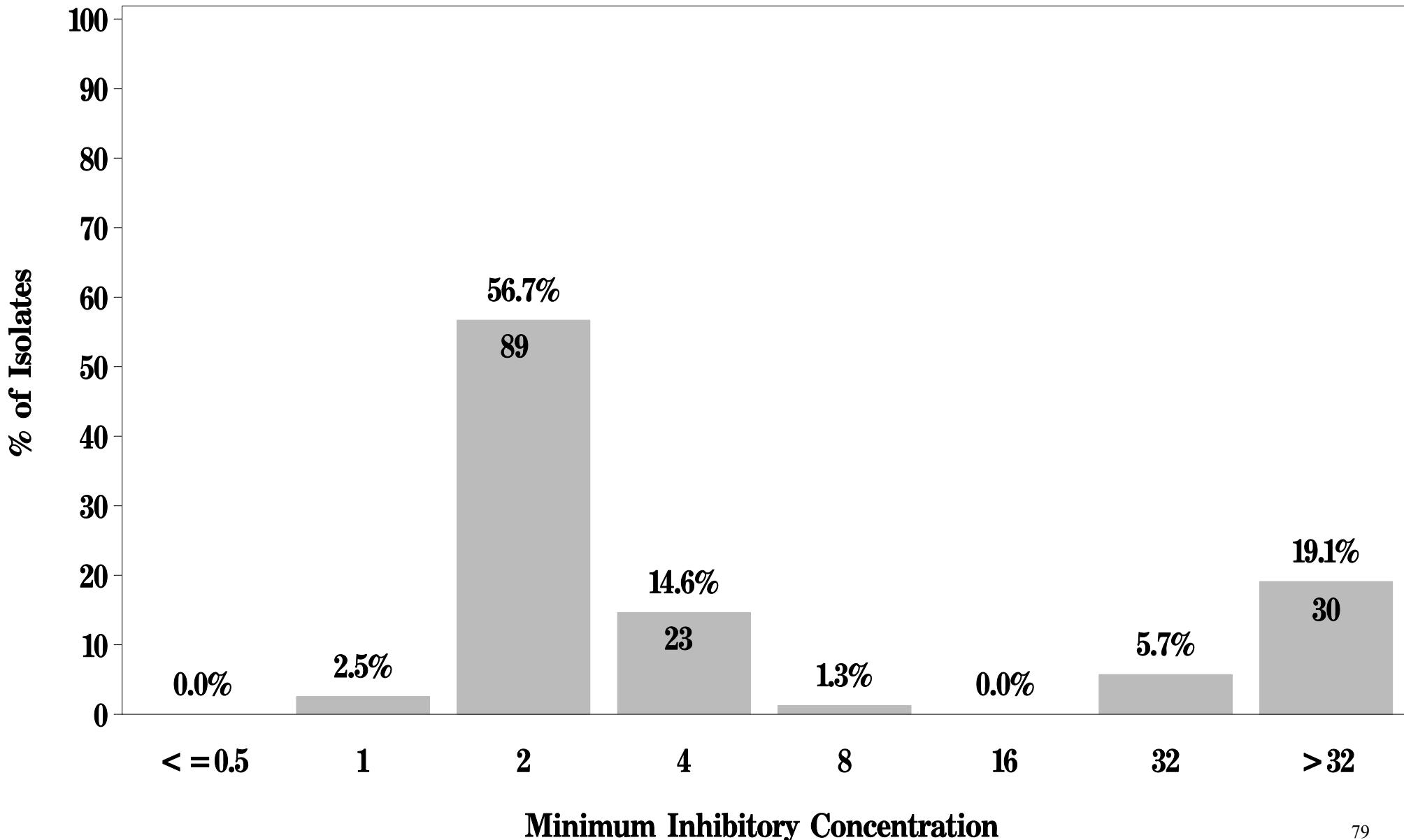
Breakpoints: Susceptible $\leq 8 \text{ } \mu\text{g/mL}$ Resistant $\geq 32 \text{ } \mu\text{g/mL}$



NARMS

**Figure 7d: Minimum Inhibitory Concentration of Cefoxitin
for *Salmonella* in Chicken Breast (N=157 Isolates)**

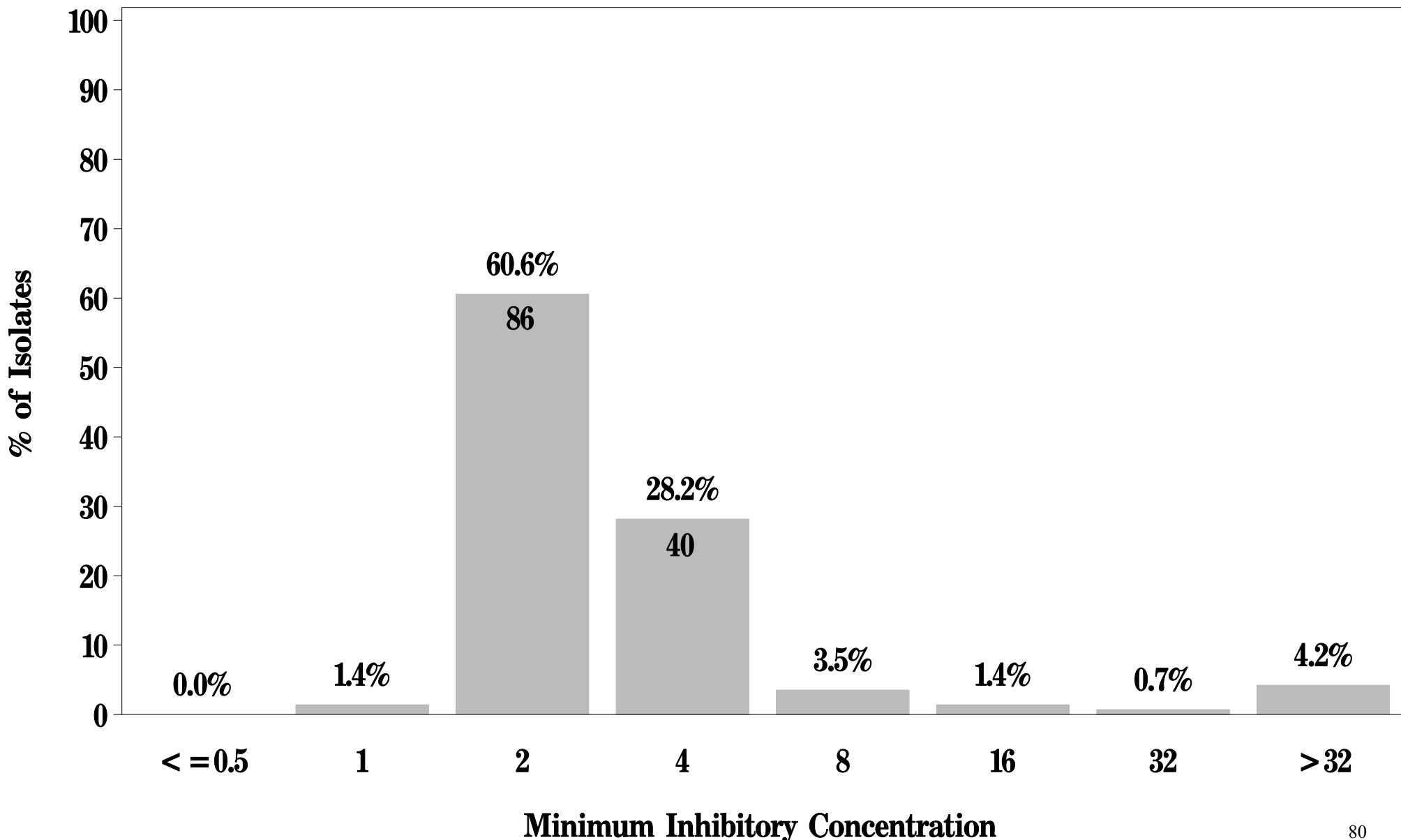
Breakpoints: Susceptible $\leq 8 \mu\text{g/mL}$ Resistant $\geq 32 \mu\text{g/mL}$



NARMS

**Figure 7d: Minimum Inhibitory Concentration of Cefoxitin
for *Salmonella* in Ground Turkey (N=142 Isolates)**

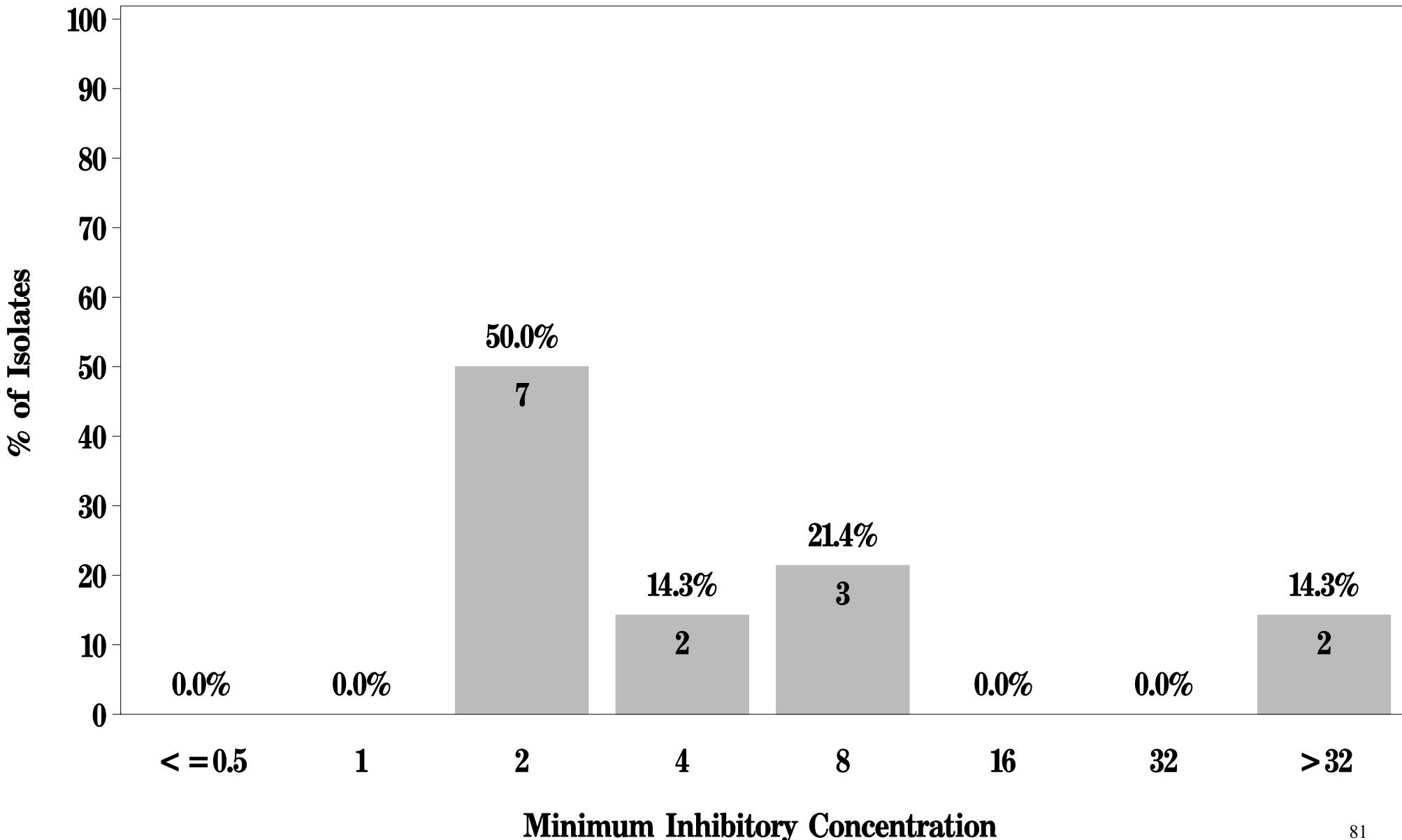
Breakpoints: Susceptible $\leq 8 \mu\text{g/mL}$ Resistant $\geq 32 \mu\text{g/mL}$



NARMS

**Figure 7d: Minimum Inhibitory Concentration of Cefoxitin
for *Salmonella* in Ground Beef (N=14 Isolates)**

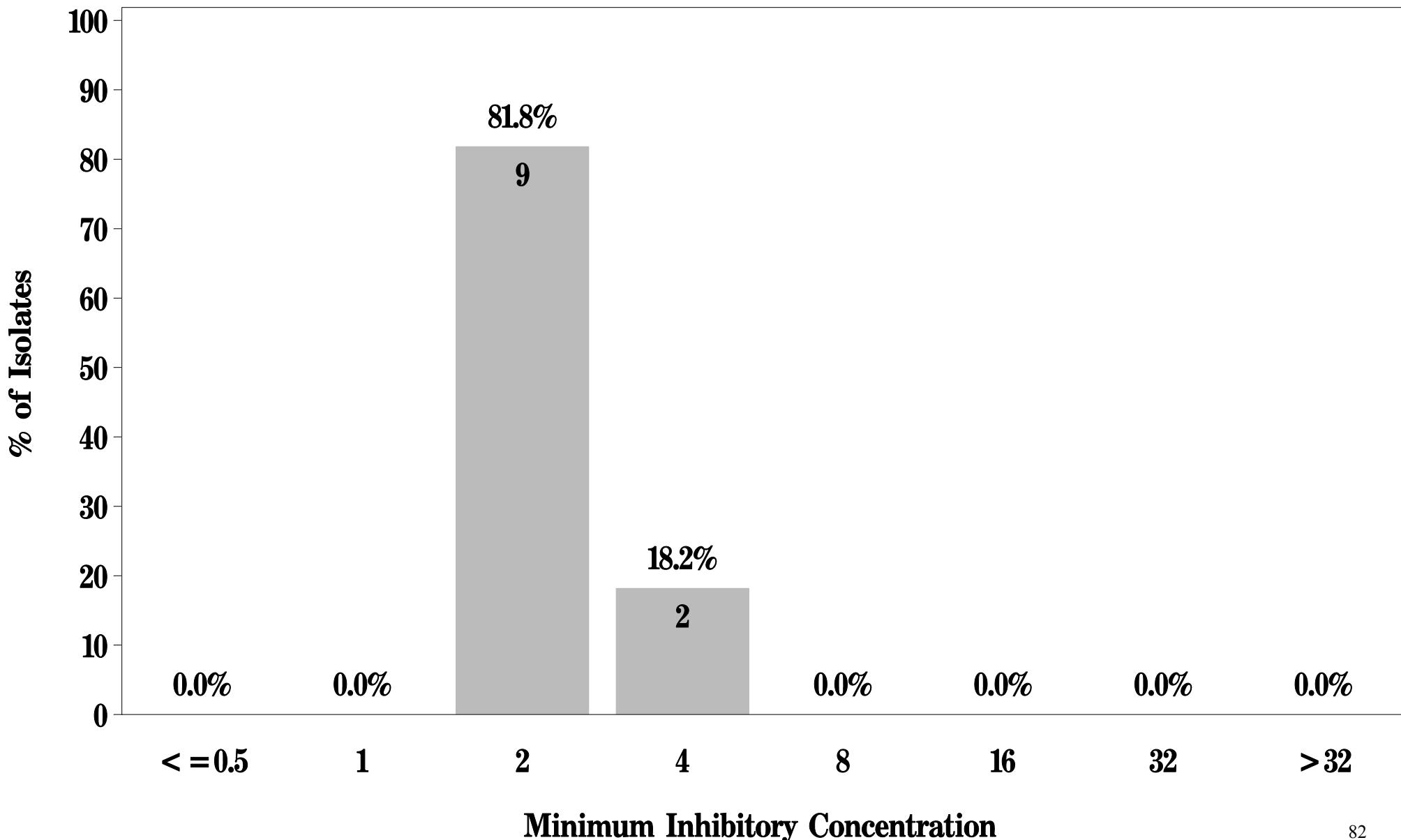
Breakpoints: Susceptible $\leq 8 \mu\text{g/mL}$ Resistant $\geq 32 \mu\text{g/mL}$



NARMS

**Figure 7d: Minimum Inhibitory Concentration of Cefoxitin
for *Salmonella* in Pork Chop (N=11 Isolates)**

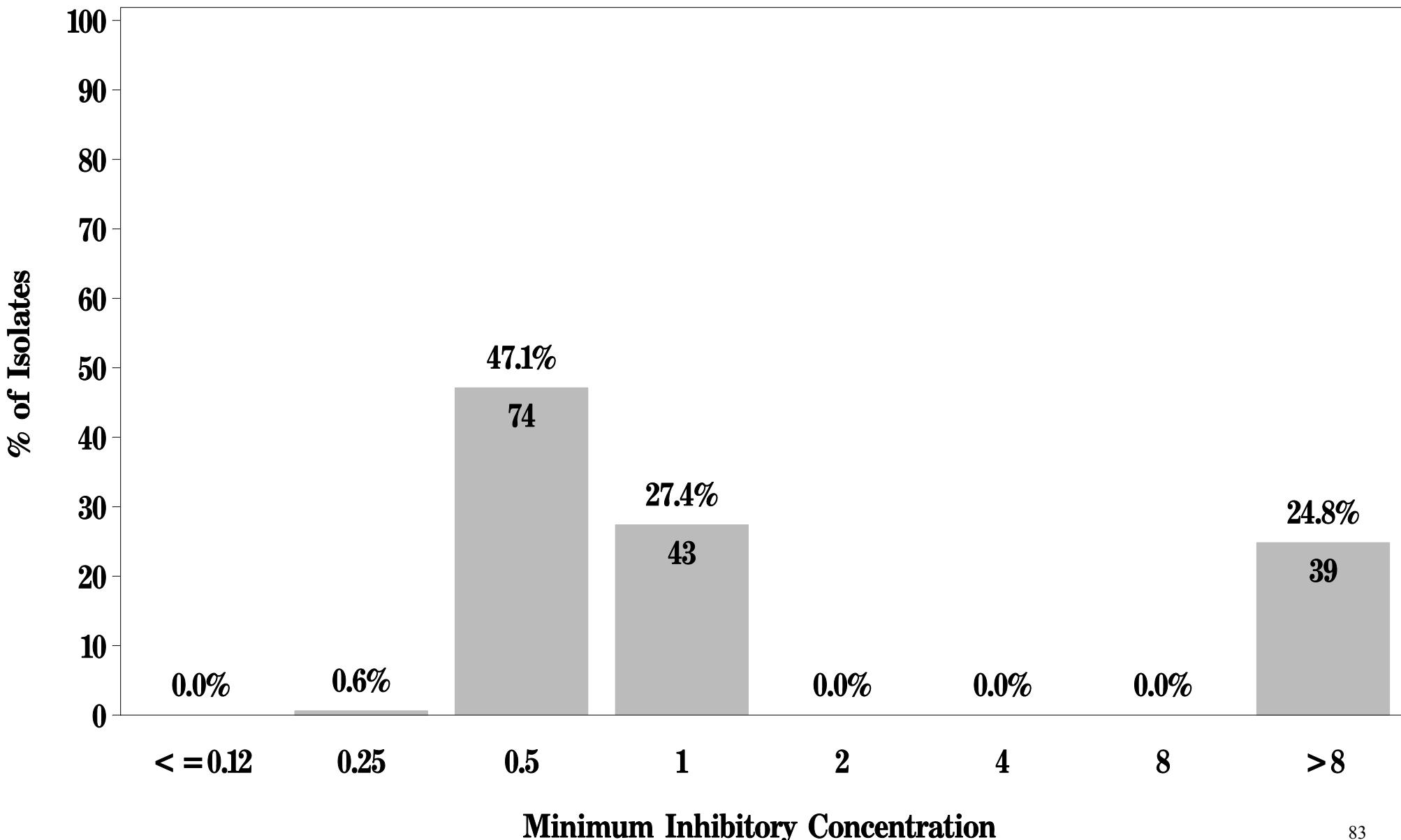
Breakpoints: Susceptible $\leq 8 \mu\text{g/mL}$ Resistant $\geq 32 \mu\text{g/mL}$



NARMS

**Figure 7e: Minimum Inhibitory Concentration of Ceftiofur
for *Salmonella* in Chicken Breast (N=157 Isolates)**

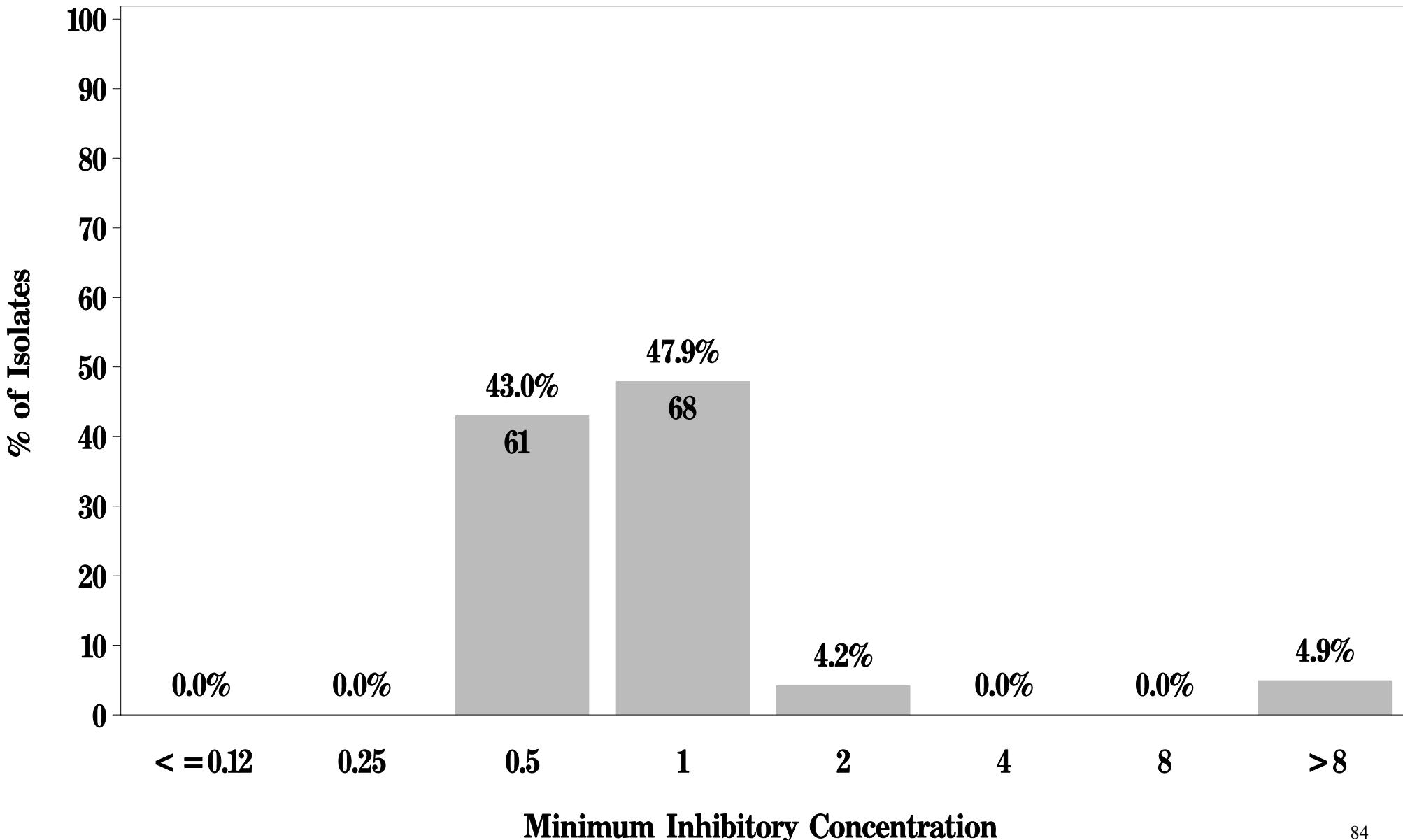
Breakpoints: Susceptible $\leq 2 \mu\text{g/mL}$ Resistant $\geq 8 \mu\text{g/mL}$



NARMS

**Figure 7e: Minimum Inhibitory Concentration of Ceftiofur
for *Salmonella* in Ground Turkey (N=142 Isolates)**

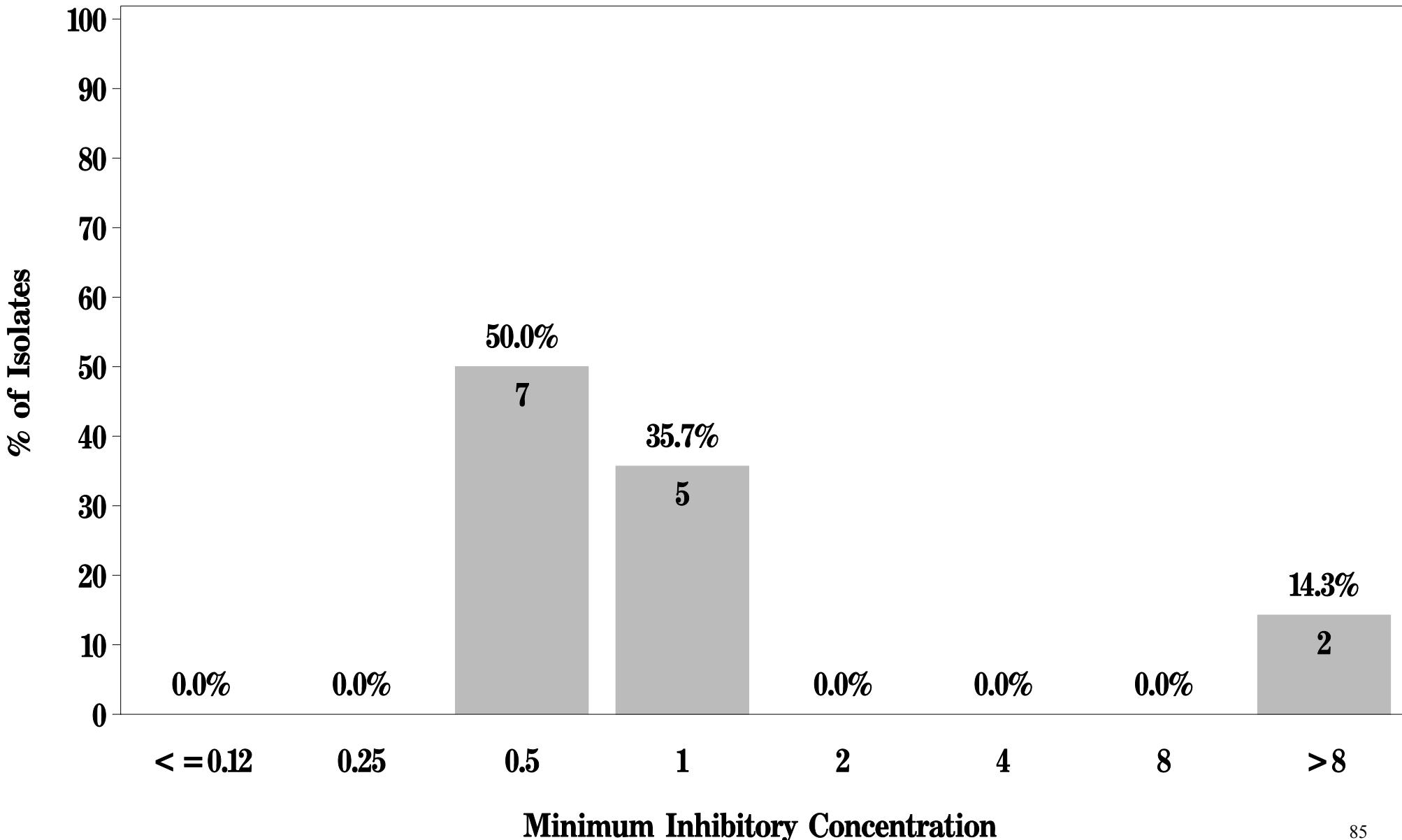
Breakpoints: Susceptible $\leq 2 \mu\text{g/mL}$ Resistant $\geq 8 \mu\text{g/mL}$



NARMS

**Figure 7e: Minimum Inhibitory Concentration of Ceftiofur
for *Salmonella* in Ground Beef (N=14 Isolates)**

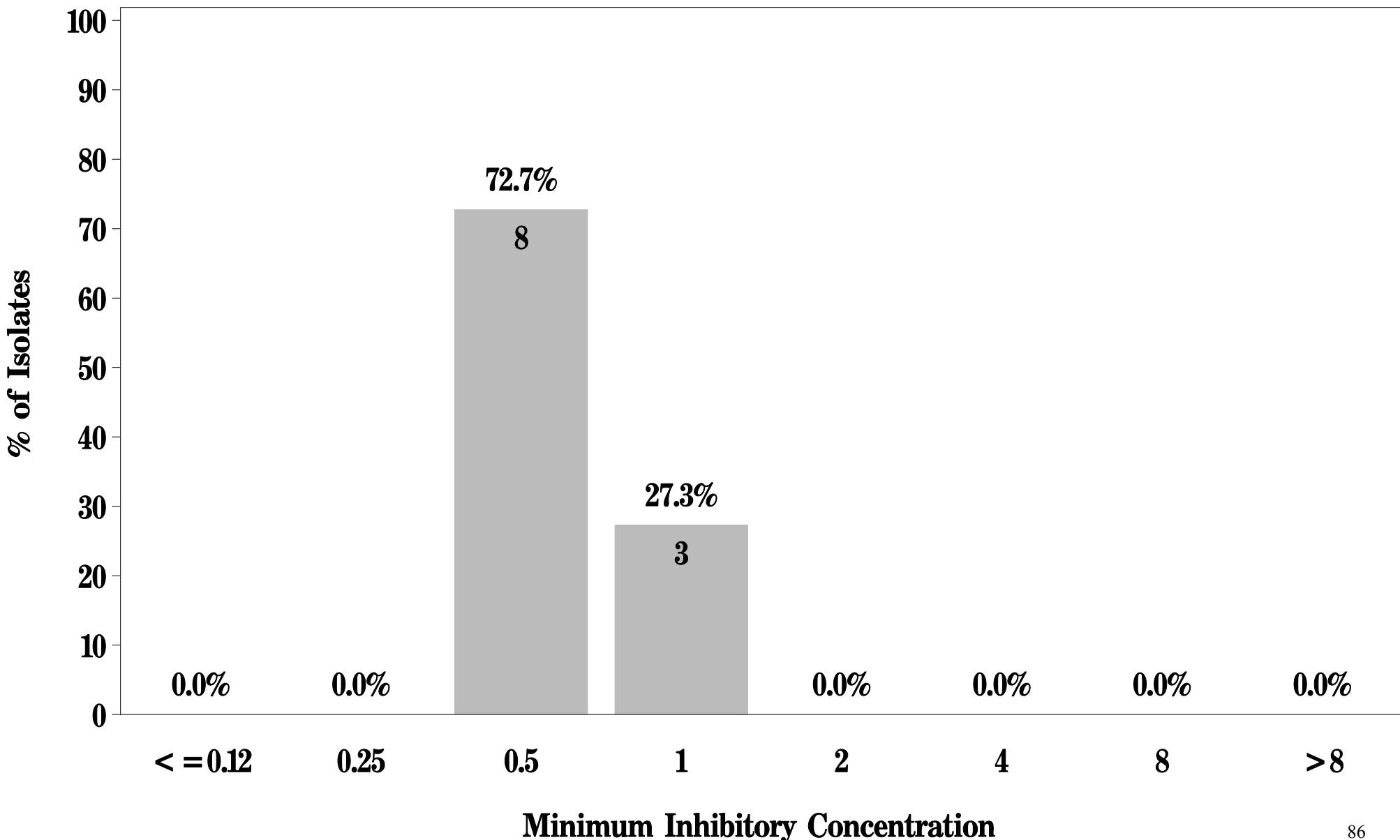
Breakpoints: Susceptible $\leq 2 \mu\text{g/mL}$ Resistant $\geq 8 \mu\text{g/mL}$



NARMS

**Figure 7e: Minimum Inhibitory Concentration of Ceftiofur
for *Salmonella* in Pork Chop (N=11 Isolates)**

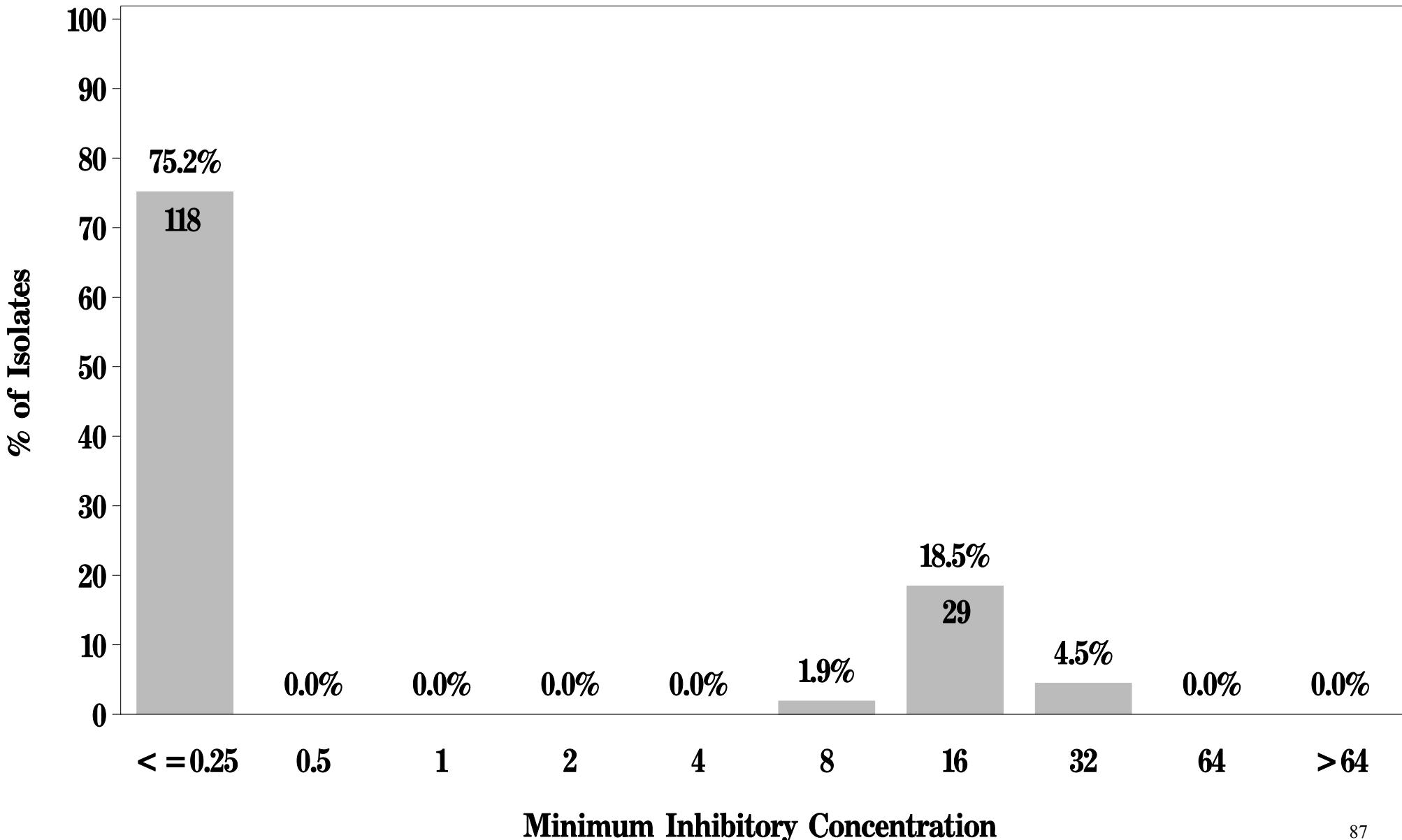
Breakpoints: Susceptible $\leq 2 \mu\text{g/mL}$ Resistant $\geq 8 \mu\text{g/mL}$



NARMS

**Figure 7f: Minimum Inhibitory Concentration of Ceftriaxone
for *Salmonella* in Chicken Breast (N=157 Isolates)**

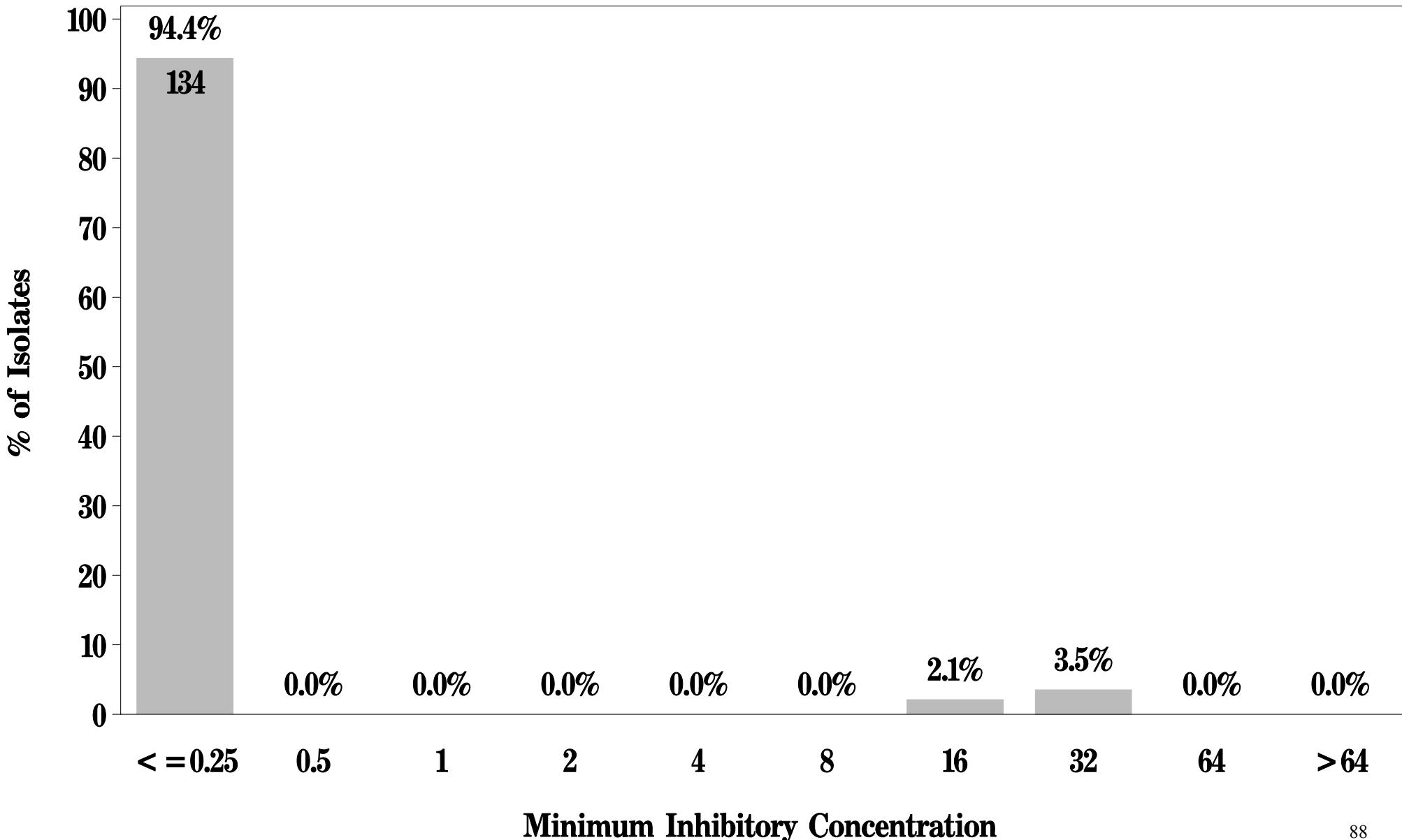
Breakpoints: Susceptible $\leq 8 \mu\text{g/mL}$ Resistant $\geq 16 \mu\text{g/mL}$



NARMS

**Figure 7f: Minimum Inhibitory Concentration of Ceftriaxone
for *Salmonella* in Ground Turkey (N=142 Isolates)**

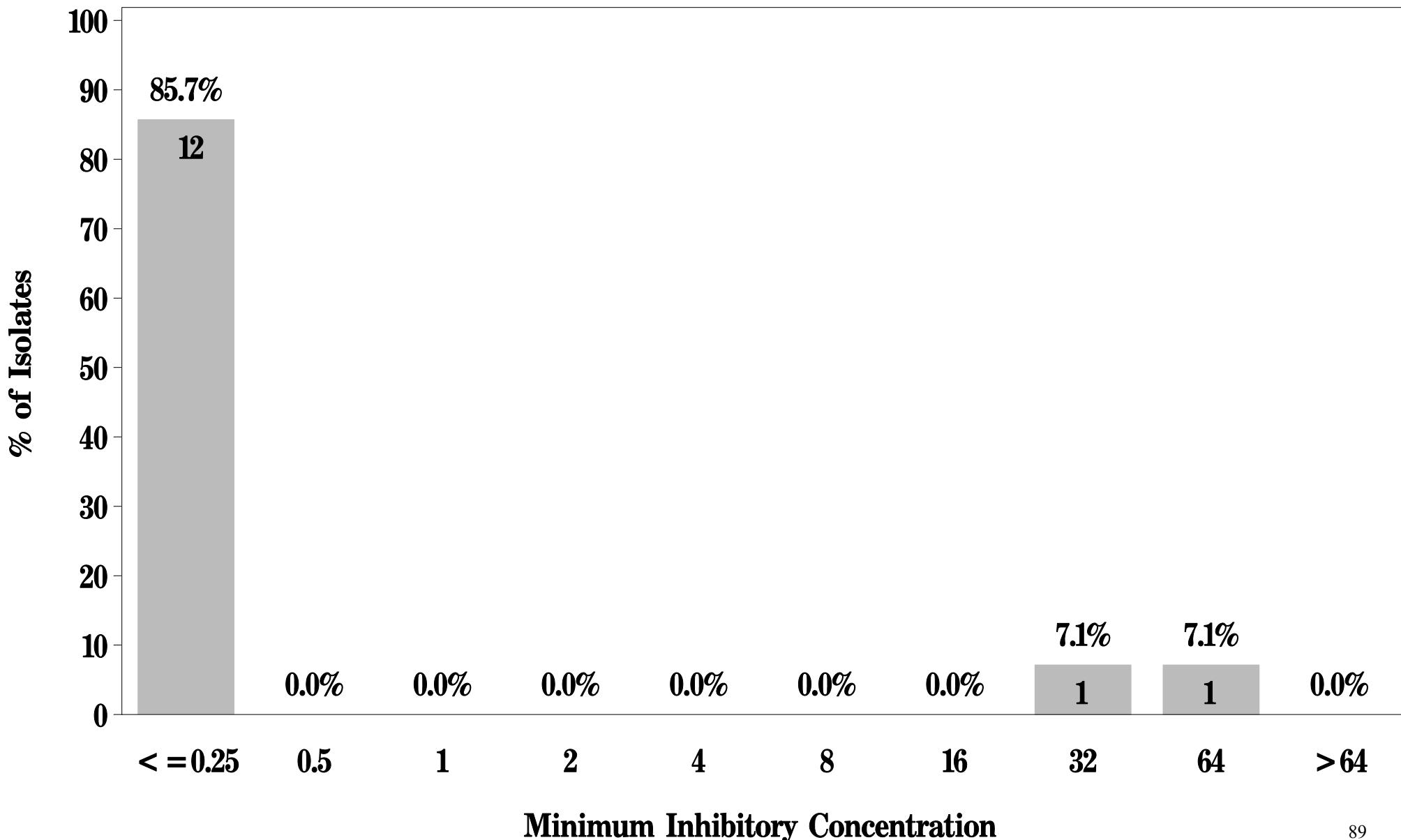
Breakpoints: Susceptible $\leq 8 \mu\text{g/mL}$ Resistant $\geq 16 \mu\text{g/mL}$



NARMS

**Figure 7f: Minimum Inhibitory Concentration of Ceftriaxone
for *Salmonella* in Ground Beef (N=14 Isolates)**

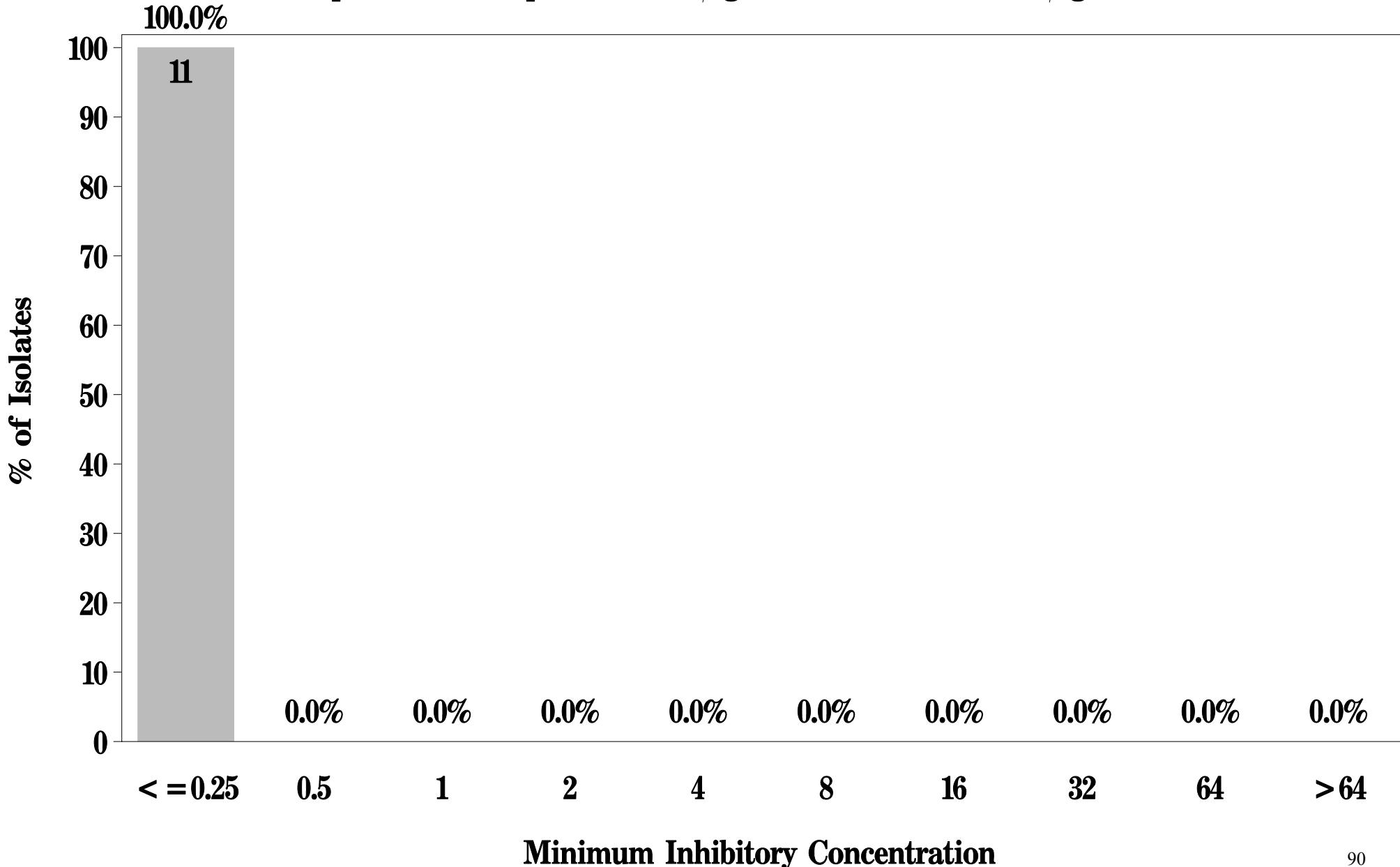
Breakpoints: Susceptible $\leq 8 \mu\text{g/mL}$ Resistant $\geq 64 \mu\text{g/mL}$



NARMS

**Figure 7f: Minimum Inhibitory Concentration of Ceftriaxone
for *Salmonella* in Pork Chop (N=11 Isolates)**

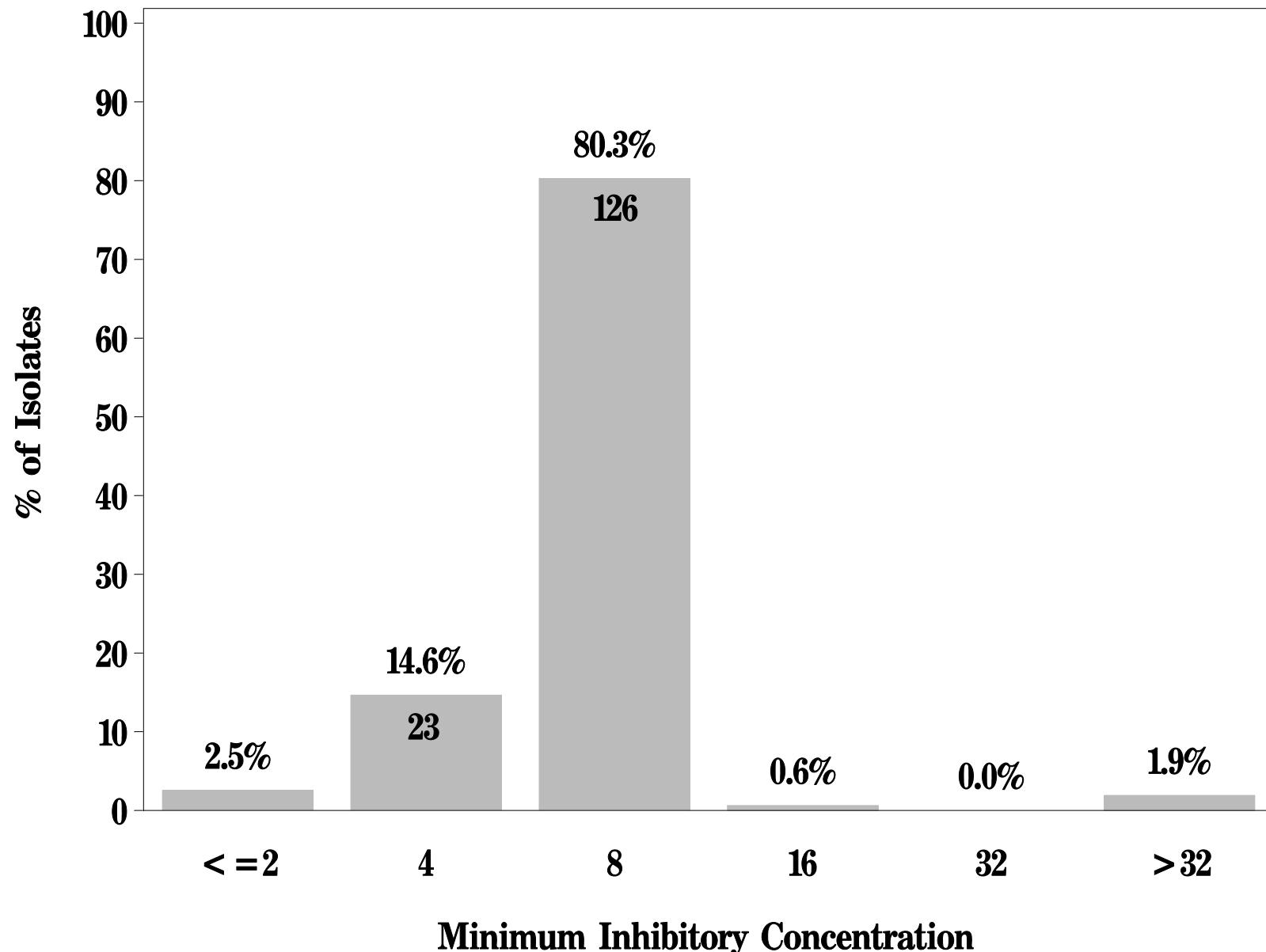
Breakpoints: Susceptible $\leq 8 \mu\text{g/mL}$ Resistant $\geq 64 \mu\text{g/mL}$



NARMS

**Figure 7g: Minimum Inhibitory Concentration of Chloramphenicol
for *Salmonella* in Chicken Breast (N=157 Isolates)**

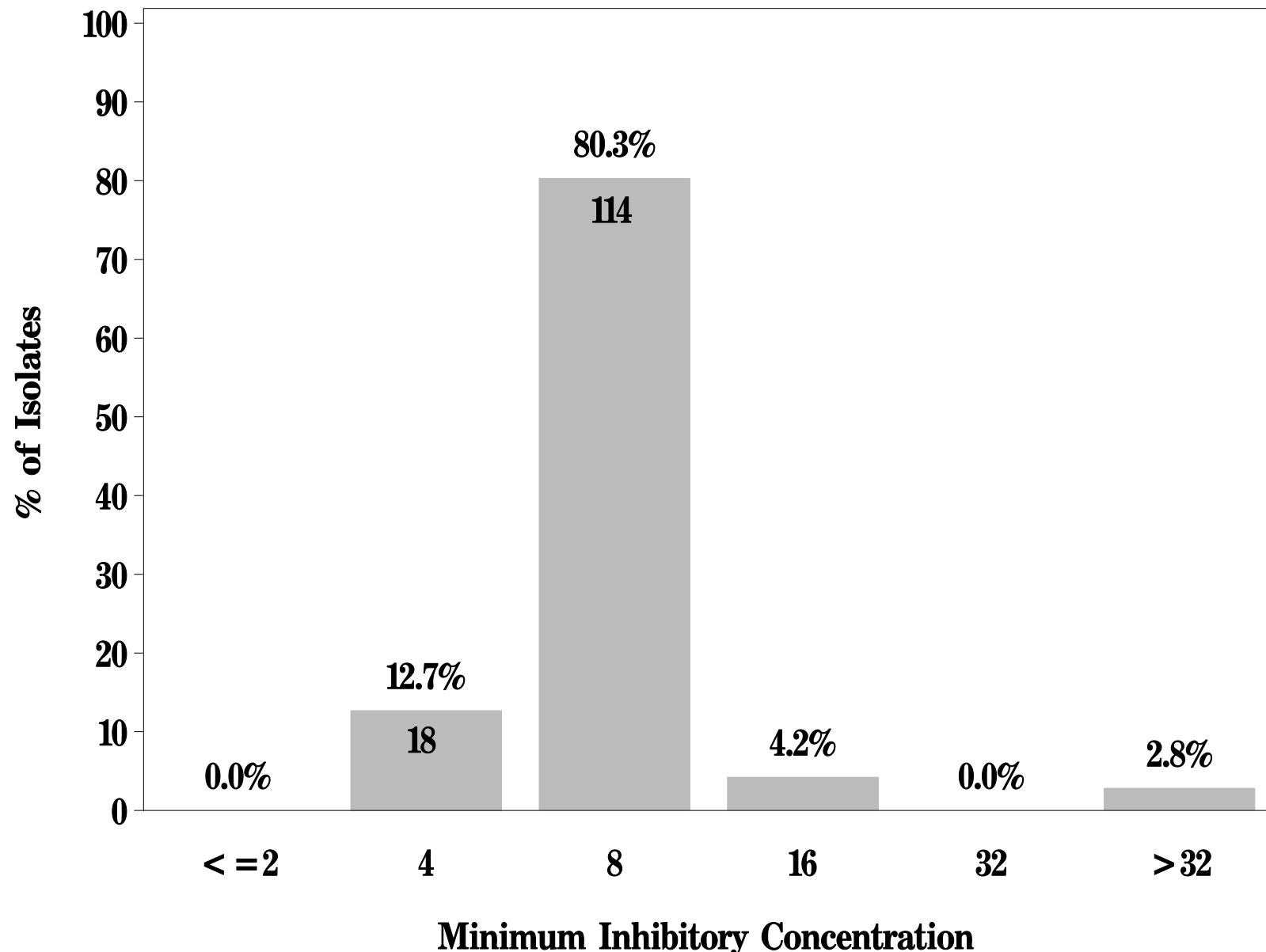
Breakpoints: Susceptible $\leq 8 \mu\text{g/mL}$ Resistant $\geq 32 \mu\text{g/mL}$



NARMS

**Figure 7g: Minimum Inhibitory Concentration of Chloramphenicol
for *Salmonella* in Ground Turkey (N=142 Isolates)**

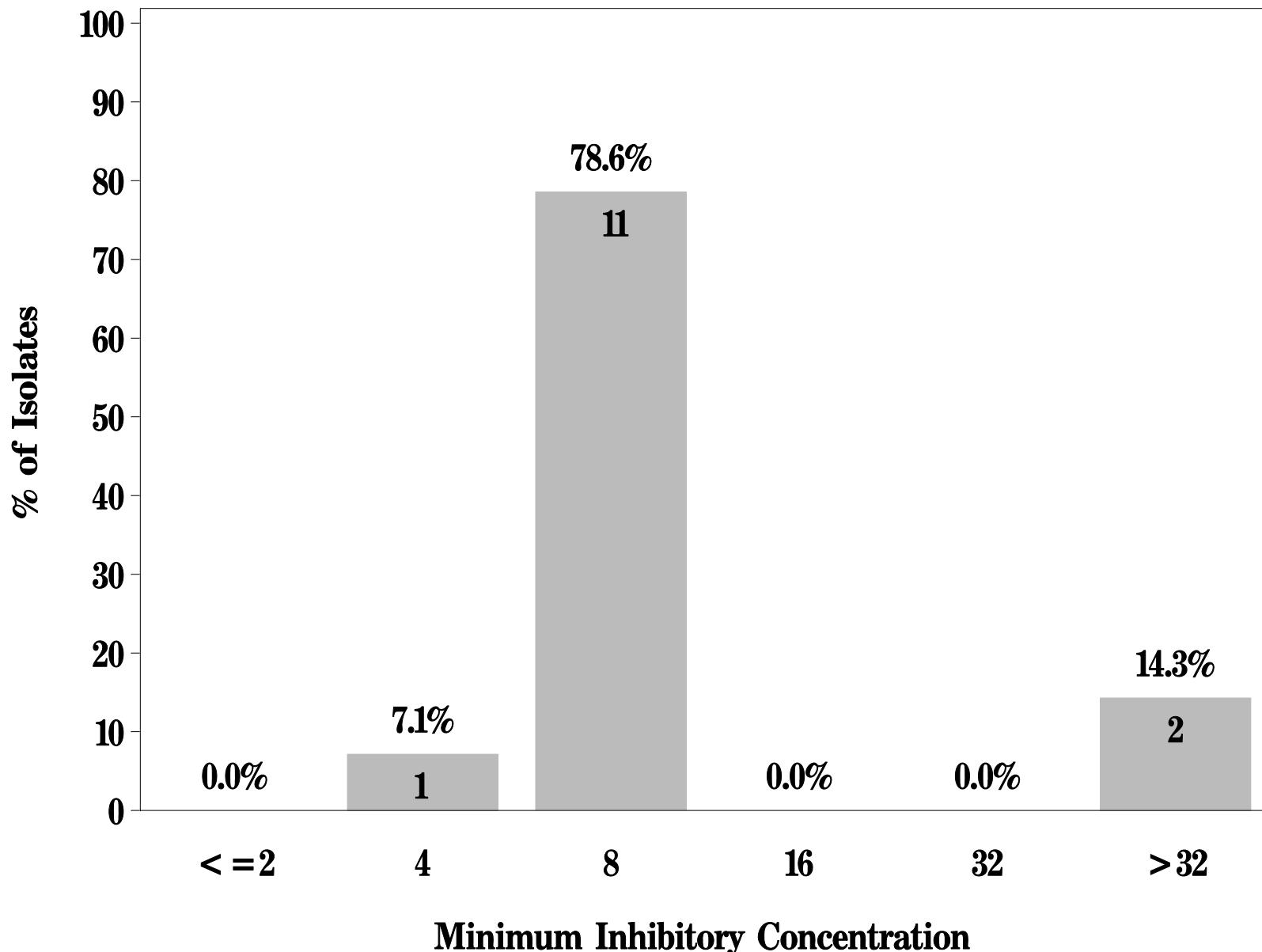
Breakpoints: Susceptible $\leq 8 \mu\text{g/mL}$ Resistant $\geq 32 \mu\text{g/mL}$



NARMS

**Figure 7g: Minimum Inhibitory Concentration of Chloramphenicol
for *Salmonella* in Ground Beef (N=14 Isolates)**

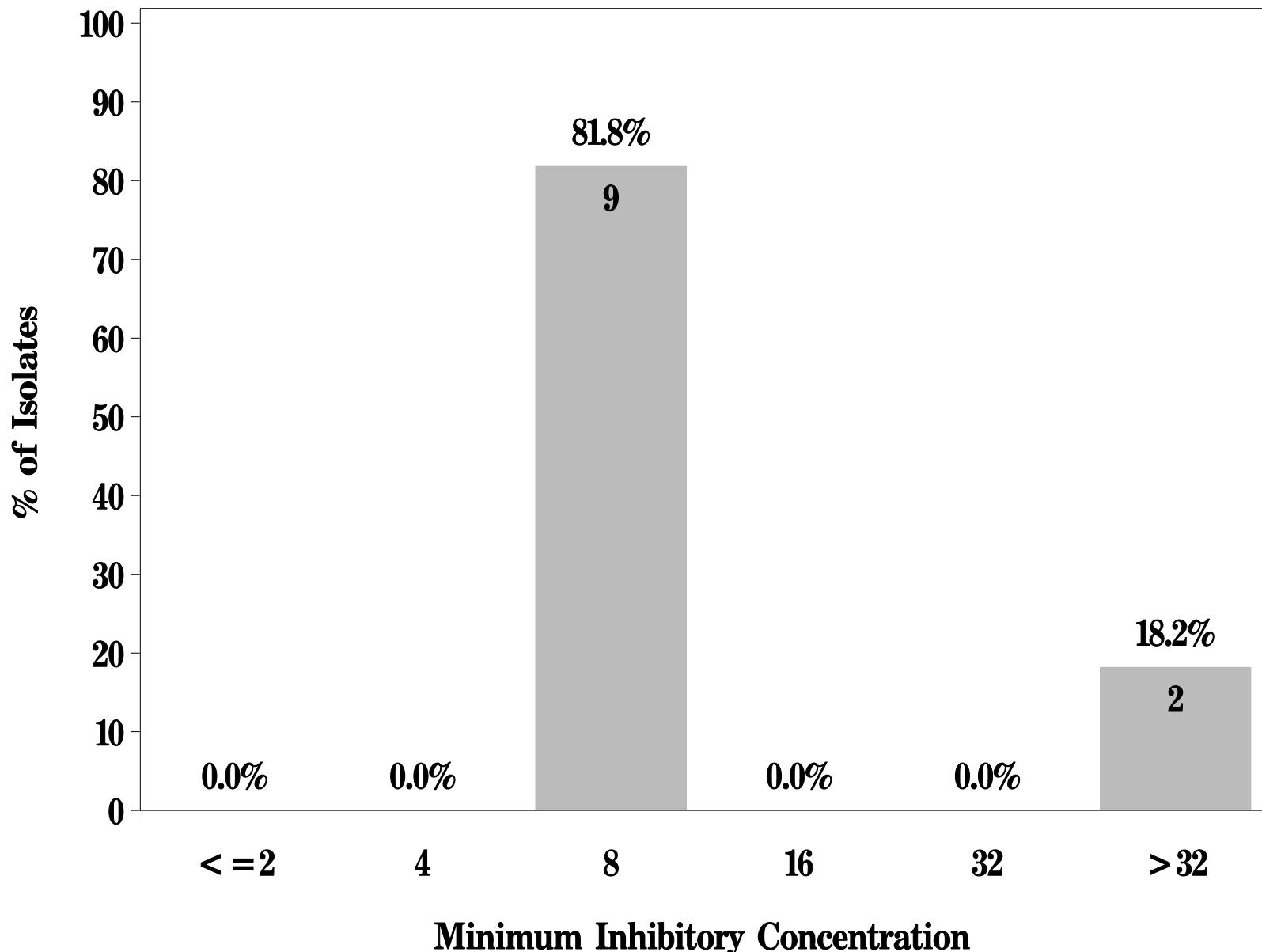
Breakpoints: Susceptible $\leq 8 \mu\text{g/mL}$ Resistant $\geq 32 \mu\text{g/mL}$



NARMS

**Figure 7g: Minimum Inhibitory Concentration of Chloramphenicol
for *Salmonella* in Pork Chop (N=11 Isolates)**

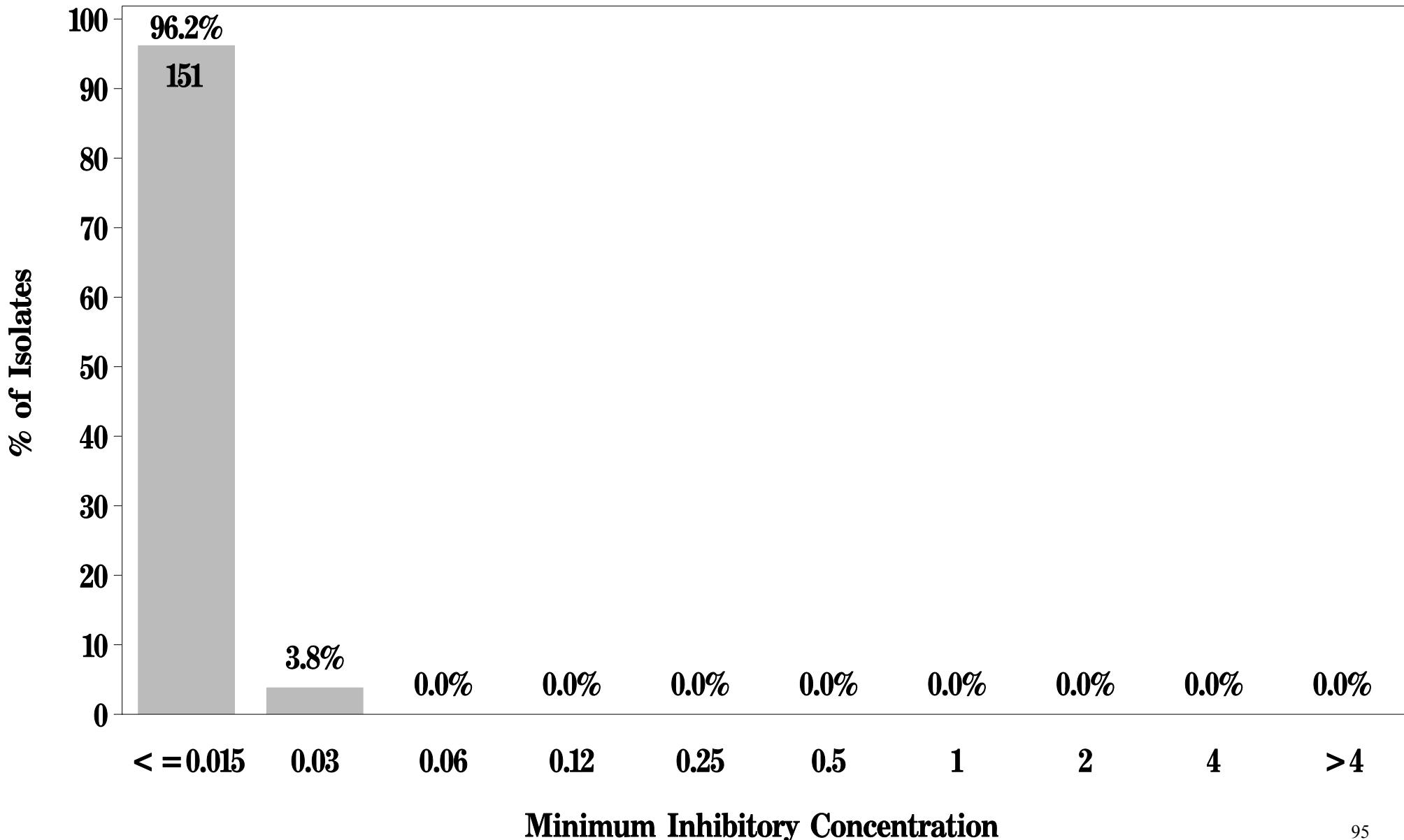
Breakpoints: Susceptible $\leq 8 \mu\text{g/mL}$ Resistant $\geq 32 \mu\text{g/mL}$



NARMS

**Figure 7h: Minimum Inhibitory Concentration of Ciprofloxacin
for *Salmonella* in Chicken Breast (N=157 Isolates)**

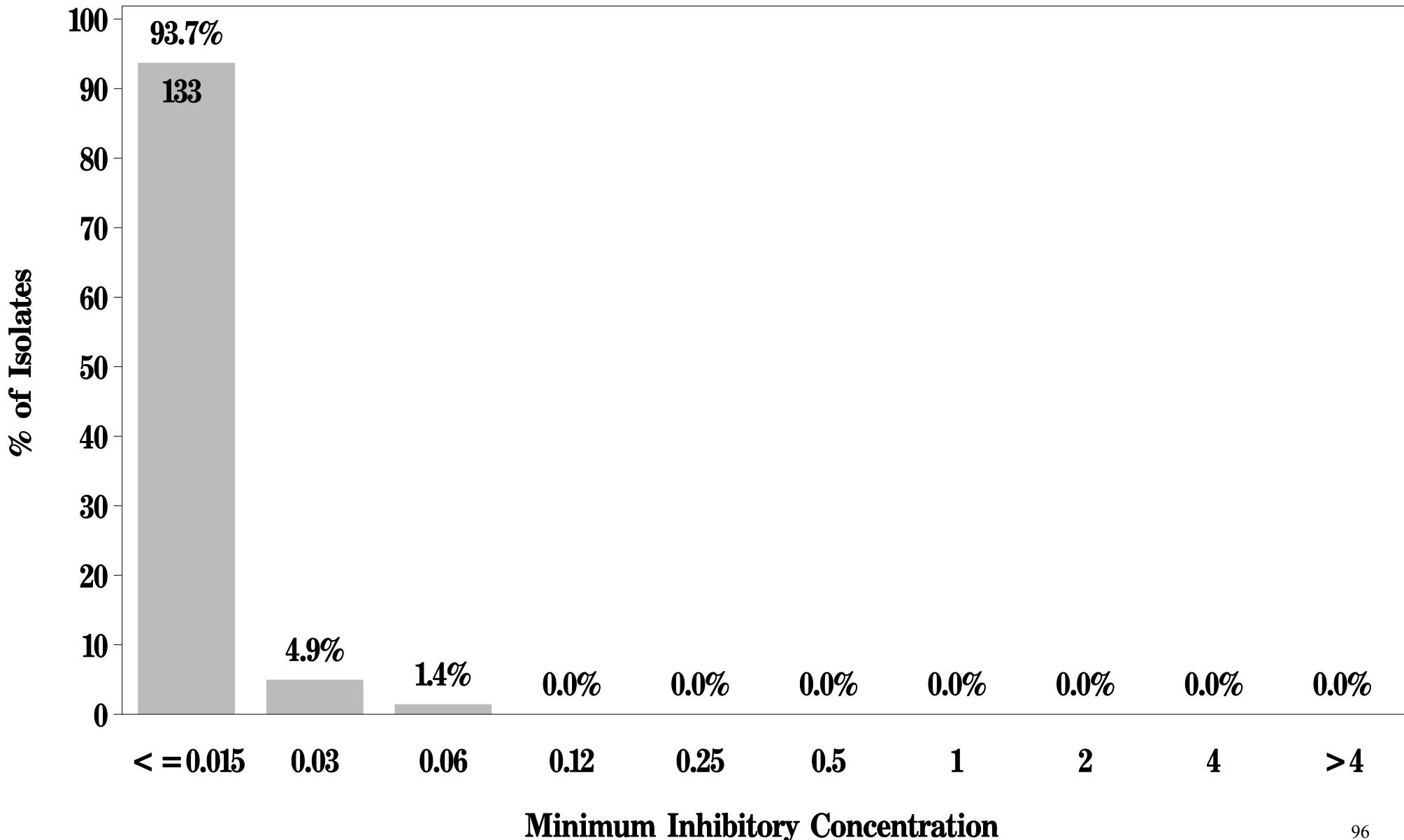
Breakpoints: Susceptible $\leq 1 \mu\text{g/mL}$ Resistant $\geq 4 \mu\text{g/mL}$



NARMS

**Figure 7h: Minimum Inhibitory Concentration of Ciprofloxacin
for *Salmonella* in Ground Turkey (N=142 Isolates)**

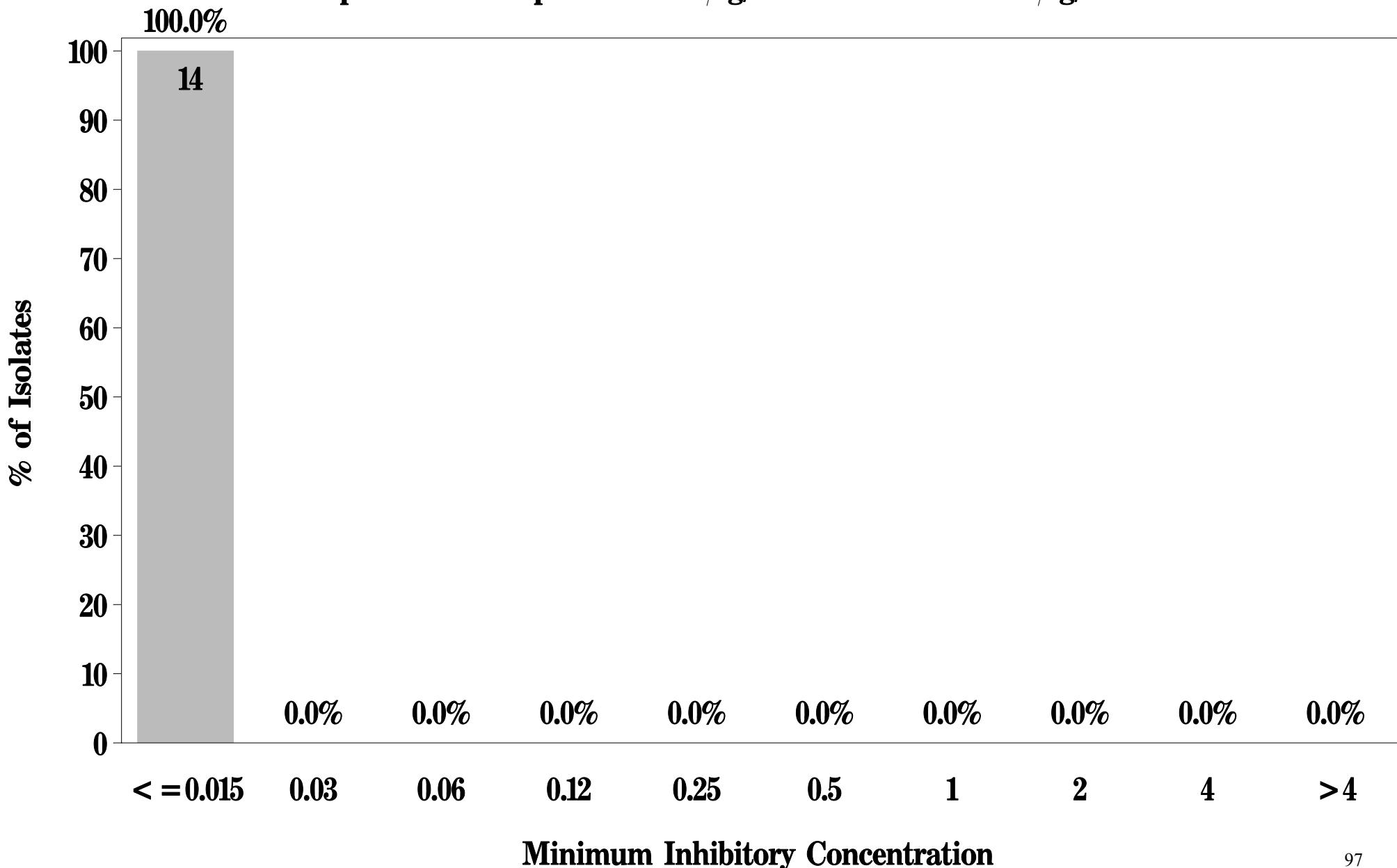
Breakpoints: Susceptible $\leq 1 \mu\text{g/mL}$ Resistant $\geq 4 \mu\text{g/mL}$



NARMS

**Figure 7h: Minimum Inhibitory Concentration of Ciprofloxacin
for *Salmonella* in Ground Beef (N=14 Isolates)**

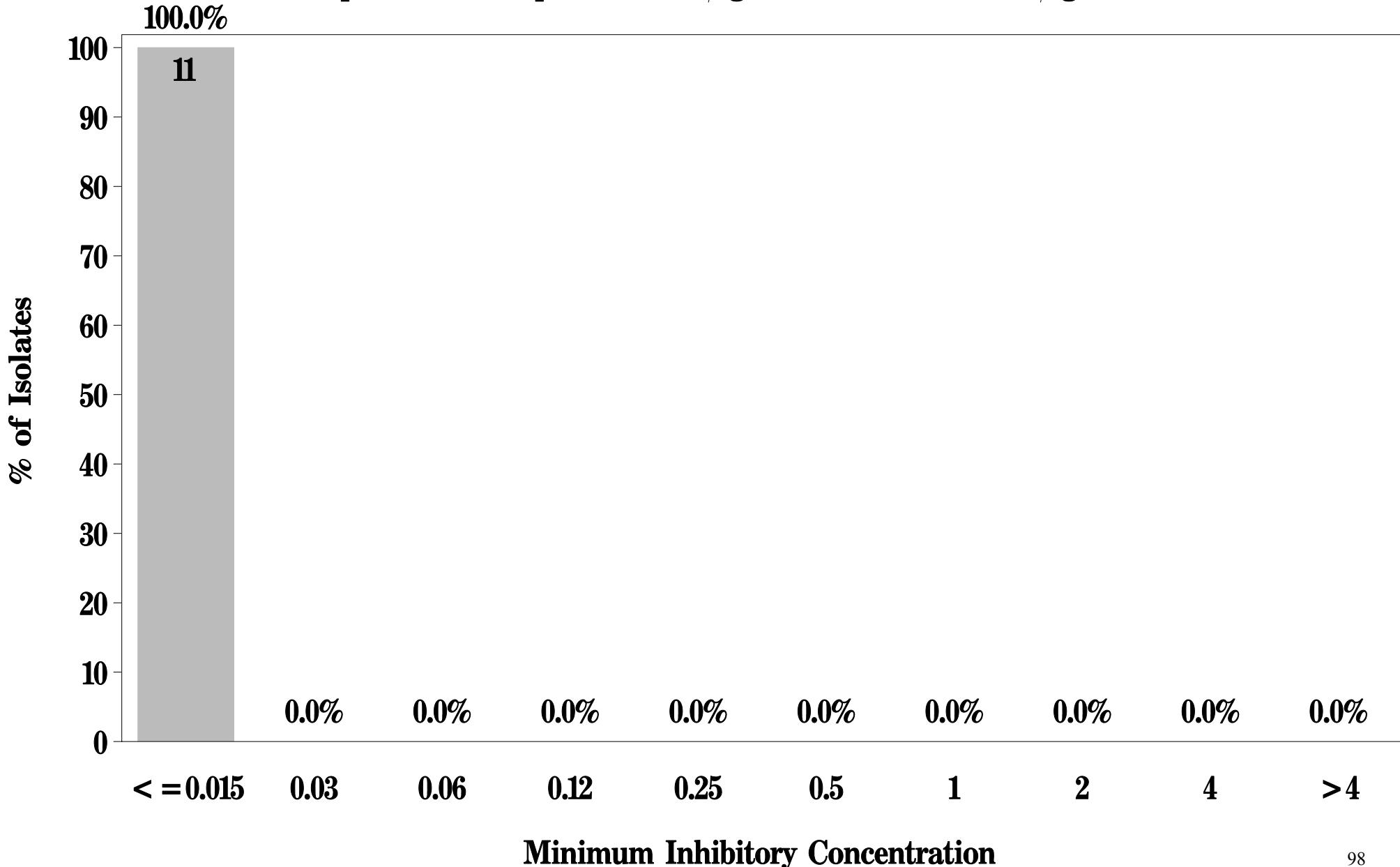
Breakpoints: Susceptible $\leq 1 \mu\text{g/mL}$ Resistant $\geq 4 \mu\text{g/mL}$



NARMS

**Figure 7h: Minimum Inhibitory Concentration of Ciprofloxacin
for *Salmonella* in Pork Chop (N=11 Isolates)**

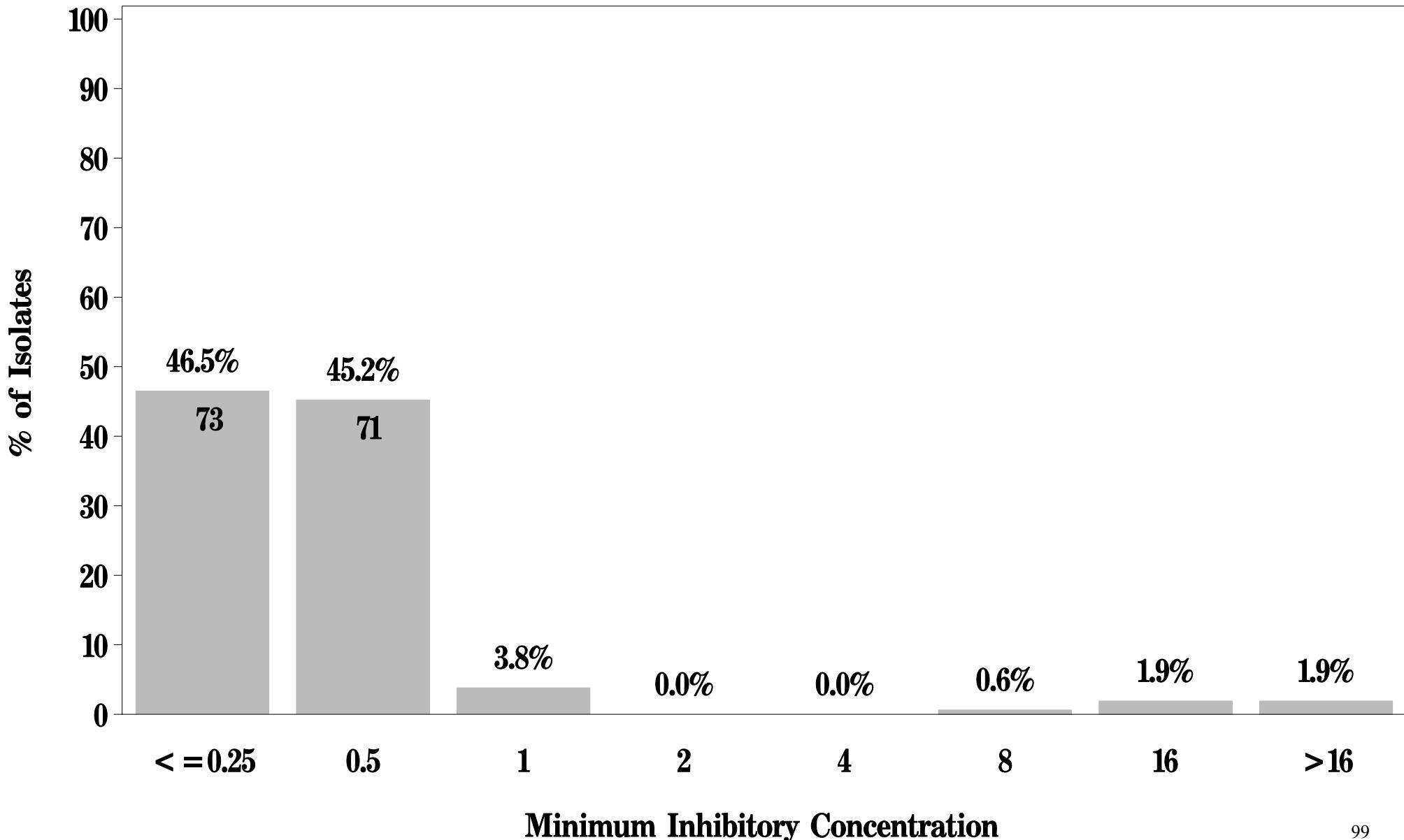
Breakpoints: Susceptible $\leq 1 \mu\text{g/mL}$ Resistant $\geq 4 \mu\text{g/mL}$



NARMS

**Figure 7i: Minimum Inhibitory Concentration of Gentamicin
for *Salmonella* in Chicken Breast (N=157 Isolates)**

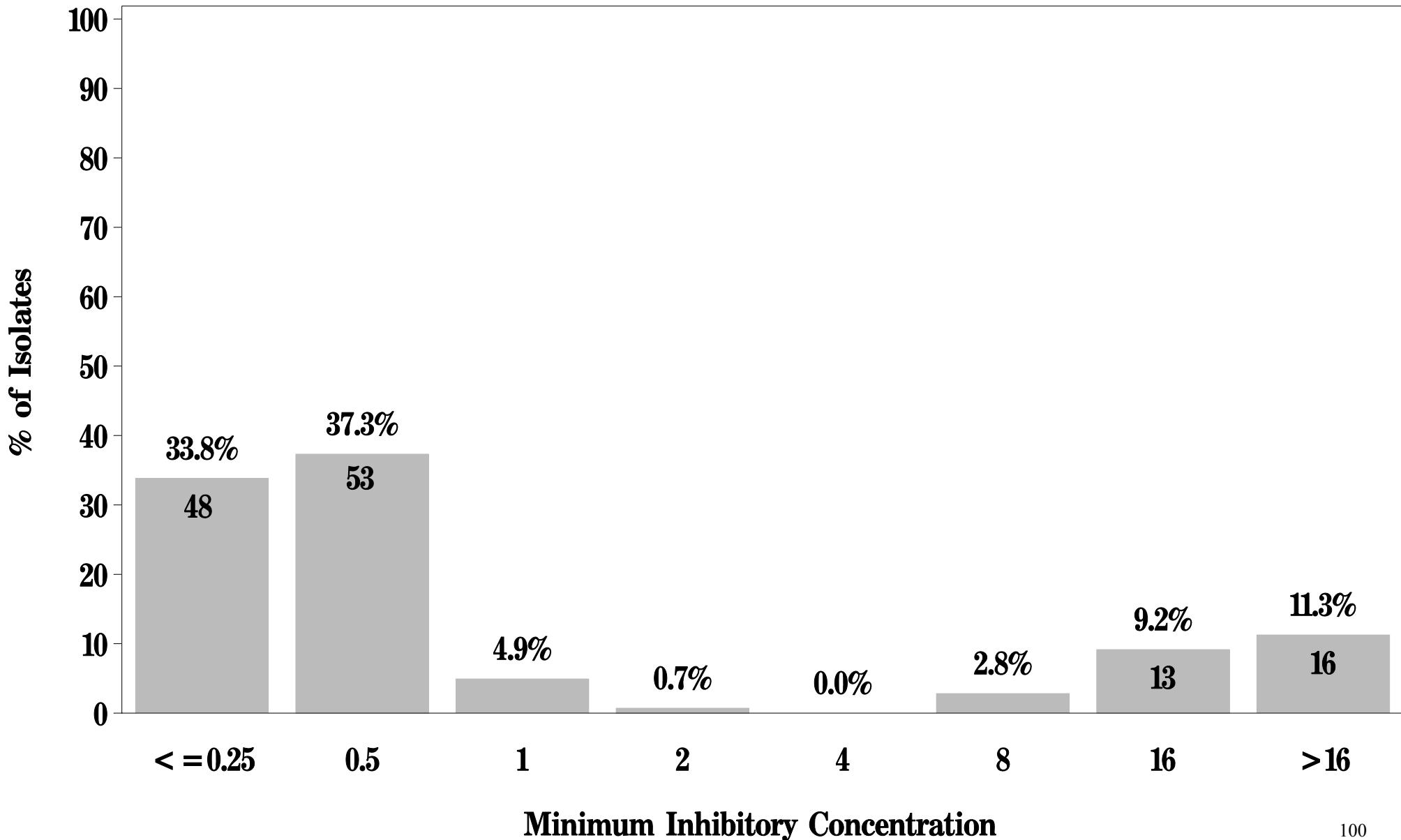
Breakpoints: Susceptible $\leq 4 \text{ } \mu\text{g/mL}$ Resistant $\geq 16 \text{ } \mu\text{g/mL}$



NARMS

**Figure 7i: Minimum Inhibitory Concentration of Gentamicin
for *Salmonella* in Ground Turkey (N=142 Isolates)**

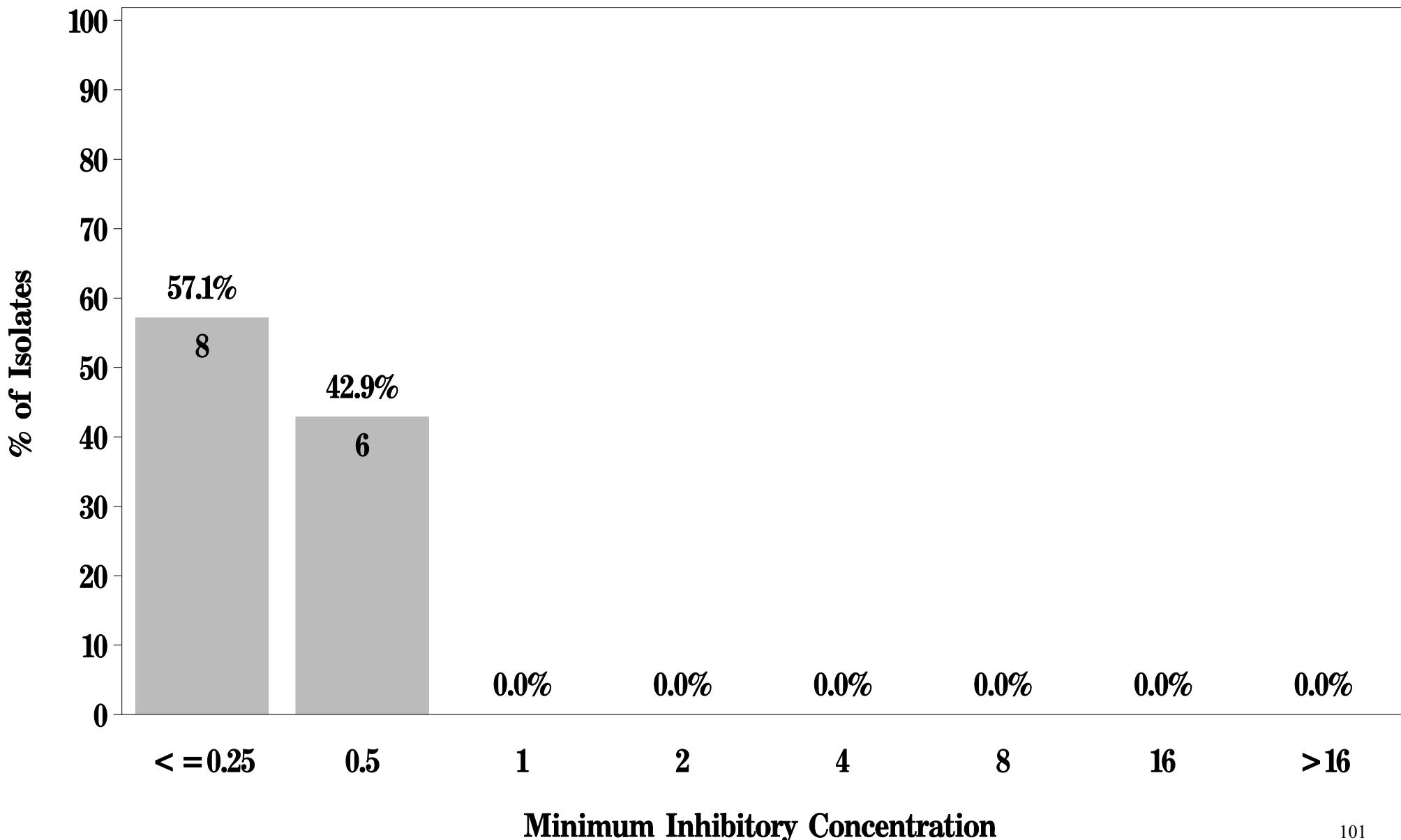
Breakpoints: Susceptible $\leq 4 \text{ } \mu\text{g/mL}$ Resistant $\geq 16 \text{ } \mu\text{g/mL}$



NARMS

**Figure 7i: Minimum Inhibitory Concentration of Gentamicin
for *Salmonella* in Ground Beef (N=14 Isolates)**

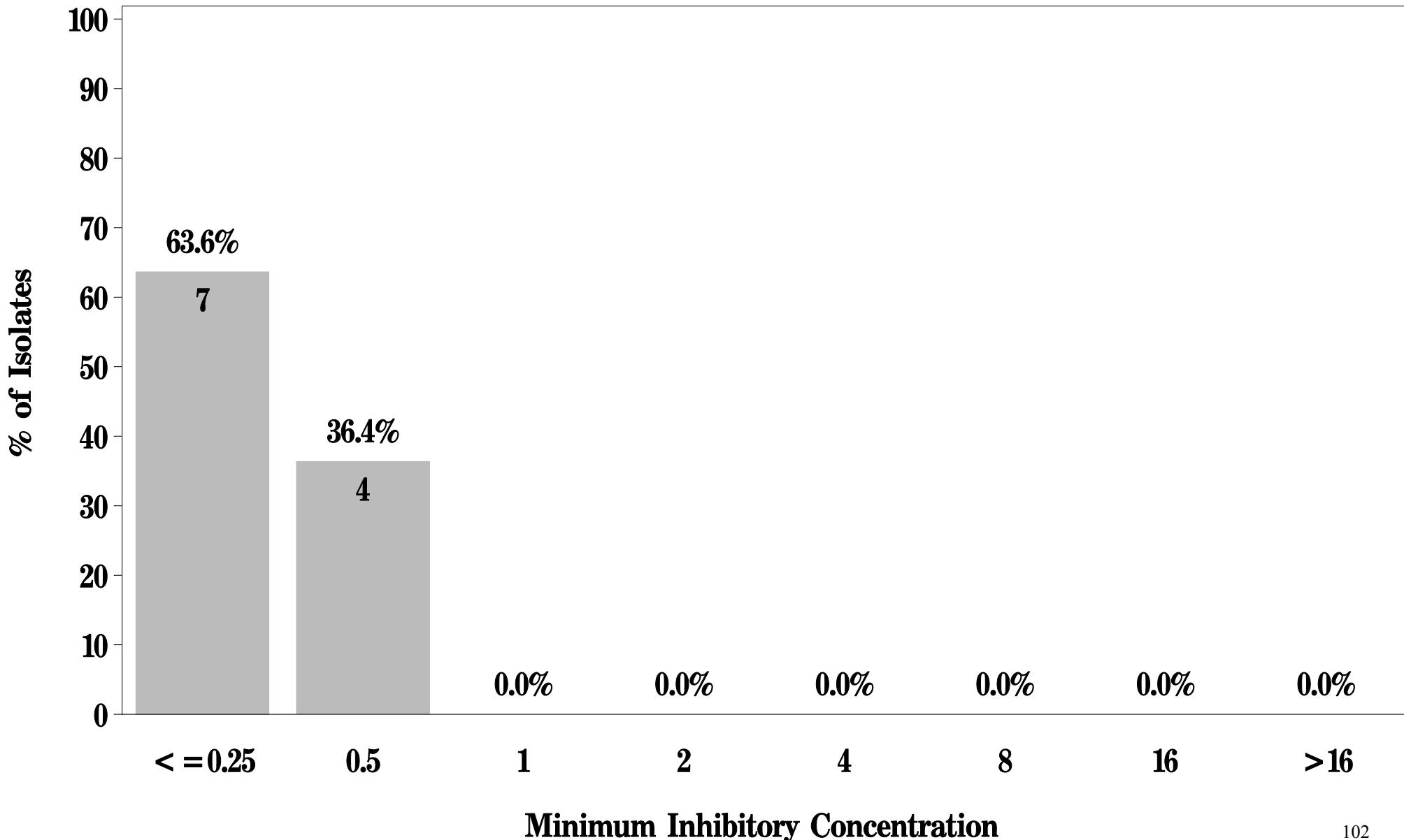
Breakpoints: Susceptible $\leq 4 \mu\text{g/mL}$ Resistant $\geq 16 \mu\text{g/mL}$



NARMS

**Figure 7i: Minimum Inhibitory Concentration of Gentamicin
for *Salmonella* in Pork Chop (N=11 Isolates)**

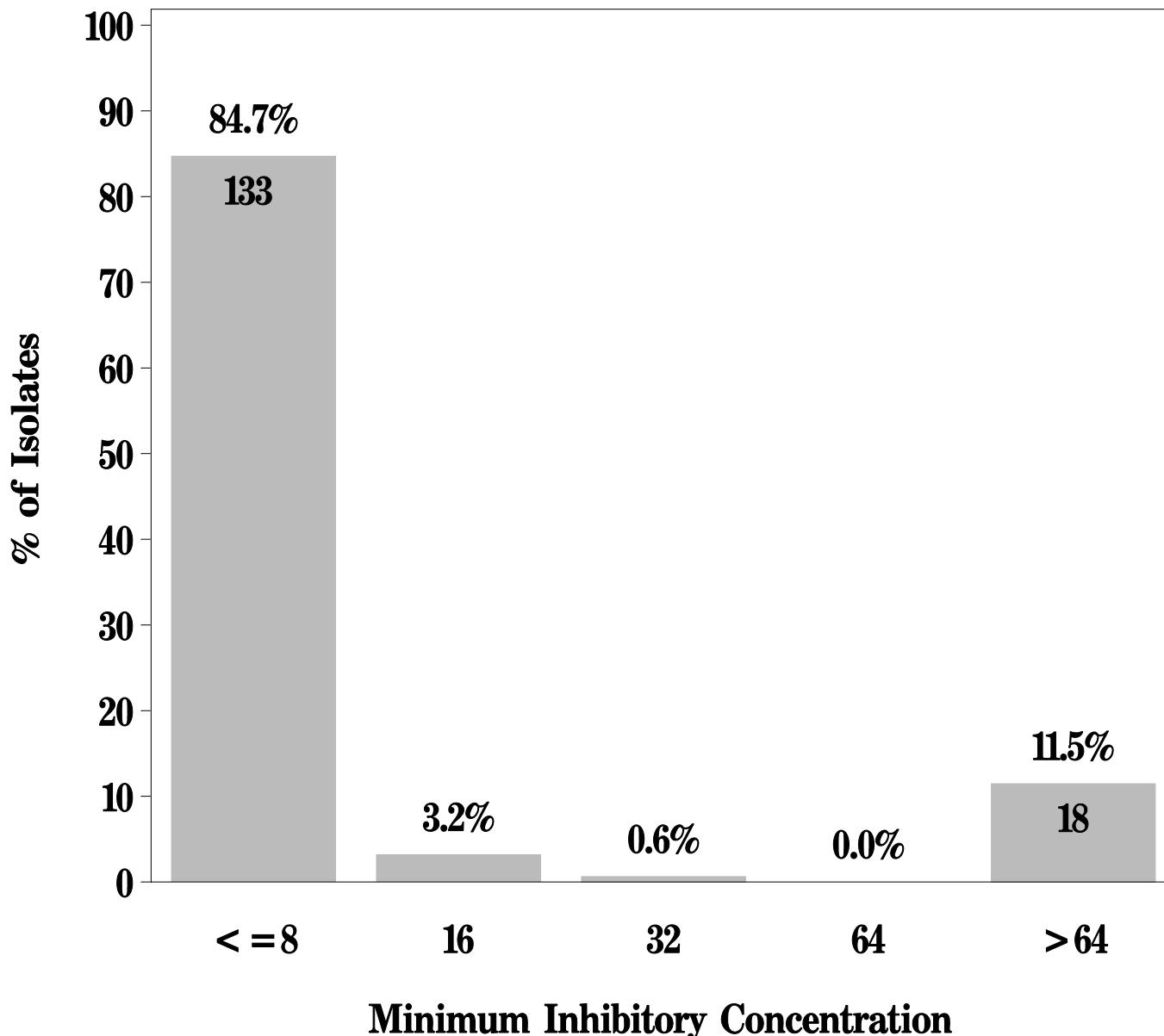
Breakpoints: Susceptible $\leq 4 \text{ } \mu\text{g/mL}$ Resistant $\geq 16 \text{ } \mu\text{g/mL}$



NARMS

**Figure 7j: Minimum Inhibitory Concentration of Kanamycin
for *Salmonella* in Chicken Breast (N=157 Isolates)**

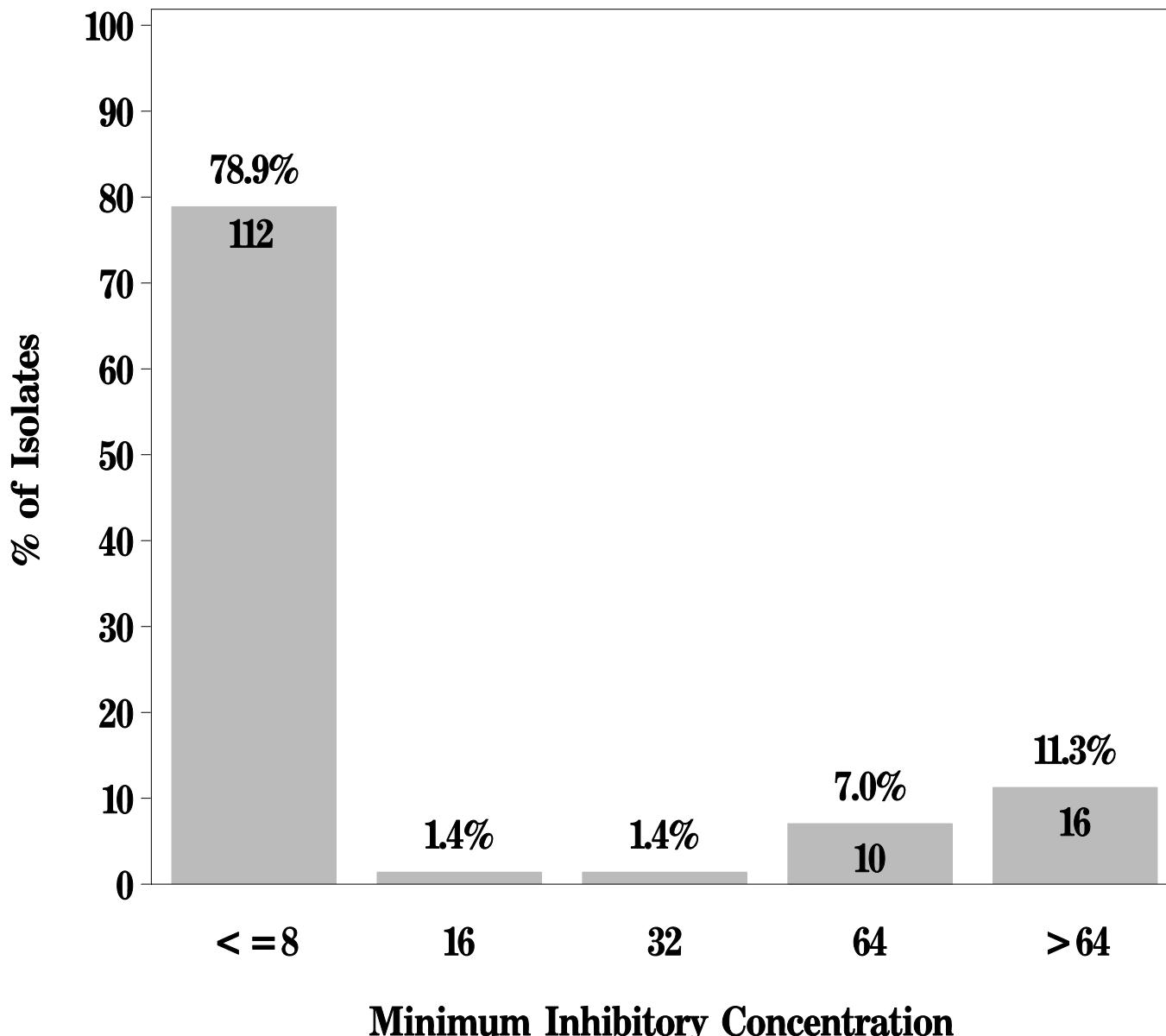
Breakpoints: Susceptible $\leq 16 \mu\text{g/mL}$ Resistant $\geq 64 \mu\text{g/mL}$



NARMS

**Figure 7j: Minimum Inhibitory Concentration of Kanamycin
for *Salmonella* in Ground Turkey (N=142 Isolates)**

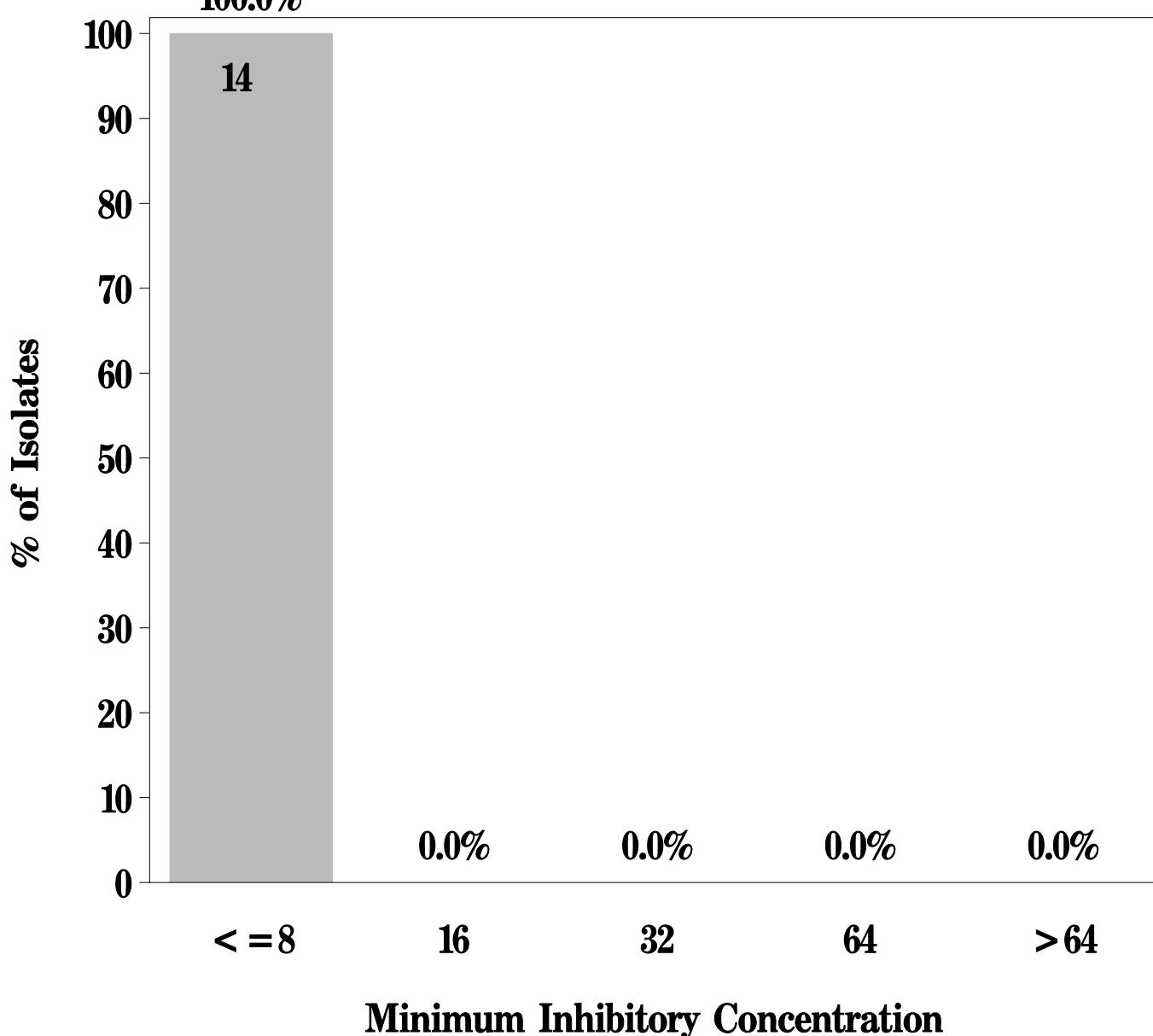
Breakpoints: Susceptible $\leq 16 \mu\text{g/mL}$ Resistant $\geq 64 \mu\text{g/mL}$



NARMS

**Figure 7j: Minimum Inhibitory Concentration of Kanamycin
for *Salmonella* in Ground Beef (N=14 Isolates)**

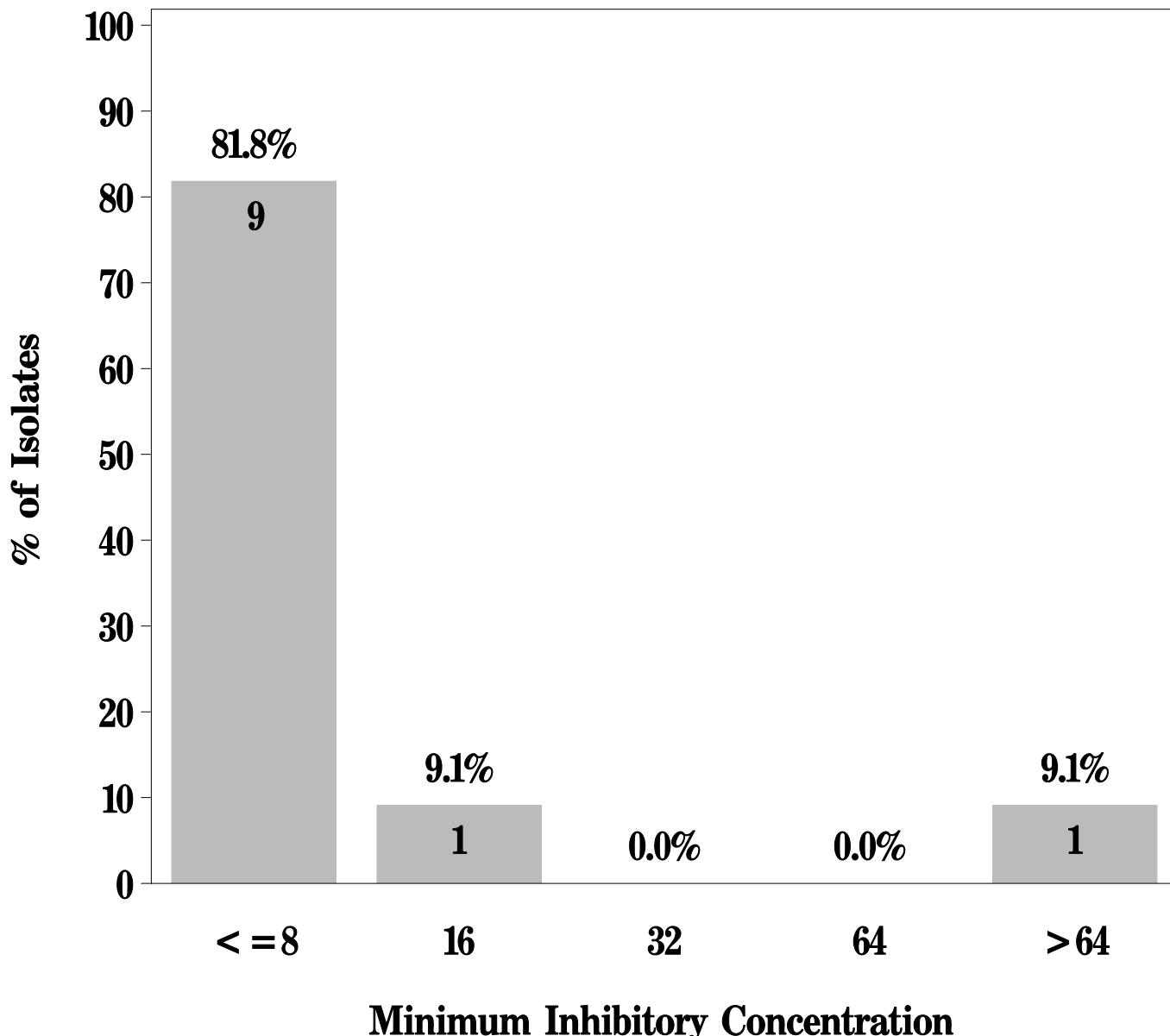
Breakpoints: Susceptible $\leq 16 \mu\text{g/mL}$ Resistant $\geq 64 \mu\text{g/mL}$



NARMS

**Figure 7j: Minimum Inhibitory Concentration of Kanamycin
for *Salmonella* in Pork Chop (N=11 Isolates)**

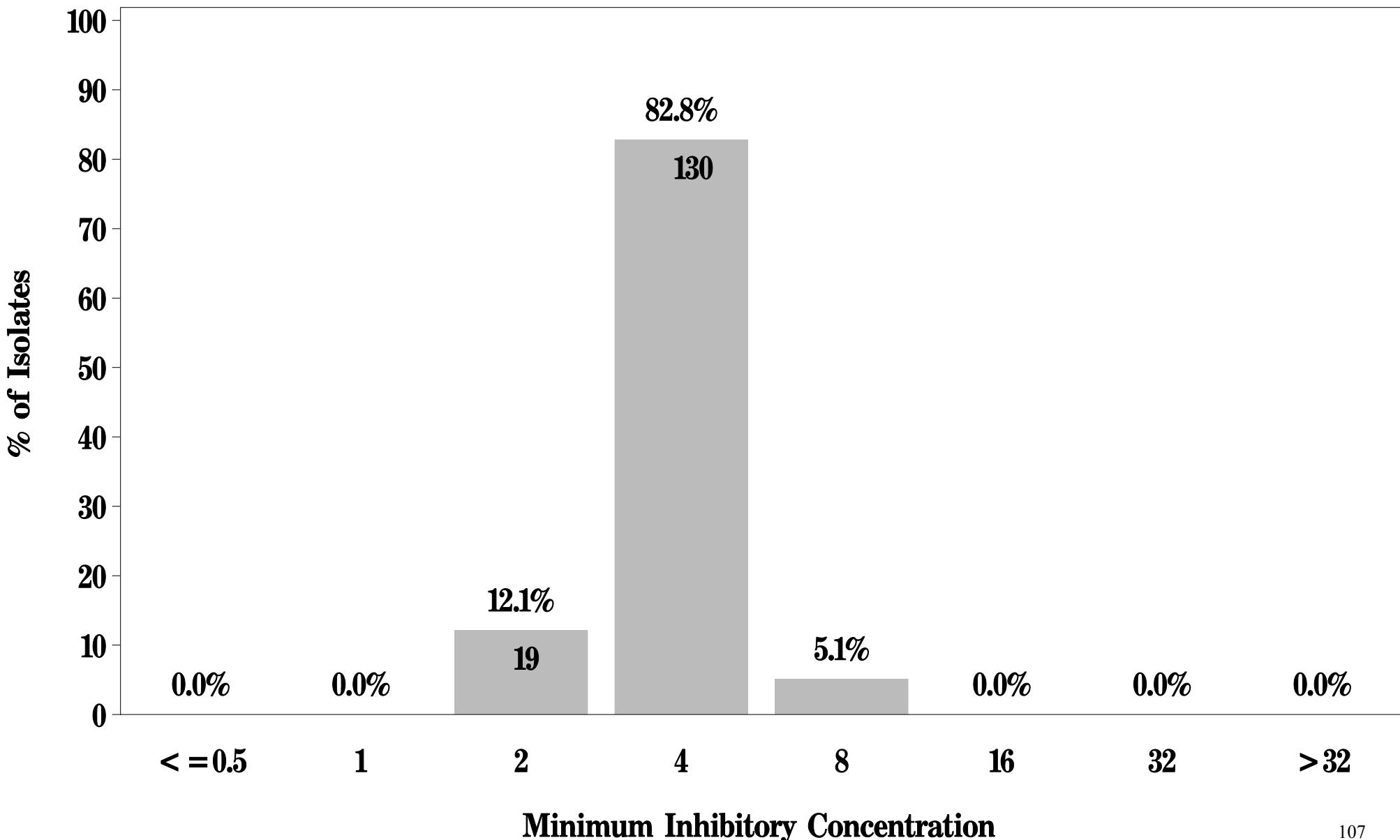
Breakpoints: Susceptible $\leq 16 \mu\text{g/mL}$ Resistant $\geq 64 \mu\text{g/mL}$



NARMS

**Figure 7k: Minimum Inhibitory Concentration of Nalidixic acid
for *Salmonella* in Chicken Breast (N=157 Isolates)**

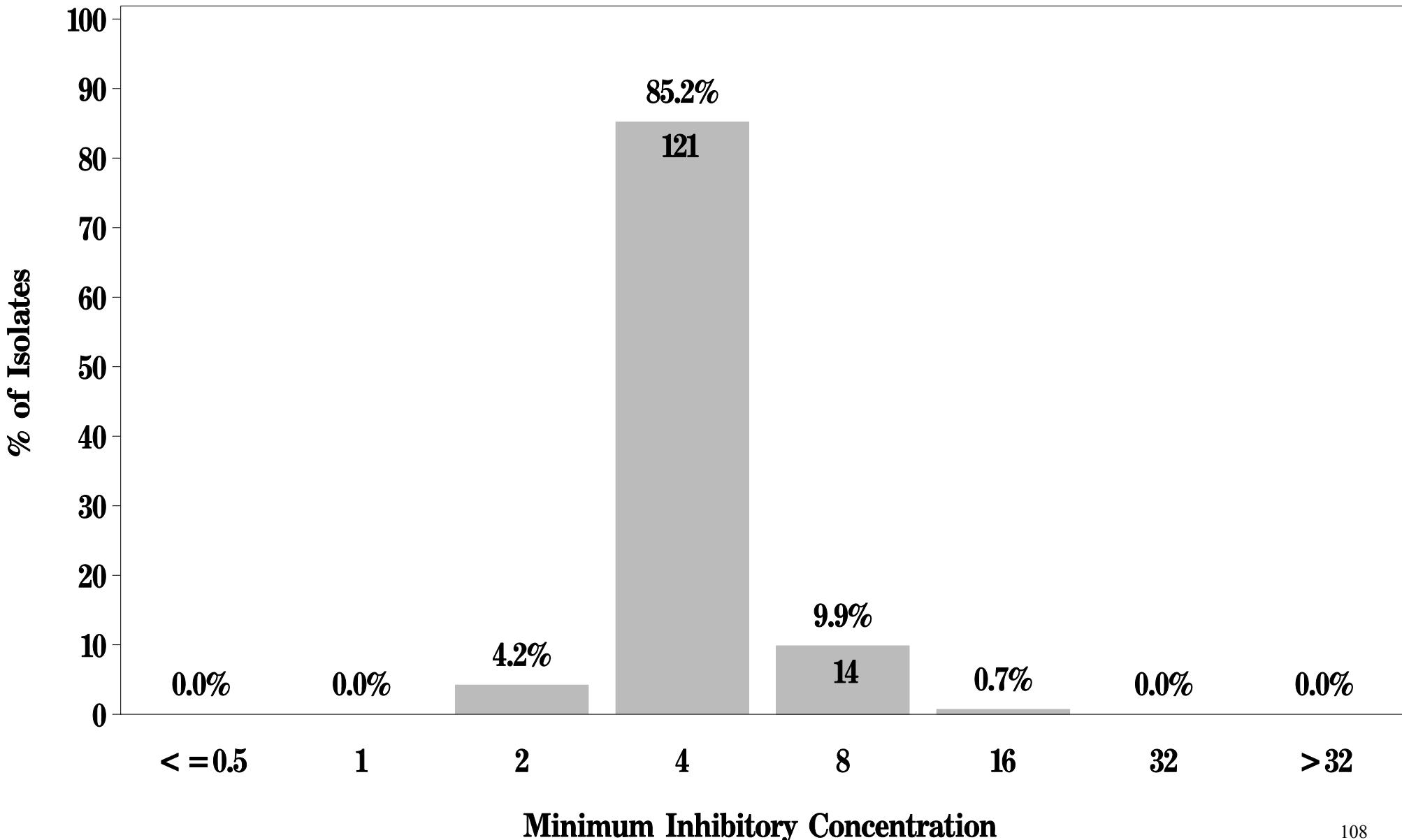
Breakpoints: Susceptible $\leq 16 \mu\text{g/mL}$ Resistant $\geq 32 \mu\text{g/mL}$



NARMS

**Figure 7k: Minimum Inhibitory Concentration of Nalidixic acid
for *Salmonella* in Ground Turkey (N=142 Isolates)**

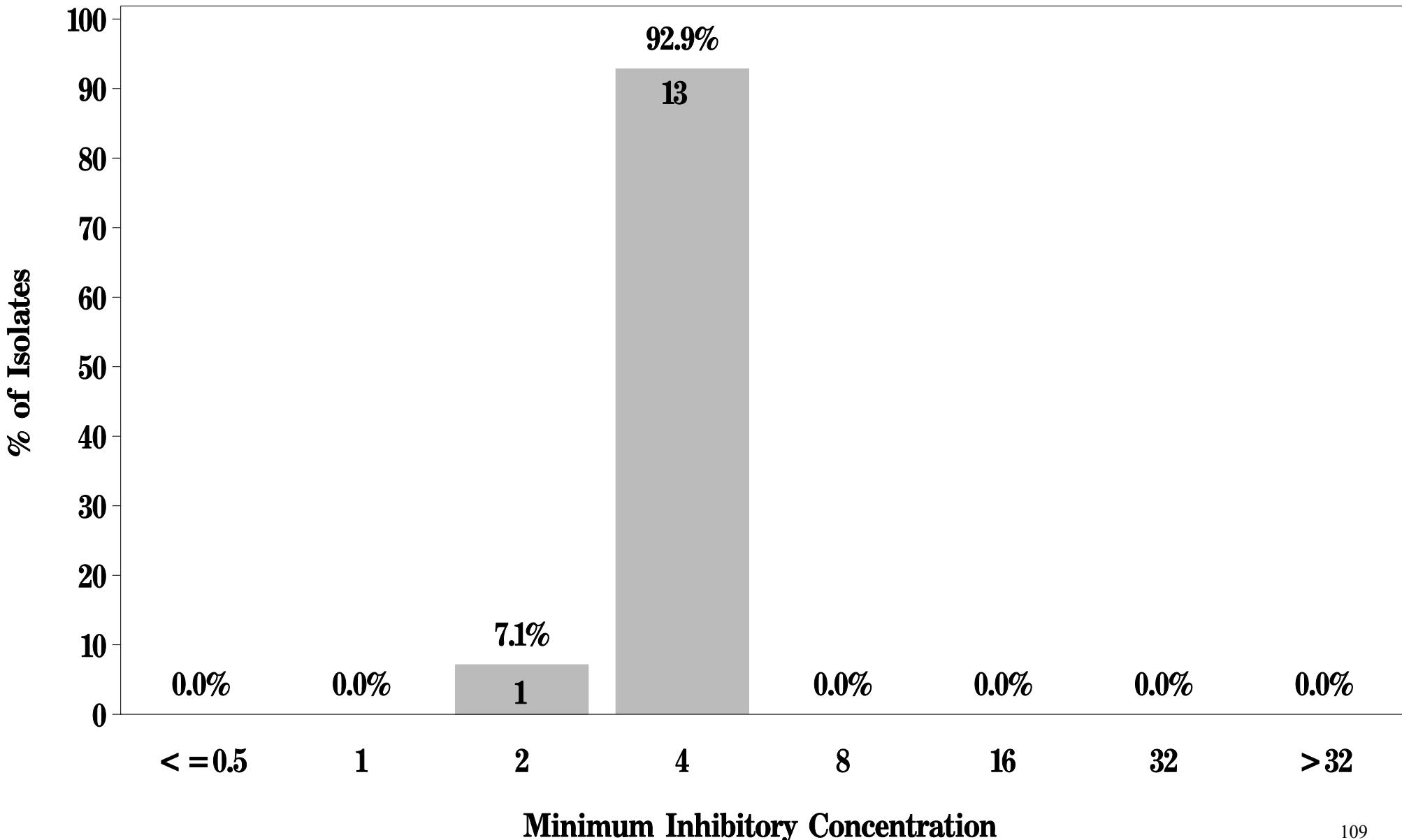
Breakpoints: Susceptible $\leq 16 \mu\text{g/mL}$ Resistant $\geq 32 \mu\text{g/mL}$



NARMS

**Figure 7k: Minimum Inhibitory Concentration of Nalidixic acid
for *Salmonella* in Ground Beef (N=14 Isolates)**

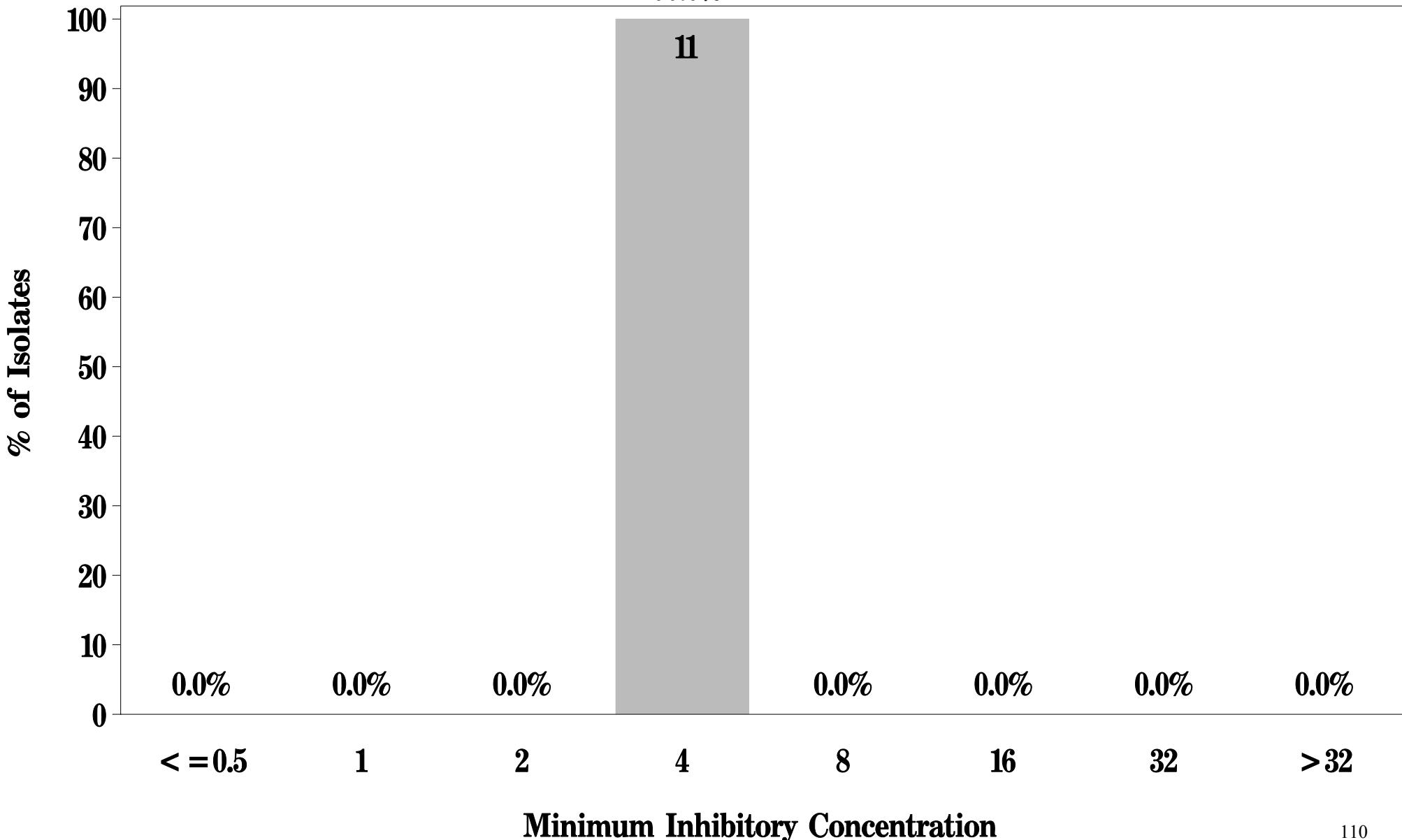
Breakpoints: Susceptible $\leq 16 \mu\text{g/mL}$ Resistant $\geq 32 \mu\text{g/mL}$



NARMS

**Figure 7k: Minimum Inhibitory Concentration of Nalidixic acid
for *Salmonella* in Pork Chop (N=11 Isolates)**

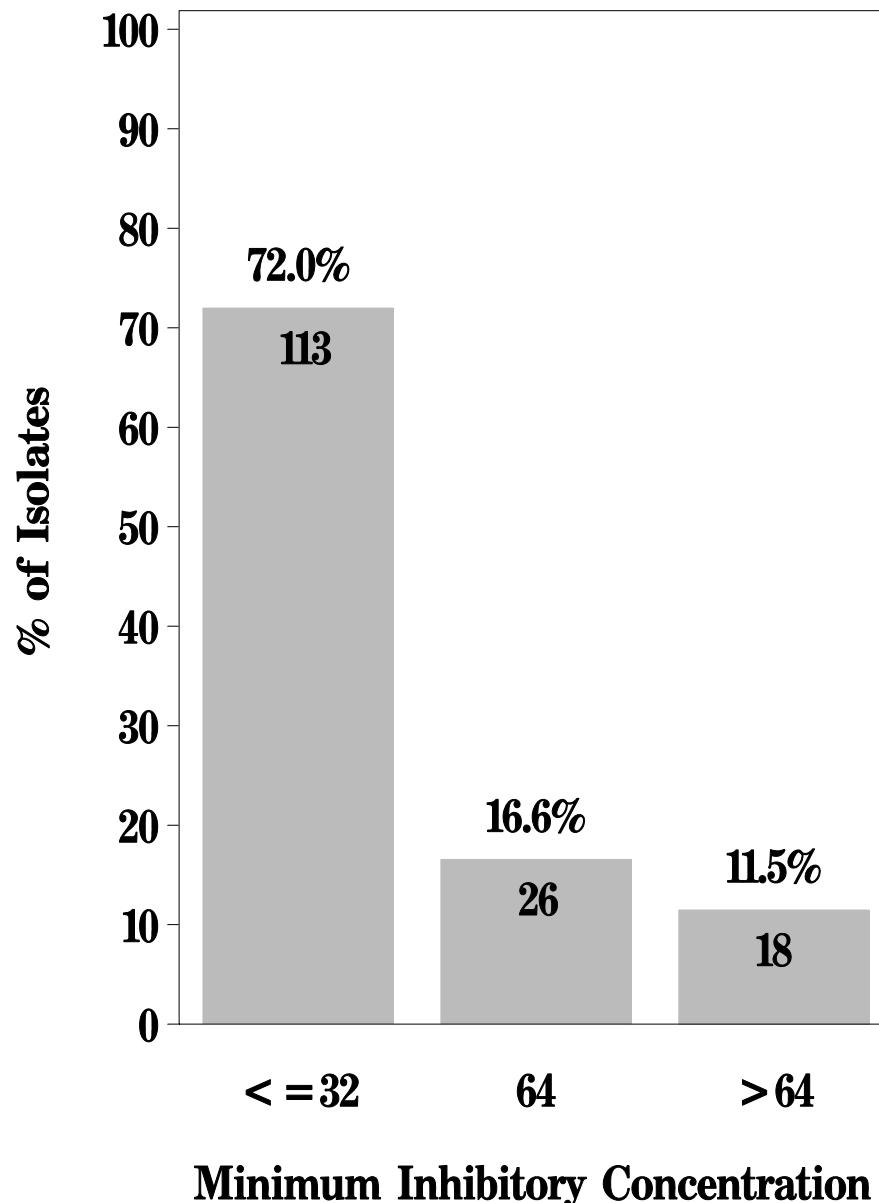
Breakpoints: Susceptible $\leq 16 \mu\text{g/mL}$ Resistant $\geq 32 \mu\text{g/mL}$
100.0%



NARMS

**Figure 7l: Minimum Inhibitory Concentration of Streptomycin
for *Salmonella* in Chicken Breast (N=157 Isolates)**

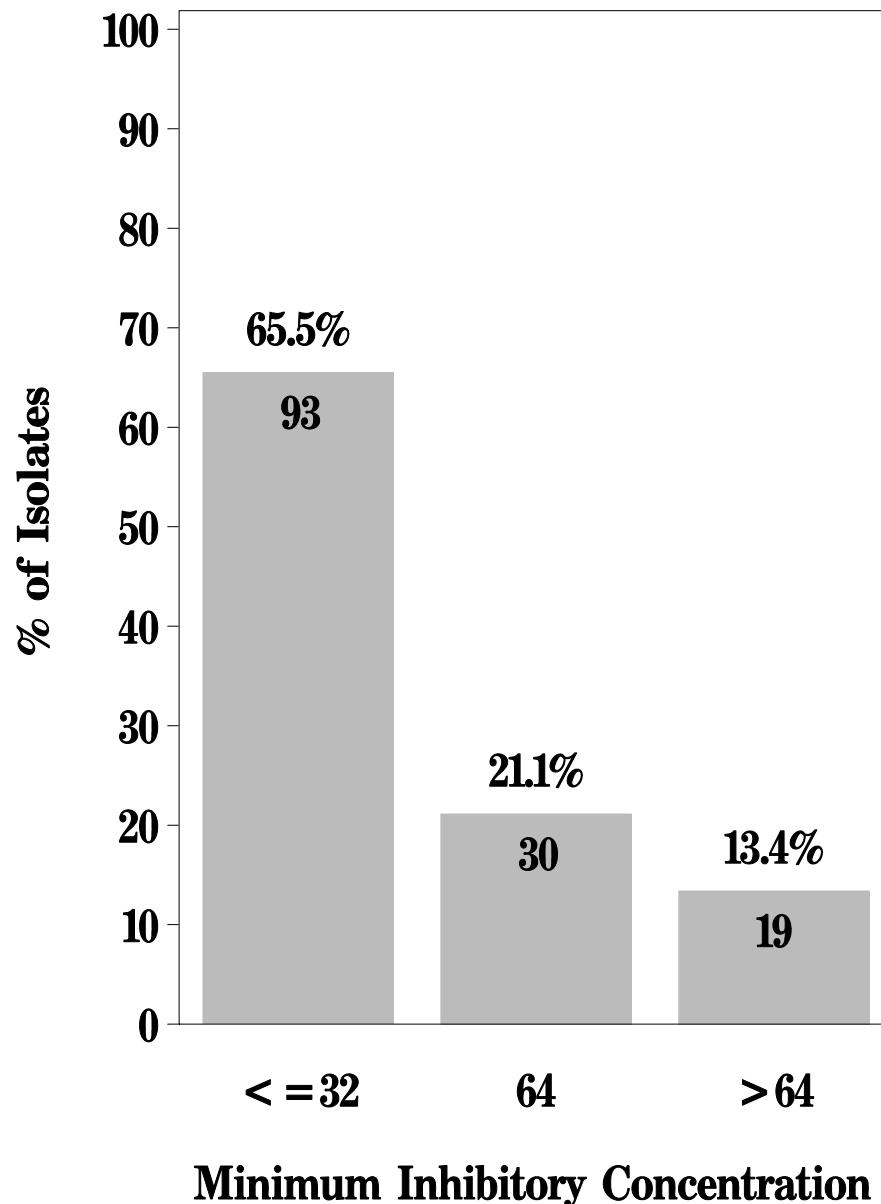
Breakpoints: Susceptible $\leq 32 \mu\text{g/mL}$ Resistant $\geq 64 \mu\text{g/mL}$



NARMS

**Figure 7l: Minimum Inhibitory Concentration of Streptomycin
for *Salmonella* in Ground Turkey (N=142 Isolates)**

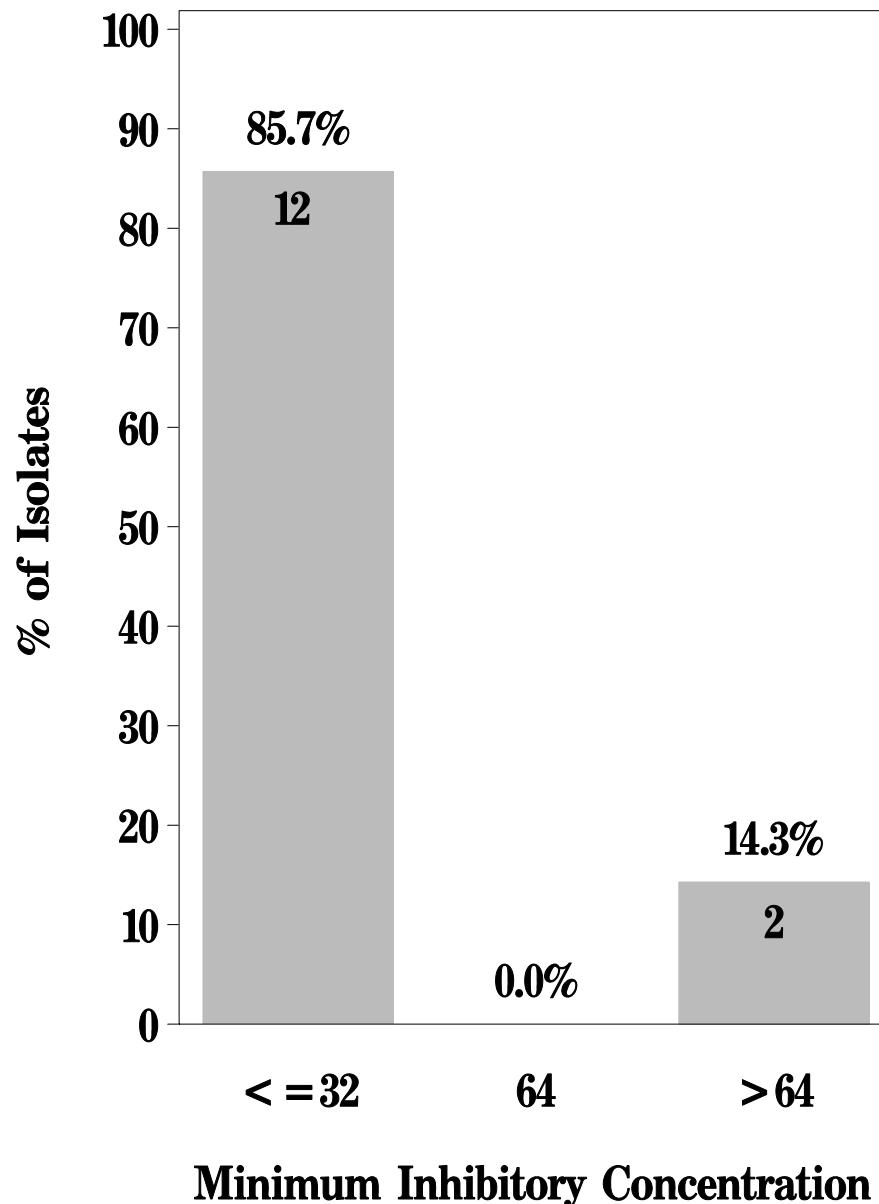
Breakpoints: Susceptible $\leq 32 \mu\text{g/mL}$ Resistant $\geq 64 \mu\text{g/mL}$



NARMS

**Figure 7l: Minimum Inhibitory Concentration of Streptomycin
for *Salmonella* in Ground Beef (N=14 Isolates)**

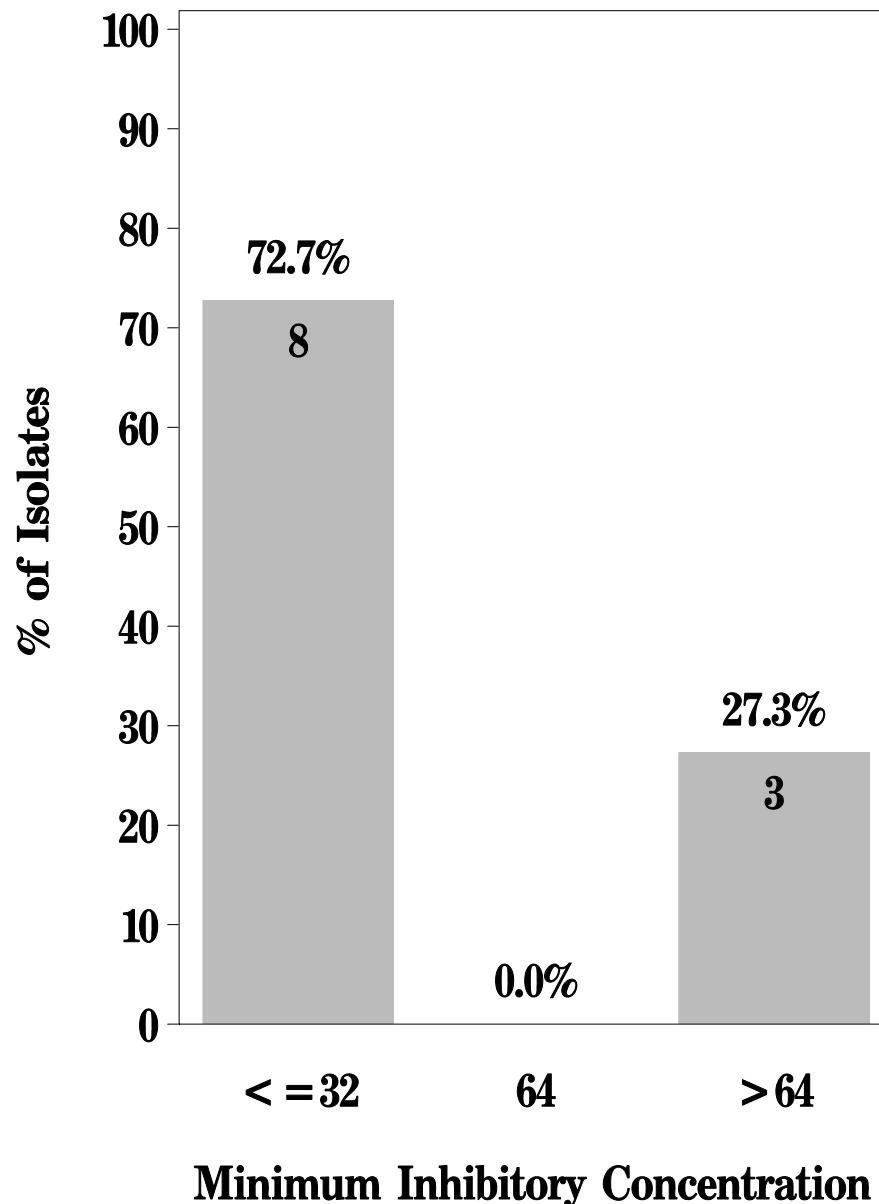
Breakpoints: Susceptible $\leq 32 \text{ } \mu\text{g/mL}$ Resistant $\geq 64 \text{ } \mu\text{g/mL}$



NARMS

**Figure 7l: Minimum Inhibitory Concentration of Streptomycin
for *Salmonella* in Pork Chop (N=11 Isolates)**

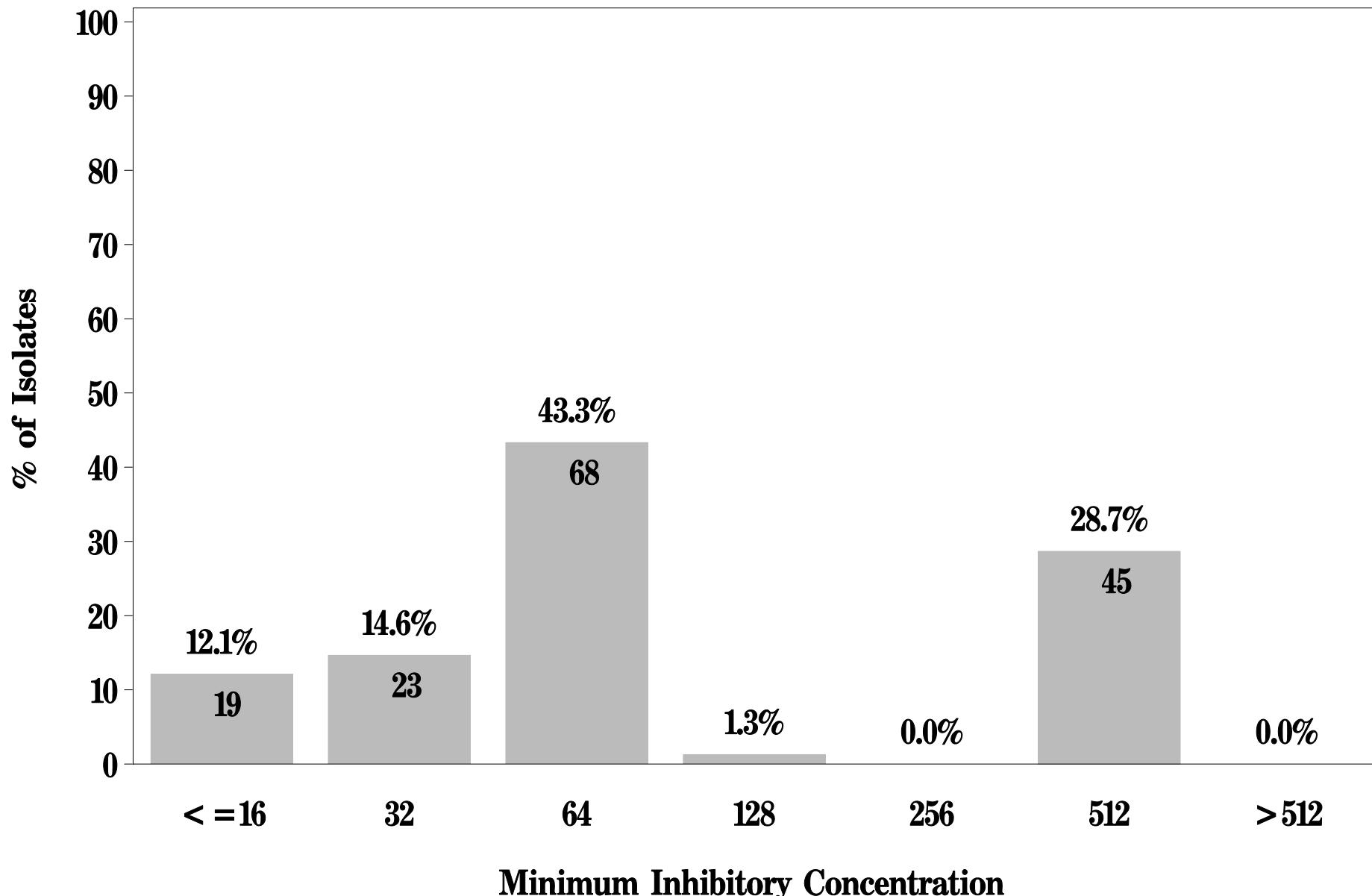
Breakpoints: Susceptible $\leq 32 \mu\text{g/mL}$ Resistant $\geq 64 \mu\text{g/mL}$



NARMS

**Figure 7m: Minimum Inhibitory Concentration of Sulfisoxazole
for *Salmonella* in Chicken Breast (N=157 Isolates)**

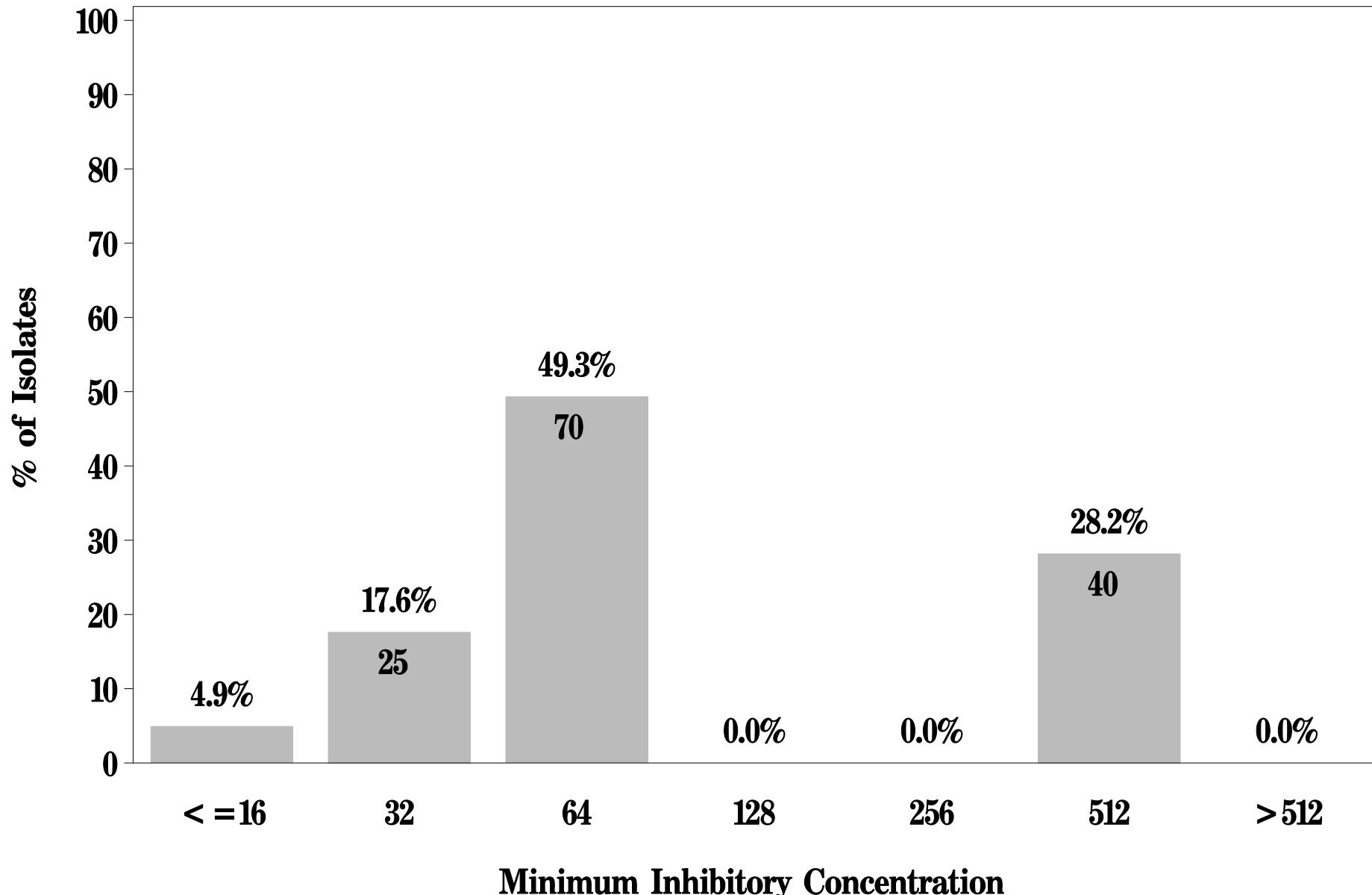
Breakpoints: Susceptible $\leq 256 \mu\text{g/mL}$ Resistant $\geq 512 \mu\text{g/mL}$



NARMS

**Figure 7m: Minimum Inhibitory Concentration of Sulfisoxazole
for *Salmonella* in Ground Turkey (N=142 Isolates)**

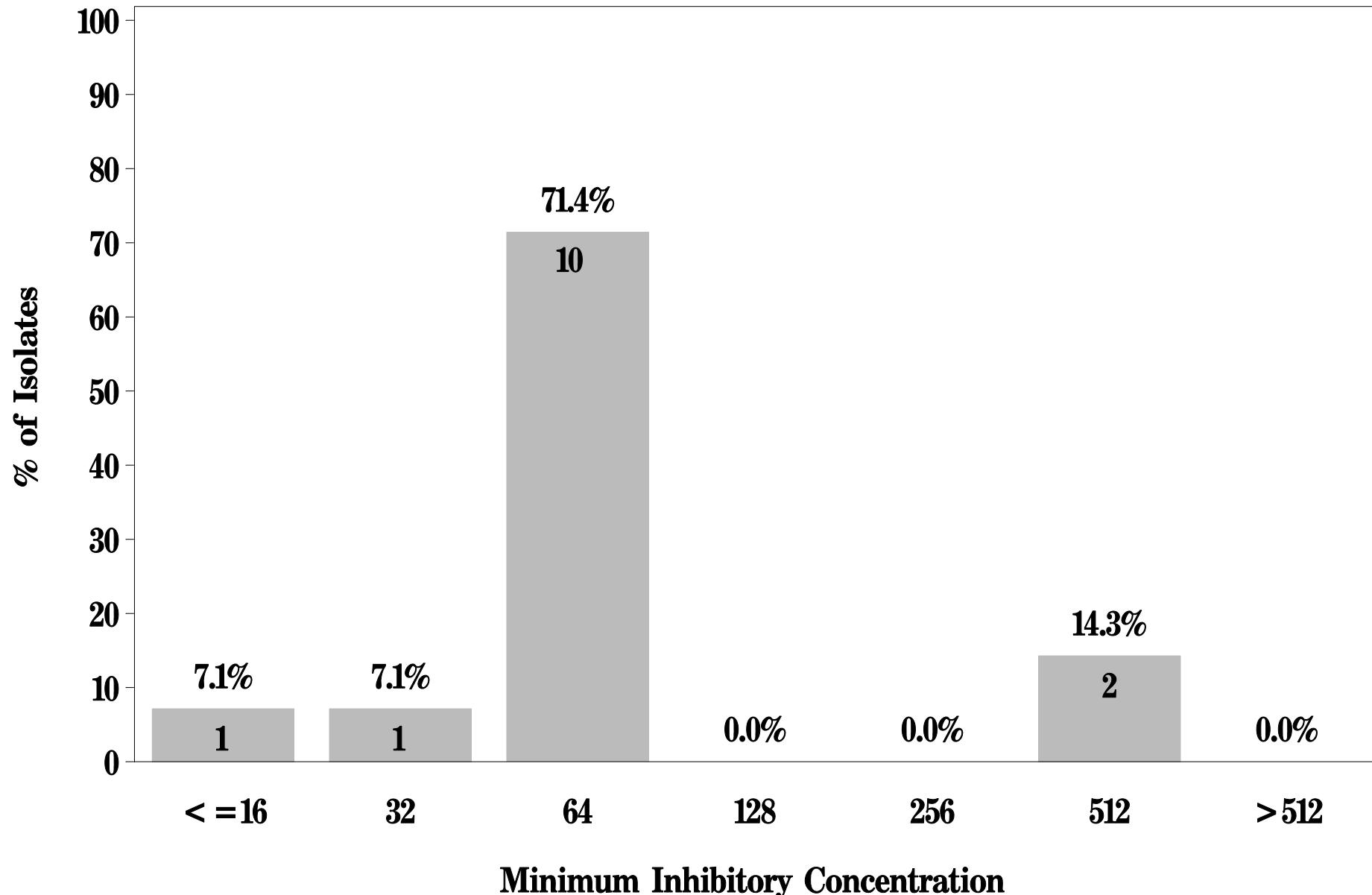
Breakpoints: Susceptible $\leq 256 \mu\text{g/mL}$ Resistant $\geq 512 \mu\text{g/mL}$



NARMS

**Figure 7m: Minimum Inhibitory Concentration of Sulfisoxazole
for *Salmonella* in Ground Beef (N=14 Isolates)**

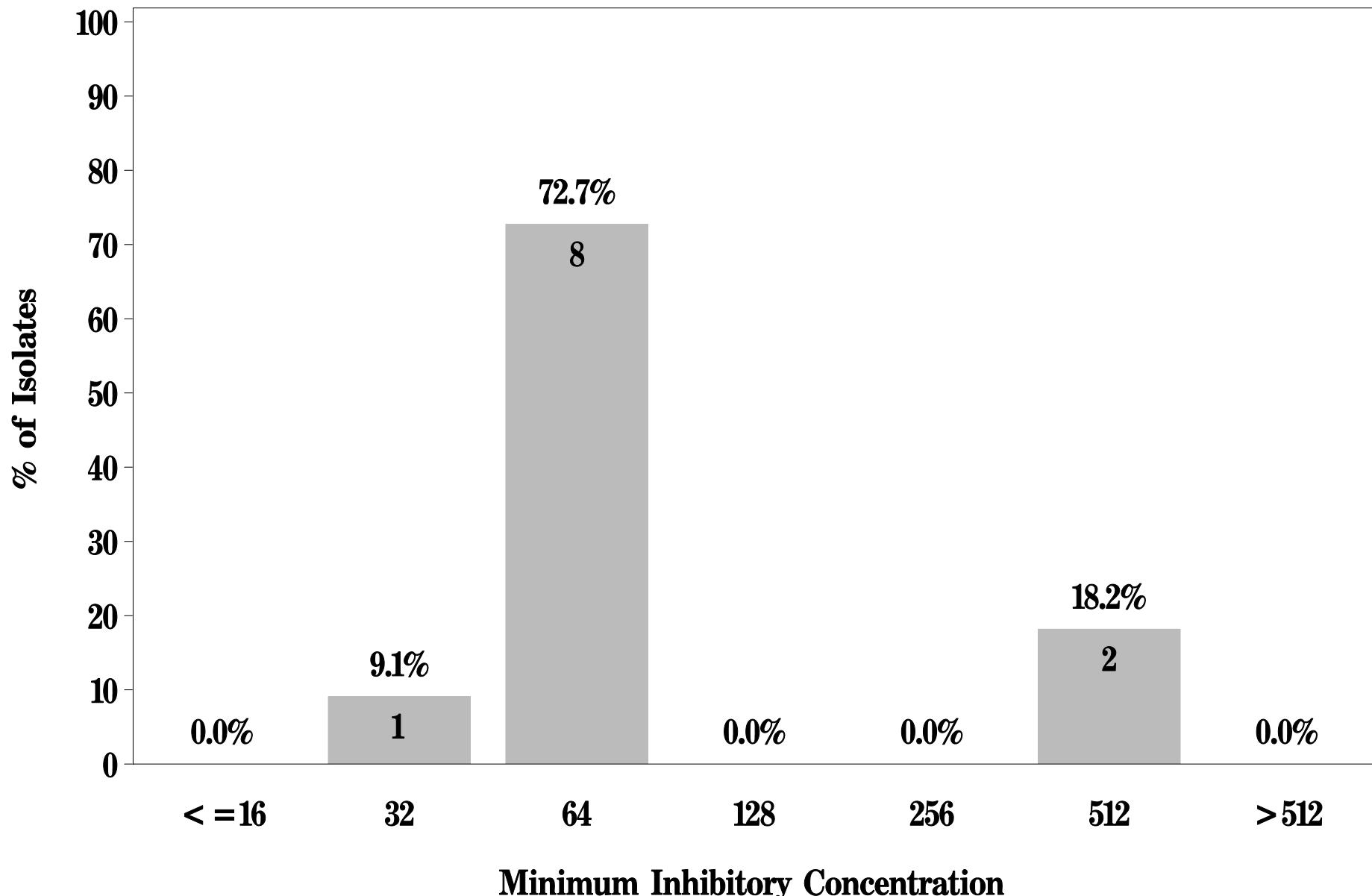
Breakpoints: Susceptible $\leq 256 \mu\text{g/mL}$ Resistant $\geq 512 \mu\text{g/mL}$



NARMS

**Figure 7m: Minimum Inhibitory Concentration of Sulfisoxazole
for *Salmonella* in Pork Chop (N=11 Isolates)**

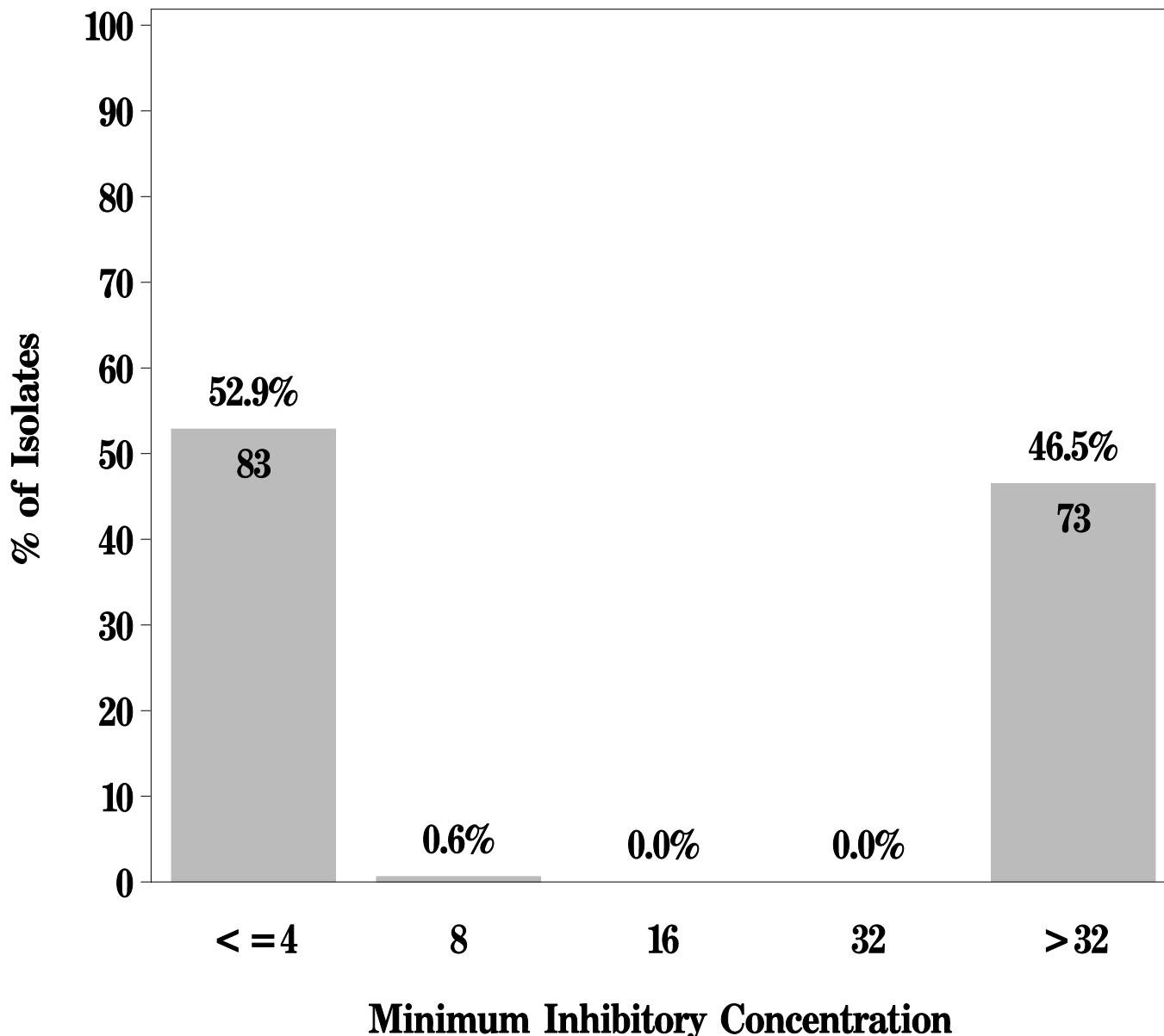
Breakpoints: Susceptible $\leq 256 \mu\text{g/mL}$ Resistant $\geq 512 \mu\text{g/mL}$



NARMS

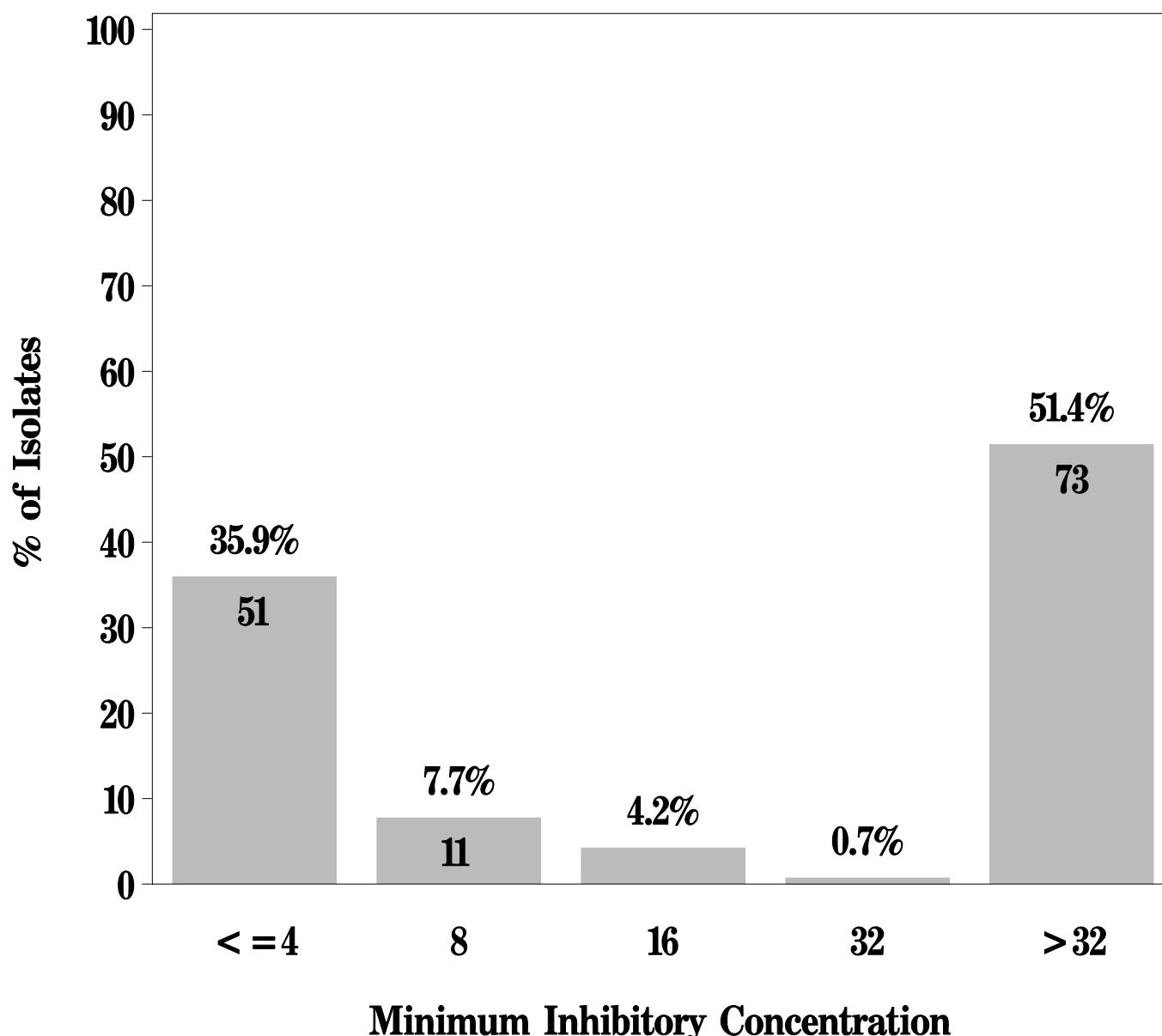
**Figure 7n: Minimum Inhibitory Concentration of Tetracycline
for *Salmonella* in Chicken Breast (N=157 Isolates)**

Breakpoints: Susceptible $\leq 4 \mu\text{g/mL}$ Resistant $\geq 16 \mu\text{g/mL}$



NARMS

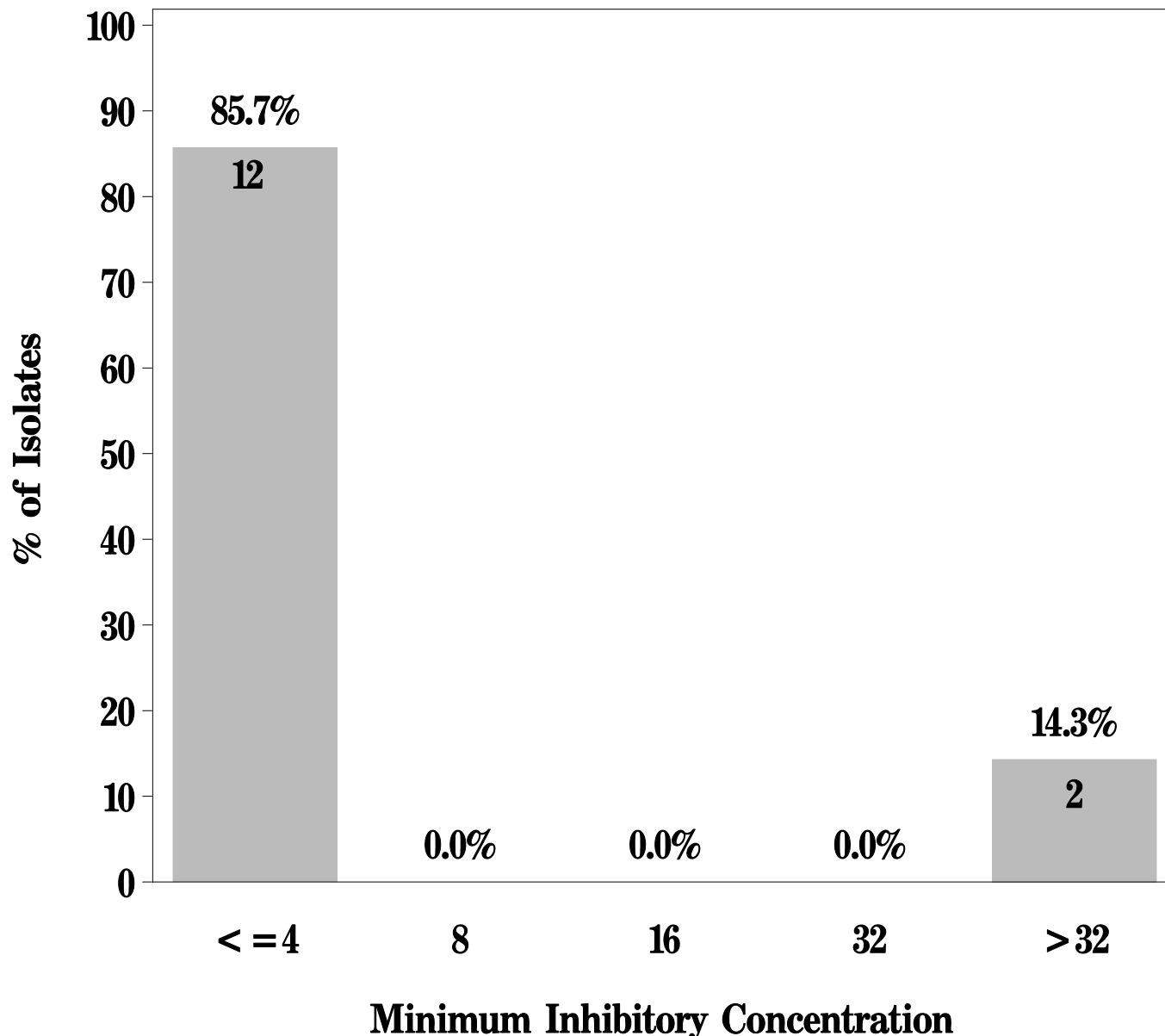
**Figure 7n: Minimum Inhibitory Concentration of Tetracycline
for *Salmonella* in Ground Turkey (N=142 Isolates)**
Breakpoints: Susceptible $\leq 4 \text{ } \mu\text{g/mL}$ Resistant $\geq 16 \text{ } \mu\text{g/mL}$



NARMS

**Figure 7n: Minimum Inhibitory Concentration of Tetracycline
for *Salmonella* in Ground Beef (N=14 Isolates)**

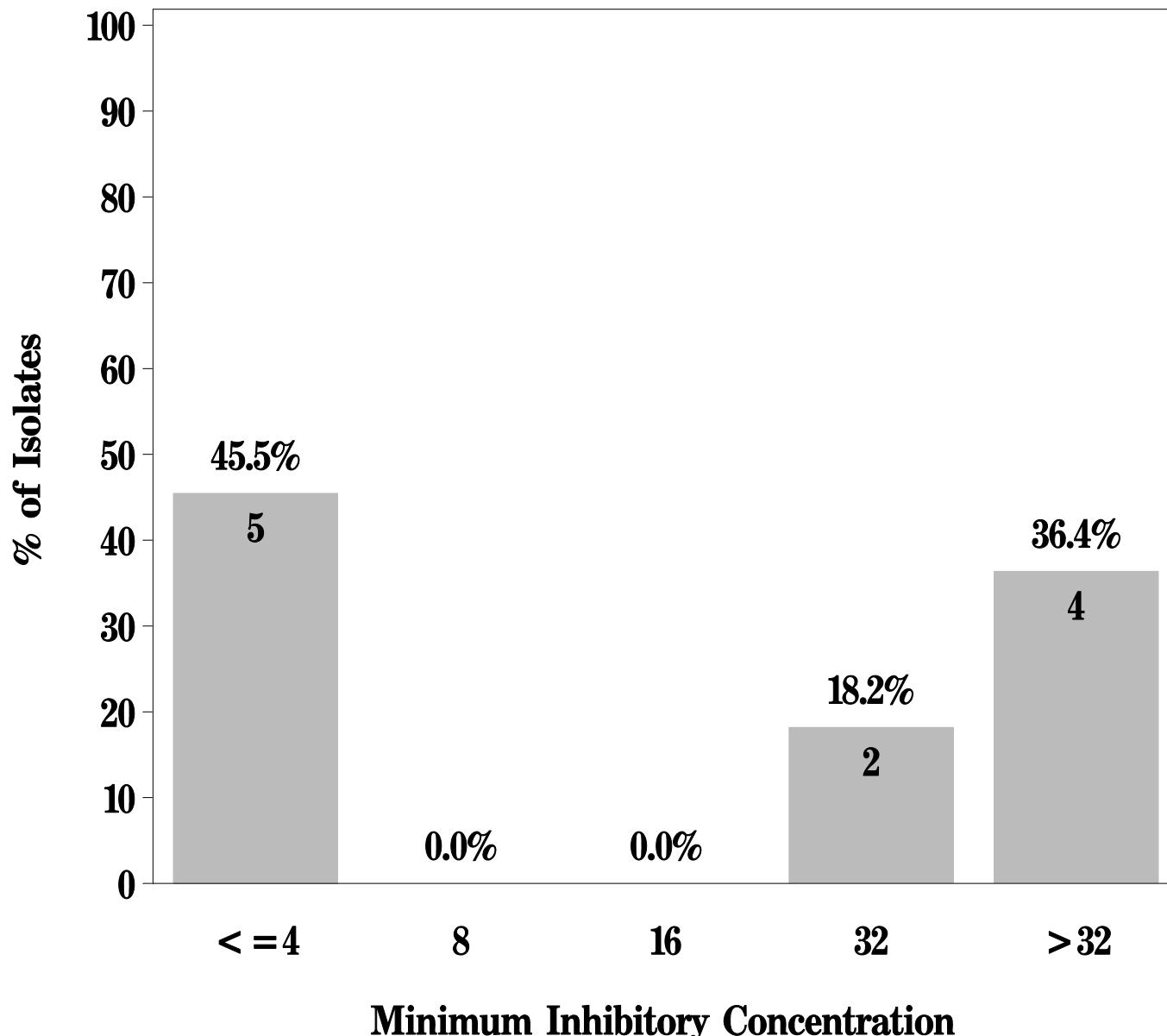
Breakpoints: Susceptible $\leq 4 \text{ } \mu\text{g/mL}$ Resistant $\geq 16 \text{ } \mu\text{g/mL}$



NARMS

**Figure 7n: Minimum Inhibitory Concentration of Tetracycline
for *Salmonella* in Pork Chop (N=11 Isolates)**

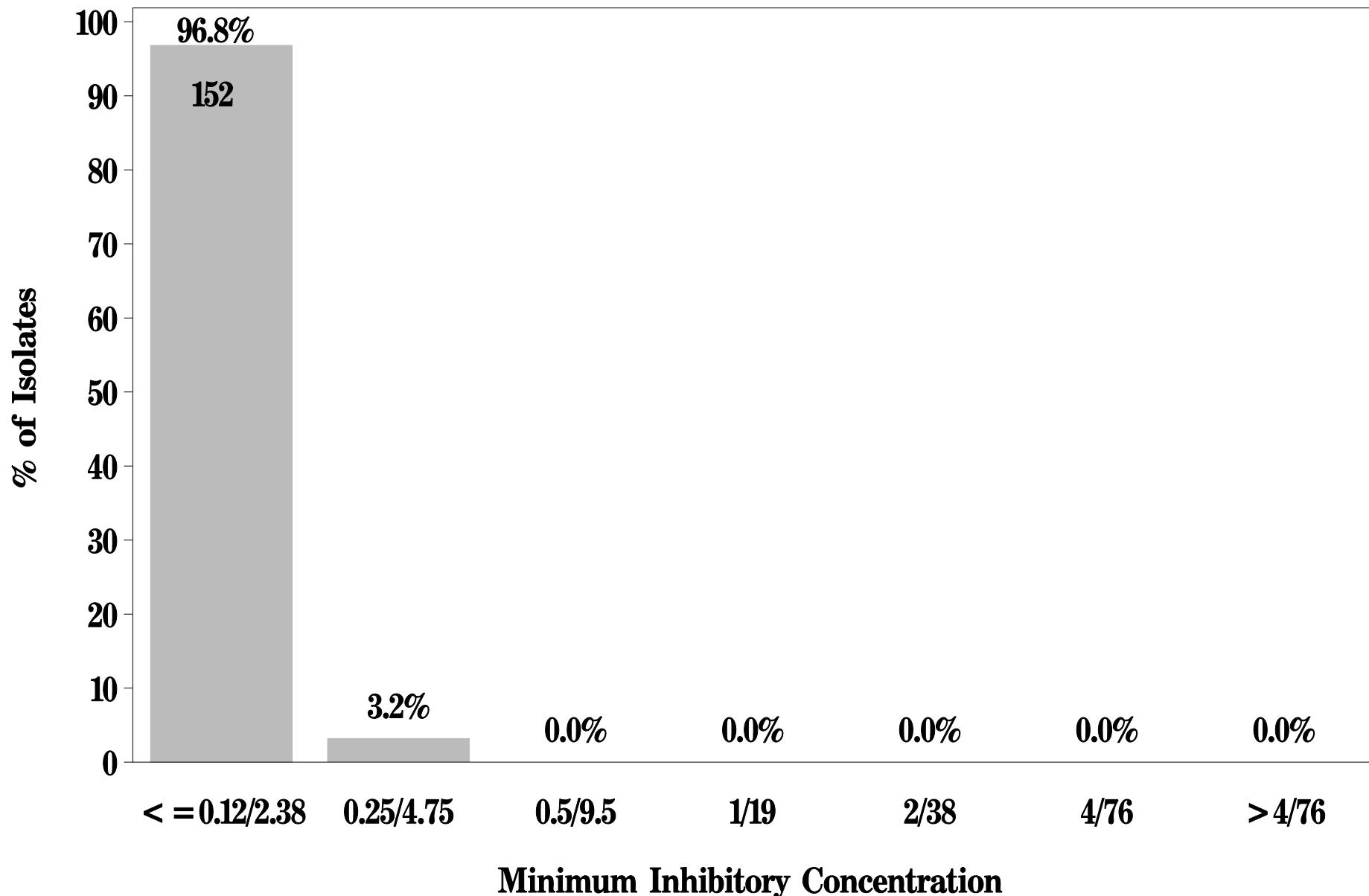
Breakpoints: Susceptible $\leq 4 \text{ } \mu\text{g/mL}$ Resistant $\geq 16 \text{ } \mu\text{g/mL}$



NARMS

Figure 7o: Minimum Inhibitory Concentration of Trimethoprim/sulfamethoxazole for *Salmonella* in Chicken Breast (N=157 Isolates)

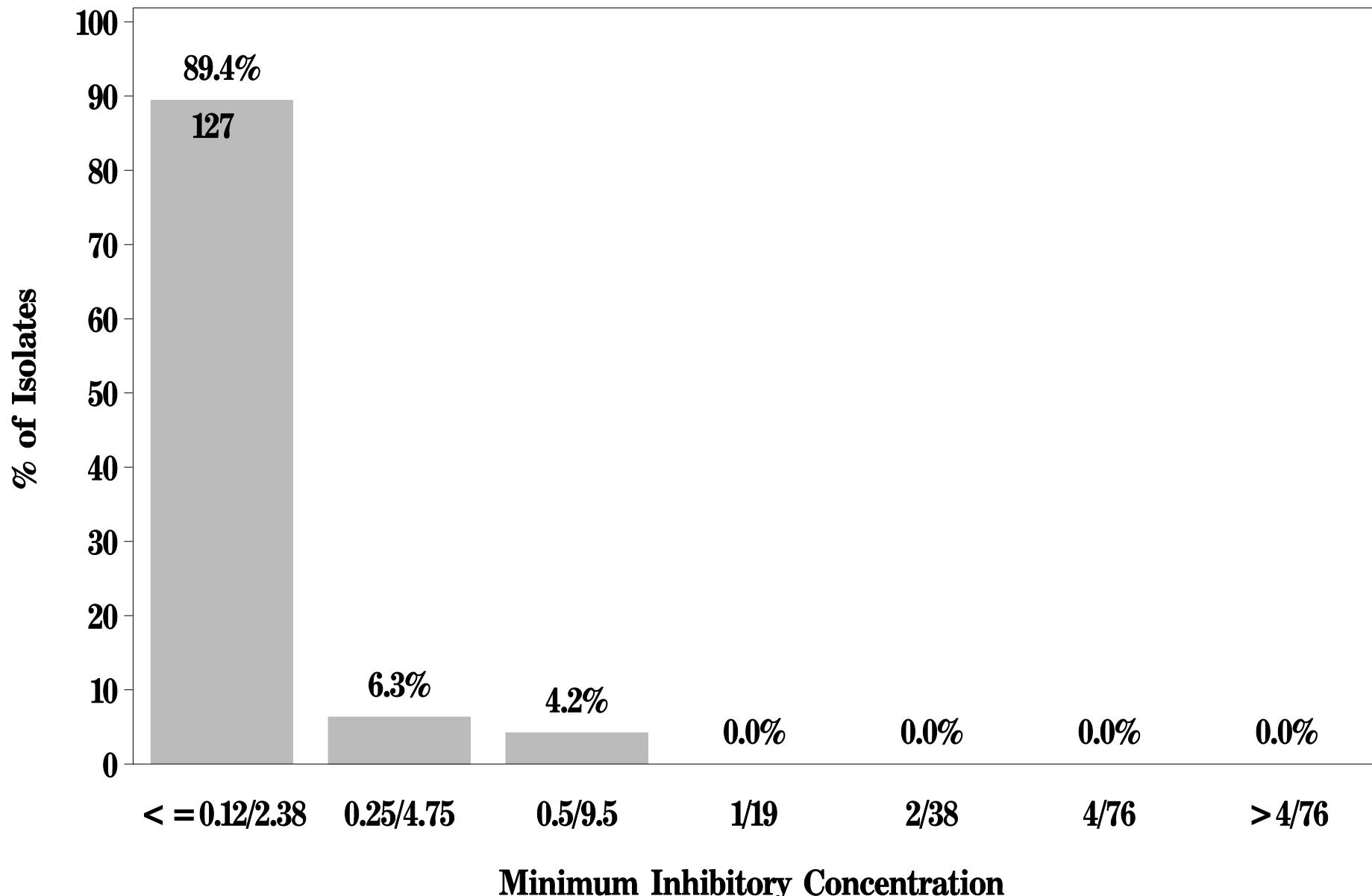
Breakpoints: Susceptible $\leq 2 \mu\text{g/mL}$ Resistant $\geq 4 \mu\text{g/mL}$



NARMS

Figure 7o: Minimum Inhibitory Concentration of Trimethoprim/sulfamethoxazole for *Salmonella* in Ground Turkey (N=142 Isolates)

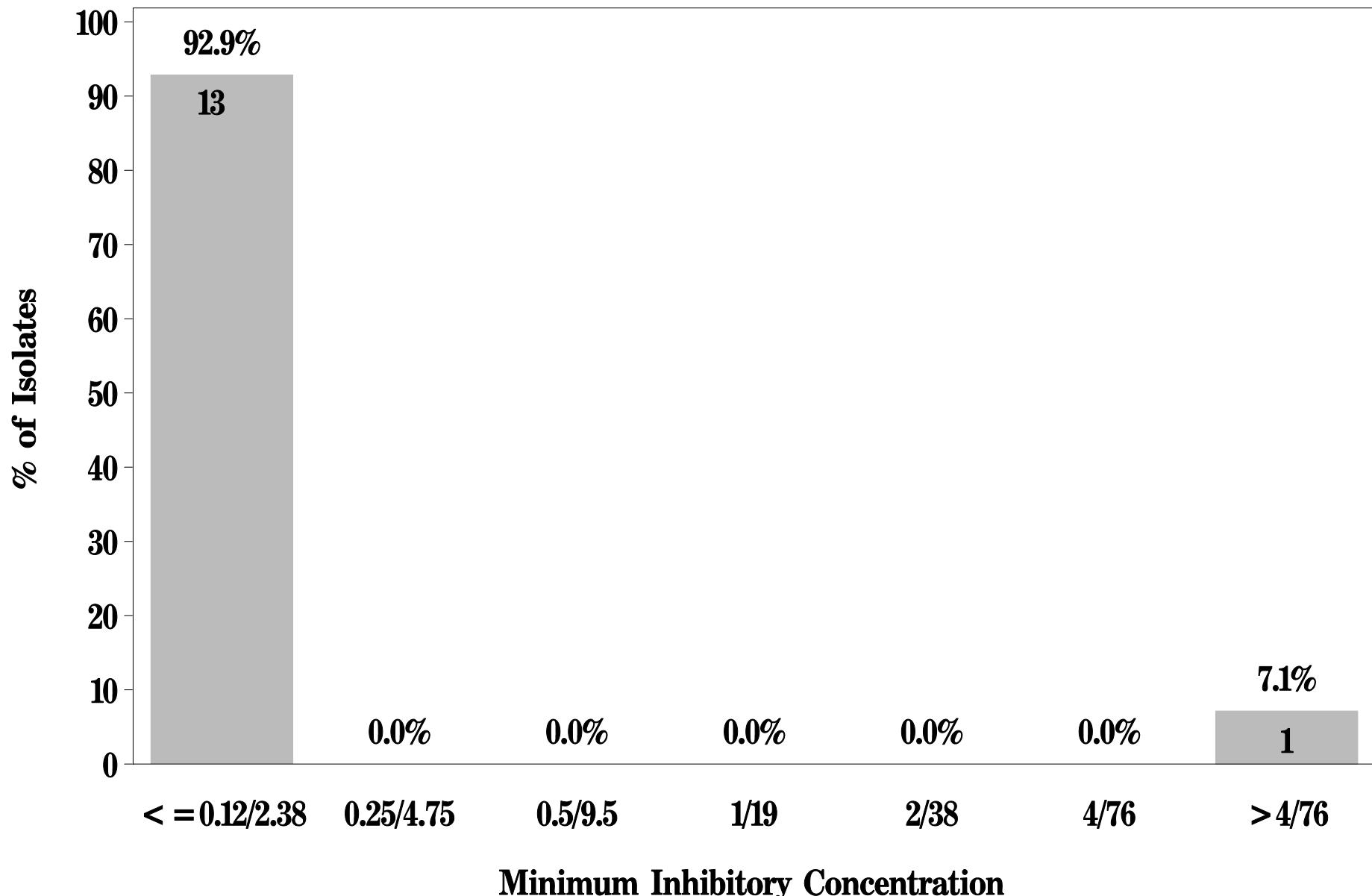
Breakpoints: Susceptible $\leq 2 \mu\text{g/mL}$ Resistant $\geq 4 \mu\text{g/mL}$



NARMS

Figure 7o: Minimum Inhibitory Concentration of Trimethoprim/sulfamethoxazole for *Salmonella* in Ground Beef (N=14 Isolates)

Breakpoints: Susceptible $\leq 2 \mu\text{g/mL}$ Resistant $\geq 4 \mu\text{g/mL}$



NARMS

Figure 7o: Minimum Inhibitory Concentration of Trimethoprim/sulfamethoxazole for *Salmonella* in Pork Chop (N=11 Isolates)

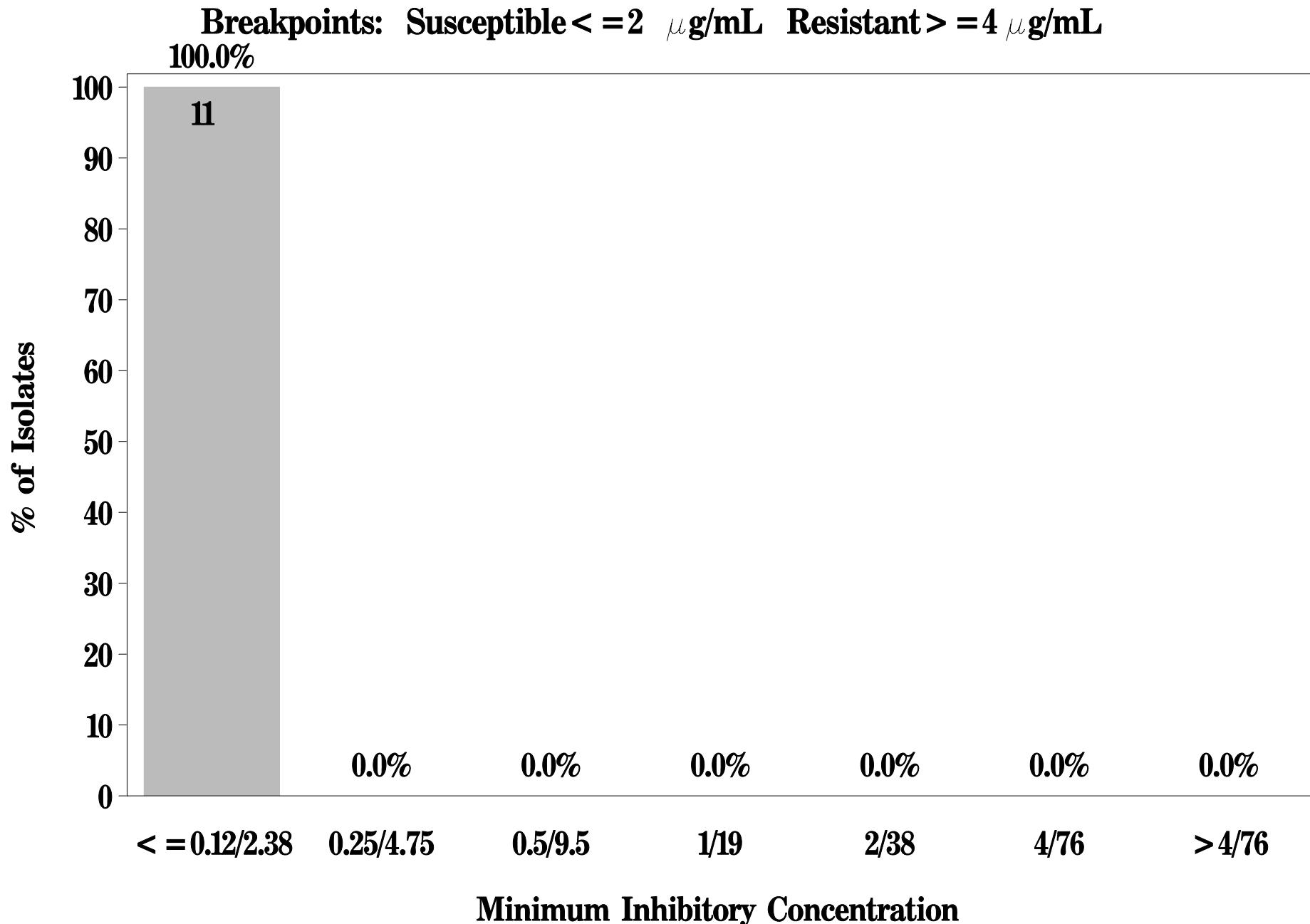


Table 10. Antimicrobial Resistance among *Salmonella* Isolates by Meat Type, 2004

Antimicrobial Agent	Chicken Breast (n=157)	Ground Turkey (n=142)	Ground Beef (n=14)	Pork Chop (n=11)
Tetracycline	46.5%*	56.3%	14.3%	54.5%
Streptomycin	28.0%	33.8%	14.3%	27.3%
Sulfisoxazole	28.7%	28.2%	14.3%	18.2%
Ampicillin	30.6%	20.4%	21.4%	9.1%
Amoxicillin/Clavulanic Acid	24.8%	7.7%	14.3%	-†
Cefoxitin	24.8%	4.9%	14.3%	-
Ceftiofur	24.8%	4.9%	14.3%	-
Kanamycin	11.5%	18.3%	-	9.1%
Gentamicin	3.8%	20.4%	-	-
Chloramphenicol	1.9%	2.8%	14.3%	18.2%
Ceftriaxone	-	-	7.1%	-
Trimethoprim/Sulfamethoxazole	-	-	7.1%	-
Amikacin	-	-	-	-
Ciprofloxacin	-	-	-	-
Nalidixic Acid	-	-	-	-

* Where % Resistance = (# isolates per meat type resistant to antimicrobial) / (total # isolates per meat type).

† Dashes indicate 0.0% resistance to antimicrobial.

Table 11. Antimicrobial Resistance among *Salmonella* Isolates by Serotype, 2004

Serotype	Antimicrobial Agent														
	TET	STR	FIS	AMP	AMC	FOX	TIO	KAN	GEN	CHL	AXO	COT	AMI	CIP	NAL
S. Heidelberg (n=71)	43.7%*	33.8%	25.4%	18.3%	7.0%	7.0%	7.0%	15.5%	22.5%	4.2%	-	-	-	-	-
S. Typhimurium† (n=53)	73.6%	18.9%	75.5%	52.8%	45.3%	45.3%	45.3%	34.0%	1.9%	9.4%	-	-	-	-	-
S. Kentucky (n=43)	53.5%	51.2%	4.7%	27.9%	25.6%	25.6%	25.6%	2.3%	2.3%	-	-	-	-	-	-
S. Saintpaul (n=24)	58.3%	54.2%	54.2%	50.0%	16.7%	4.2%	4.2%	45.8%	37.5%	4.2%	-	-	-	-	-
S. Schwarzengrund (n=21)	28.6%	-‡	14.3%	4.8%	-	-	-	-	4.8%	-	-	-	-	-	-
S. Hadar (n=19)	94.7%	89.5%	-	5.3%	-	-	-	-	-	-	-	-	-	-	-
S. Reading (n=16)	6.3%	6.3%	6.3%	-	-	-	-	-	6.3%	-	-	-	-	-	-
S. Braenderup (n=11)	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
S. Muenster (n=10)	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
S. Agona (n=9)	88.9%	44.4%	55.6%	44.4%	22.2%	11.1%	11.1%	22.2%	11.1%	-	-	-	-	-	-
S. III 18a: z4, z32: - (n=6)	83.3%	-	-	-	-	-	-	-	-	-	-	-	-	-	-
S. Berta (n=5)	-	-	-	60.0%	-	-	-	-	-	-	-	-	-	-	-
S. Montevideo (n=5)	-	40.0%	-	-	-	-	-	20.0%	60.0%	-	-	-	-	-	-
S. Mbandaka (n=4)	100.0%	-	25.0%	25.0%	-	-	-	-	-	-	-	-	-	-	-
S. Newport (n=4)	50.0%	50.0%	50.0%	50.0%	50.0%	50.0%	50.0%	-	-	50.0%	25.0%	25.0%	-	-	-
S. I 4, 12 : i : - (n=4)	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
S. Derby (n=3)	100.0%	-	-	66.7%	66.7%	66.7%	66.7%	-	-	-	-	-	-	-	-
S. Enteritidis (n=3)	33.3%	-	33.3%	33.3%	33.3%	33.3%	33.3%	-	-	-	-	-	-	-	-
S. III 18a: z4, z23: - (n=2)	-	50.0%	50.0%	-	-	-	-	-	50.0%	-	-	-	-	-	-
S. I 4, 12 : r : - (n=2)	100.0%	-	-	-	-	-	-	-	-	-	-	-	-	-	-
S. Senftenberg (n=2)	50.0%	-	-	-	-	-	-	-	-	-	-	-	-	-	-
S. Bredeney (n=1)	100.0%	100.0%	100.0%	100.0%	100.0%	100.0%	100.0%	-	100.0%	-	-	-	-	-	-
S. Dublin (n=1)	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
S. Livingstone (n=1)	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
S. Minnesota (n=1)	100.0%	100.0%	100.0%	-	-	-	-	-	100.0%	-	-	-	-	-	-
S. Muenchen (n=1)	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
S. I 4, 12 : d : - (n=1)	100.0%	-	-	-	-	-	-	-	-	-	-	-	-	-	-
S. Urbana (n=1)	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
• Total (N=324)	49.7%	30.2%	27.5%	25.0%	16.0%	14.8%	14.8%	13.9%	10.8%	3.4%	0.3%	0.3%	0.0%	0.0%	0.0%

* Where % Resistance = (# isolates per serotype resistant to antimicrobial) / (total # isolates per serotype).

† Includes Typhimurium var. 5-.

‡ Where dashes indicate 0.0% resistance to antimicrobial.

Table 12. Antimicrobial Resistance among *Salmonella* by Meat Type in Overall Top 6 Serotypes, 2004

Meat Type	Serotype	Antimicrobial Agent														
		TET	STR	FIS	AMP	AMC	FOX	TIO	KAN	GEN	CHL	AXO	COT	AMI	CIP	NAL
Chicken Breast	<i>S. Heidelberg</i> (n=31)	6.5%*	22.6%	12.9%	25.8%	9.7%	9.7%	9.7%	-	9.7%	3.2%	-	-	-	-	-
	<i>S. Typhimurium</i> [†] (n=49)	71.4%	14.3%	73.5%	53.1%	49.0%	49.0%	49.0%	34.7%	2.0%	4.1%	-	-	-	-	-
	<i>S. Kentucky</i> (n=42)	54.8%	52.4%	4.8%	28.6%	26.2%	26.2%	26.2%	2.4%	2.4%	-	-	-	-	-	-
	<i>S. Saintpaul</i> (n=0)	‡														
	<i>S. Schwarzengrund</i> (n=5)	-	-	20.0%	-	-	-	-	-	-	-	-	-	-	-	-
	<i>S. Hadar</i> (n=8)	87.5%	87.5%	-	-	-	-	-	-	-	-	-	-	-	-	-
Ground Turkey	<i>S. Heidelberg</i> (n=37)	70.3%	43.2%	37.8%	13.5%	5.4%	5.4%	5.4%	27.0%	35.1%	5.4%	-	-	-	-	-
	<i>S. Typhimurium</i> (n=2)	100.0%	50.0%	100.0%	50.0%	-	-	-	50.0%	-	50.0%	-	-	-	-	-
	<i>S. Kentucky</i> (n=1)	§	-	-	-	-	-	-	-	-	-	-	-	-	-	-
	<i>S. Saintpaul</i> (n=24)	58.3%	54.2%	54.2%	50.0%	16.7%	4.2%	4.2%	45.8%	37.5%	4.2%	-	-	-	-	-
	<i>S. Schwarzengrund</i> (n=16)	37.5%	-	12.5%	6.3%	-	-	-	-	6.3%	-	-	-	-	-	-
	<i>S. Hadar</i> (n=11)	100.0%	90.9	-	9.1%	-	-	-	-	-	-	-	-	-	-	-
Ground Beef	<i>S. Heidelberg</i> (n=0)															
	<i>S. Typhimurium</i> (n=0)															
	<i>S. Kentucky</i> (n=0)															
	<i>S. Saintpaul</i> (n=0)															
	<i>S. Schwarzengrund</i> (n=0)															
	<i>S. Hadar</i> (n=0)															
Pork Chop	<i>S. Heidelberg</i> (n=3)	100.0%	33.3%	-	-	-	-	-	33.3%	-	-	-	-	-	-	-
	<i>S. Typhimurium</i> (n=2)	100.0%	100.0%	100.0%	50.0%	-	-	-	-	-	100.0%	-	-	-	-	-
	<i>S. Kentucky</i> (n=0)															
	<i>S. Saintpaul</i> (n=0)															
	<i>S. Schwarzengrund</i> (n=0)															
	<i>S. Hadar</i> (n=0)															

* Where % Resistance = (# isolates per serotype resistant to antimicrobial) / (total # isolates per serotype).

† Includes Typhimurium var. 5-.

‡ Grey areas indicate serotype not isolated from that meat type.

§ Where dashes indicate 0.0% resistance to antimicrobial.

Table 13. Antimicrobial Resistance among *Salmonella* by Top 6 Serotypes within Meat Type, 2004

Meat Type	Serotype	Antimicrobial Agent														
		TET	STR	FIS	AMP	AMC	FOX	TIO	KAN	GEN	CHL	AXO	COT	AMI	CIP	NAL
Chicken Breast	<i>S. Typhimurium</i> * (n=49)	71.4% [†]	14.3%	73.5%	53.1%	49.0%	49.0%	49.0%	34.7%	2.0%	4.1%	-	-	-	-	-
	<i>S. Kentucky</i> (n=42)	54.8%	52.4%	4.8%	28.6%	26.2%	26.2%	26.2%	2.4%	2.4%	-	-	-	-	-	-
	<i>S. Heidelberg</i> (n=31)	6.5%	22.6%	12.9%	25.8%	9.7%	9.7%	9.7%	-	9.7%	3.2%	-	-	-	-	-
	<i>S. Hadar</i> (n=8)	87.5%	87.5%	-	-	-	-	-	-	-	-	-	-	-	-	-
	<i>S. Schwarzengrund</i> (n=5)	- [‡]	-	20.0%	-	-	-	-	-	-	-	-	-	-	-	-
	<i>S. I 4, 12 : i : -</i> (n=4)	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
Ground Turkey	<i>S. Heidelberg</i> (n=37)	70.3%	43.2%	37.8%	13.5%	5.4%	5.4%	5.4%	27.0%	35.1%	5.4%	-	-	-	-	-
	<i>S. Saintpaul</i> (n=24)	58.3%	54.2%	54.2%	50.0%	16.7%	4.2%	4.2%	45.8%	37.5%	4.2%	-	-	-	-	-
	<i>S. Schwarzengrund</i> (n=16)	37.5%	-	12.5%	6.3%	-	-	-	-	6.3%	-	-	-	-	-	-
	<i>S. Reading</i> (n=16)	6.3%	6.3%	6.3%	-	-	-	-	-	6.3%	-	-	-	-	-	-
	<i>S. Hadar</i> (n=11)	100.0%	90.9%	-	9.1%	-	-	-	-	-	-	-	-	-	-	-
	<i>S. Agona</i> (n=6)	100.0%	66.7%	83.3%	66.7%	33.3%	16.73%	16.7%	33.3%	16.7%	-	-	-	-	-	-
Ground Beef	<i>S. Muenster</i> (n=5)	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
	<i>S. Braenderup</i> (n=5)	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
	<i>S. Newport</i> (n=2)	100.0%	100.0%	100.0%	100.0%	100.0%	100.0%	100.0%	-	-	100.0%	50.0%	50.0%	-	-	-
	<i>S. Berta</i> (n=1)	-	-	-	100.0%	-	-	-	-	-	-	-	-	-	-	-
	<i>S. Dublin</i> (n=1)	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
	§															
Pork Chop	<i>S. Braenderup</i> (n=5)	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
	<i>S. Heidelberg</i> (n=3)	100.0%	33.3%	-	-	-	-	-	33.3%	-	-	-	-	-	-	-
	<i>S. Typhimurium</i> (n=2)	100.0%	100.0%	100.0%	50.0%	-	-	-	-	-	100.0%	-	-	-	-	-
	<i>S. Agona</i> (n=1)	100.0%	-	-	-	-	-	-	-	-	-	-	-	-	-	-

* Includes Typhimurium var. 5-.

† Where % Resistance = (# isolates per serotype resistant to antimicrobial) / (total # isolates per serotype).

‡ Where dashes indicate 0.0% resistance to antimicrobial.

§ Grey areas indicate six serotypes not recovered from meat type.

Table 14. Antimicrobial Resistance among *Salmonella* by Site, Meat Type, and Antimicrobial Agent, 2004

Site	Meat Type	Antimicrobial Agent														
		TET	STR	FIS	AMP	AMC	FOX	TIO	KAN	GEN	CHL	AXO	COT	AMI	CIP	NAL
CA	CB (n=17)	11.8%*	11.8%	11.8%	5.9%	-	-	-	-	11.8%	-	-	-	-	-	-
	GT (n=9)	44.4%	55.6%	44.4%	22.2%	-	-	-	-	11.1%	22.2%	11.1%	-	-	-	-
	GB (n=1)	-†	-	-	-	-	-	-	-	-	-	-	-	-	-	-
	PC (n=1)	100.0%	100.0%	100.0%	-	-	-	-	-	-	100.0%	-	-	-	-	-
Total (n=28)		25.0%	28.6%	25.0%	10.7%	0.0%	0.0%	0.0%	3.6%	14.3%	7.1%	0.0%	0.0%	0.0%	0.0%	0.0%
CO	CB (n=0)	‡	-	-	-	-	-	-	-	-	-	-	-	-	-	-
	GT (n=9)	55.6%	33.3%	33.3%	11.1%	-	-	-	-	11.1%	22.2%	-	-	-	-	-
	GB (n=0)	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
	PC (n=0)	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
Total (n=28)		55.6%	33.3%	33.3%	11.1%	0.0%	0.0%	0.0%	11.1%	22.2%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%
CT	CB (n=30)	83.3%	36.7%	70.0%	53.3%	46.7%	46.7%	46.7%	23.3%	3.3%	6.7%	-	-	-	-	-
	GT (n=26)	61.5%	15.4%	23.1%	15.4%	11.5%	3.8%	3.8%	15.4%	15.4%	-	-	-	-	-	-
	GB (n=5)	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
	PC (n=5)	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
Total (n=66)		62.1%	22.7%	40.9%	30.3%	25.8%	22.7%	22.7%	16.7%	7.6%	3.0%	0.0%	0.0%	0.0%	0.0%	0.0%
GA	CB (n=6)	50.0%	33.3%	33.3%	33.3%	-	-	-	-	16.7%	-	-	-	-	-	-
	GT (n=38)	57.9%	42.1%	36.8%	18.4%	5.3%	-	-	31.6%	28.9%	-	-	-	-	-	-
	GB (n=1)	100.0%	100.0%	100.0%	100.0%	100.0%	100.0%	100.0%	-	-	100.0%	100.0%	-	-	-	-
	PC (n=0)	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
Total (n=45)		57.8%	42.2%	37.8%	22.2%	6.7%	2.2%	2.2%	26.7%	26.7%	2.2%	2.2%	0.0%	0.0%	0.0%	0.0%
MD	CB (n=24)	62.5%	20.8%	50.0%	45.8%	45.8%	45.8%	45.8%	25.0%	4.2%	-	-	-	-	-	-
	GT (n=13)	23.1%	-	7.7%	38.5%	15.4%	15.4%	15.4%	-	7.7%	-	-	-	-	-	-
	GB (n=1)	-	-	-	100.0%	-	-	-	-	-	-	-	-	-	-	-
	PC (n=0)	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
Total (n=38)		47.4%	13.2%	34.2%	44.7%	34.2%	34.2%	34.2%	15.8%	5.3%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%

* Where % Resistance = (# isolates resistant to antimicrobial per meat type per site) / (total # isolates per meat type per site).

† Where dashes indicate 0.0% resistance to antimicrobial.

‡ Grey areas indicate no isolates from meat type for that site.

Table 14_(cont'd). Percent Resistance among *Salmonella* Isolates by Site, Meat Type, and Antimicrobial Agent, 2004

Site	Meat Type	Antimicrobial Agent														
		TET	STR	FIS	AMP	AMC	FOX	TIO	KAN	GEN	CHL	AXO	COT	AMI	CIP	NAL
MN	CB (n=20)	15.0%	5.0%	-	25.0%	5.0%	5.0%	5.0%	-	-	-	-	-	-	-	-
	GT (n=14)	50.0%	35.7%	14.3%	-	-	-	-	7.1%	14.3%	-	-	-	-	-	-
	GB (n=0)															
	PC (n=0)															
Total (n=34)		29.4%	17.6%	5.9%	14.7%	2.9%	2.9%	2.9%	2.9%	5.9%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%
NM	CB (n=3)	33.3%	66.7%	33.3%	-	-	-	-	-	-	-	-	-	-	-	-
	GT (n=9)	44.4%	33.3%	33.3%	33.3%	33.3%	33.3%	33.3%	22.2%	33.3%	33.3%	-	-	-	-	-
	GB (n=0)															
	PC (n=0)															
Total (n=12)		41.7%	41.7%	33.3%	25.0%	25.0%	25.0%	25.0%	16.7%	25.0%	25.0%	0.0%	0.0%	0.0%	0.0%	0.0%
NY	CB (n=16)	93.8%	56.3%	31.3%	68.8%	68.8%	68.8%	68.8%	18.8%	-	-	-	-	-	-	-
	GT (n=11)	72.7%	54.5%	36.4%	45.5%	9.1%	9.1%	9.1%	36.4%	27.3%	-	-	-	-	-	-
	GB (n=0)															
	PC (n=3)	100.0%	66.7%	33.3%	33.3%	-	-	-	33.3%	-	33.3%	-	-	-	-	-
Total (n=30)		86.7%	56.7%	33.3%	56.7%	40.0%	40.0%	40.0%	26.7%	10.0%	3.3%	0.0%	0.0%	0.0%	0.0%	0.0%
OR	CB (n=25)	28.0%	28.0%	8.0%	8.0%	8.0%	8.0%	8.0%	-	4.0%	4.0%	-	-	-	-	-
	GT (n=6)	66.7%	16.7%	33.3%	33.3%	-	-	-	16.7%	16.7%	-	-	-	-	-	-
	GB (n=6)	16.7%	16.7%	16.7%	16.7%	16.7%	16.7%	16.7%	-	-	16.7%	-	16.7%	-	-	-
	PC (n=2)	100.0%	-	-	-	-	-	-	-	-	-	-	-	-	-	-
Total (n=39)		35.9%	23.1%	12.8%	12.8%	7.7%	7.7%	7.7%	2.6%	5.1%	5.1%	0.0%	2.6%	0.0%	0.0%	0.0%
TN	CB (n=16)	12.5%	37.5%	-	-	-	-	-	12.5%	-	-	-	-	-	-	-
	GT (n=7)	100.0%	71.4%	14.3%	-	-	-	-	-	-	-	-	-	-	-	-
	GB (n=0)															
	PC (n=0)															
Total (n=23)		39.1%	47.8%	4.3%	0.0%	0.0%	0.0%	0.0%	8.7%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%
Total %R (N=324)		49.7%	30.2%	27.5%	25.0%	16.0%	14.8%	14.8%	13.9%	10.8%	3.4%	0.3%	0.3%	0.0%	0.0%	0.0%

Table 15. Number of *Salmonella* (N=324) Resistant to Multiple Antimicrobial Agents, 2004

Meat Type	Number of Antimicrobials				
	0	1	2-4	5-7	<u>≥8</u>
Chicken Breast	63	16	42	33	3
Ground Turkey	41	43	35	19	4
Ground Beef	11	1	0	0	2
Pork Chop	5	3	2	1	0
Total	120	63	79	53	9

Table 16. Overall *Campylobacter* Species Identified, 2004

Species	N
<i>C. coli</i>	204
<i>C. jejuni</i>	517
Total	721

Table 17. *Campylobacter* Species by Meat Type, 2004

Species	Chicken Breast		Ground Turkey		Ground Beef		Pork Chop	
	n	%*	n	%	n	%	n	%
<i>C. coli</i> (n=204)	196	96.1%	5	2.5%	0	- [†]	3	1.5%
<i>C. jejuni</i> (n=517)	510	98.6%	7	1.4%	0	-	0	-
Total (N=721)	706	97.9%	12	1.7%	0	0.0%	3	0.4%

* Where % = (# of isolates per species per meat type) / (total # of isolates per species).

† Dashes indicate no isolates from that species per meat type.

Table 18. *Campylobacter* Species by Site and Meat Type*, 2004

Site	Species	Chicken Breast		Ground Turkey		Pork Chop	
		n	% [†]	n	%	n	%
CA	<i>C. coli</i> (n=13)	12	92.3%	0	-‡	1	7.7%
	<i>C. jejuni</i> (n=84)	84	100.0%	0	-	0	-
	Total (n=97)	96	99.0%	0	-	1	1.0%
CO	<i>C. coli</i> (n=11)	11	100.0%	0	-	0	-
	<i>C. jejuni</i> (n=10)	10	100.0%	0	-	0	-
	Total (n=21)	21	100.0%	0	-	0	-
CT	<i>C. coli</i> (n=17)	16	94.1%	0	-	1	5.9%
	<i>C. jejuni</i> (n=72)	70	97.2%	2	2.8%	0	-
	Total (n=89)	86	96.6%	2	2.2%	1	1.1%
GA	<i>C. coli</i> (n=25)	25	100.0%	0	-	0	-
	<i>C. jejuni</i> (n=37)	36	97.3%	1	2.7%	0	-
	Total (n=62)	61	98.4%	1	1.6%	0	-
MD	<i>C. coli</i> (n=26)	26	100.0%	0	-	0	-
	<i>C. jejuni</i> (n=52)	50	96.2%	2	3.8%	0	-
	Total (n=78)	76	97.4%	2	2.6%	0	-
MN	<i>C. coli</i> (n=18)	13	72.2%	5	27.8%	0	-
	<i>C. jejuni</i> (n=61)	60	98.4%	1	1.6%	0	-
	Total (n=79)	73	92.4%	6	7.6%	0	-
NM	<i>C. coli</i> (n=23)	22	95.7%	0	-	1	4.3%
	<i>C. jejuni</i> (n=31)	31	100.0%	0	-	0	-
	Total (n=54)	53	98.1%	0	-	1	1.9%
NY	<i>C. coli</i> (n=39)	39	100.0%	0	-	0	-
	<i>C. jejuni</i> (n=57)	57	100.0%	0	-	0	-
	Total (n=96)	96	100.0%	0	-	0	-
OR	<i>C. coli</i> (n=5)	5	100.0%	0	-	0	-
	<i>C. jejuni</i> (n=68)	68	100.0%	0	-	0	-
	Total (n=73)	73	100.0%	0	-	0	-
TN	<i>C. coli</i> (n=27)	27	100.0%	0	-	0	-
	<i>C. jejuni</i> (n=45)	44	97.8%	1	2.2%	0	-
	Total (n=72)	71	98.6%	1	1.4%	0	-
Grand Total (N=721)		706	97.9%	12	1.7%	3	0.4%

* No *Campylobacter* recovered from ground beef.

† Where % = (# isolates per species per meat type per site) / (total # isolates per species per site).

‡ Dashes indicate no isolates from that species per meat type isolated from that site.

Table 19. *Campylobacter* Isolates by Month for All Sites, 2004

Month	n	%[*]
January	61	8.5%
February	59	8.2%
March	49	6.8%
April	35	4.9%
May	51	7.1%
June	62	8.6%
July	67	9.3%
August	62	8.6%
September	73	10.1%
October	74	10.3%
November	64	8.9%
December	64	8.9%
Total (N)	721	100.0%

* Where % = (n/N).

Table 20. Antimicrobial Resistance among *Campylobacter* Isolates (N=721), 2004

Antimicrobial Agent	n	%R*
Tetracycline	352	48.8%
Nalidixic Acid	111	15.4%
Ciprofloxacin	111	15.4%
Azithromycin	23	3.2%
Erythromycin	23	3.2%
Telithromycin	18	2.5%
Clindamycin	17	2.4%
Florfenicol	0	0.0%
Gentamicin	0	0.0%

* Where %R = (n/N).

Figure 8. Antimicrobial Resistance among *Campylobacter* isolates (n =721), 2004

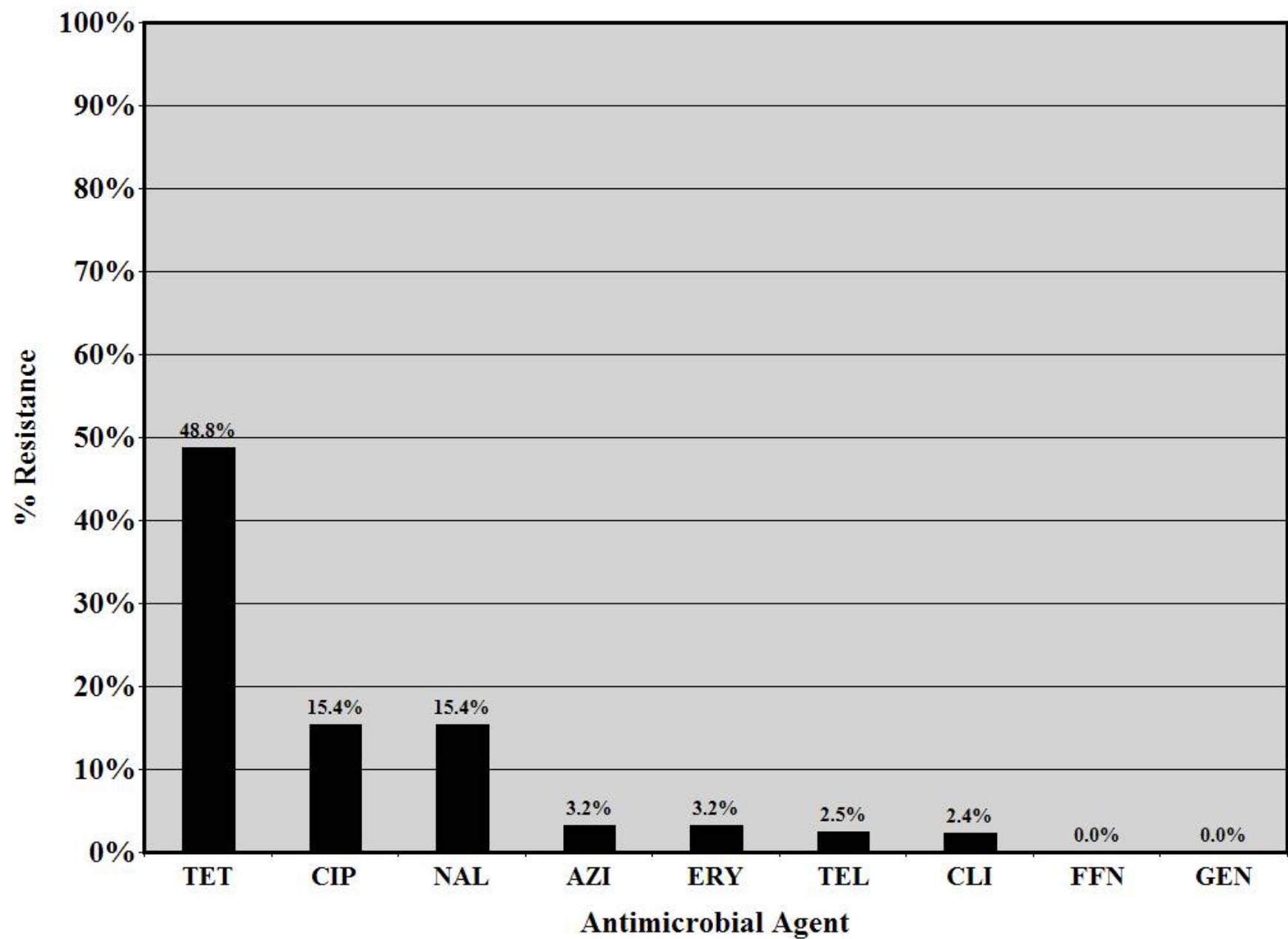


Figure 9. MIC Distribution among all Antimicrobial Agents, 2004

<i>Campylobacter</i> from All Meats (N=721)		Distribution (%) of MICs (in µg/ml)													
Antimicrobial Agent	%R [†]	0.015	0.03	0.06	0.12	0.25	0.5	1	2	4	8	16	32	64	>64
Nalidixic Acid*	15.5%									58.9	25.0	0.6	0.1	1.2	14.1
Ciprofloxacin	15.4%		0.1	34.8	37.7	11.8		0.1		0.3	6.9	6.7	1.5		
Azithromycin*	3.2%	3.7	39.1	39.4	12.6	1.0	0.3	0.6	0.1						3.2
Clindamycin*	2.4%		0.4	7.6	44.9	35.5	5.4	2.8	0.4	0.6	0.8	1.5			
Erythromycin	3.2%			0.3	2.2	43.8	30.0	17.6	2.4	0.4	0.1				3.2
Telithromycin*	2.5%	0.3		0.3	0.4	15.1	41.7	23.3	13.2	2.1	1.1	2.5			
Gentamicin*	0.0%				1.4	4.9	84.6	9.2							
Florfenicol [‡]	§				0.4		4.3	78.8	16.0	0.6					
Tetracycline	48.8%		0.4	19.1	16.9	8.3	4.3	1.8	0.1	0.1	1.5	3.7	19.1	24.4	

[†]Discrepancies between %R and sums of distribution %s are due to rounding.

Vertical bars show the CLSI Susceptible/Resistant breakpoints for each drug.

Unshaded areas indicate the dilution ranges used to test the 2004 isolates.

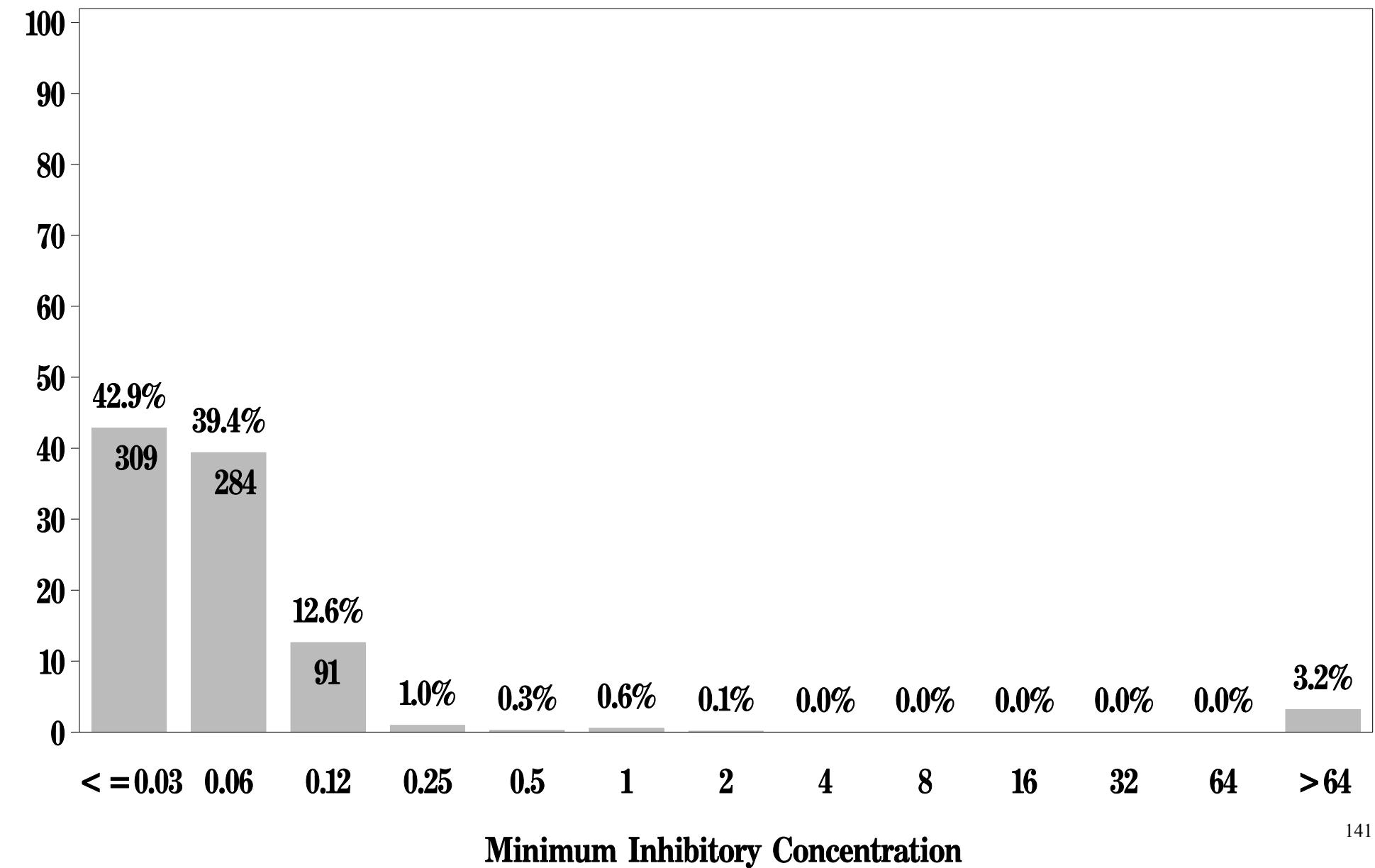
*Currently no CLSI breakpoints have been established for this organism/antimicrobial combination. Indicated breakpoints were established by NARMS.

§Absence of resistant strains precludes defining any results category other than "susceptible."

NARMS

**Figure 9a: Minimum Inhibitory Concentration of Azithromycin
for *Campylobacter* (N = 721 Isolates)**

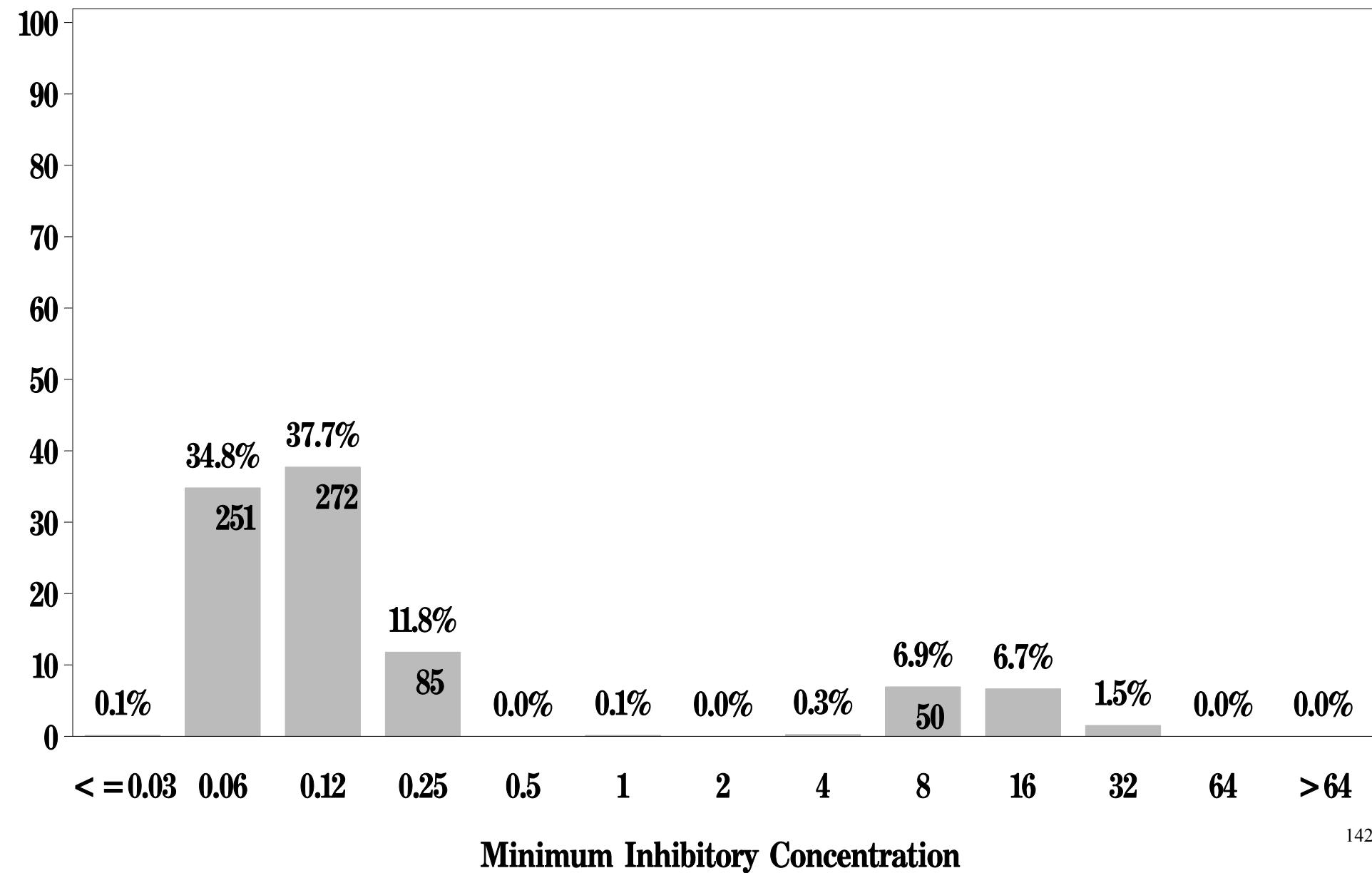
Breakpoints: Susceptible $\leq 2 \text{ } \mu\text{g/mL}$ Resistant $\geq 8 \text{ } \mu\text{g/mL}$



NARMS

**Figure 9b: Minimum Inhibitory Concentration of Ciprofloxacin
for *Campylobacter* (N = 721 Isolates)**

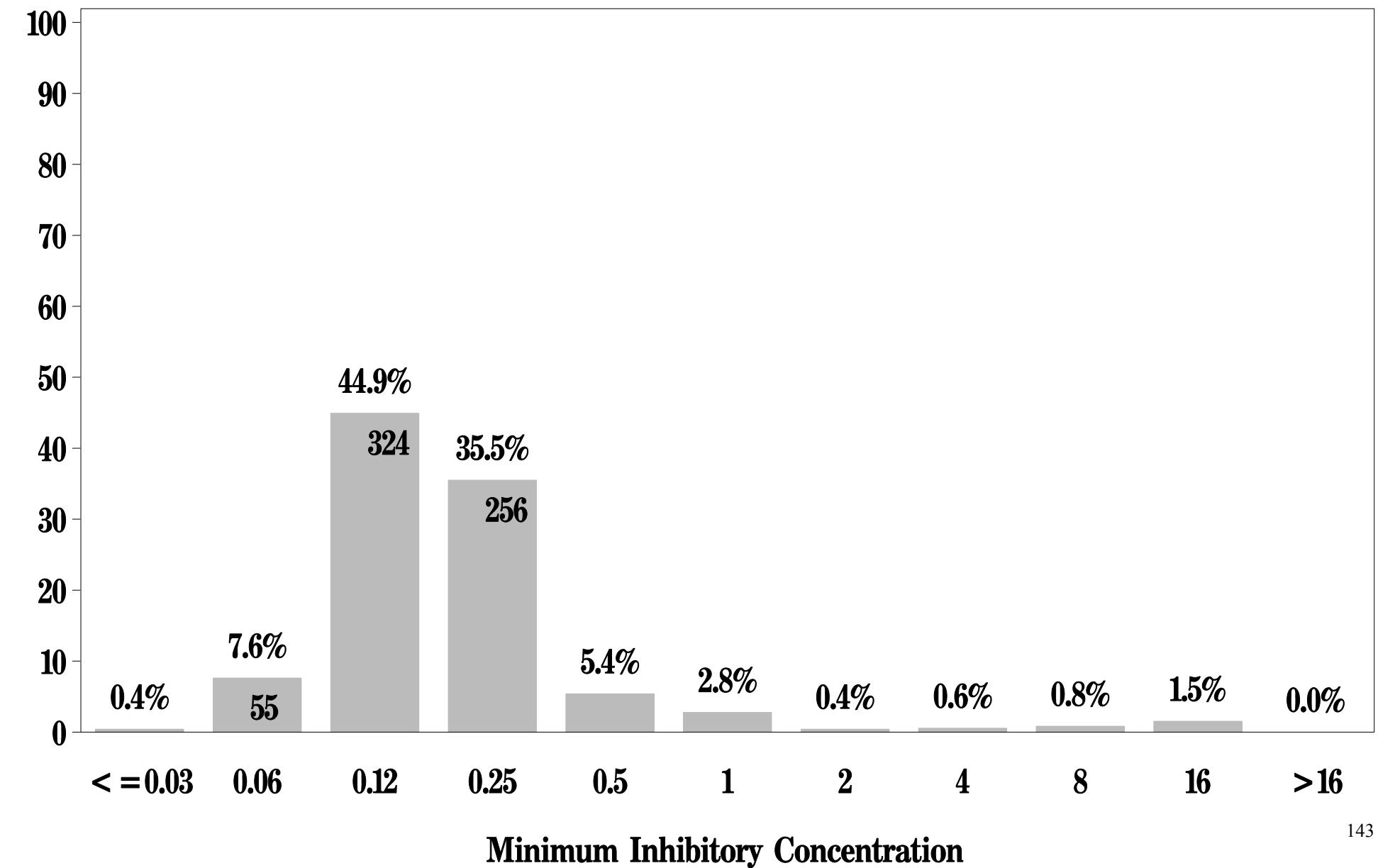
Breakpoints: Susceptible $\leq 1 \mu\text{g/mL}$ Resistant $\geq 4 \mu\text{g/mL}$



NARMS

**Figure 9c: Minimum Inhibitory Concentration of Clindamycin
for *Campylobacter* (N=721 Isolates)**

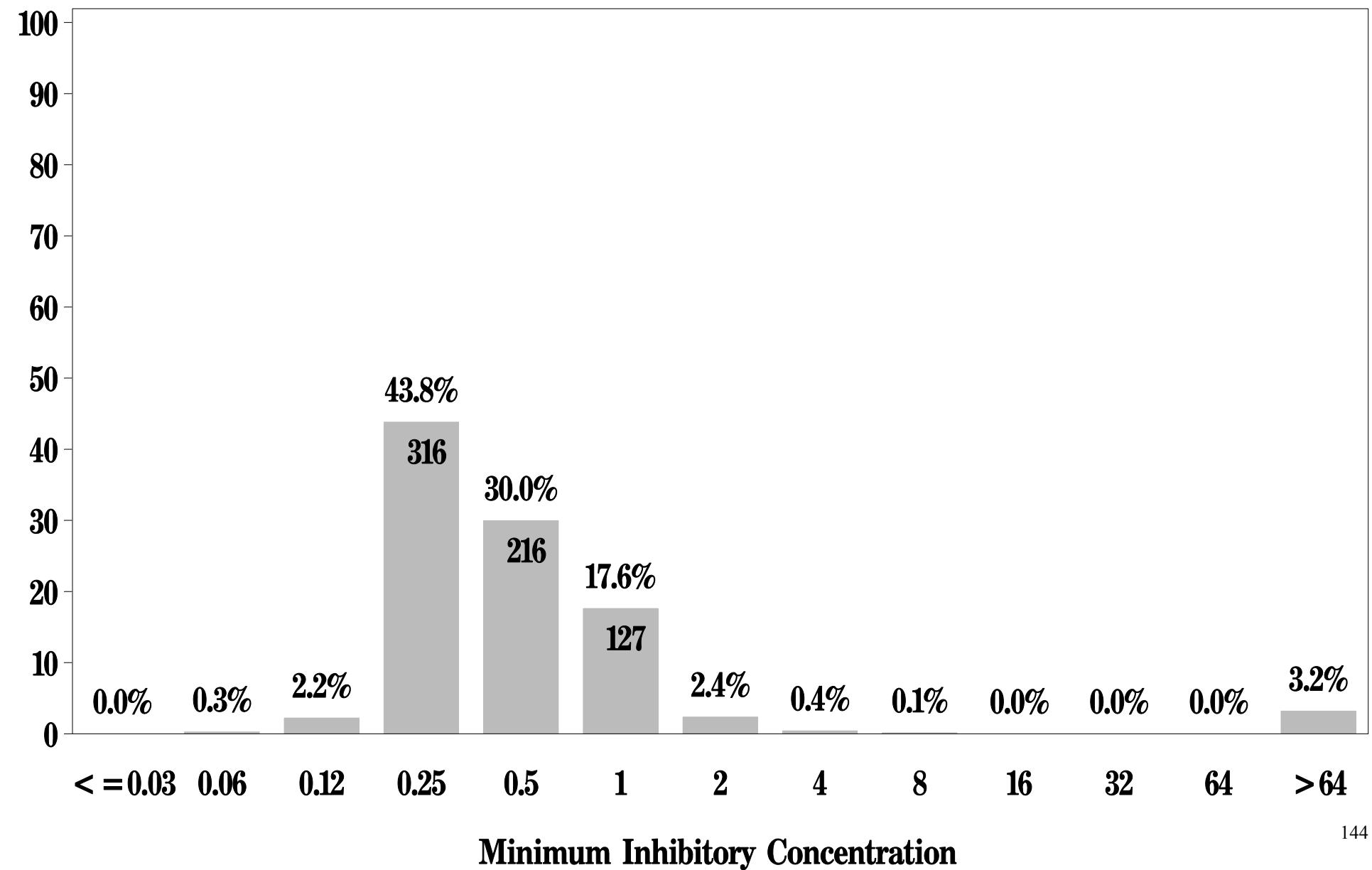
Breakpoints: Susceptible $\leq 2 \text{ } \mu\text{g/mL}$ Resistant $\geq 8 \text{ } \mu\text{g/mL}$



NARMS

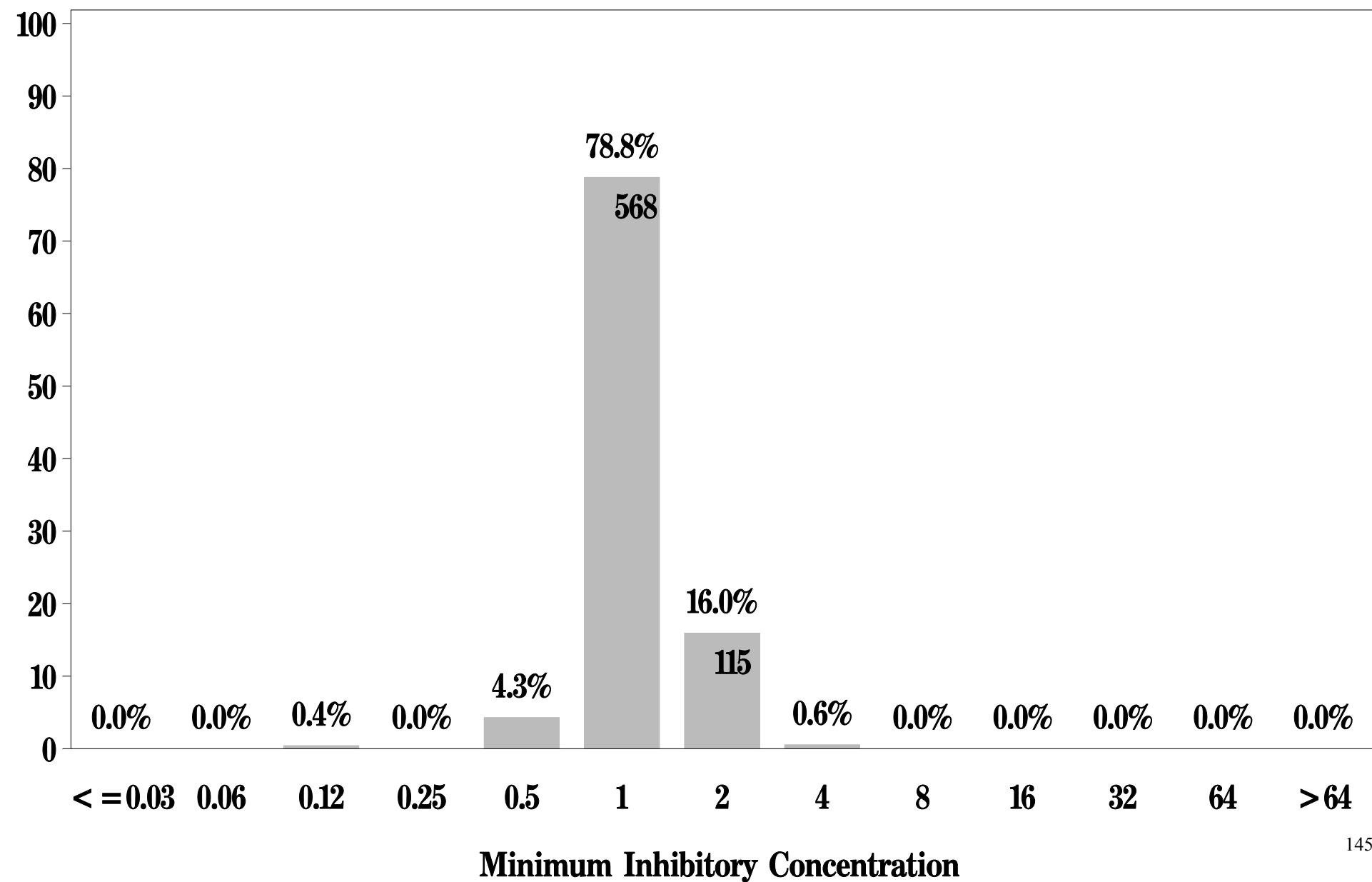
Figure 9d: Minimum Inhibitory Concentration of Erythromycin
for *Campylobacter* (N=721 Isolates)

Breakpoints: Susceptible $\leq 8 \text{ } \mu\text{g/mL}$ Resistant $\geq 32 \text{ } \mu\text{g/mL}$



NARMS

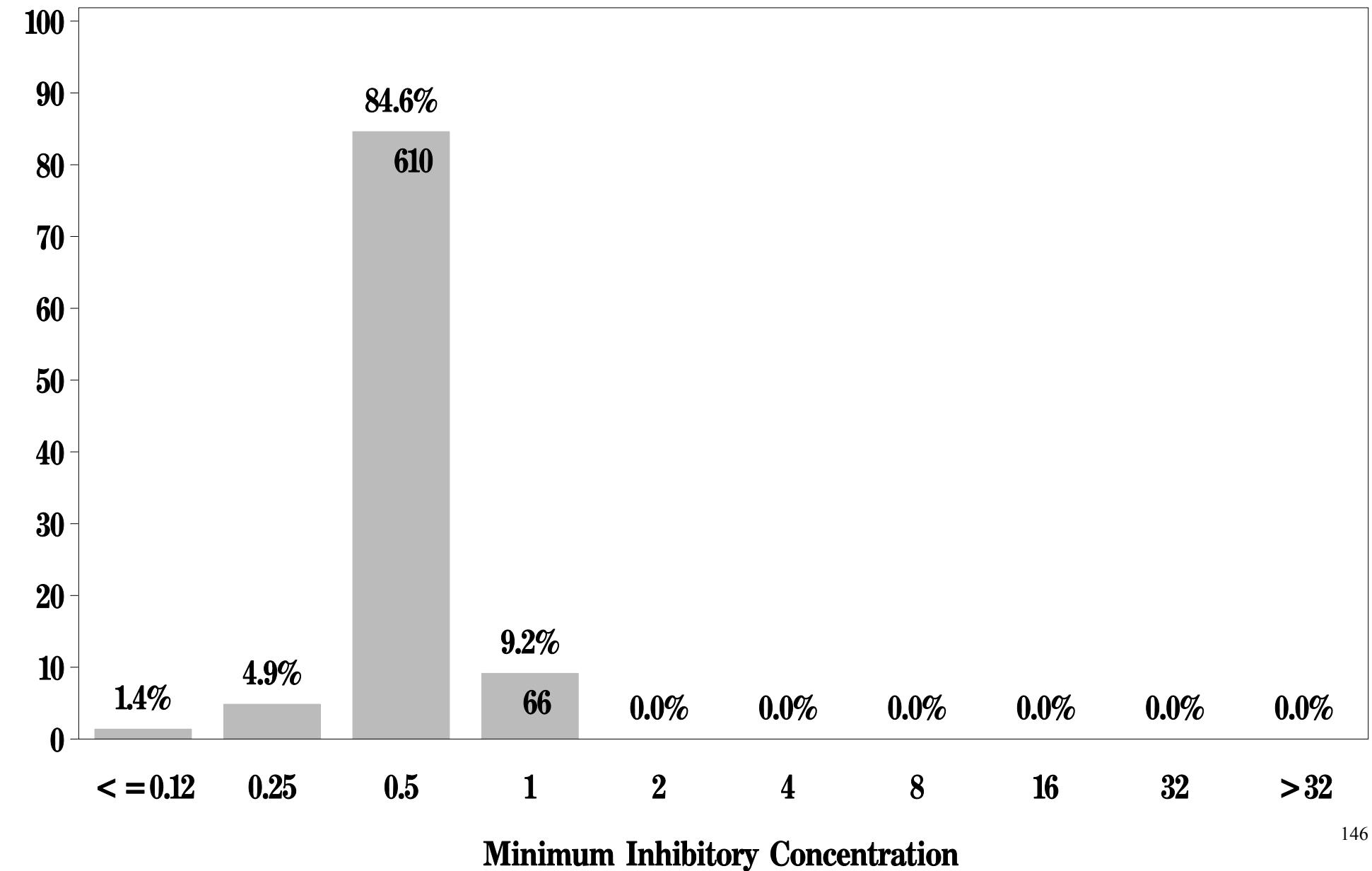
**Figure 9e: Minimum Inhibitory Concentration of Florfenicol
for *Campylobacter* (N=721 Isolates)**
Breakpoint: Susceptible $\leq 4 \mu\text{g/mL}$



NARMS

Figure 9f: Minimum Inhibitory Concentration of Gentamicin
for *Campylobacter* (N=721 Isolates)

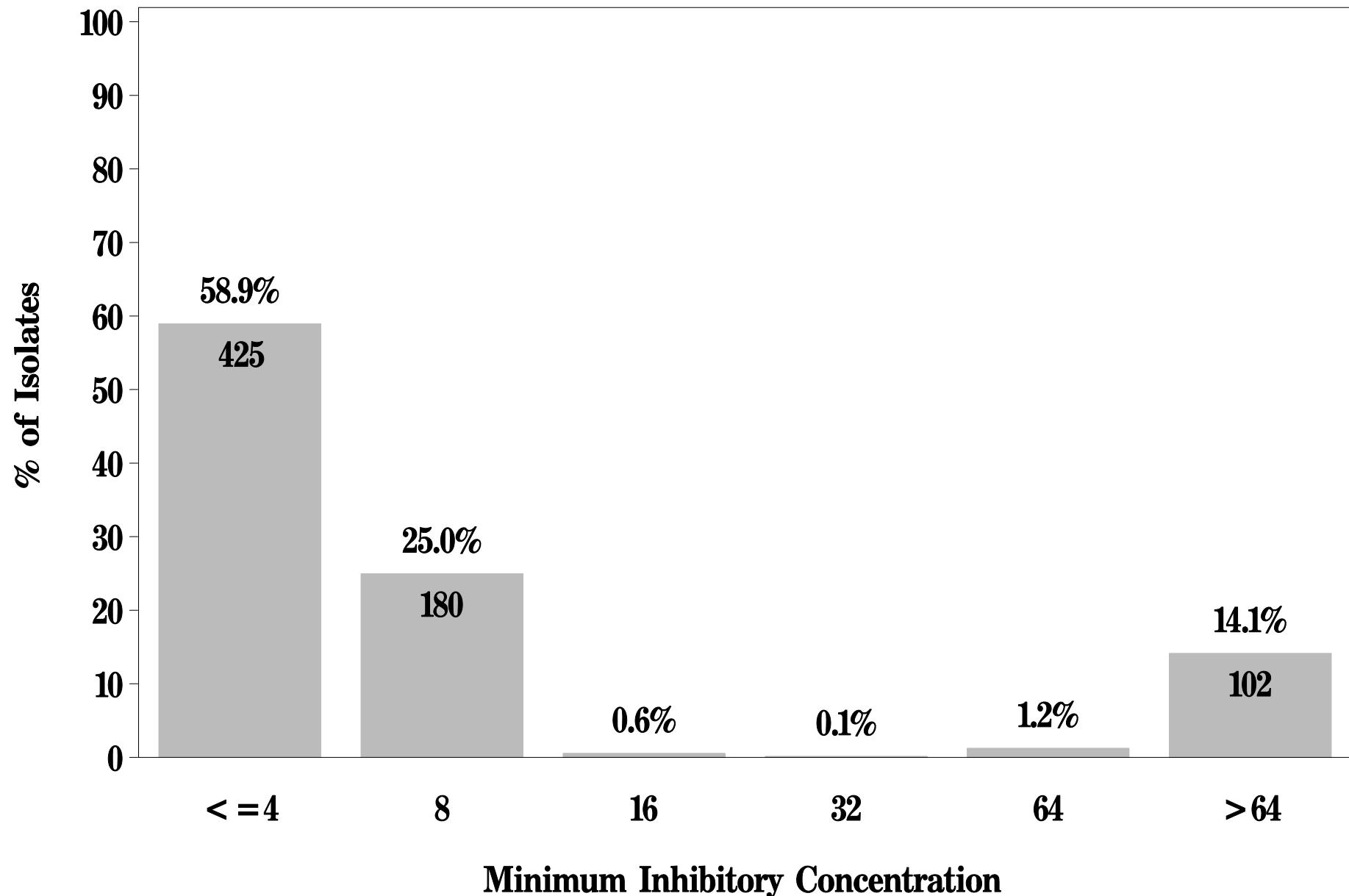
Breakpoints: Susceptible $\leq 2 \text{ } \mu\text{g/mL}$ Resistant $\geq 8 \text{ } \mu\text{g/mL}$



NARMS

**Figure 9g: Minimum Inhibitory Concentration of Nalidixic acid
for *Campylobacter* (N=721 Isolates)**

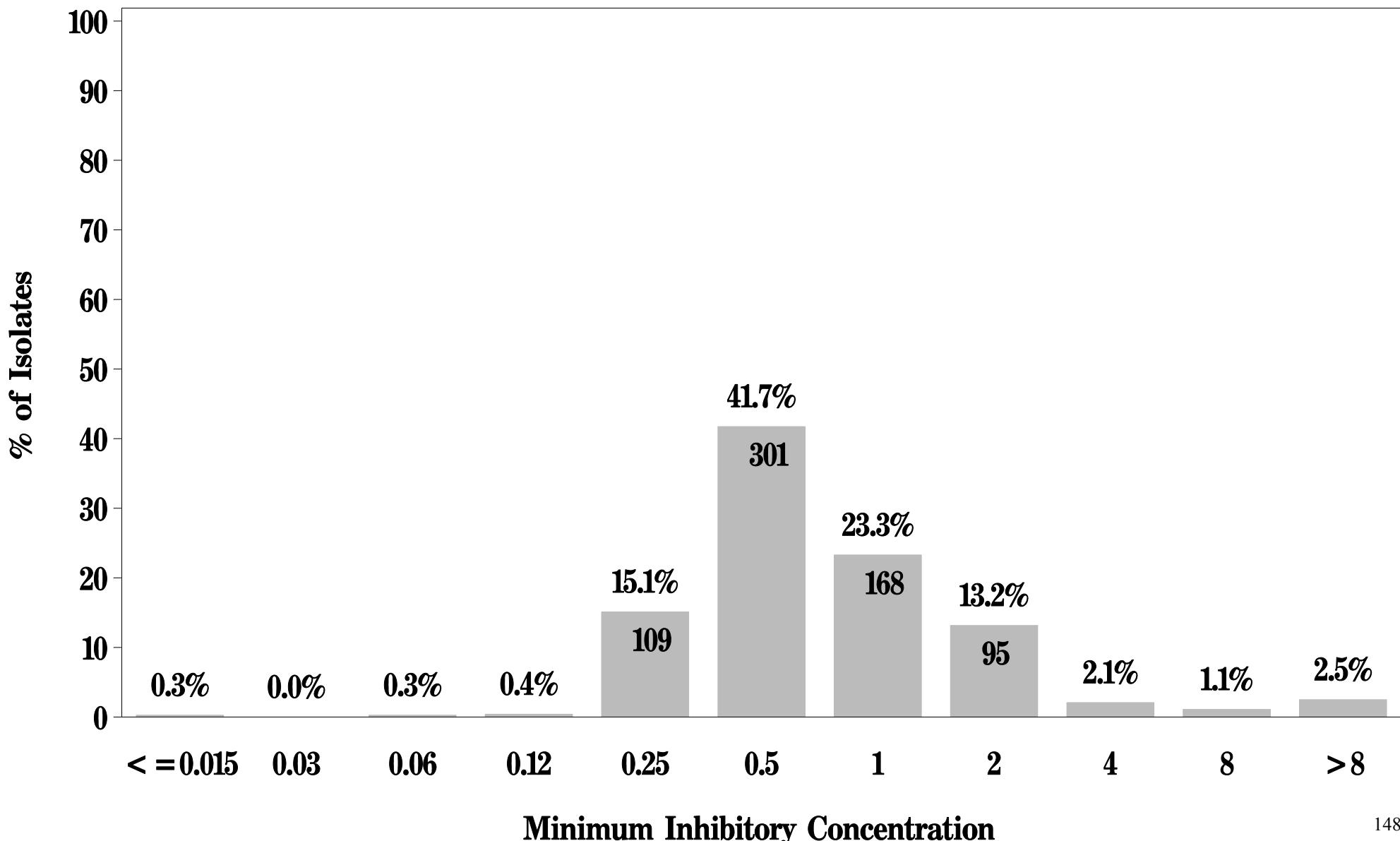
Breakpoints: Susceptible $\leq 16 \text{ } \mu\text{g/mL}$ Resistant $\geq 64 \text{ } \mu\text{g/mL}$



NARMS

**Figure 9h: Minimum Inhibitory Concentration of Telithromycin
for *Campylobacter* (N = 721 Isolates)**

Breakpoints: Susceptible $\leq 4 \text{ } \mu\text{g/mL}$ Resistant $\geq 16 \text{ } \mu\text{g/mL}$



NARMS

**Figure 9i: Minimum Inhibitory Concentration of Tetracycline
for *Campylobacter* (N=721 Isolates)**

Breakpoints: Susceptible $\leq 4 \text{ } \mu\text{g/mL}$ Resistant $\geq 16 \text{ } \mu\text{g/mL}$

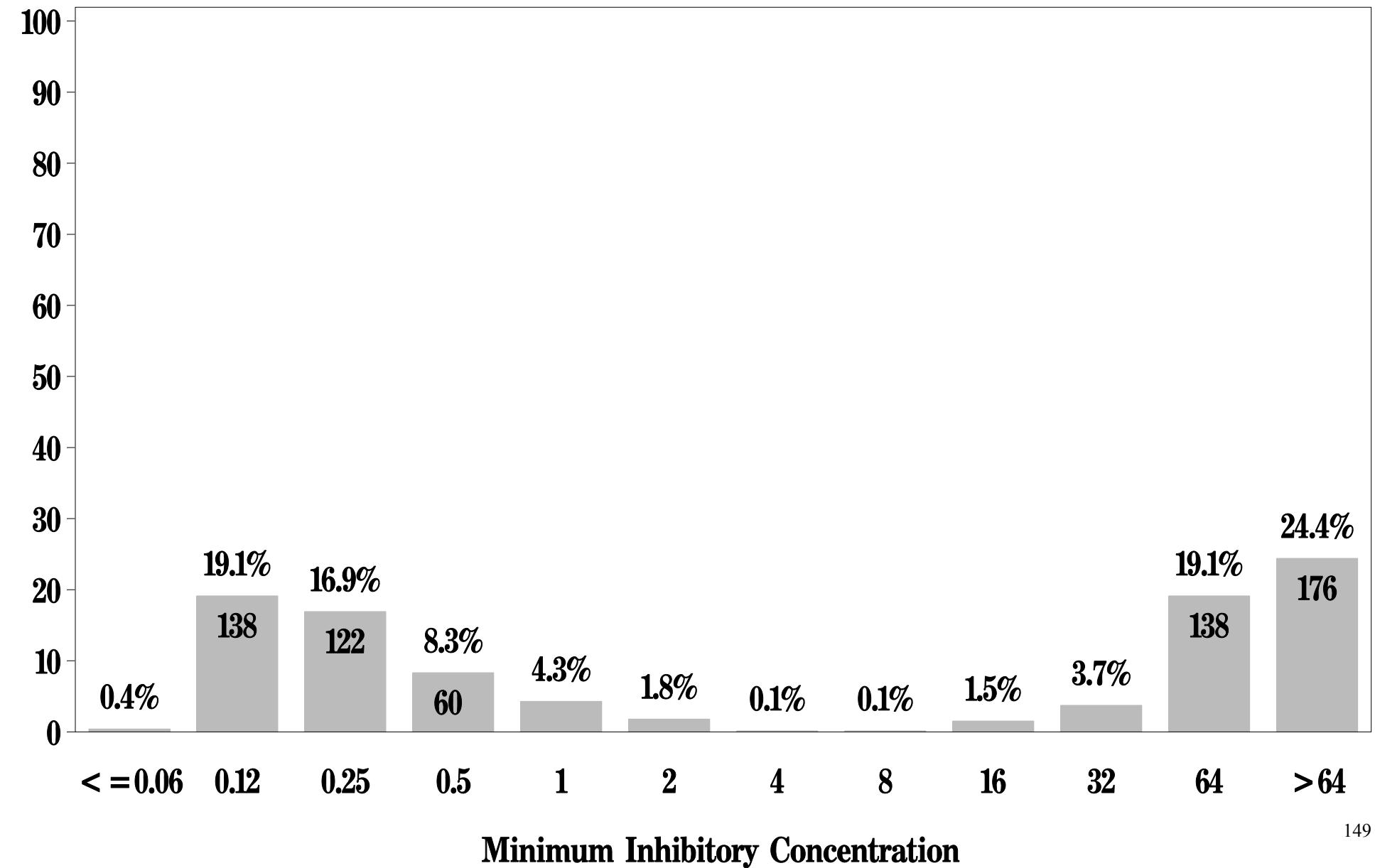


Figure 10a. MIC Distribution among *Campylobacter* Isolates from Chicken Breast, 2004

<i>Campylobacter</i> from Chicken Breast (N=706)		Distribution (%) of MICs (in µg/ml)													
Antimicrobial Agent	%R [†]	0.015	0.03	0.06	0.12	0.25	0.5	1	2	4	8	16	32	64	>64
Nalidixic Acid	15.6%									59.6	24.4	0.4	0.1	1.3	14.2
Ciprofloxacin	15.4%		0.1	35.1	37.1	12.0		0.1		0.3	7.1	6.8	1.3		
Azithromycin	3.1%	3.5	39.8	39.5	12.0	1.0	0.3	0.6	0.1						3.1
Clindamycin	2.3%		0.4	7.8	45.5	35.6	5.4	2.1	0.4	0.6	0.8	1.4			
Erythromycin	3.1%			0.3	2.1	44.5	30.3	16.7	2.4	0.4	0.1				3.1
Telithromycin	3.5%	0.3		0.3	0.4	15.2	42.4	22.4	13.5	2.1	1.0	2.5			
Gentamicin*	0.0%				1.4	4.8	85.3	8.5							
Florfenicol [§]	§				0.4		4.1	79.9	15.2	0.4					
Tetracycline	49.2%			0.4	19.4	17.0	8.2	4.0	1.6	0.1	0.1	1.6	3.8	19.4	24.4

[†]Discrepancies between %R and sums of distribution %s are due to rounding.

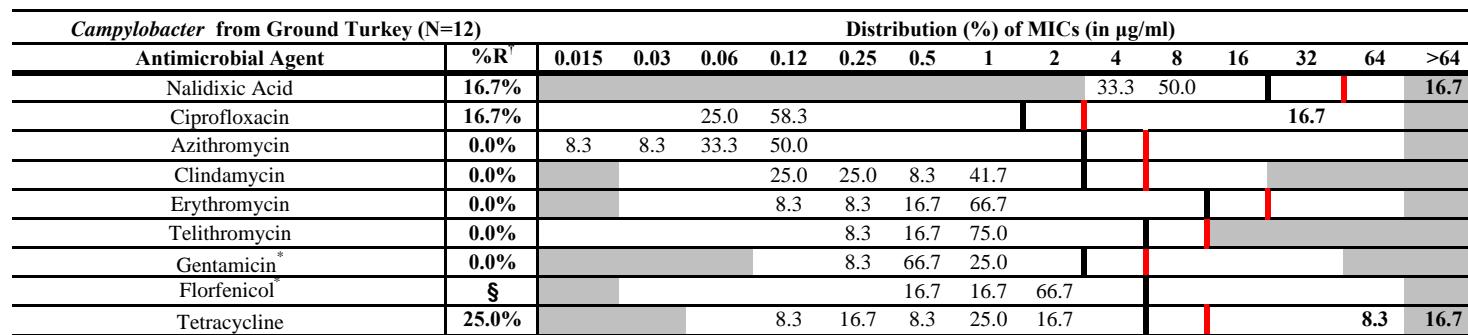
Vertical bars show the CLSI Susceptible/Resistant breakpoints for each drug.

Unshaded areas indicate the dilution ranges used to test the 2004 isolates.

*Currently no CLSI breakpoints have been established for this organism/antimicrobial combination. Indicated breakpoints were established by NARMS.

§Absence of resistant strains precludes defining any results category other than "susceptible."

Figure 10b. MIC Distribution among *Campylobacter* Isolates from Ground Turkey, 2004



[†]Discrepancies between %R and sums of distribution %s are due to rounding.

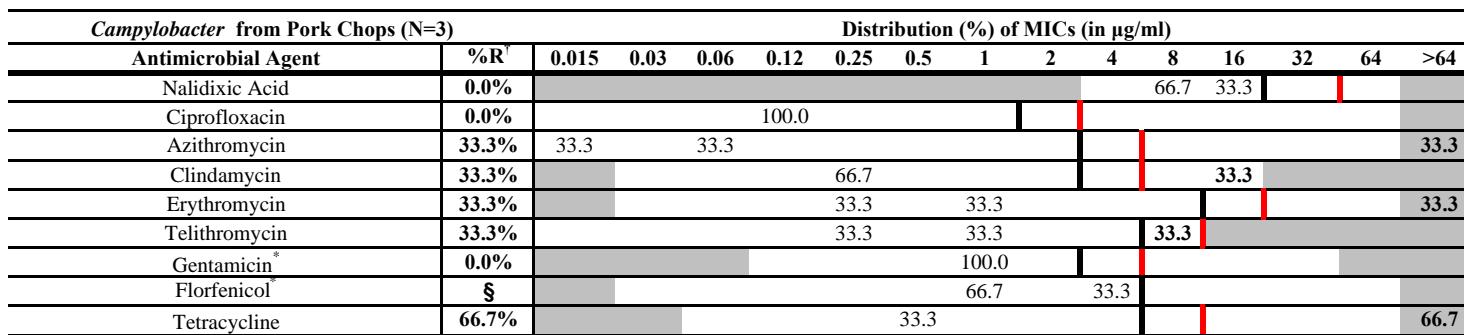
Vertical bars show the CLSI Susceptible/Resistant breakpoints for each drug.

Unshaded areas indicate the dilution ranges used to test the 2004 isolates.

*Currently no CLSI breakpoints have been established for this organism/antimicrobial combination. Indicated breakpoints were established by NARMS.

§Absence of resistant strains precludes defining any results category other than "susceptible."

Figure 10c. MIC Distribution among *Campylobacter* Isolates from Pork Chops, 2004



[†]Discrepancies between %R and sums of distribution %s are due to rounding.

Vertical bars show the CLSI Susceptible/Resistant breakpoints for each drug.

Unshaded areas indicate the dilution ranges used to test the 2004 isolates.

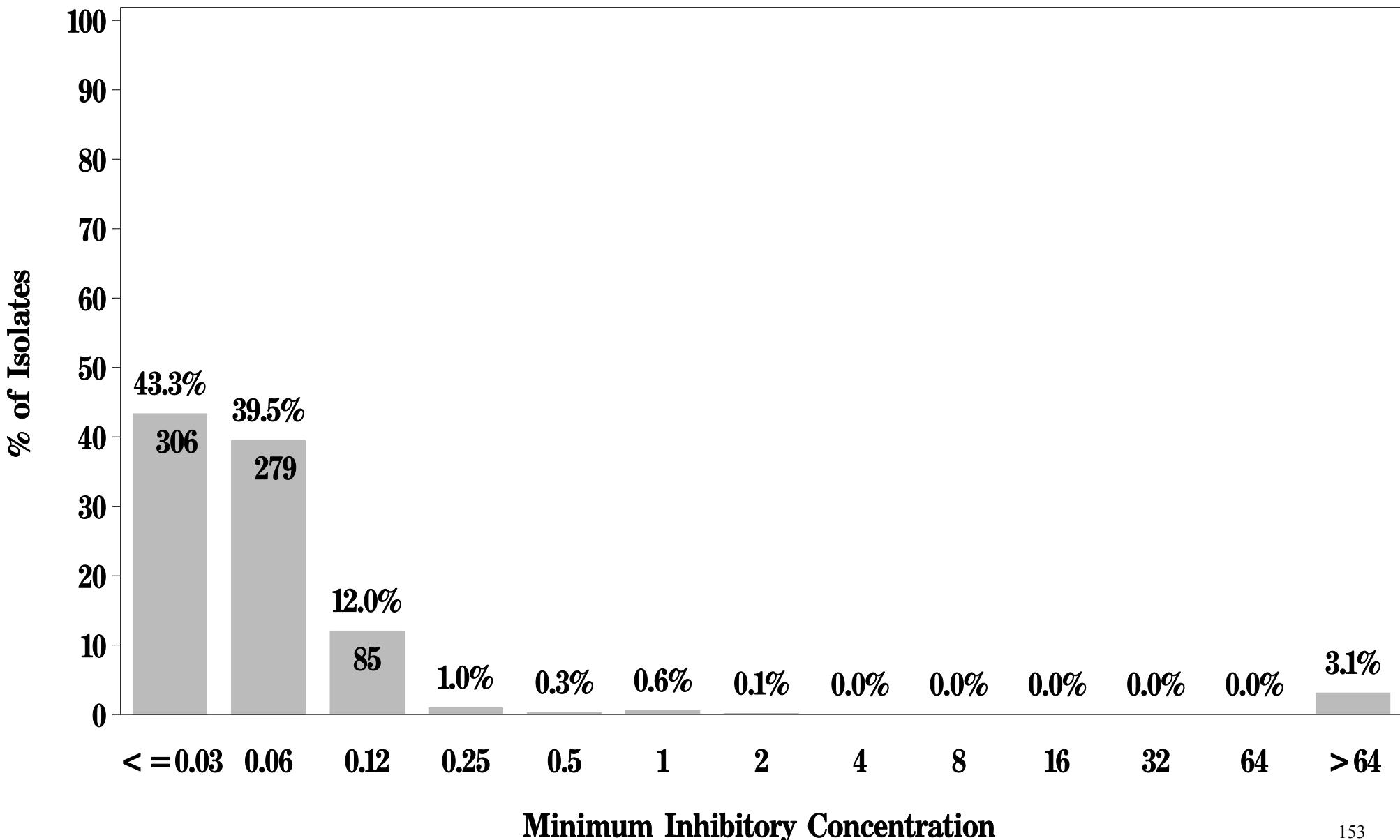
*Currently no CLSI breakpoints have been established for this organism/antimicrobial combination. Indicated breakpoints were established by NARMS.

[§]Absence of resistant strains precludes defining any results category other than "susceptible."

NARMS

**Figure 11a: Minimum Inhibitory Concentration of Azithromycin
for *Campylobacter* in Chicken Breast (N = 706 Isolates)**

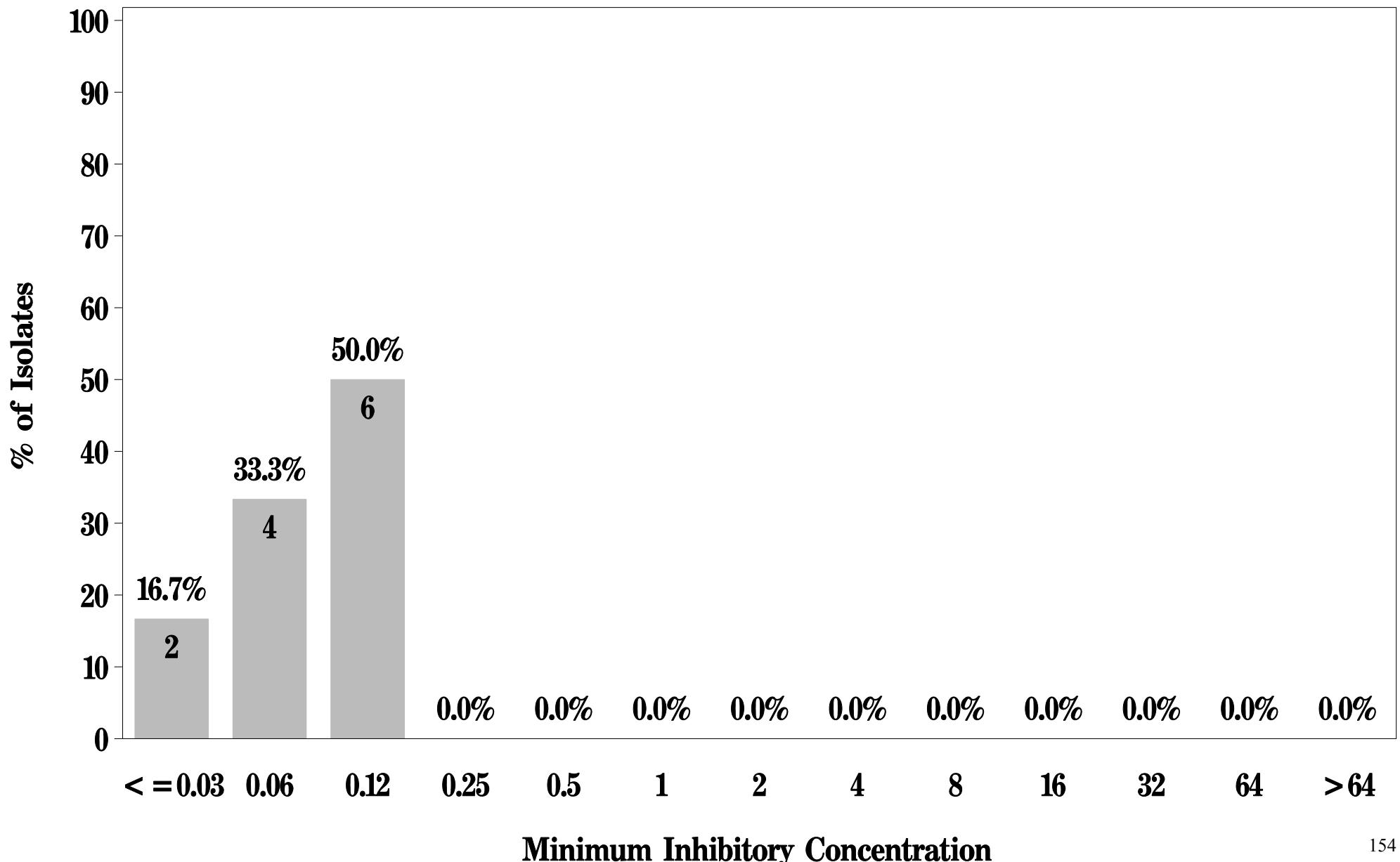
Breakpoints: Susceptible $\leq 2 \mu\text{g/mL}$ Resistant $\geq 8 \mu\text{g/mL}$



NARMS

**Figure 11a: Minimum Inhibitory Concentration of Azithromycin
for *Campylobacter* in Ground Turkey (N=12 Isolates)**

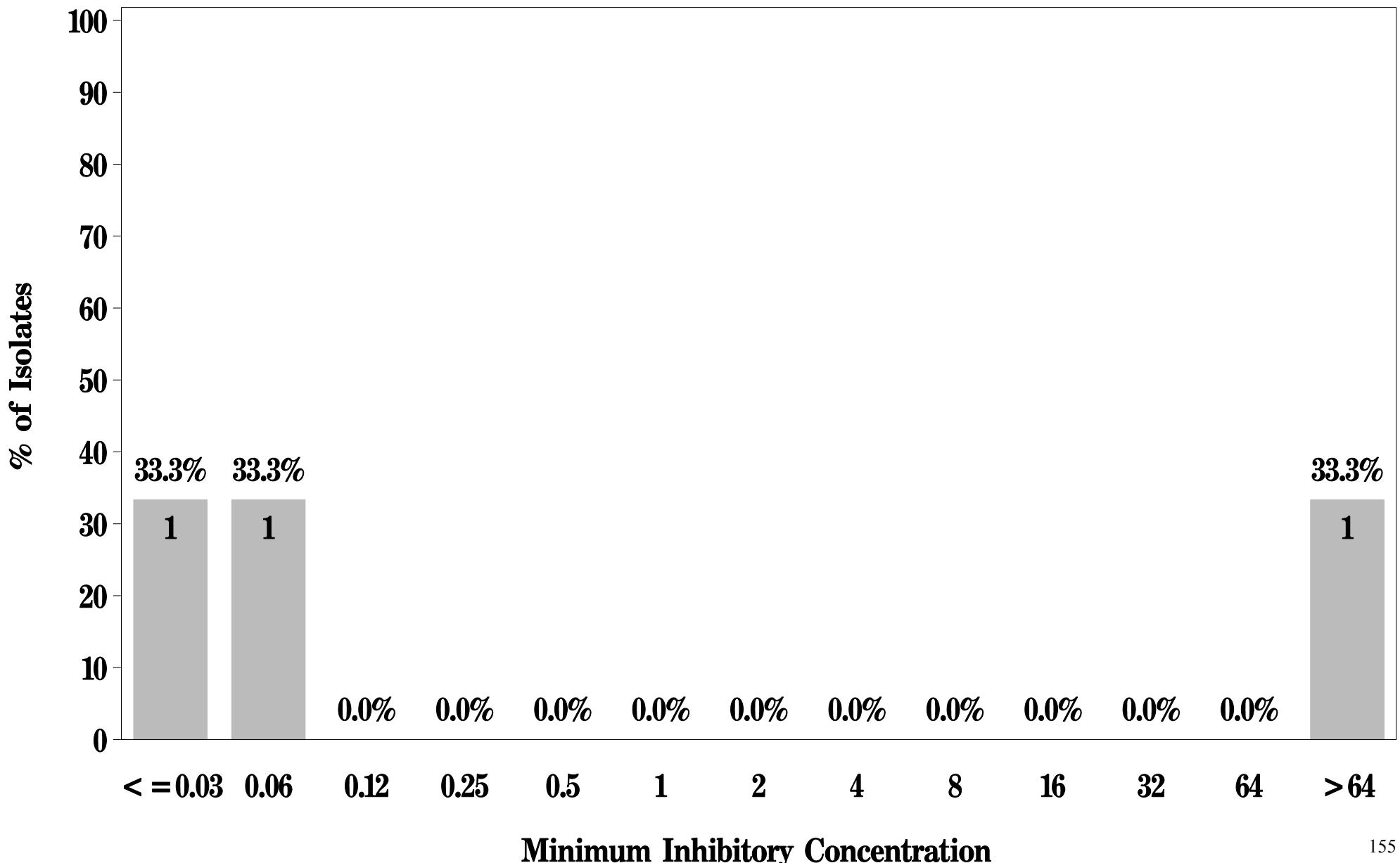
Breakpoints: Susceptible $\leq 2 \mu\text{g/mL}$ Resistant $\geq 8 \mu\text{g/mL}$



NARMS

**Figure 11a: Minimum Inhibitory Concentration of Azithromycin
for *Campylobacter* in Pork Chop (N=3 Isolates)**

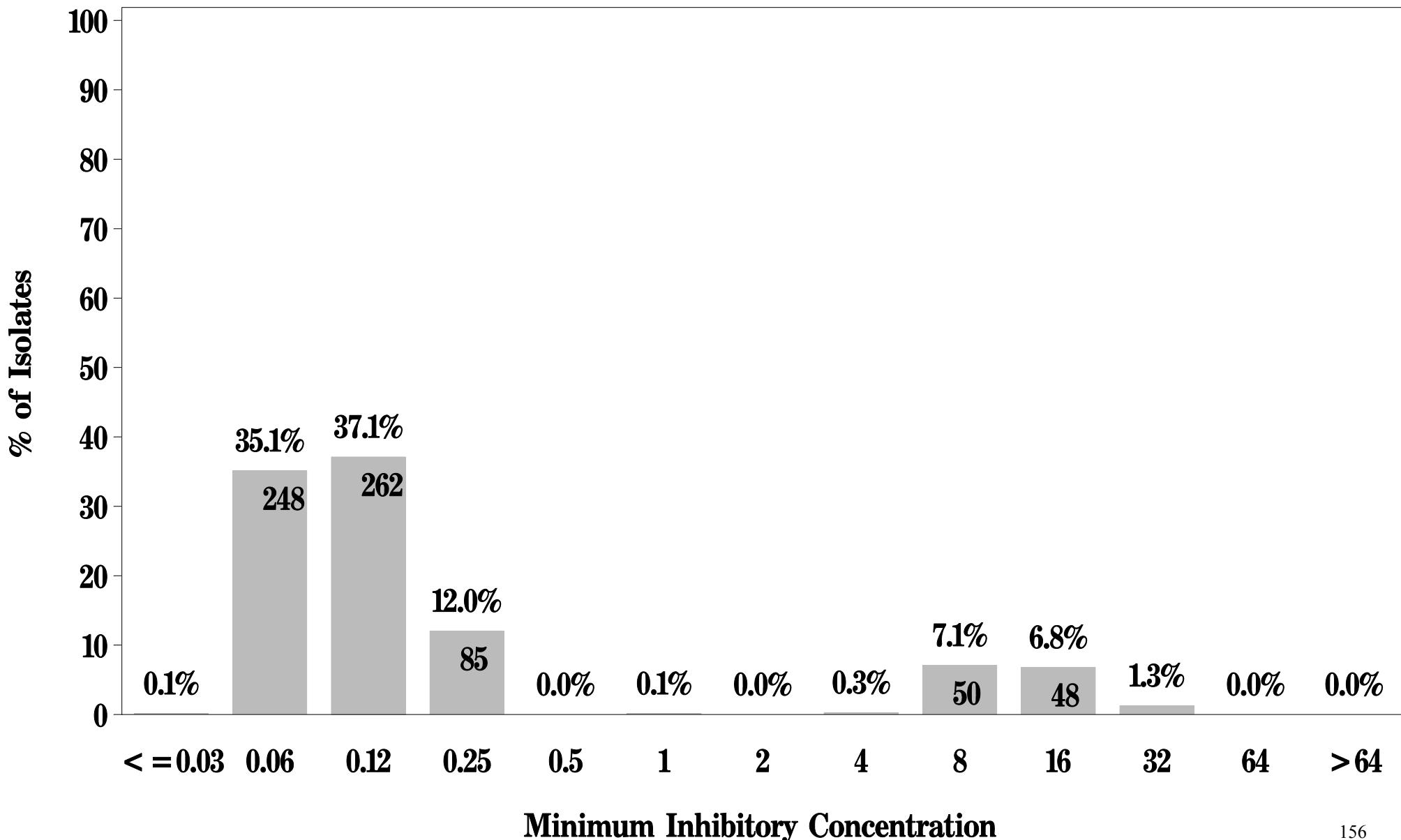
Breakpoints: Susceptible $\leq 2 \mu\text{g/mL}$ Resistant $\geq 8 \mu\text{g/mL}$



NARMS

**Figure 11b: Minimum Inhibitory Concentration of Ciprofloxacin
for *Campylobacter* in Chicken Breast (N = 706 Isolates)**

Breakpoints: Susceptible $\leq 1 \mu\text{g/mL}$ Resistant $\geq 4 \mu\text{g/mL}$

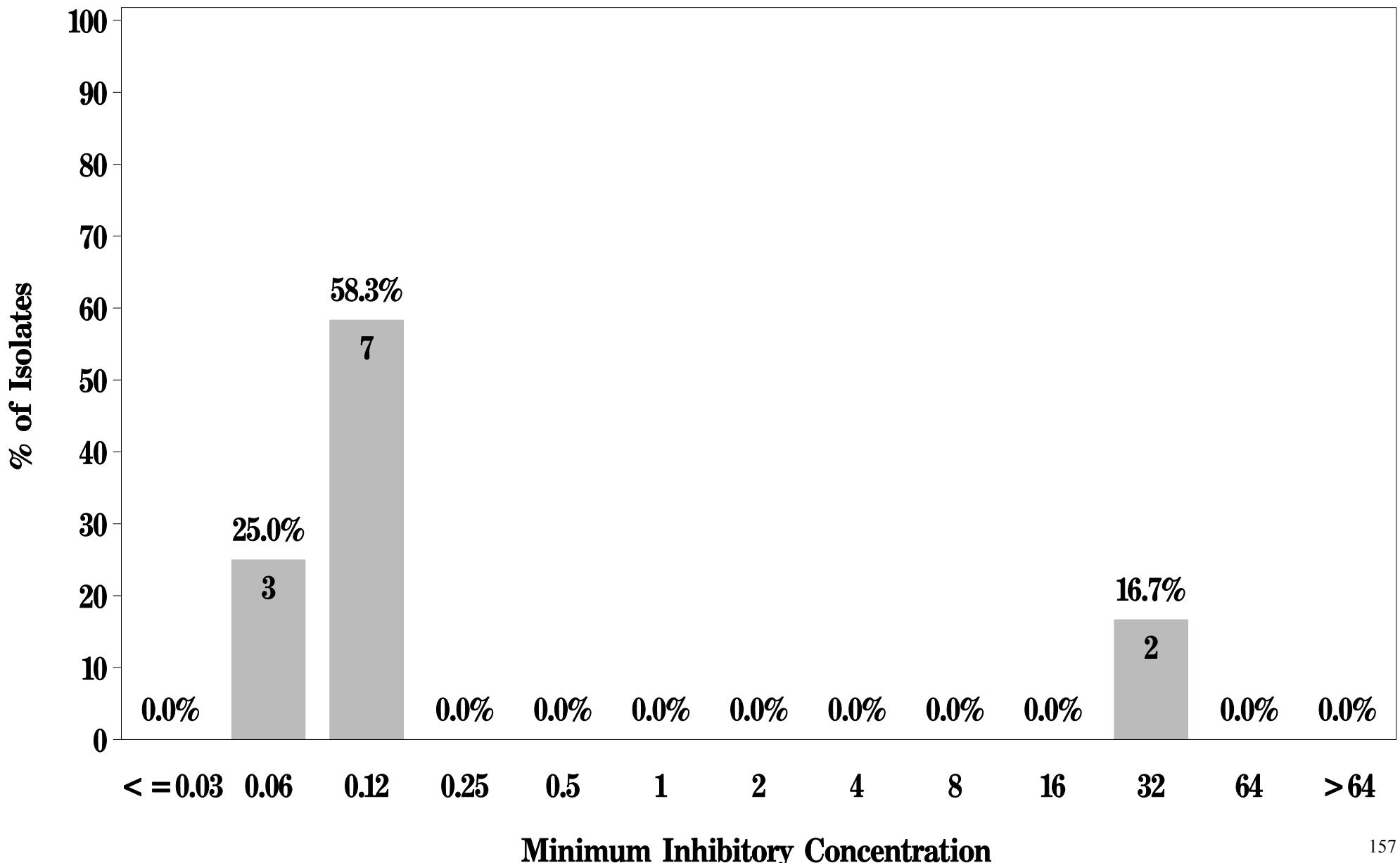


NARMS

Figure 11b: Minimum Inhibitory Concentration of Ciprofloxacin

for *Campylobacter* in Ground Turkey (N=12 Isolates)

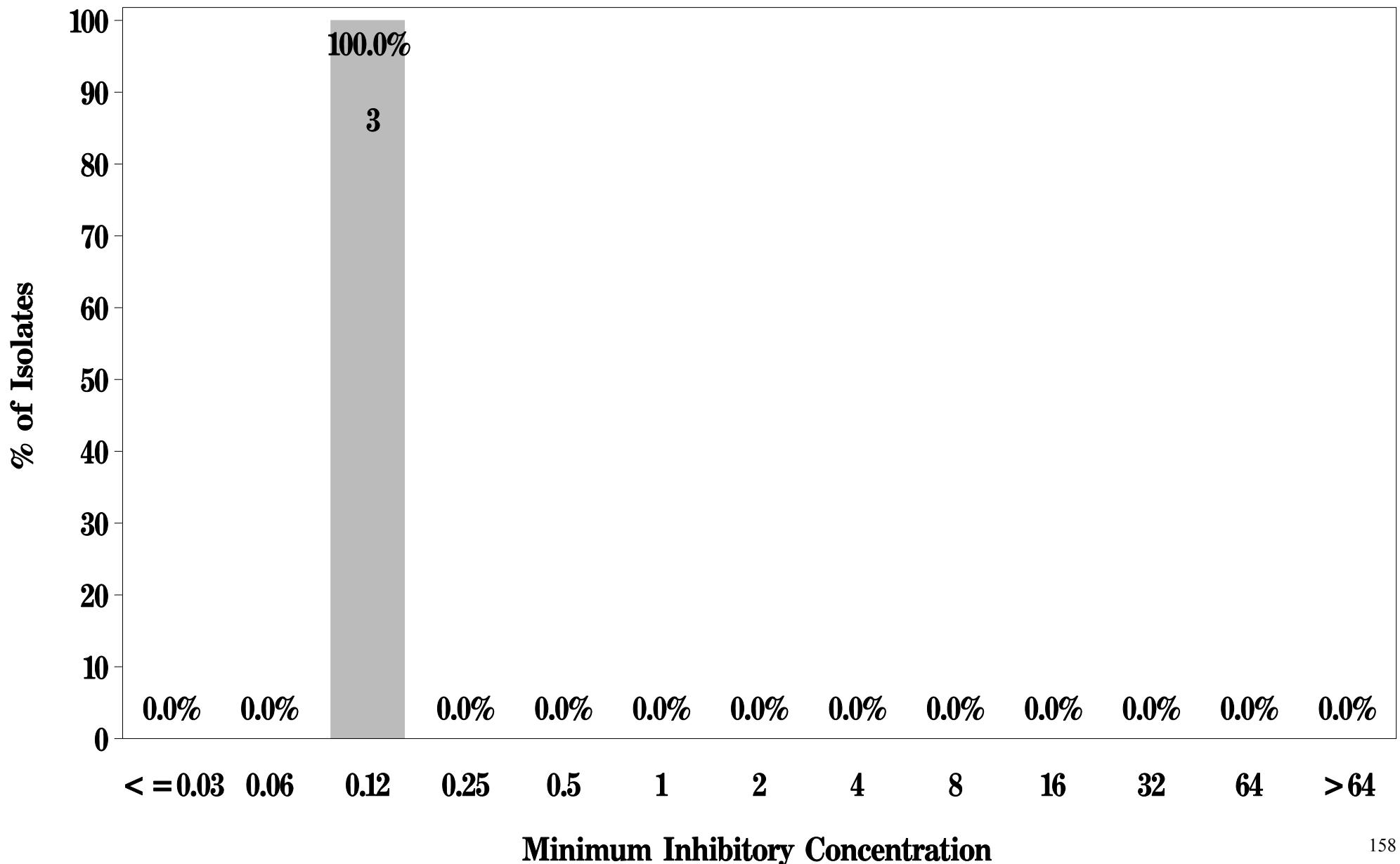
Breakpoints: Susceptible $\leq 1 \mu\text{g/mL}$ Resistant $\geq 4 \mu\text{g/mL}$



NARMS

Figure 11b: Minimum Inhibitory Concentration of Ciprofloxacin
for *Campylobacter* in Pork Chop (N=3 Isolates)

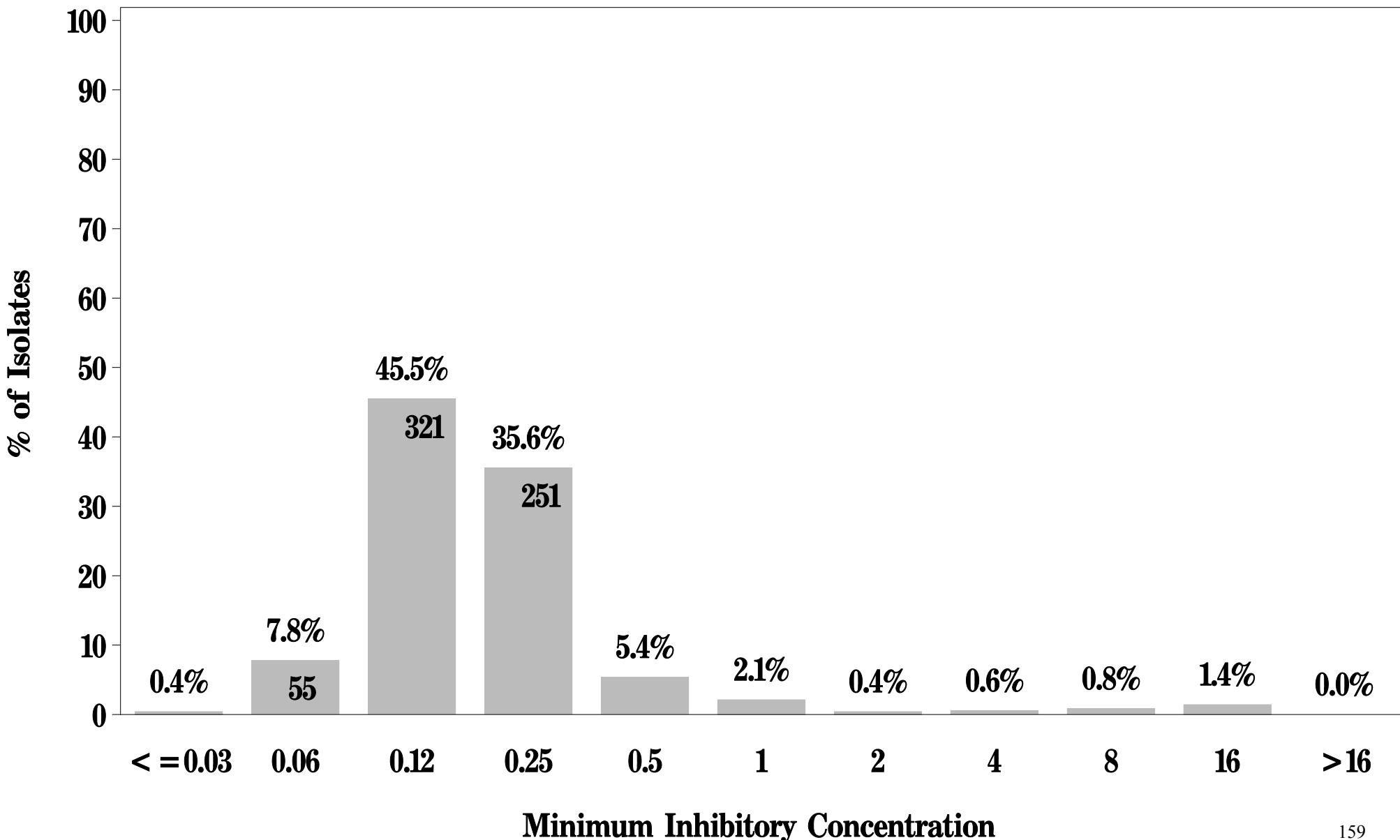
Breakpoints: Susceptible $\leq 1 \mu\text{g/mL}$ Resistant $\geq 4 \mu\text{g/mL}$



NARMS

**Figure 11c: Minimum Inhibitory Concentration of Clindamycin
for *Campylobacter* in Chicken Breast (N = 706 Isolates)**

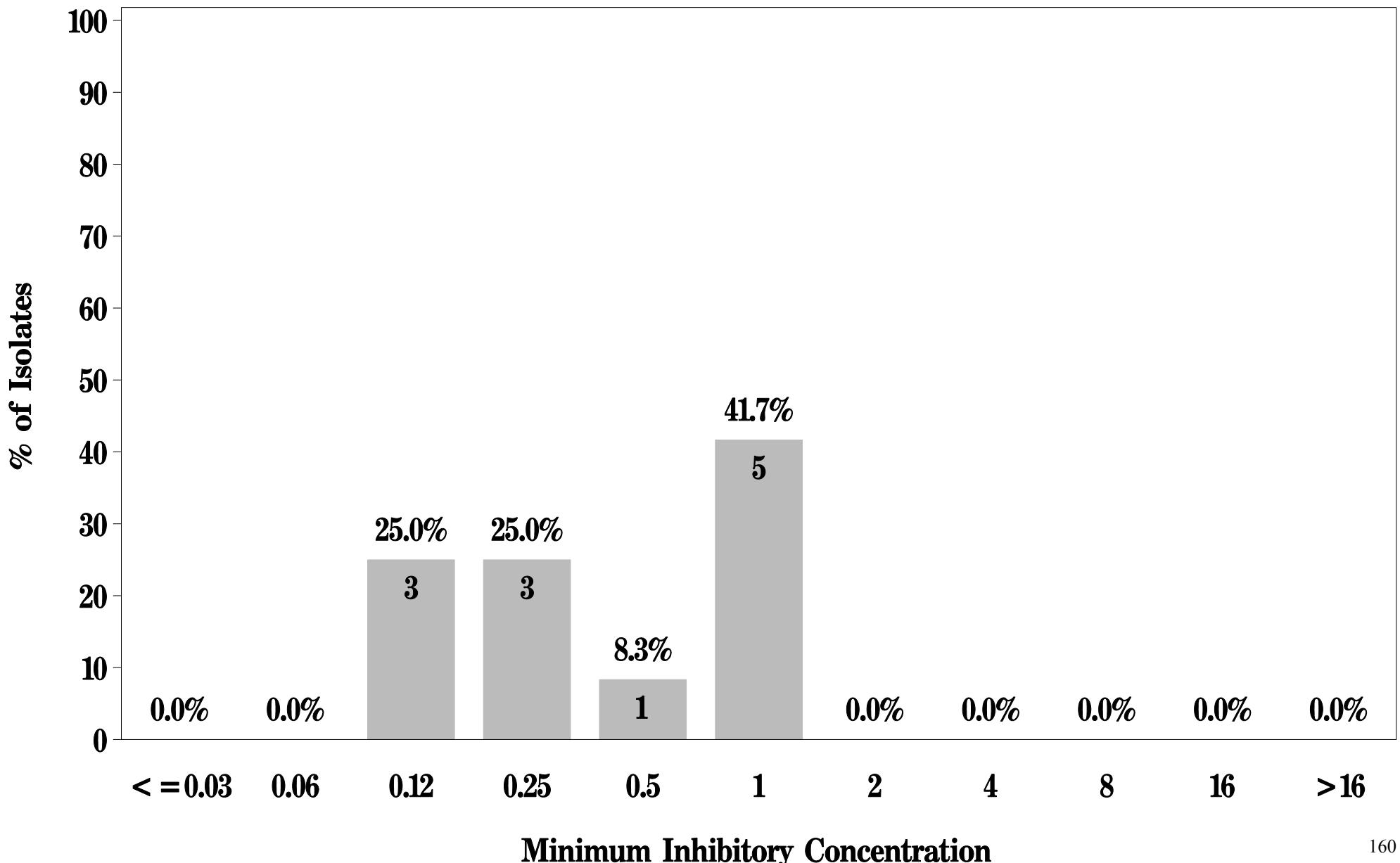
Breakpoints: Susceptible $\leq 2 \mu\text{g/mL}$ Resistant $\geq 8 \mu\text{g/mL}$



NARMS

Figure 11c: Minimum Inhibitory Concentration of Clindamycin
for *Campylobacter* in Ground Turkey (N=12 Isolates)

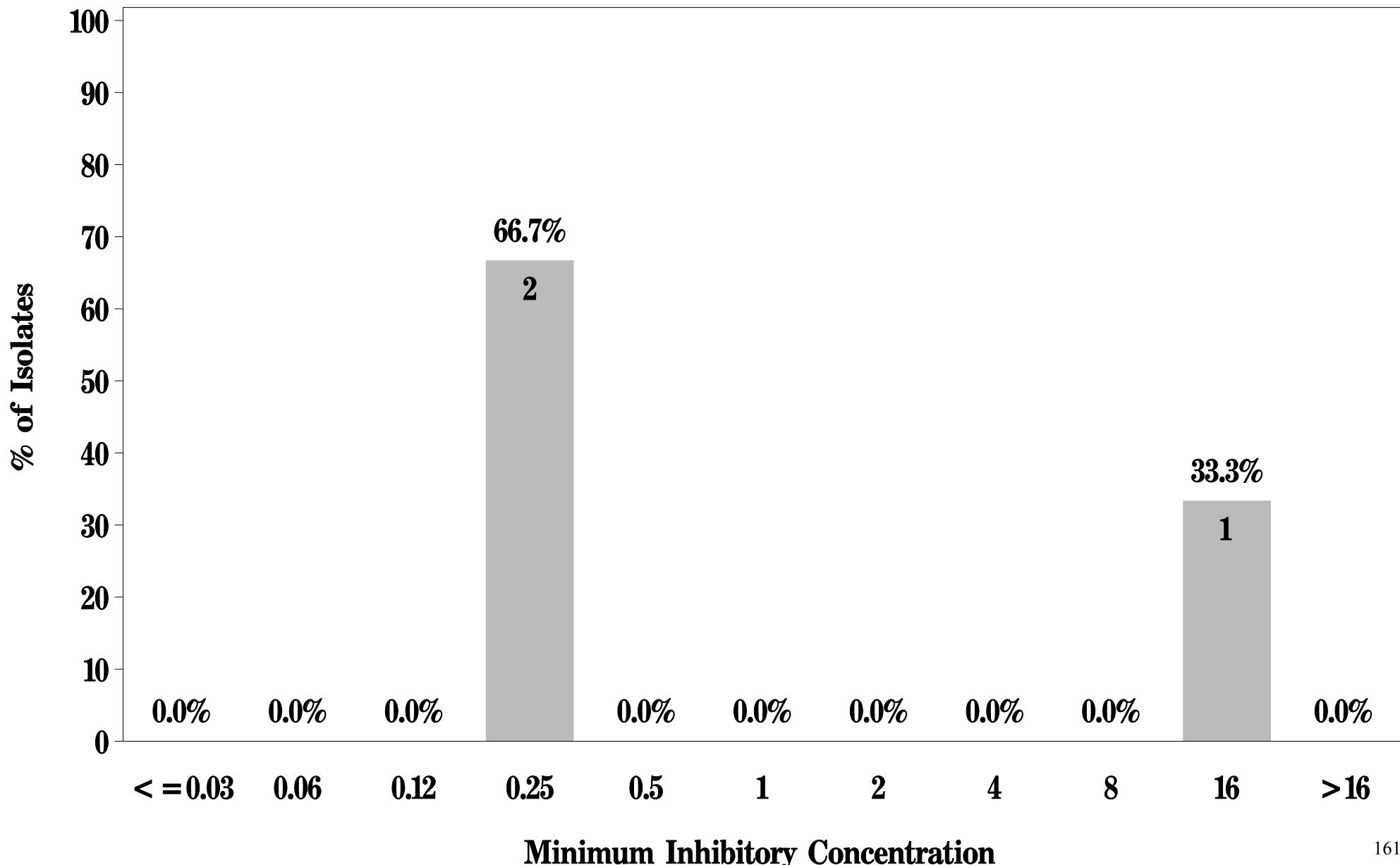
Breakpoints: Susceptible $\leq 2 \mu\text{g/mL}$ Resistant $\geq 8 \mu\text{g/mL}$



NARMS

Figure 11c: Minimum Inhibitory Concentration of Clindamycin
for *Campylobacter* in Pork Chop (N=3 Isolates)

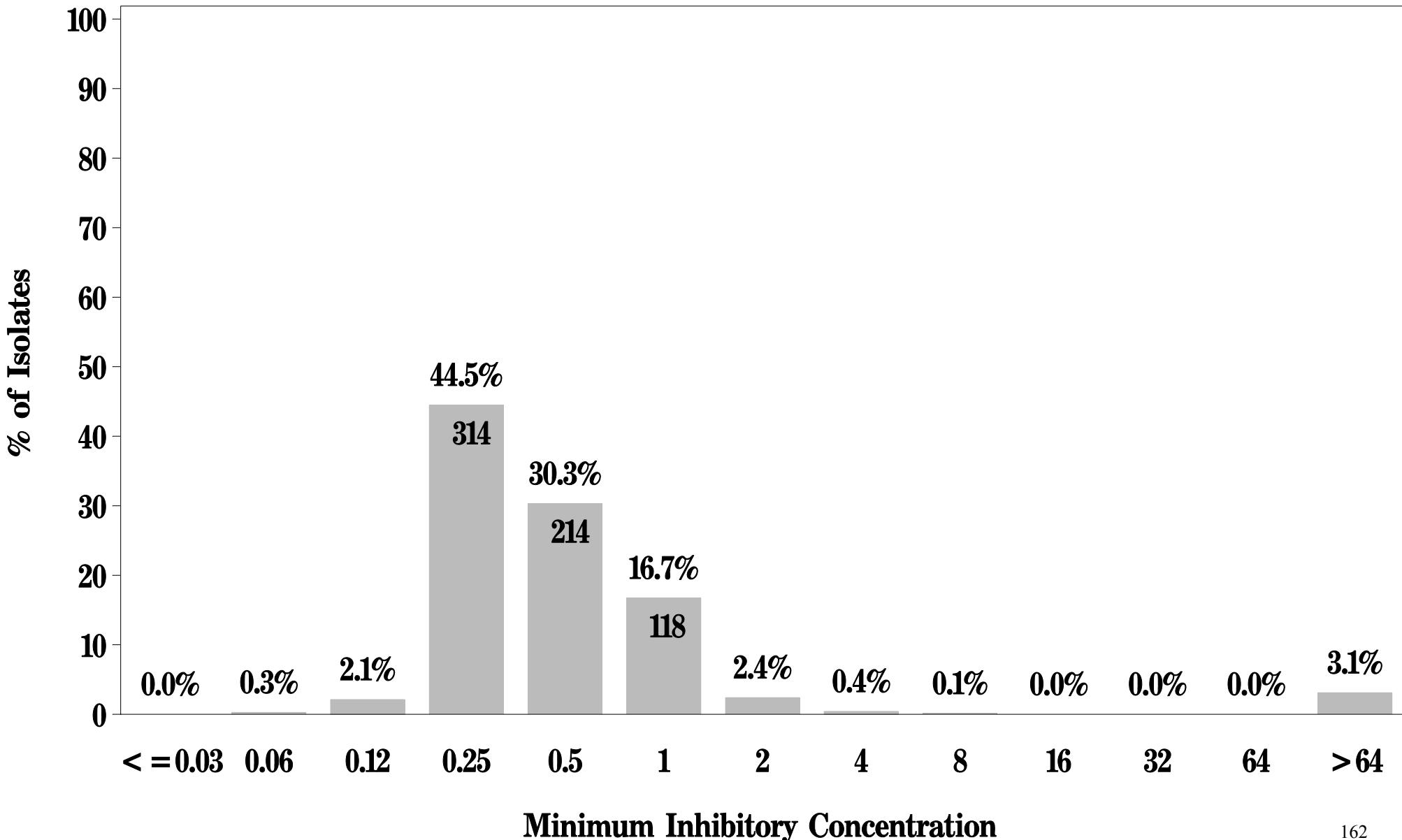
Breakpoints: Susceptible $\leq 2 \mu\text{g/mL}$ Resistant $\geq 8 \mu\text{g/mL}$



NARMS

**Figure 11d: Minimum Inhibitory Concentration of Erythromycin
for *Campylobacter* in Chicken Breast (N = 706 Isolates)**

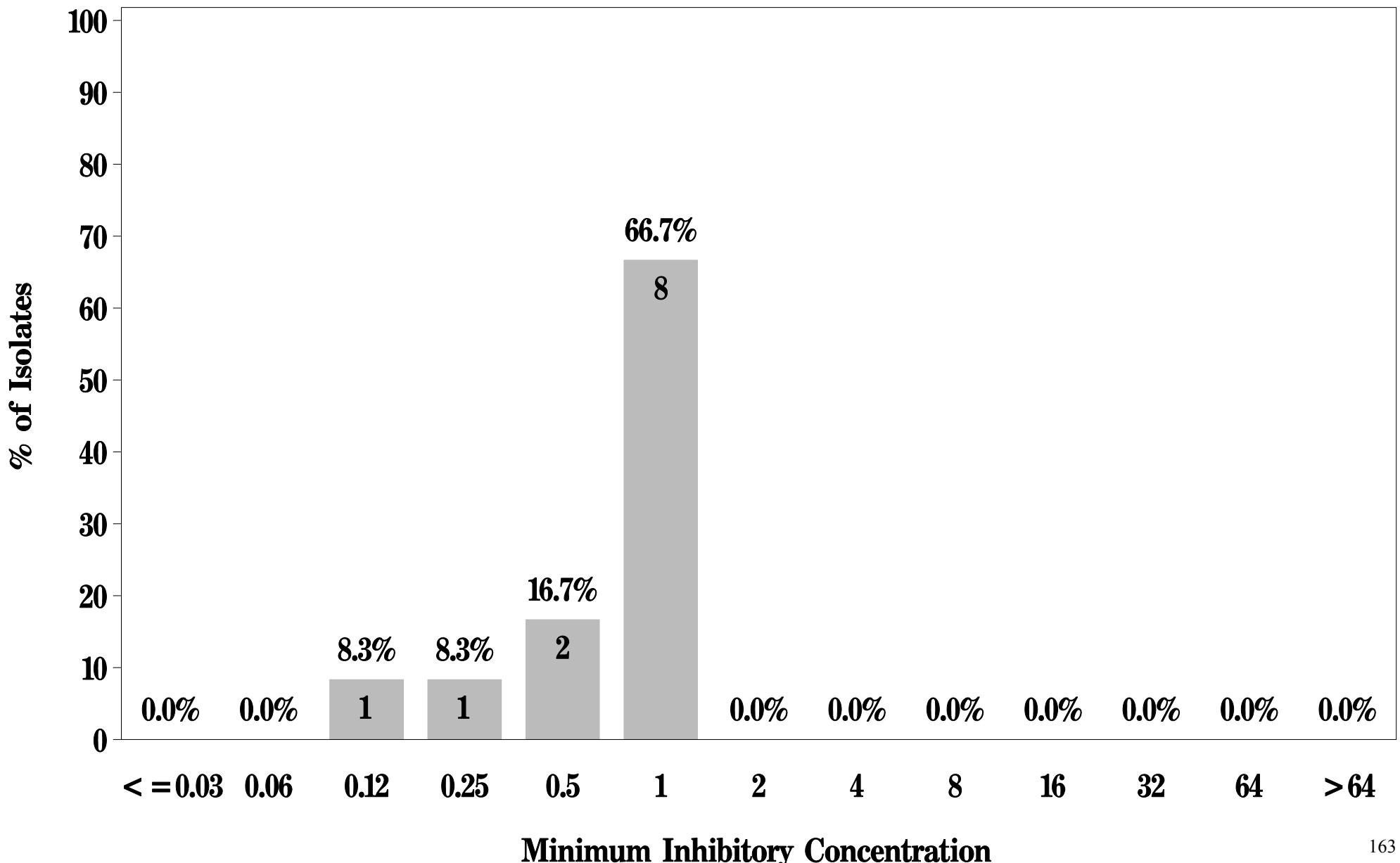
Breakpoints: Susceptible $\leq 8 \mu\text{g/mL}$ Resistant $\geq 32 \mu\text{g/mL}$



NARMS

Figure 11d: Minimum Inhibitory Concentration of Erythromycin
for *Campylobacter* in Ground Turkey (N=12 Isolates)

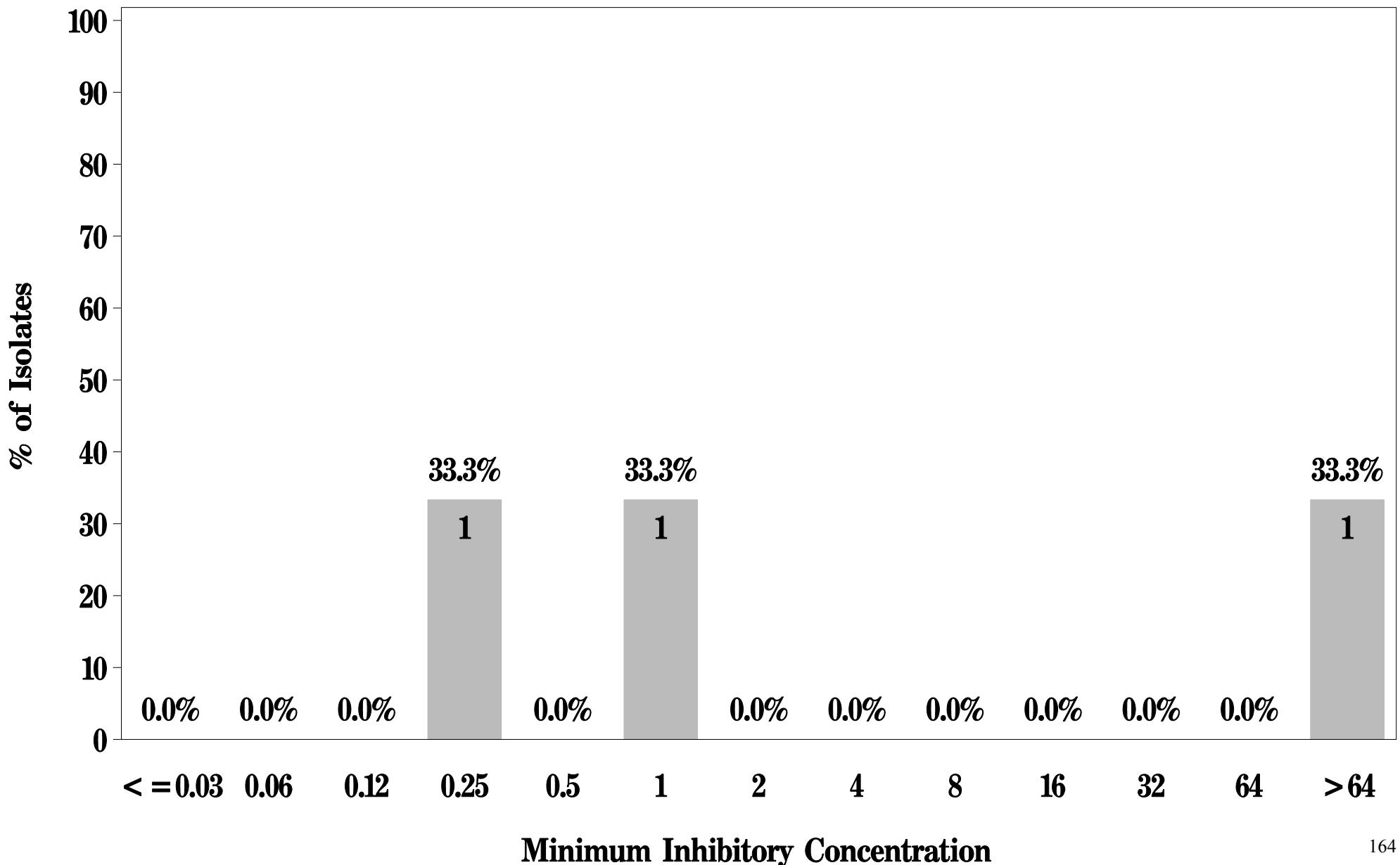
Breakpoints: Susceptible $\leq 8 \mu\text{g/mL}$ Resistant $\geq 32 \mu\text{g/mL}$



NARMS

Figure 11d: Minimum Inhibitory Concentration of Erythromycin
for *Campylobacter* in Pork Chop (N=3 Isolates)

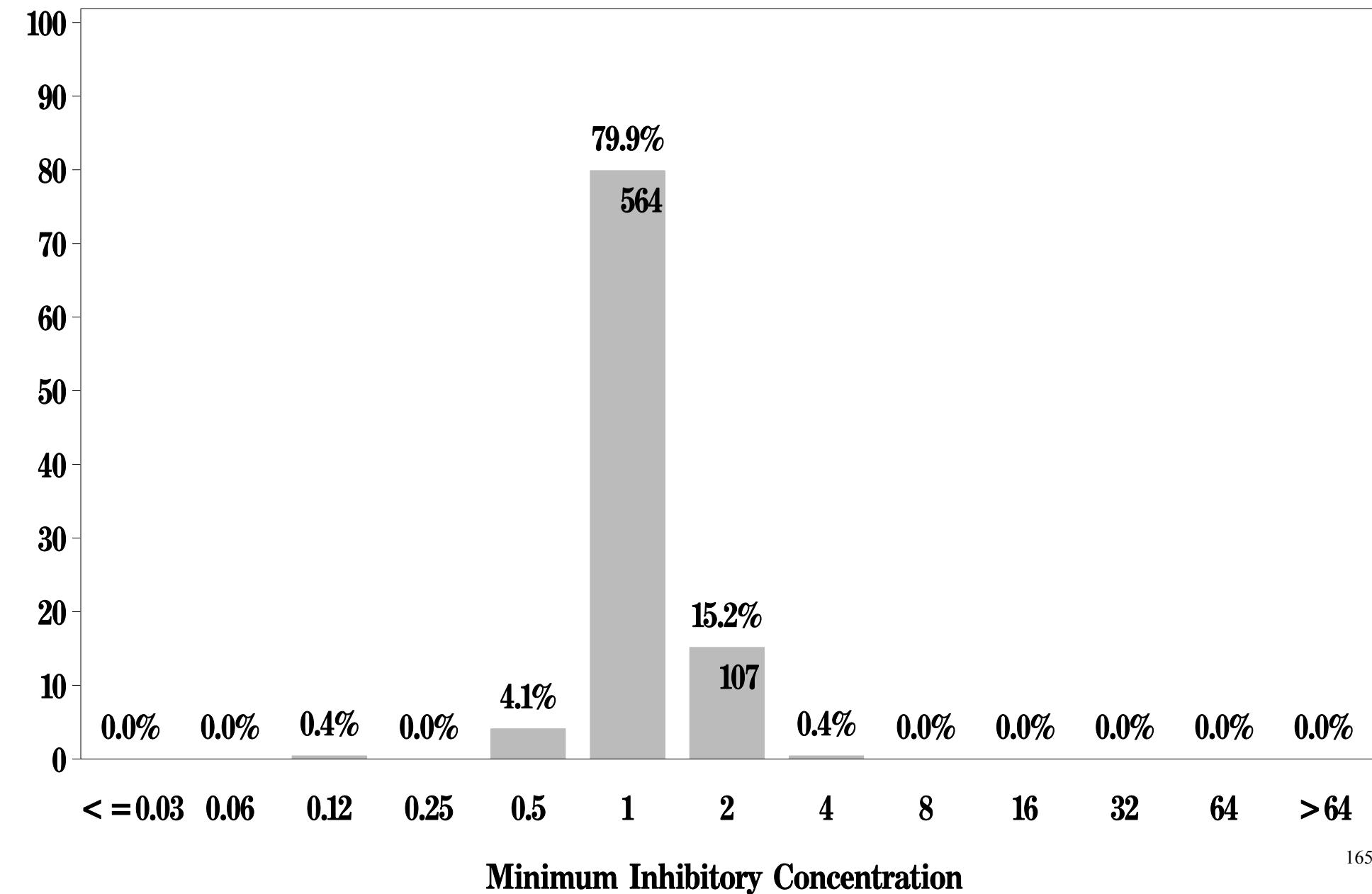
Breakpoints: Susceptible $\leq 8 \text{ } \mu\text{g/mL}$ Resistant $\geq 32 \text{ } \mu\text{g/mL}$



NARMS

Figure 11e: Minimum Inhibitory Concentration of Florfenicol
for *Campylobacter* in Chicken Breast (N=706 Isolates)

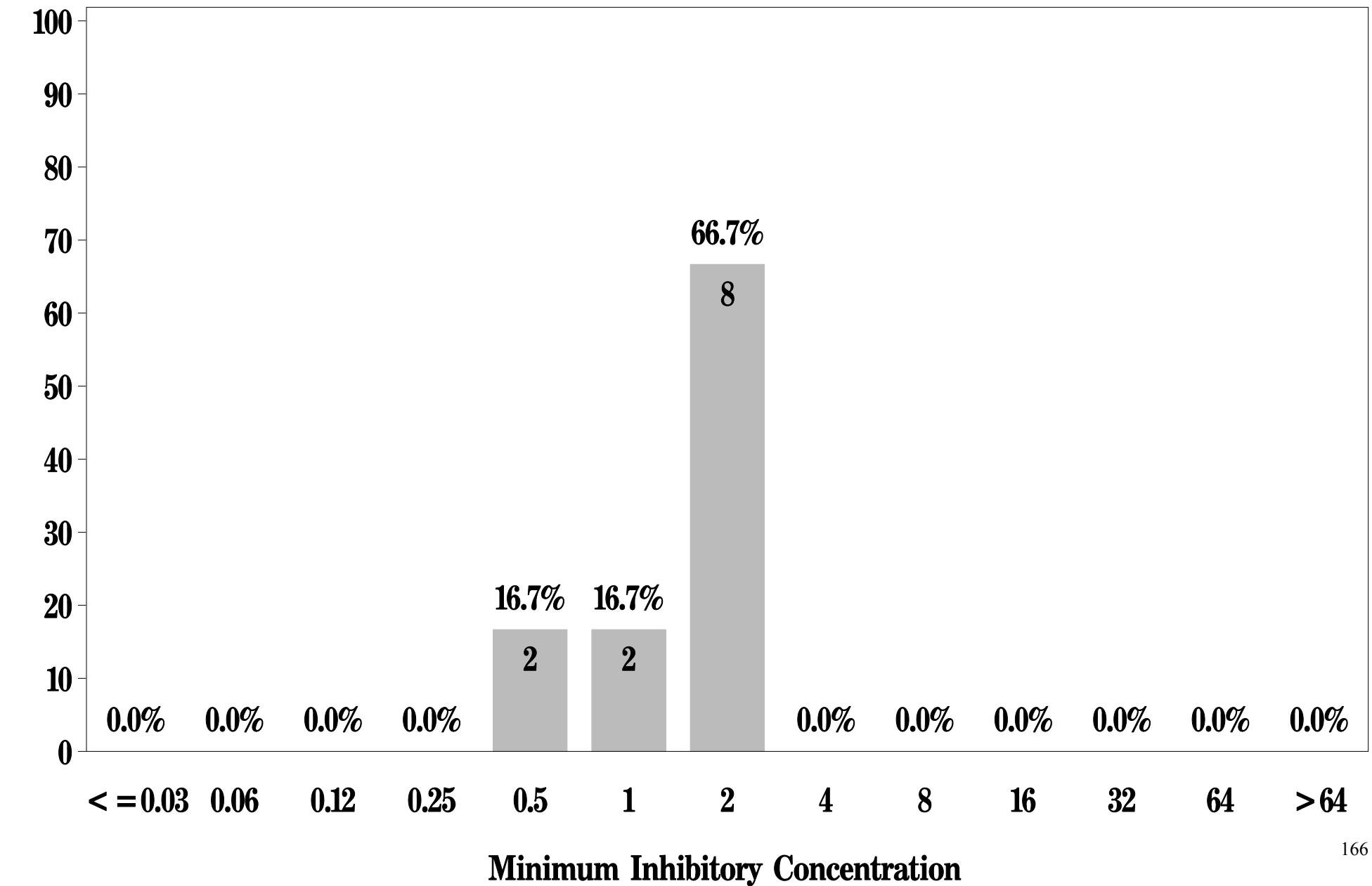
Breakpoints: Susceptible $\leq 4 \mu\text{g/mL}$



NARMS

**Figure 11e: Minimum Inhibitory Concentration of Florfenicol
for *Campylobacter* in Ground Turkey (N=12 Isolates)**

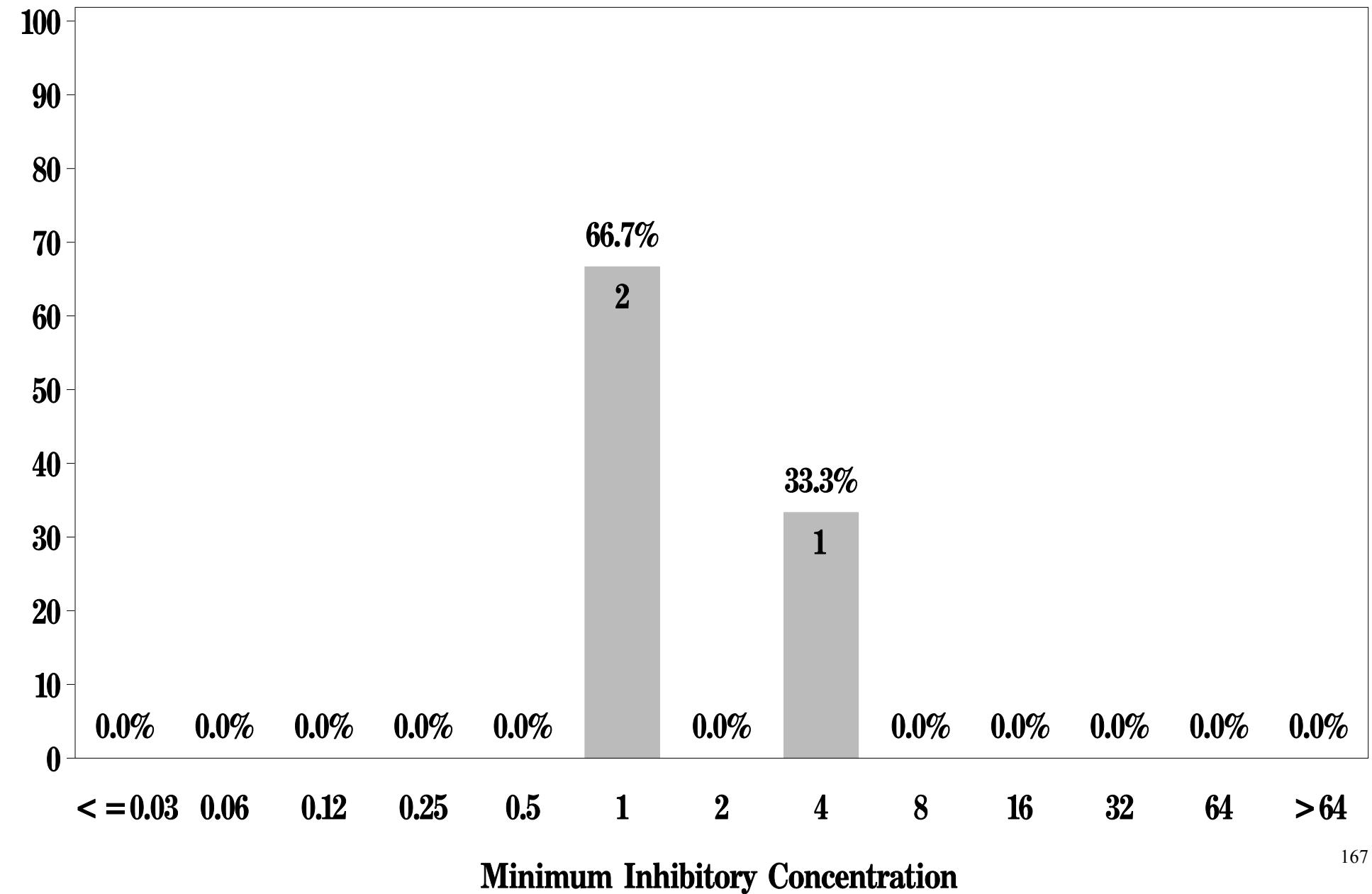
Breakpoints: Susceptible $\leq 4 \mu\text{g/mL}$



NARMS

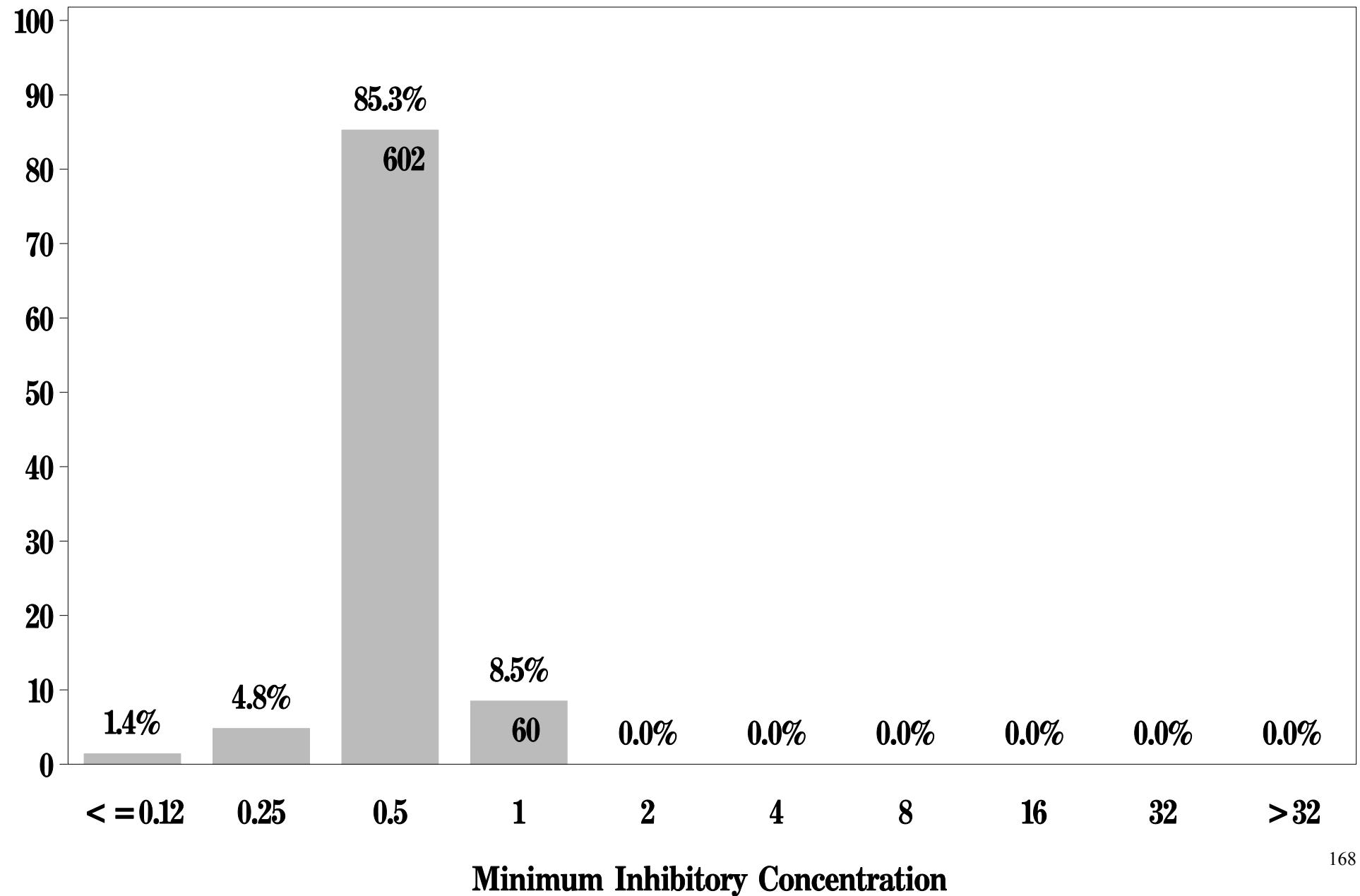
Figure 11e: Minimum Inhibitory Concentration of Florfenicol
for *Campylobacter* in Pork Chop (N=3 Isolates)

Breakpoints: Susceptible $\leq 4 \mu\text{g/mL}$



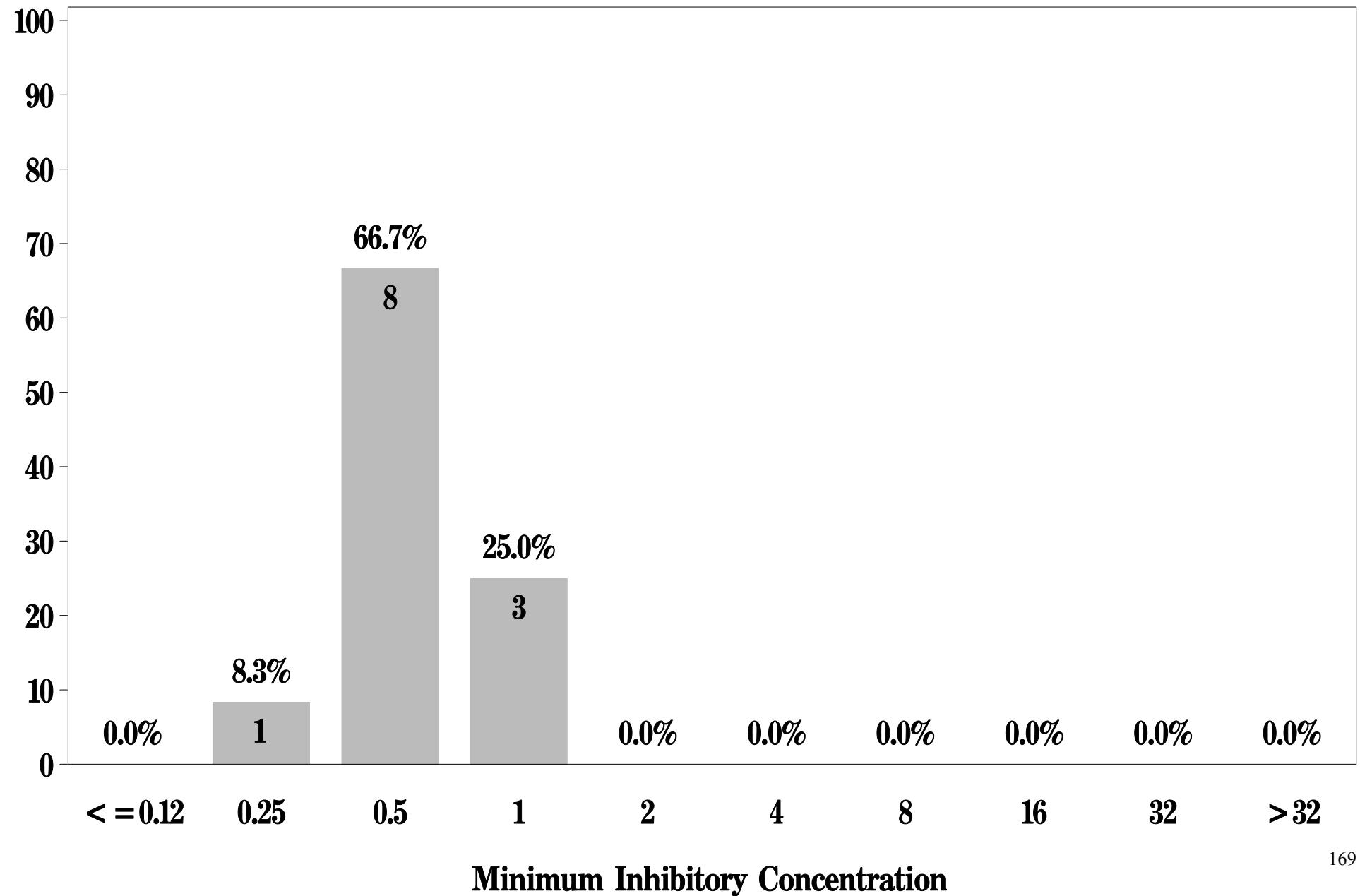
NARMS

**Figure 11f: Minimum Inhibitory Concentration of Gentamicin
for *Campylobacter* in Chicken Breast (N=706 Isolates)**
Breakpoints: Susceptible <= 2 $\mu\text{g/mL}$ Resistant >= 8 $\mu\text{g/mL}$



NARMS

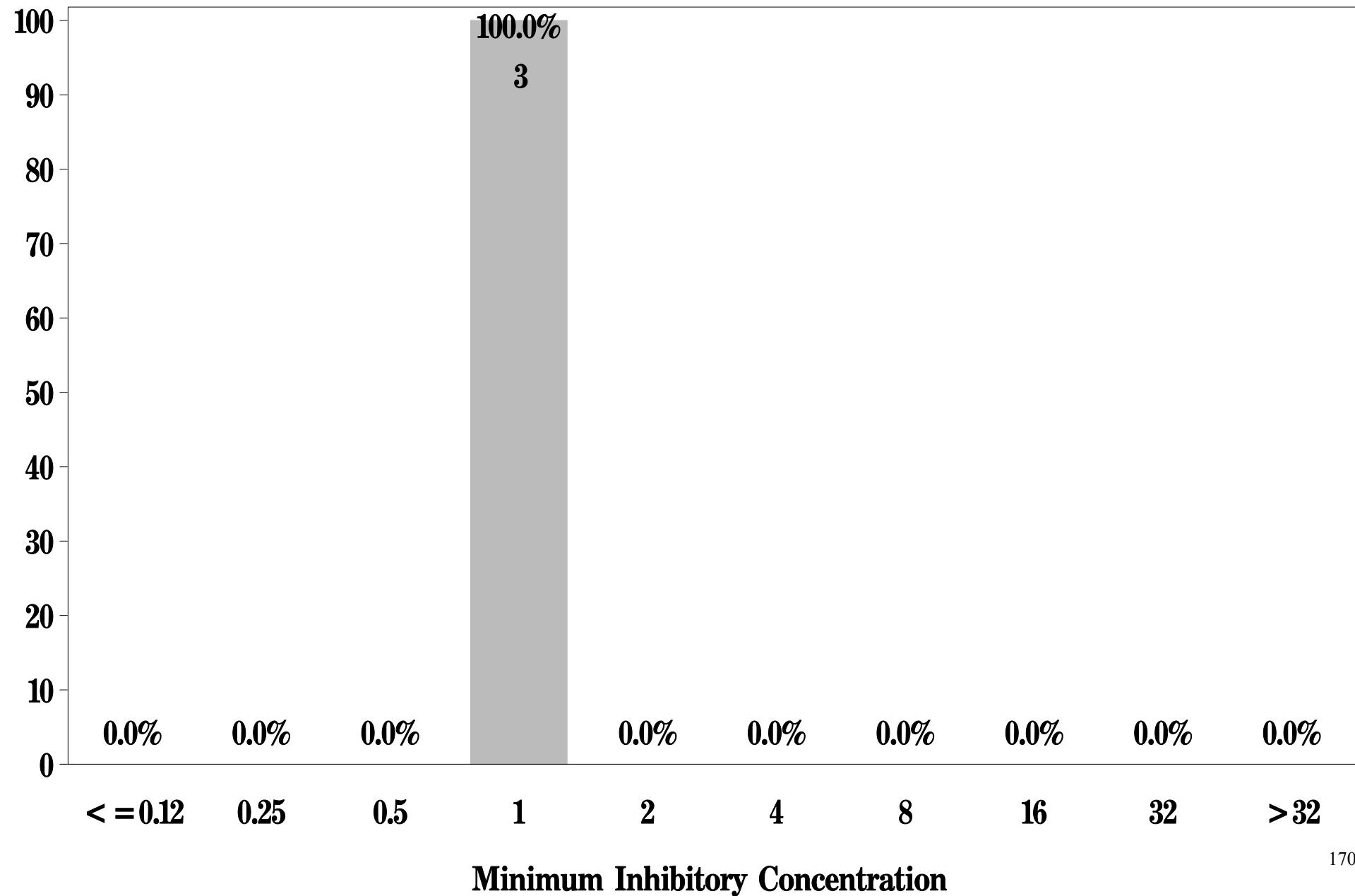
**Figure 11f: Minimum Inhibitory Concentration of Gentamicin
for *Campylobacter* in Ground Turkey (N=12 Isolates)**
Breakpoints: Susceptible < = 2 $\mu\text{g}/\text{mL}$ Resistant > = 8 $\mu\text{g}/\text{mL}$



NARMS

**Figure 11f: Minimum Inhibitory Concentration of Gentamicin
for *Campylobacter* in Pork Chop (N=3 Isolates)**

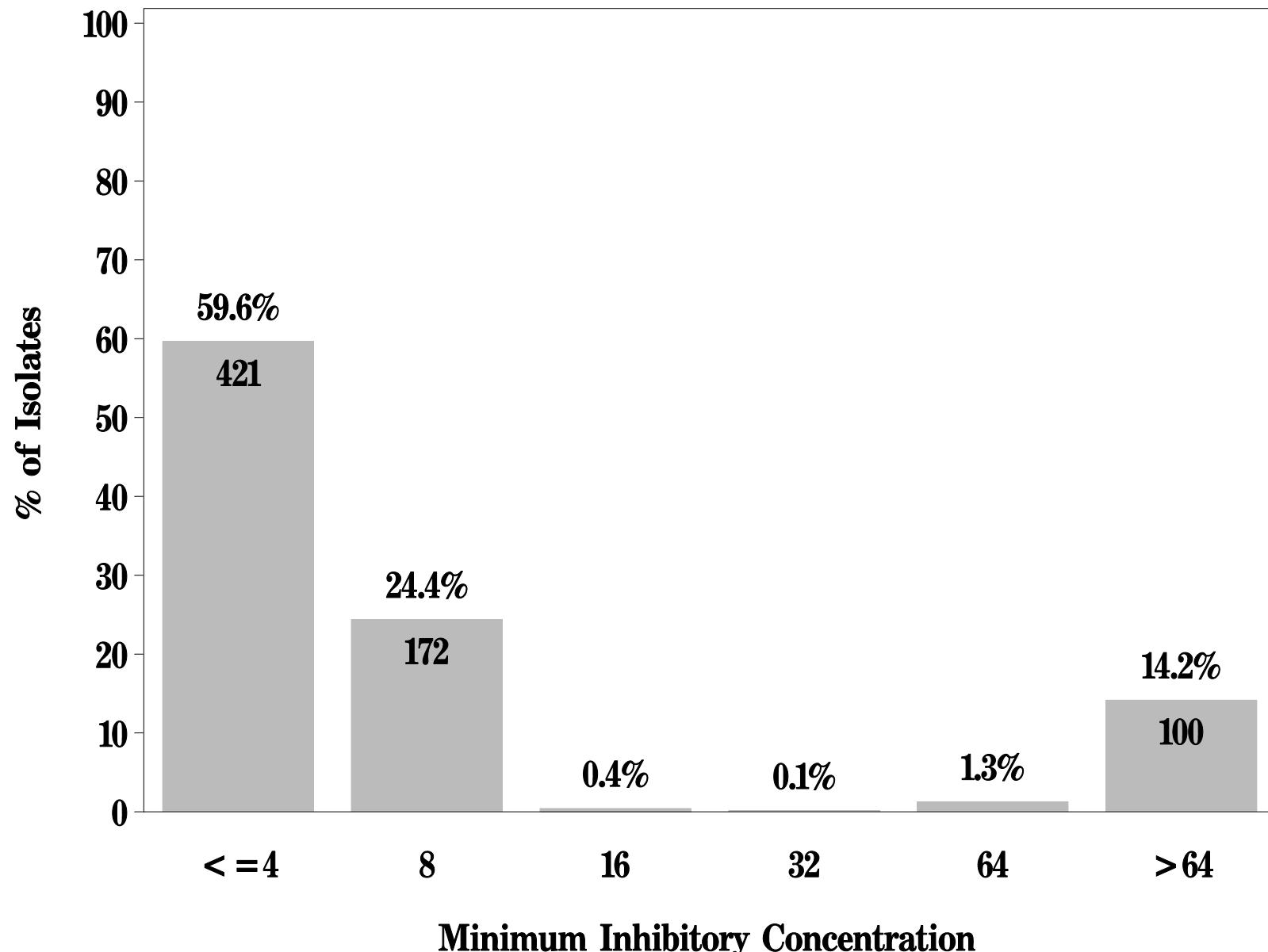
Breakpoints: Susceptible $\leq 2 \mu\text{g/mL}$ Resistant $\geq 8 \mu\text{g/mL}$



NARMS

**Figure 11g: Minimum Inhibitory Concentration of Nalidixic acid
for *Campylobacter* in Chicken Breast (N = 706 Isolates)**

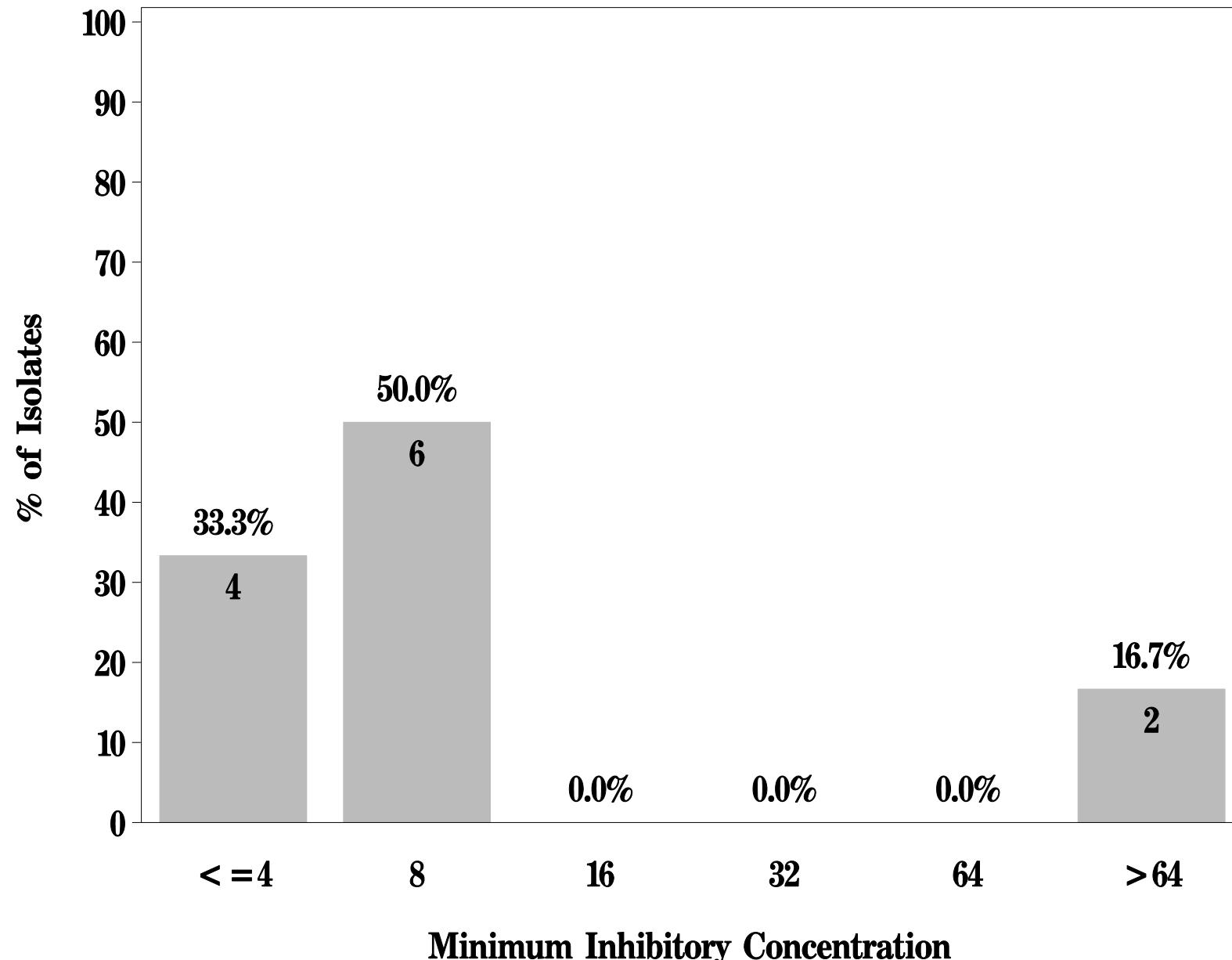
Breakpoints: Susceptible $\leq 16 \mu\text{g/mL}$ Resistant $\geq 64 \mu\text{g/mL}$



NARMS

**Figure 11g: Minimum Inhibitory Concentration of Nalidixic acid
for *Campylobacter* in Ground Turkey (N=12 Isolates)**

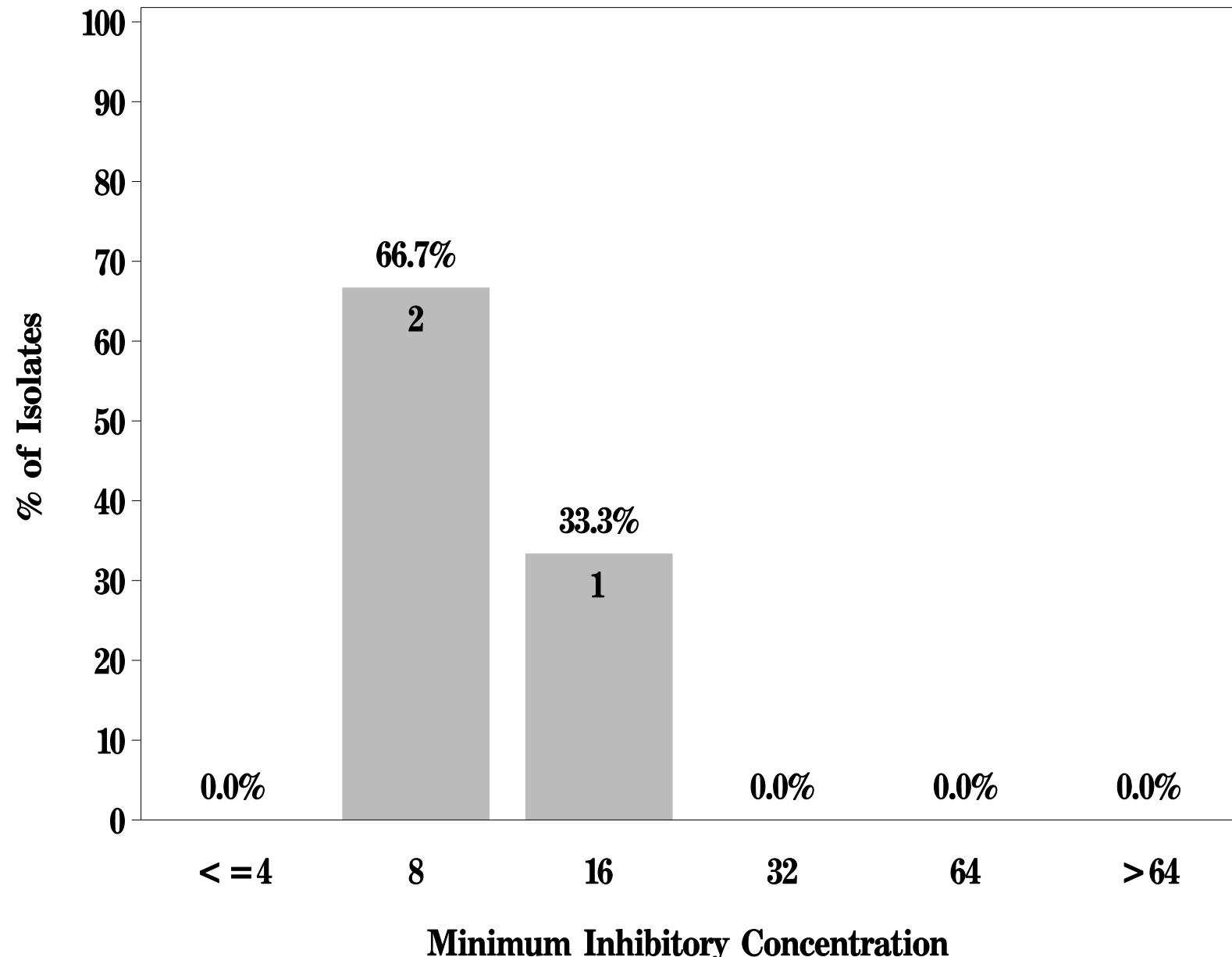
Breakpoints: Susceptible $\leq 16 \mu\text{g/mL}$ Resistant $\geq 64 \mu\text{g/mL}$



NARMS

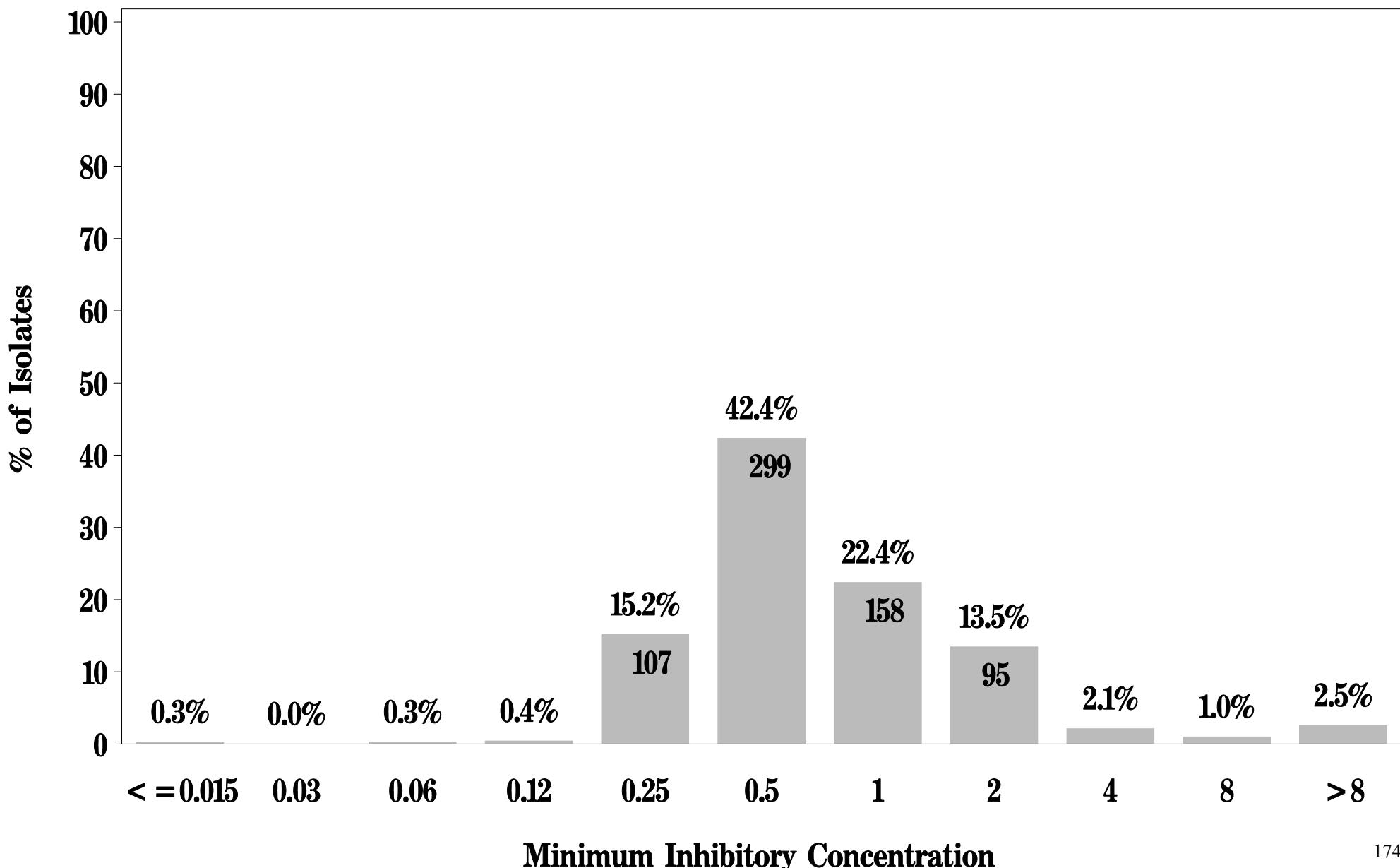
**Figure 11g: Minimum Inhibitory Concentration of Nalidixic acid
for *Campylobacter* in Pork Chop (N=3 Isolates)**

Breakpoints: Susceptible $\leq 16 \mu\text{g/mL}$ Resistant $\geq 64 \mu\text{g/mL}$



NARMS

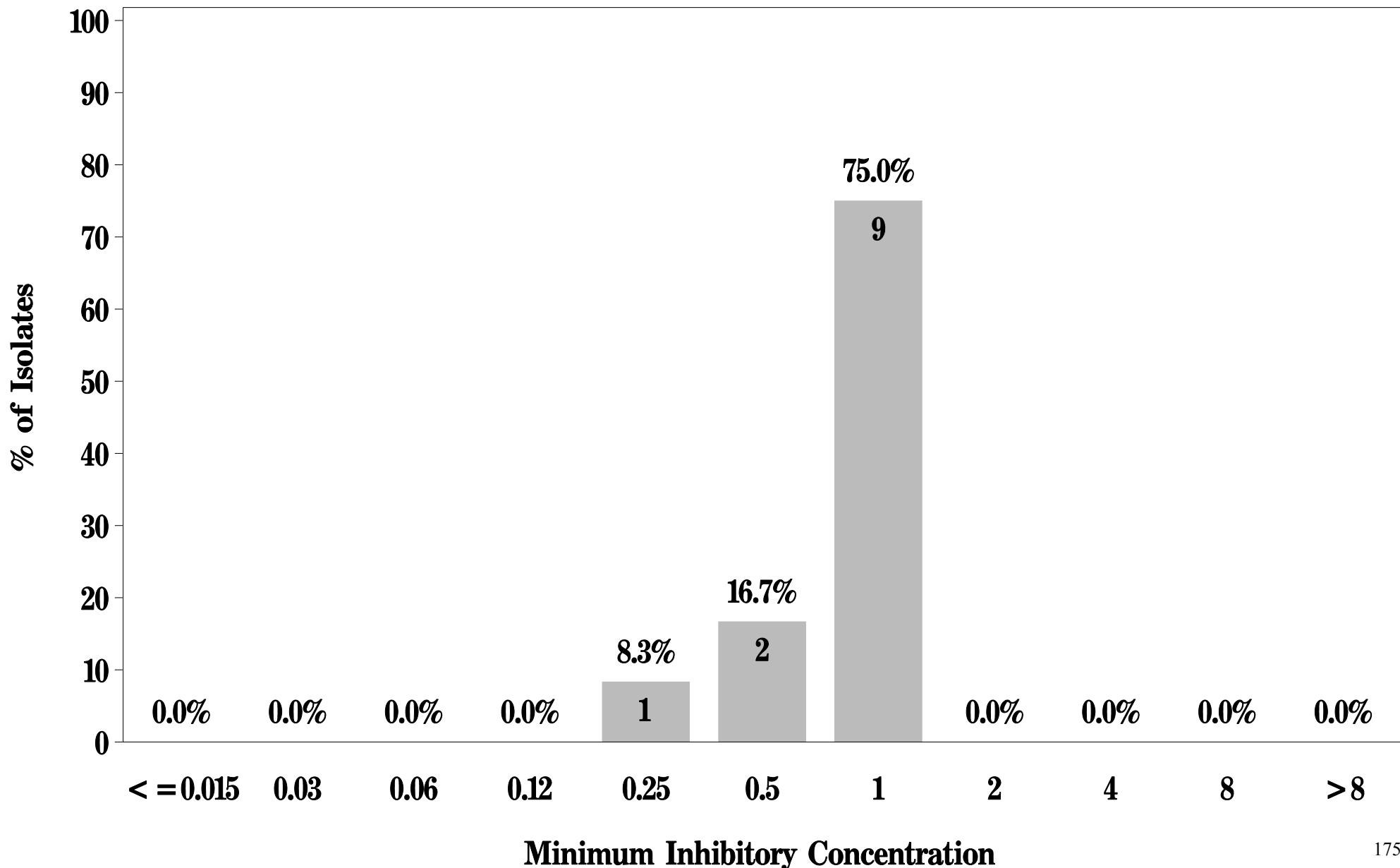
**Figure 11h: Minimum Inhibitory Concentration of Telithromycin
for *Campylobacter* in Chicken Breast (N=706 Isolates)**
Breakpoints: Susceptible <= 4 µg/mL Resistant >= 16 µg/mL



NARMS

**Figure 11h: Minimum Inhibitory Concentration of Telithromycin
for *Campylobacter* in Ground Turkey (N=12 Isolates)**

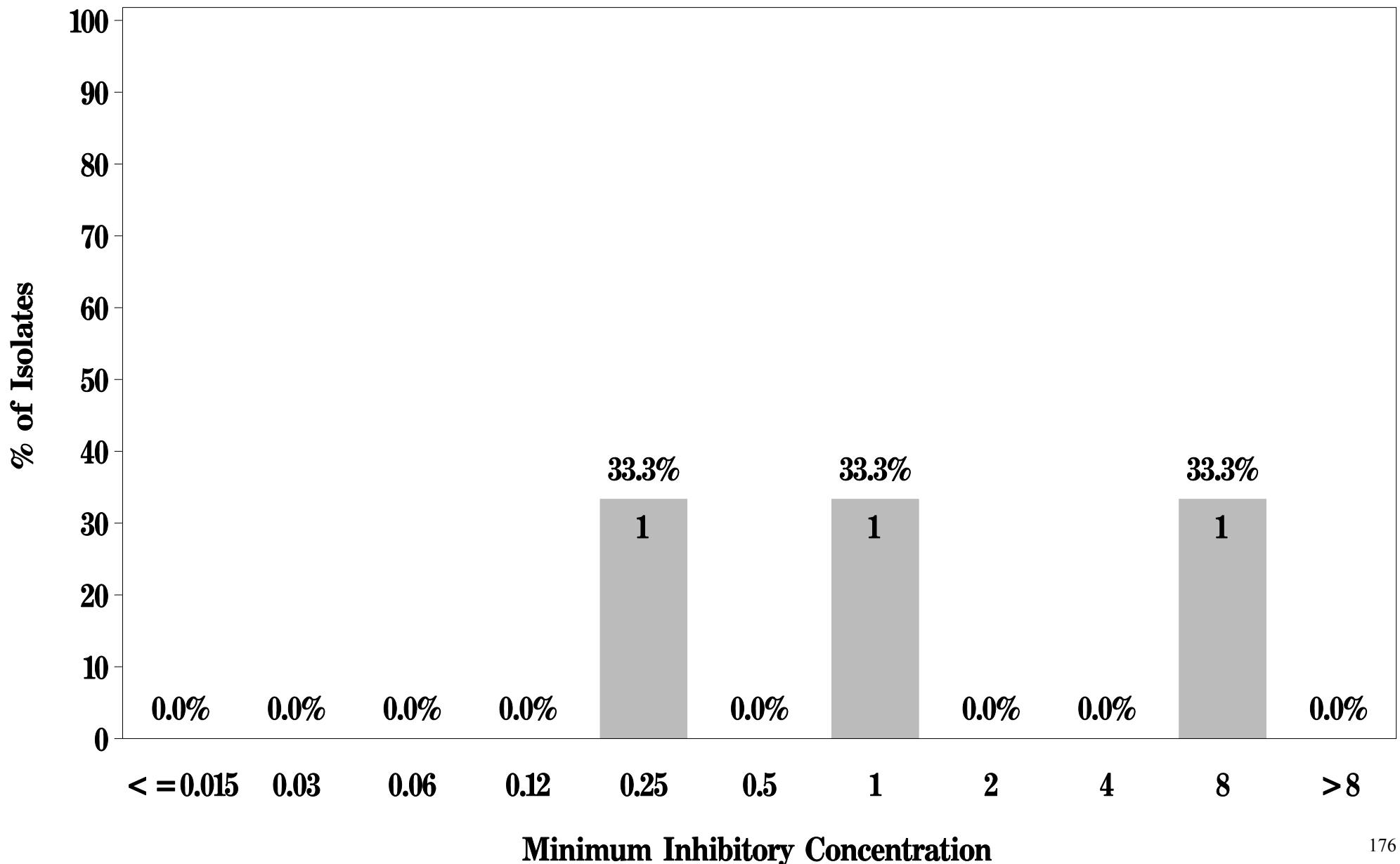
Breakpoints: Susceptible $\leq 4 \mu\text{g/mL}$ Resistant $\geq 16 \mu\text{g/mL}$



NARMS

**Figure 11h: Minimum Inhibitory Concentration of Telithromycin
for *Campylobacter* in Pork Chop (N=3 Isolates)**

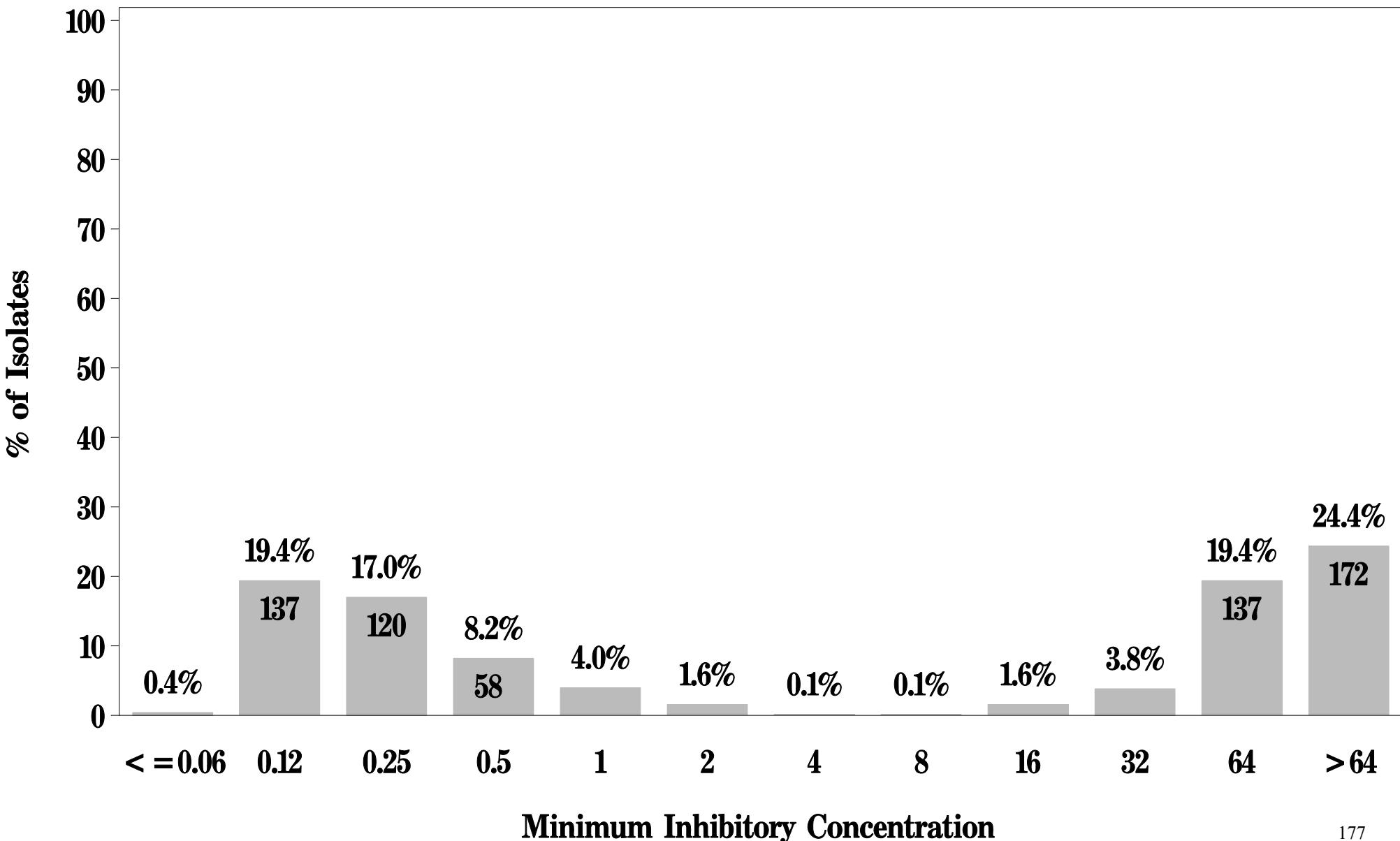
Breakpoints: Susceptible $\leq 4 \mu\text{g/mL}$ Resistant $\geq 16 \mu\text{g/mL}$



NARMS

**Figure 11i: Minimum Inhibitory Concentration of Tetracycline
for *Campylobacter* in Chicken Breast (N = 706 Isolates)**

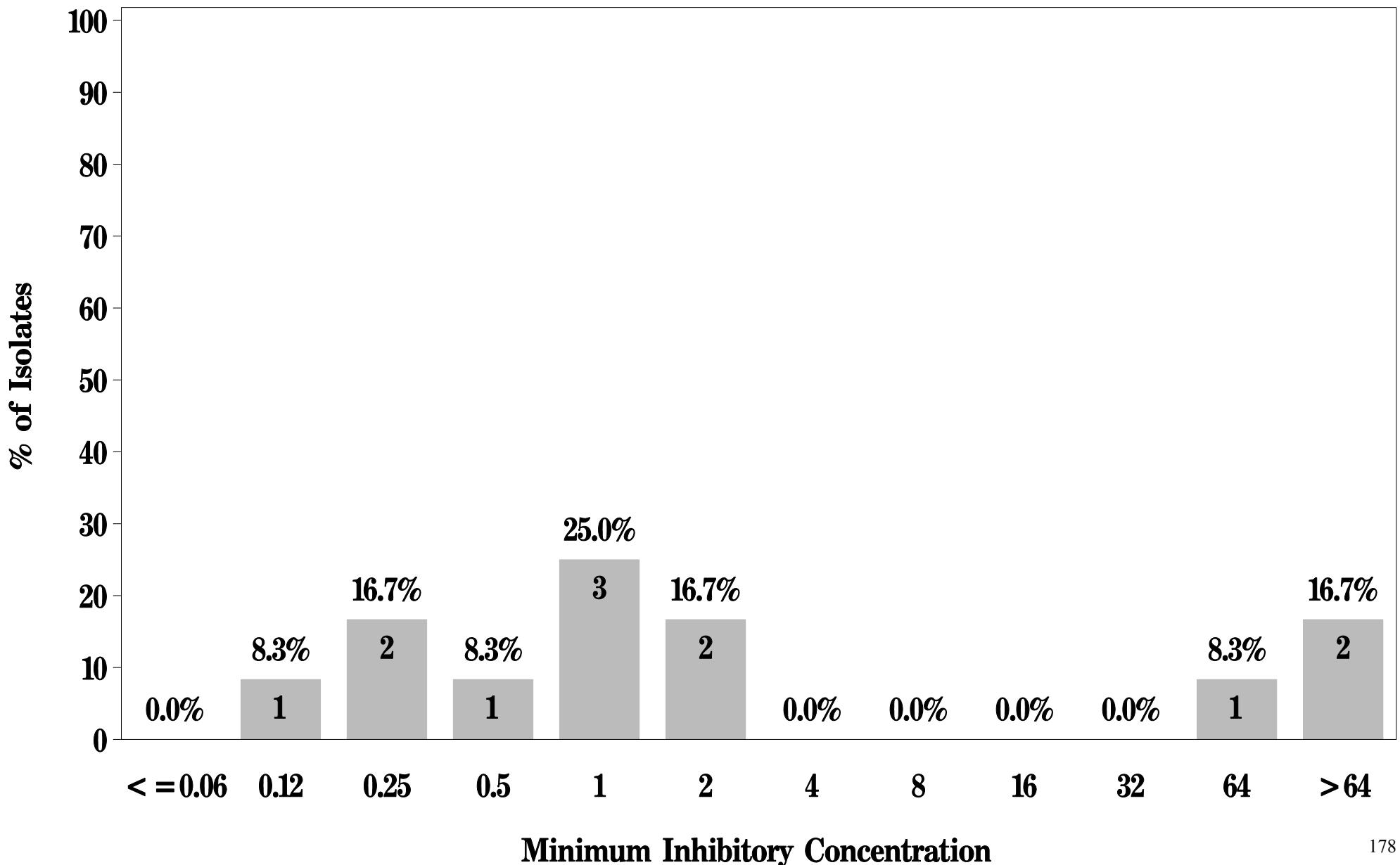
Breakpoints: Susceptible $\leq 4 \mu\text{g/mL}$ Resistant $\geq 16 \mu\text{g/mL}$



NARMS

**Figure 11i: Minimum Inhibitory Concentration of Tetracycline
for *Campylobacter* in Ground Turkey (N=12 Isolates)**

Breakpoints: Susceptible $\leq 4 \mu\text{g/mL}$ Resistant $\geq 16 \mu\text{g/mL}$



NARMS

**Figure 11i: Minimum Inhibitory Concentration of Tetracycline
for *Campylobacter* in Pork Chop (N=3 Isolates)**

Breakpoints: Susceptible $\leq 4 \mu\text{g/mL}$ Resistant $\geq 16 \mu\text{g/mL}$

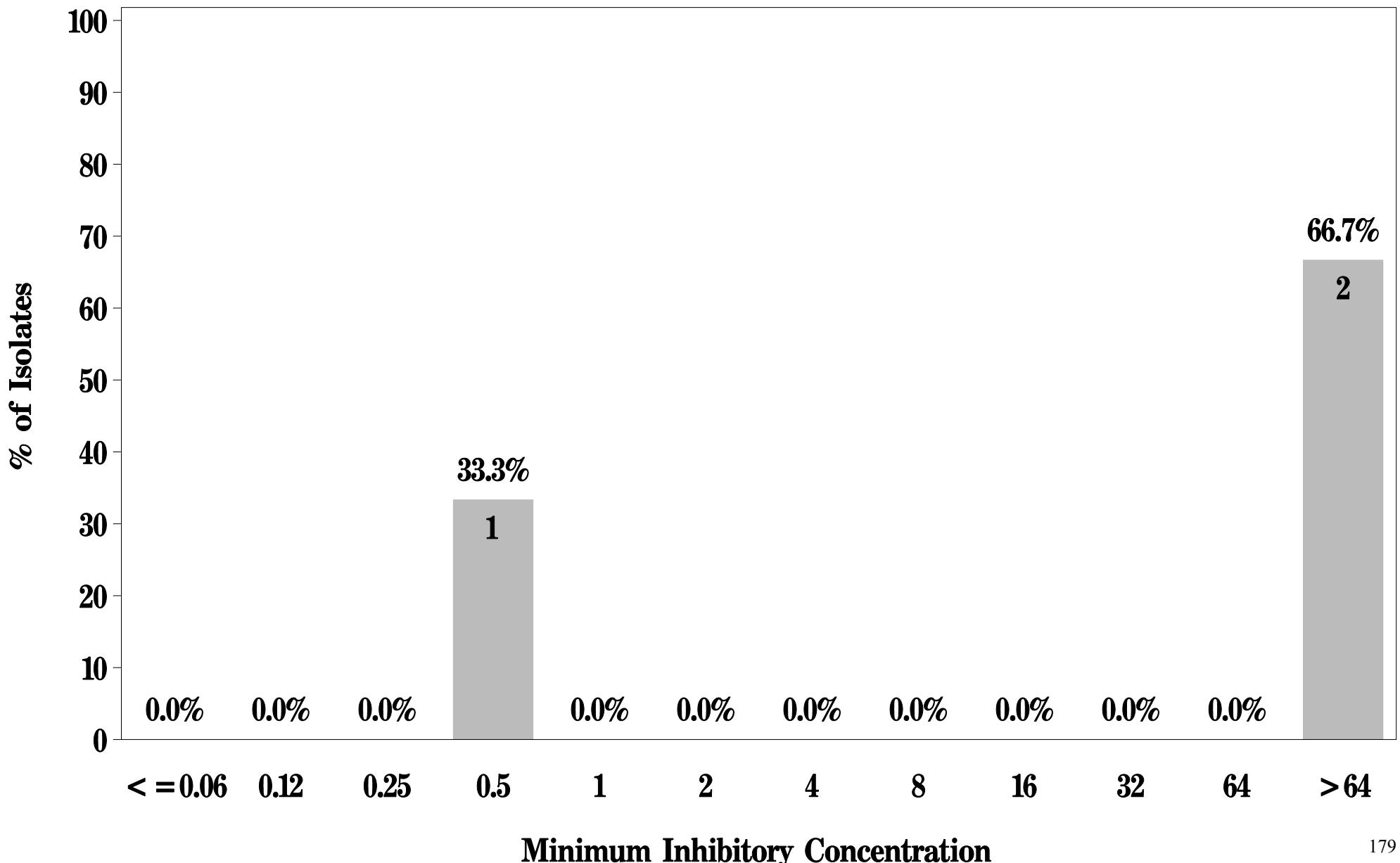


Table 21. Antimicrobial Resistance among *Campylobacter* by Meat Type^{*}, 2004

Antimicrobial Agent	Chicken Breast (n=706)	Ground Turkey (n=12)	Pork Chop (n=3)
Tetracycline	49.2% [†]	25.0%	66.7%
Nalidixic Acid	15.4%	16.7%	- [‡]
Ciprofloxacin	15.4%	16.7%	-
Telithromycin	2.5%	-	-
Azithromycin	3.1%	-	33.3%
Erythromycin	3.1%	-	33.3%
Clindamycin	2.3%	-	33.3%
Florfenicol	-	-	-
Gentamicin	-	-	-

* No *Campylobacter* recovered from ground beef.

† Where % Resistance = (# isolates per meat type resistant to antimicrobial) / (total # isolates per meat type).

‡ Dashes indicate 0.0% resistance to antimicrobial.

Table 22. Antimicrobial Resistance among *Campylobacter* by Species, 2004

Species	Antimicrobial Agent								
	TET	NAL	CIP	TEL	AZI	ERY	CLI	FFN	GEN
<i>C. coli</i> (n=204)	45.6%*	15.7%	15.7%	7.8%	9.3%	9.3%	7.4%	-	-†
<i>C. jejuni</i> (n=517)	50.1%	15.3%	15.3%	0.4%	0.8%	0.8%	0.4%	-	-
Total %R (N=721)	48.8%	15.4%	15.4%	2.5%	3.2%	3.2%	2.4%	0.0%	0.0%

* Where % Resistance = (# isolates per species resistant to antimicrobial) / (total # isolates per species).

† Dashes indicate 0.0% resistance to antimicrobial.

Table 23. Antimicrobial Resistance among *Campylobacter* Species by Meat Type, 2004

Meat Type*	Species	Antimicrobial Agent								
		TET	NAL	CIP	TEL	AZI	ERY	CLI	FFN	GEN
Chicken Breast	<i>C. coli</i> (n=196)	46.4% [†]	16.3%	16.3%	8.2%	9.2%	9.2%	7.1%	-	-
	<i>C. jejuni</i> (n=510)	50.2%	15.1%	15.1%	0.4%	0.8%	0.8%	0.4%	-	-
Ground Turkey	<i>C. coli</i> (n=5)	- [‡]	-	-	-	-	-	-	-	
	<i>C. jejuni</i> (n=7)	42.9%	28.6%	28.6%	-	-	-	-	-	
Pork Chop	<i>C. coli</i> (n=3)	66.7%	-	-	-	33.3%	33.3%	33.3%	-	-
	<i>C. jejuni</i> (n=0)	§								

* No *Campylobacter* recovered from ground beef.

† Where % Resistance = (# isolates per species resistant to antimicrobial within meat type) / (total # isolates per species within meat type).

‡ Where dashes indicate 0.0% resistance to antimicrobial.

§ Grey areas indicate species not isolated from that meat type.

Table 24. Antimicrobial Resistance among *Campylobacter* by Site, Meat Type, and Antimicrobial Agent, 2004

Site	Meat Type*	Antimicrobial Agent								
		TET	NAL	CIP	TEL	AZI	ERY	CLI	FFN	GEN
CA	CB (n=96)	54.2% [†]	6.3%	6.3%	-	-	-	-	-	-
	GT (n=0)	‡								
	PC (n=1)	100.0%	§	-	-	-	-	-	-	-
	Total (n=97)	54.6%	6.2%	6.2%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%
CO	CB (n=21)	38.1%	19.0%	19.0%	4.8%	4.8%	4.8%	4.8%	-	-
	GT (n=0)									
	PC (n=0)									
	Total (n=21)	38.1%	19.0%	19.0%	4.8%	4.8%	4.8%	4.8%	0.0%	0.0%
CT	CB (n=86)	65.1%	27.9%	27.9%	2.3%	2.3%	2.3%	2.3%	-	-
	GT (n=2)	100.0%	50.0%	50.0%	-	-	-	-		
	PC (n=1)	100.0%	-	-	-	100.0%	100.0%	100.0%	-	-
	Total (n=89)	66.3%	28.1%	28.1%	2.3%	3.4%	3.4%	3.4%	0.0%	0.0%
GA	CB (n=61)	39.3%	13.1%	13.1%	6.6%	8.2%	8.2%	6.6%	-	-
	GT (n=0)									
	PC (n=0)									
	Total (n=62)	38.7%	12.9%	12.9%	6.6%	8.1%	8.1%	6.5%	0.0%	0.0%
MD	CB (n=76)	43.4%	27.6%	27.6%	-	1.3%	1.3%	-	-	-
	GT (n=2)	-	50.0%	50.0%	-	-	-	-		
	PC (n=0)									
	Total (n=78)	42.3%	28.2%	28.2%	0.0%	1.3%	1.3%	0.0%	0.0%	0.0%

* No *Campylobacter* recovered from ground beef.

† Where % Resistance = (# isolates resistant to antimicrobial per meat type per site) / (total # isolates per meat type per site).

‡ Grey areas indicate species not isolated from that meat type.

§ Where dashes indicate 0.0% resistance to antimicrobial.

Table 24_(cont'd). Antimicrobial Resistance among *Campylobacter* by Site, Meat Type, and Antimicrobial Agent, 2004

Site	Meat Type	Antimicrobial Agent								
		TET	NAL	CIP	TEL	AZI	ERY	CLI	FFN	GEN
MN	CB (n=73)	68.5%	4.1%	4.1%	2.7%	2.7%	2.7%	2.7%	-	-
	GT (n=6)	16.7%	-	-	-	-	-	-	-	-
	PC (n=0)									
	Total (n=79)	64.6%	3.8%	3.8%	2.5%	2.5%	2.5%	2.5%	0.0%	0.0%
NM	CB (n=53)	35.8%	5.7%	5.7%	11.3%	11.3%	11.3%	9.4%	-	-
	GT (n=0)									
	PC (n=1)									
	Total (n=54)	35.2%	5.6%	5.6%	11.1%	11.1%	11.1%	9.4%	0.0%	0.0%
NY	CB (n=96)	40.6%	35.4%	35.4%	-	-	-	-	-	-
	GT (n=0)									
	PC (n=0)									
	Total (n=96)	40.6	35.4%	35.4%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%
OR	CB (n=73)	52.1%	-	-	-	1.4%	1.4%	-	-	-
	GT (n=0)									
	PC (n=0)									
	Total (n=73)	52.1%	0.0%	0.0%	0.0%	1.4%	1.4%	0.0%	0.0%	0.0%
TN	CB (n=71)	39.4%	8.5%	8.5%	4.2%	5.6%	5.6%	2.8%	-	-
	GT (n=0)									
	PC (n=0)									
	Total (n=72)	38.9%	8.3%	8.3%	4.2%	5.6%	5.6%	2.8%	0.0%	0.0%
Total %R (N=721)		48.8%	15.5%	15.4%	3.6%	3.2%	3.2%	2.4%	0.0%	0.0%

Table 25. Number of *Campylobacter* (N=721) Resistant to Multiple Antimicrobial Agents, 2004

Meat Type	Number of Antimicrobials			
	0	1	2-4	≥5
Chicken Breast	284	292	129	1
Ground Turkey	8	2	2	0
Pork Chop	1	1	1	0
Total	293	295	132	1

Table 26. Overall *Enterococcus* Species Identified, 2004

Species	n
1. <i>E. faecalis</i>	855
2. <i>E. faecium</i>	757
3. <i>E. hirae</i>	129
4. <i>E. gallinarum</i>	7
5. <i>E. durans</i>	3
6. <i>E. casseliflavus</i>	3
7. <i>E. mundtii</i>	1
Total	1755

Table 27. *Enterococcus* Species by Meat Type, 2004

Species	Chicken Breast		Ground Turkey		Ground Beef		Pork Chop	
	n	%*	n	%	n	%	n	%
<i>E. faecalis</i> (n=855)	88	10.3%	260	30.4%	194	22.7%	313	36.6%
<i>E. faecium</i> (n=757)	348	46.0%	172	22.7%	162	21.4%	75	9.9%
<i>E. hirae</i> (n=129)	27	20.9%	-	-	88	68.2%	14	10.9%
<i>E. gallinarium</i> (n=7)	-†	-	4	57.1%	2	28.6%	1	14.3%
<i>E. durans</i> (n=3)	2	66.7%	1	33.3%	-	-	-	-
<i>E. casseliflavus</i> (n=3)	-	-	-	-	2	66.7%	1	33.3%
<i>E. mundtii</i> (n=1)	1	100.0%	-	-	-	-	-	-
Total (N=1755)	466	26.6%	437	24.9%	448	25.5%	404	23.0%

* Where % = (# isolates per species per meat) / (total # isolates per species).

† Dashes indicate no isolates of that species were isolated from that meat type.

Table 28. *Enterococcus* Species by Site and Meat Type, 2004

Site	Species	Chicken Breast		Ground Turkey		Ground Beef		Pork Chop	
		n	%*	n	%	n	%	n	%
GA	<i>E. faecalis</i> (n=346)	54	15.6%	108	31.2%	78	22.5%	106	30.6%
	<i>E. faecium</i> (n=100)	59	59.0%	11	11.0%	22	22.2%	8	8.0%
	<i>E. hirae</i> (n=23)	5	21.2%	- [†]	-	16	69.6%	2	8.7%
	<i>E. durans</i> (n=2)	1	50.0%	1	50.0%	-	-	-	-
	<i>E. casseliflavus</i> (n=1)	-	-	-	-	1	100.0%	-	-
	<i>E. mundtii</i> (n=1)	1	100.0%	-	-	-	-	-	-
	Total (n=473)	120	25.1%	120	25.3%	117	24.7%	116	24.5%
MD	<i>E. faecalis</i> (n=110)	3	2.9%	30	29.4%	21	20.6%	48	47.1%
	<i>E. faecium</i> (n=233)	106	41.1%	75	29.1%	54	20.9%	23	8.9%
	<i>E. hirae</i> (n=43)	5	14.3%	-	-	24	68.6%	6	17.1%
	<i>E. gallinarum</i> (n=5)	-	-	1	50.0%	1	50.0%	-	-
	Total (n=397)	114	28.7%	106	26.7%	100	25.2%	77	19.4%
OR	<i>E. faecalis</i> (n=201)	18	9.0%	67	33.3%	37	18.4%	79	39.3%
	<i>E. faecium</i> (n=181)	85	47.0%	35	19.3%	37	20.4%	24	13.3%
	<i>E. hirae</i> (n=58)	15	25.9%	-	-	39	67.2%	4	6.9%
	<i>E. gallinarum</i> (n=4)	-	-	3	75.0%	1	25.0%	-	-
	<i>E. casseliflavus</i> (n=2)	-	-	-	-	1	50.0%	1	50.0%
	Total (n=446)	118	26.5%	105	23.5%	115	25.8%	108	24.2%
TN	<i>E. faecalis</i> (n=206)	13	6.3%	55	26.7%	58	28.2%	80	38.8%
	<i>E. faecium</i> (n=218)	98	45.0%	51	23.4%	49	22.5%	20	9.2%
	<i>E. hirae</i> (n=13)	2	15.4%	-	-	9	69.2%	2	15.4%
	<i>E. gallinarum</i> (n=1)	-	-	-	-	-	-	1	100.0%
	<i>E. durans</i> (n=1)	1	100.0%	-	-	-	-	-	-
	Total (n=439)	114	26.0%	106	24.1%	116	26.4%	103	23.5%

* Where % = (# isolates per species per meat type per site) / (total # isolates per species per site).

† Dashes indicate no isolates for that species were isolated from that meat type.

Table 29. *Enterococcus* Isolates by Month for All Sites, 2004

Month	n	%*
January	148	8.4%
February	144	8.2%
March	140	8.0%
April	156	8.9%
May	153	8.7%
June	136	7.7%
July	135	7.7%
August	148	8.4%
September	154	8.8%
October	141	8.0%
November	148	8.4%
December	152	8.7%
Total (N)	1755	100.0%

* Where % = (n / N).

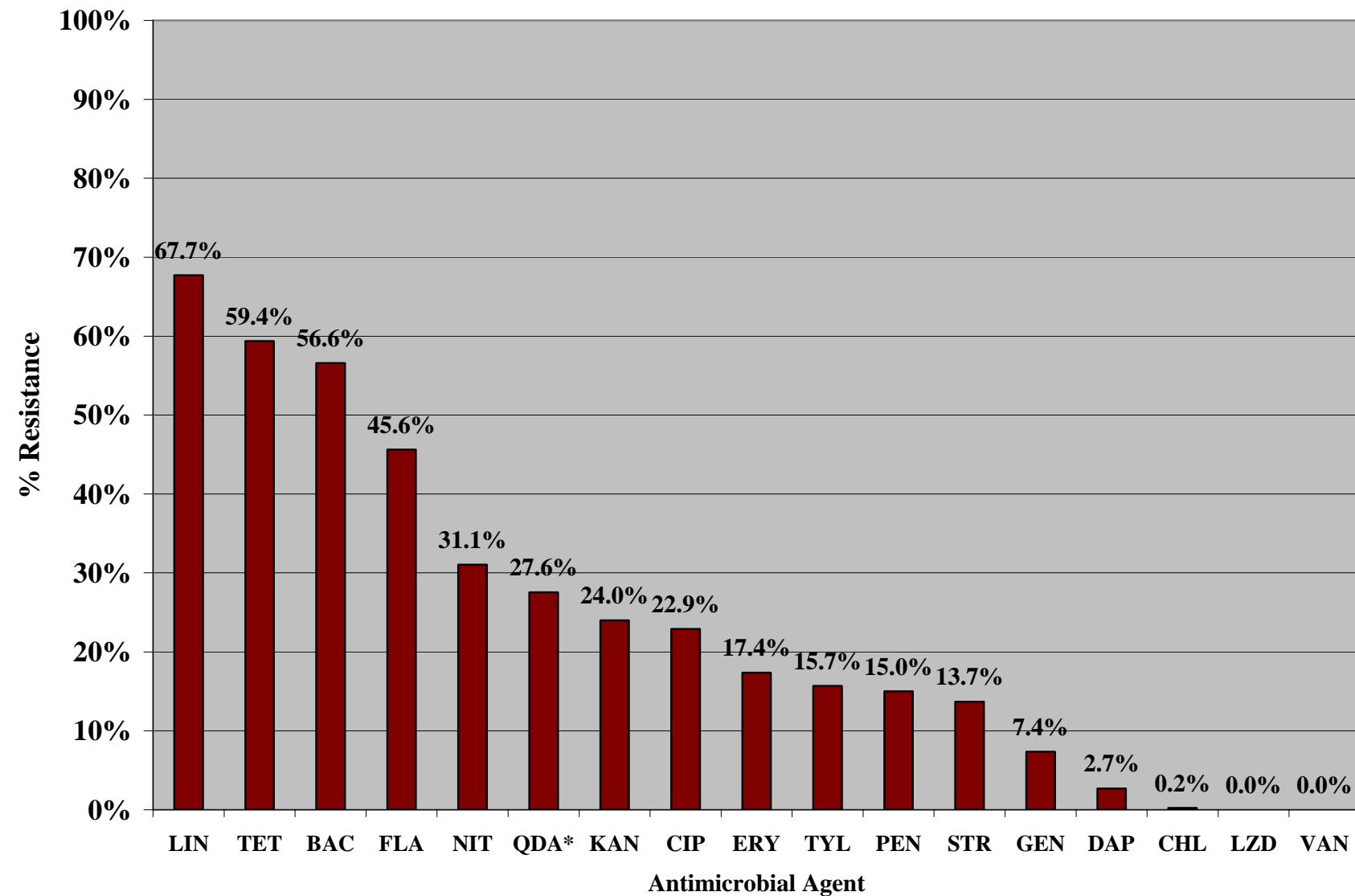
Table 30. Antimicrobial Resistance among *Enterococcus* Isolates (N=1755), 2004

Antimicrobial Agent	n	%R*
Lincomycin	1188	67.7%
Tetracycline	1042	59.4%
Bacitracin	993	56.6%
Flavomycin	801	45.6%
Quinupristin-Dalfopristin†	248	27.6%
Nitrofurantoin	545	31.1%
Kanamycin	421	24.0%
Ciprofloxacin	402	22.9%
Erythromycin	305	17.4%
Tylosin	275	15.7%
Penicillin	263	15.0%
Streptomycin	240	13.7%
Gentamicin	129	7.4%
Daptomycin	48	2.7%
Chloramphenicol	4	0.2%
Linezolid	0	0.0%
Vancomycin	0	0.0%

* Where % R = (n / N).

† Presented for all species except *E. faecalis* (n = 855).

Figure 12. Antimicrobial Resistance among *Enterococcus* Isolates (N=1755), 2004



* Presented for all species except *E. faecalis* in QDA (N=1755-855=900 non-*faecalis*)

Figure 13. MIC Distribution among all Antimicrobial Agents

<i>Enterococcus</i> from All Meats (N=1755)		Distribution (%) of MICs (in µg/ml)																		
Antimicrobial Agent	%R†	0.015	0.03	0.06	0.125	0.25	0.5	1	2	4	8	16	32	64	128	256	512	1024	2048	>2048
Bacitracin*	56.6										7.3	3.1	7.5	25.6	17.4	39.1				
Chloramphenicol	0.2								0.1	4.6	88.4	6.8	0.2	0.1						
Ciprofloxacin	22.9				0.3	0.1	5.3	30.3	41.1	18.3	4.6									
Daptomycin*	§							4.1	37.8	24.2	31.2	2.1	0.6							
Erythromycin	17.4							40.2	23.7	12.0	6.8	1.2	16.2							
Tylosin*	15.7							0.2	4.2	29.1	41.7	8.9	0.3	0.1	15.6					
Gentamicin	7.4													92.2	0.5	0.6	0.5	6.4		
Kanamycin*	24.0													64.2	11.8	11.2	1.1	11.7		
Streptomycin*	13.7														86.4	3.5	3.8	6.5		
Lincomycin*	67.7							9.5	0.5	0.7	5.7	16.0	34.5	33.2						
Linezolid	0.0							0.2	1.5	88.5	9.9									
Nitrofurantoin	31.1									0.1	18.6	29.1	4.3	17.0	31.0					
Flavomycin*	45.6							45.5	2.8	1.0	2.3	2.7	1.5	44.2						
Penicillin	15.0							7.0	3.7	26.4	43.8	4.2	7.8	7.2						
Tetracycline	59.4									38.8	1.8	1.2	4.4	53.8						
Quinupristin/Dalfopristin†	27.4							24.7	47.7	7.4	13.1	5.9	1.1							
Vancomycin	0.0							32.7	44.3	21.2	1.4	0.5								

Vertical bars show the CLSI Susceptible/Resistant breakpoints for each drug where appropriate.

*Currently no CLSI breakpoints have been established for this organism/antimicrobial combination. Indicated breakpoints were established by NARMS.

†Discrepancies between %R and sums of distribution %s are due to rounding.

Unshaded areas indicate the dilution ranges of the Sensititre plate used to test the 2004 isolates.

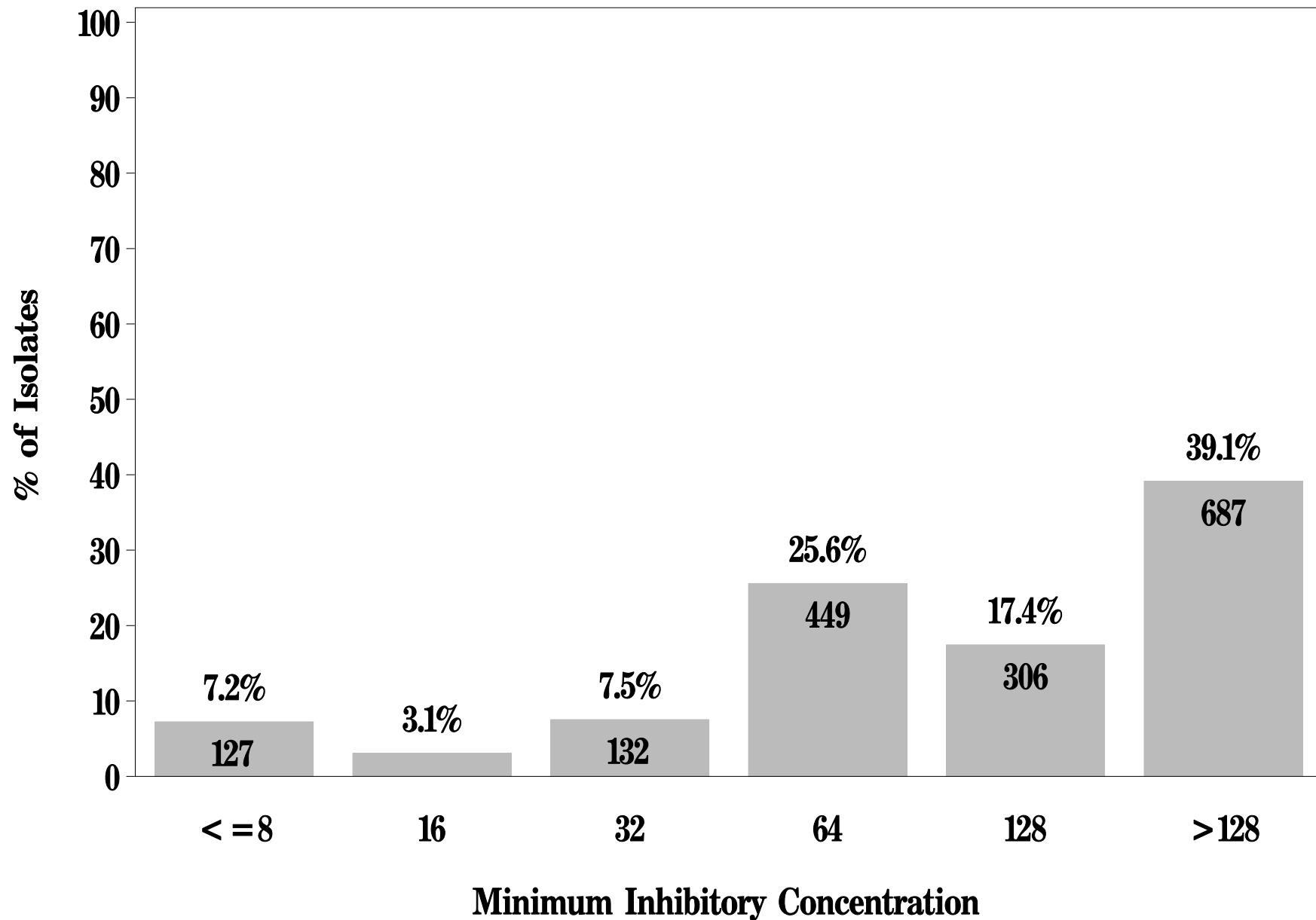
↑ Presented for all species except *E. faecalis* in QDA (n=1755-855= 900 non *E. faecalis*)

§Absence of resistant strains precludes defining any results category other than “susceptible.”

NARMS

**Figure 13a: Minimum Inhibitory Concentration of Bacitracin
for *Enterococcus* (N=1755 Isolates)**

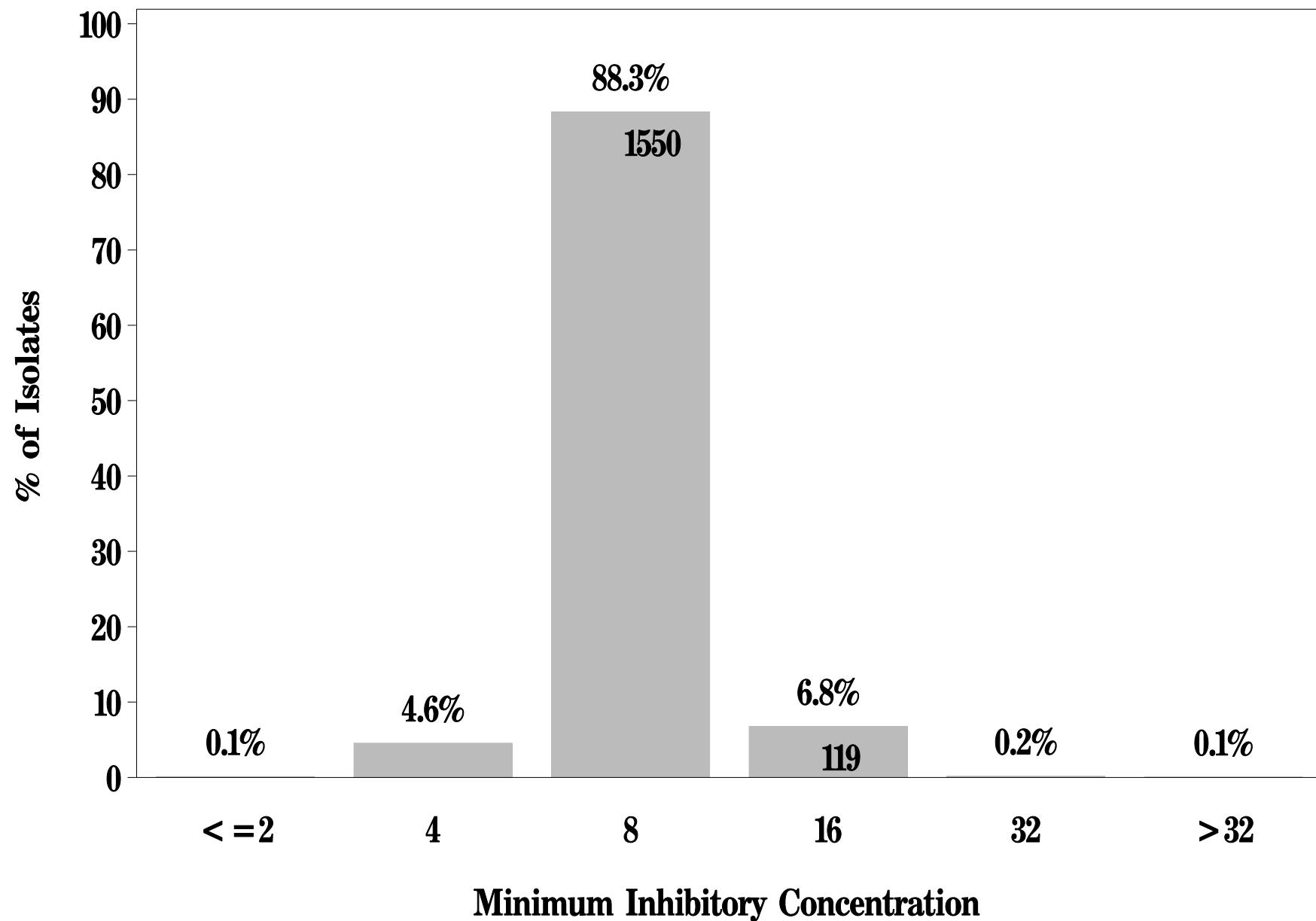
Breakpoints: Susceptible $\leq 32 \text{ } \mu\text{g/mL}$ Resistant $\geq 128 \text{ } \mu\text{g/mL}$



NARMS

**Figure 13b: Minimum Inhibitory Concentration of Chloramphenicol
for *Enterococcus* (N=1755 Isolates)**

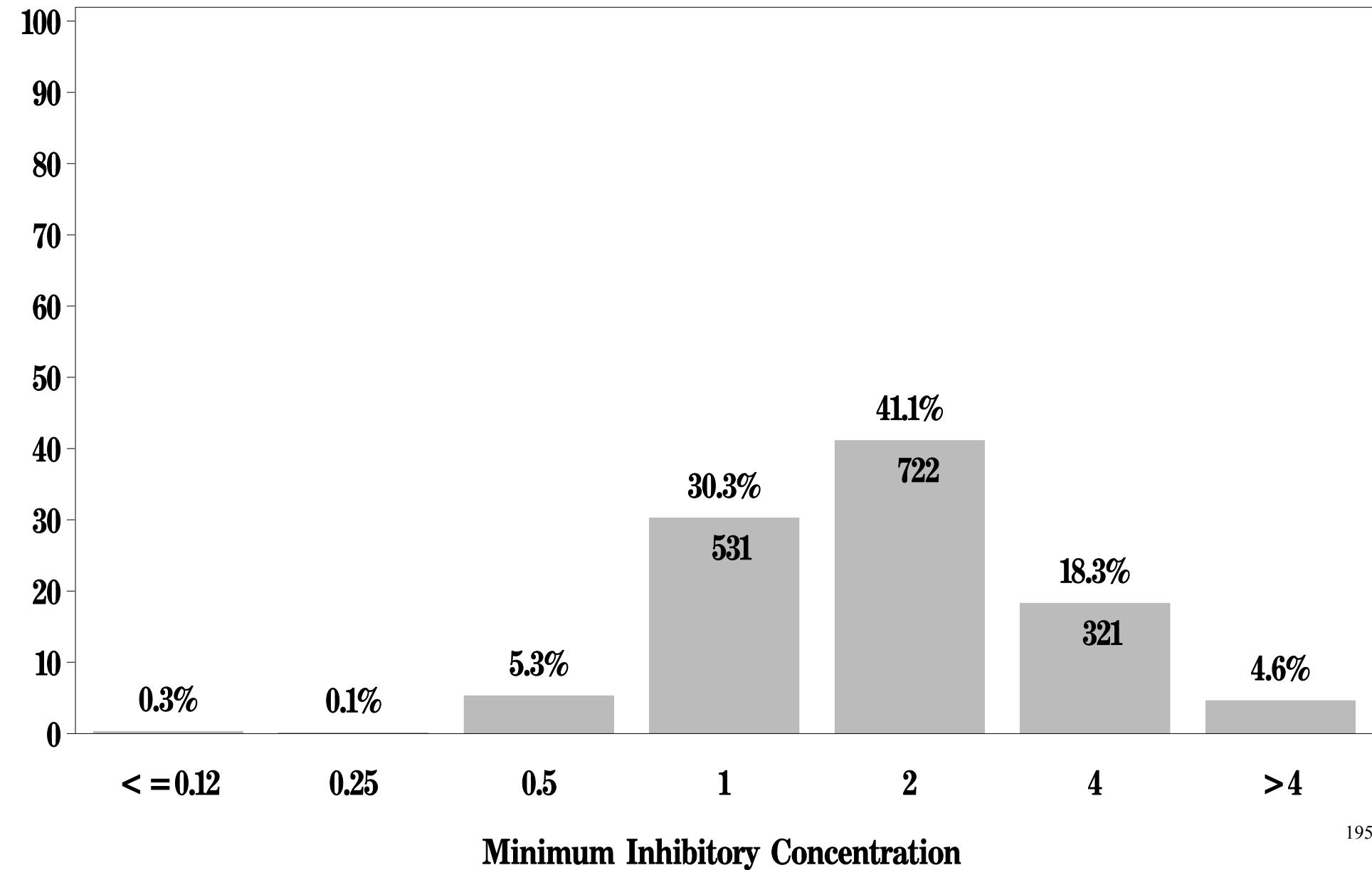
Breakpoints: Susceptible $\leq 8 \text{ } \mu\text{g/mL}$ Resistant $\geq 32 \text{ } \mu\text{g/mL}$



NARMS

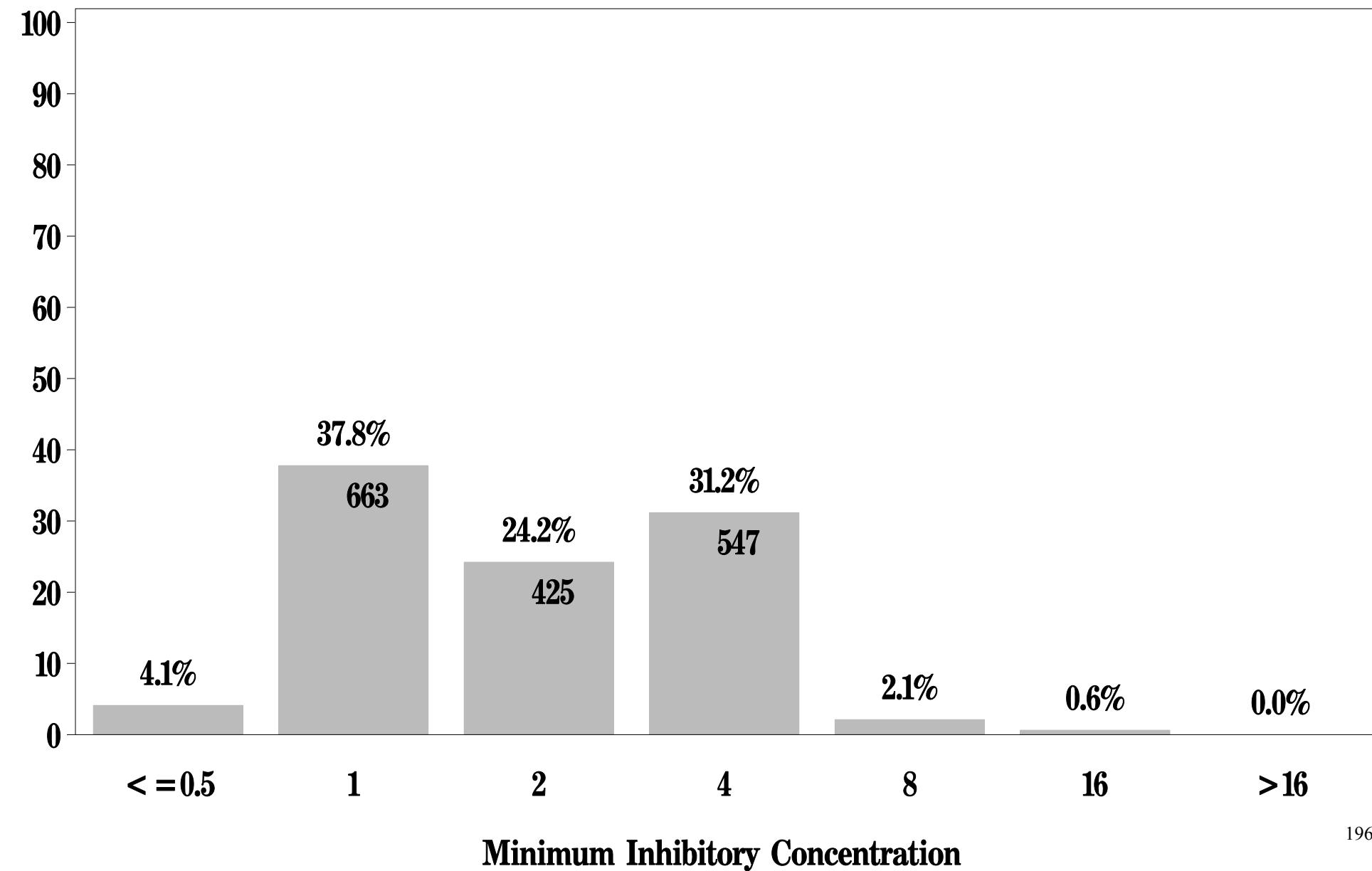
**Figure 13c: Minimum Inhibitory Concentration of Ciprofloxacin
for *Enterococcus* (N=1755 Isolates)**

Breakpoints: Susceptible $\leq 1 \text{ } \mu\text{g/mL}$ Resistant $\geq 4 \text{ } \mu\text{g/mL}$



NARMS

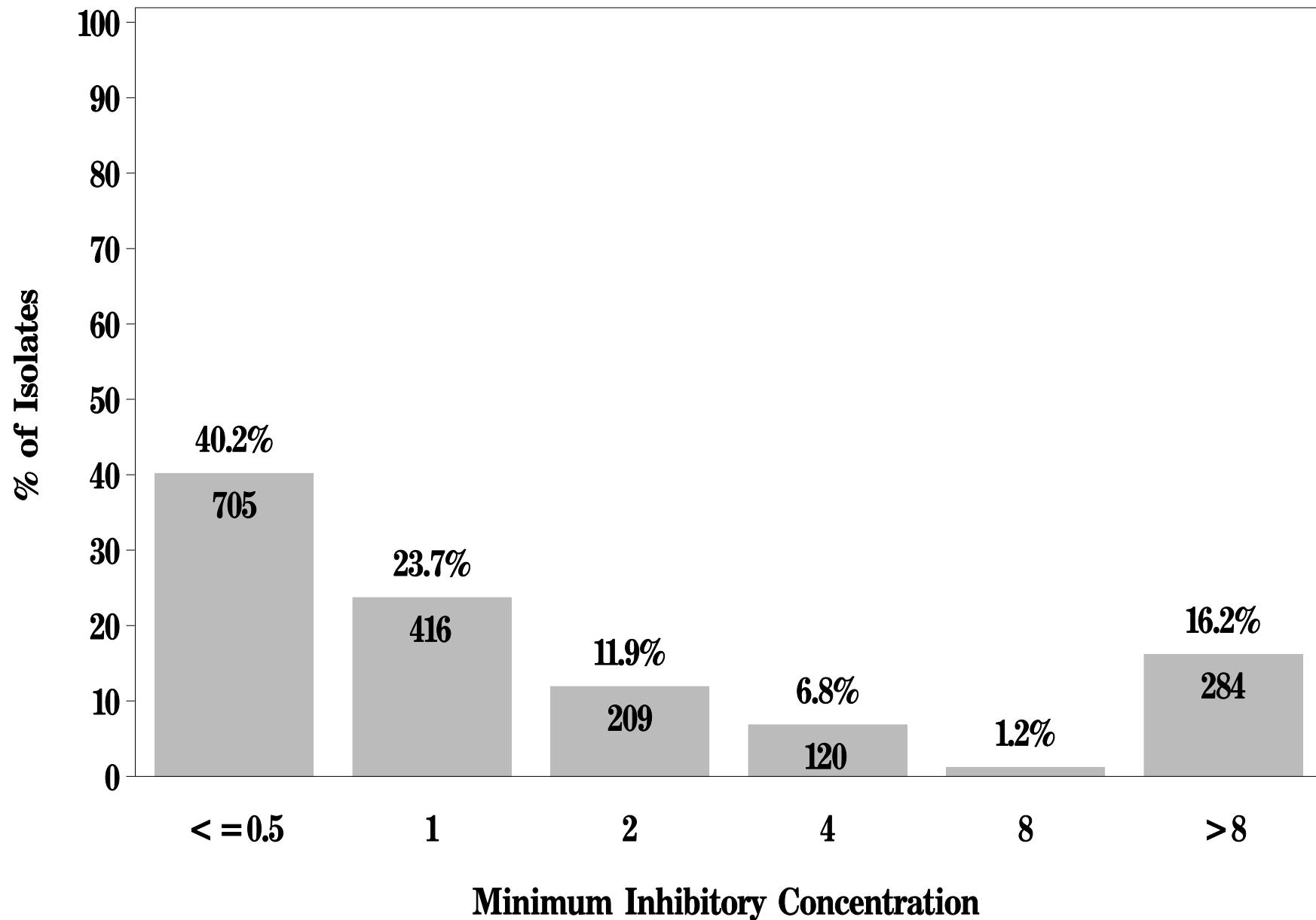
**Figure 13d: Minimum Inhibitory Concentration of Daptomycin
for *Enterococcus* (N=1755 Isolates)**
Breakpoint: Susceptible $\leq 4 \mu\text{g/mL}$



NARMS

Figure 13e: Minimum Inhibitory Concentration of Erythromycin
for *Enterococcus* (N=1755 Isolates)

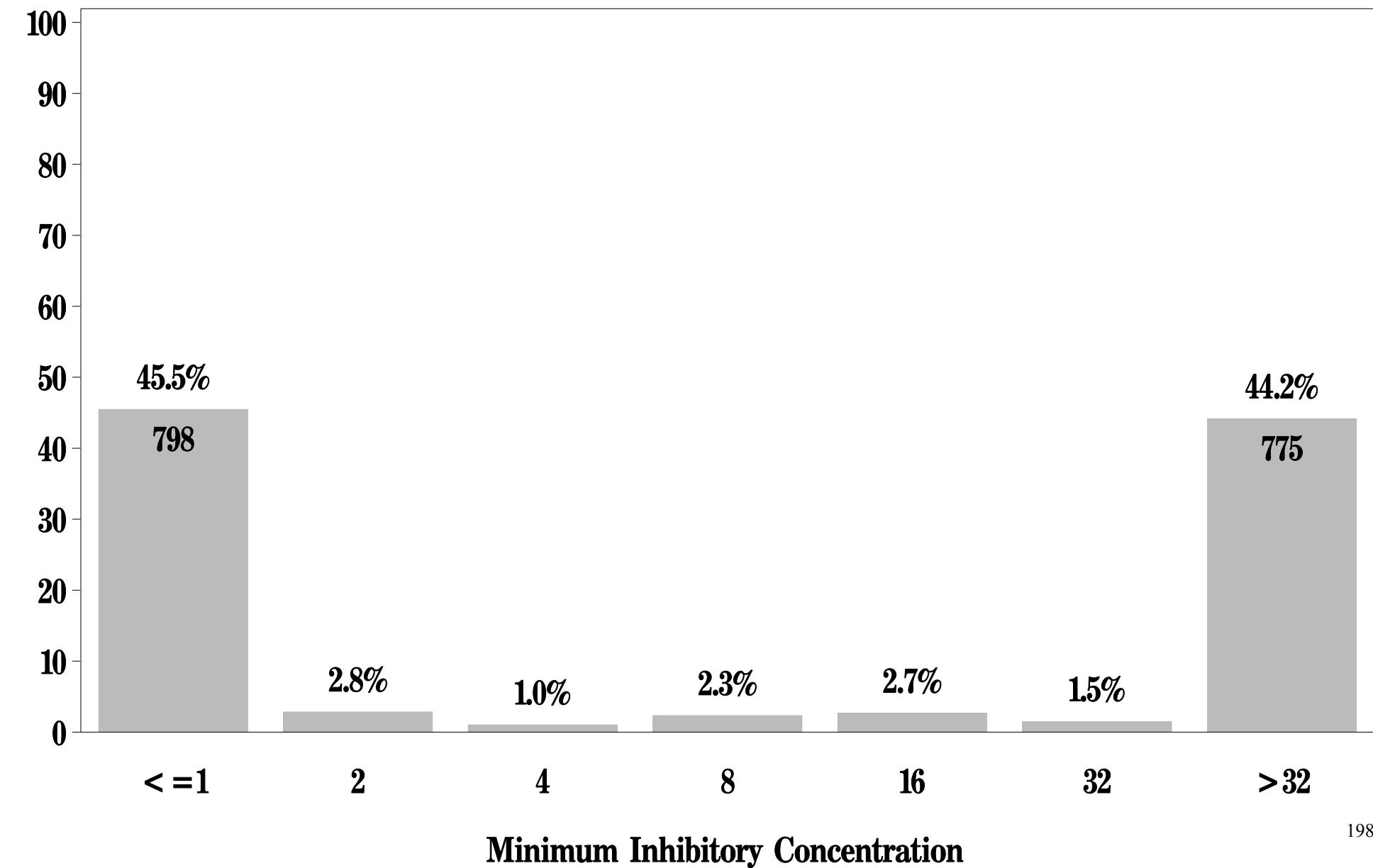
Breakpoints: Susceptible $\leq 0.5 \text{ } \mu\text{g/mL}$ Resistant $\geq 8 \text{ } \mu\text{g/mL}$



NARMS

Figure 13f: Minimum Inhibitory Concentration of Flavomycin
for *Enterococcus* (N=1755 Isolates)

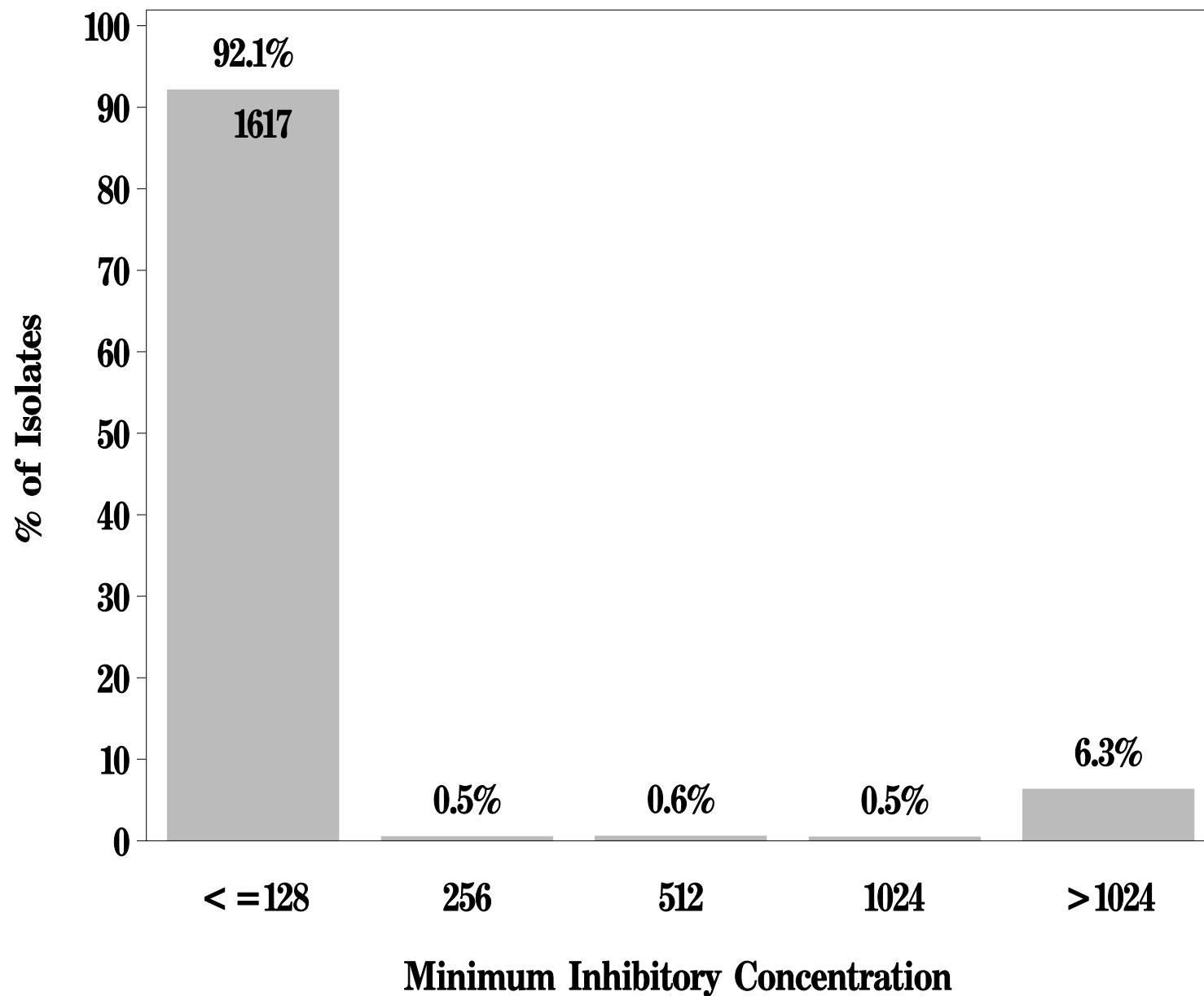
Breakpoints: Susceptible $\leq 8 \text{ } \mu\text{g/mL}$ Resistant $\geq 32 \text{ } \mu\text{g/mL}$



NARMS

**Figure 13g: Minimum Inhibitory Concentration of Gentamicin
for *Enterococcus* (N=1755 Isolates)**

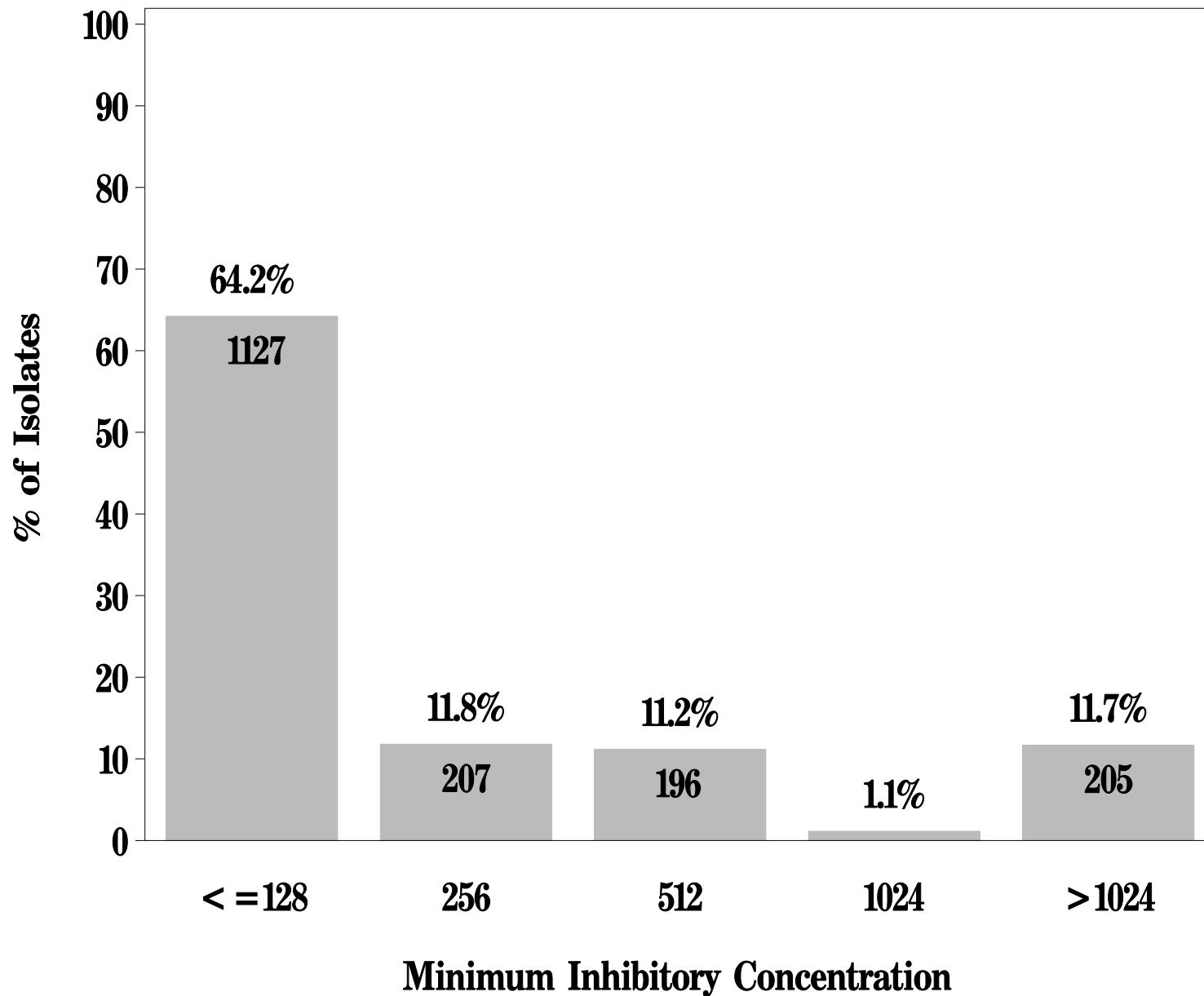
Breakpoints: Susceptible $\leq 500 \mu\text{g/mL}$ Resistant $\geq 500 \mu\text{g/mL}$



NARMS

Figure 13h: Minimum Inhibitory Concentration of Kanamycin
for *Enterococcus* (N=1755 Isolates)

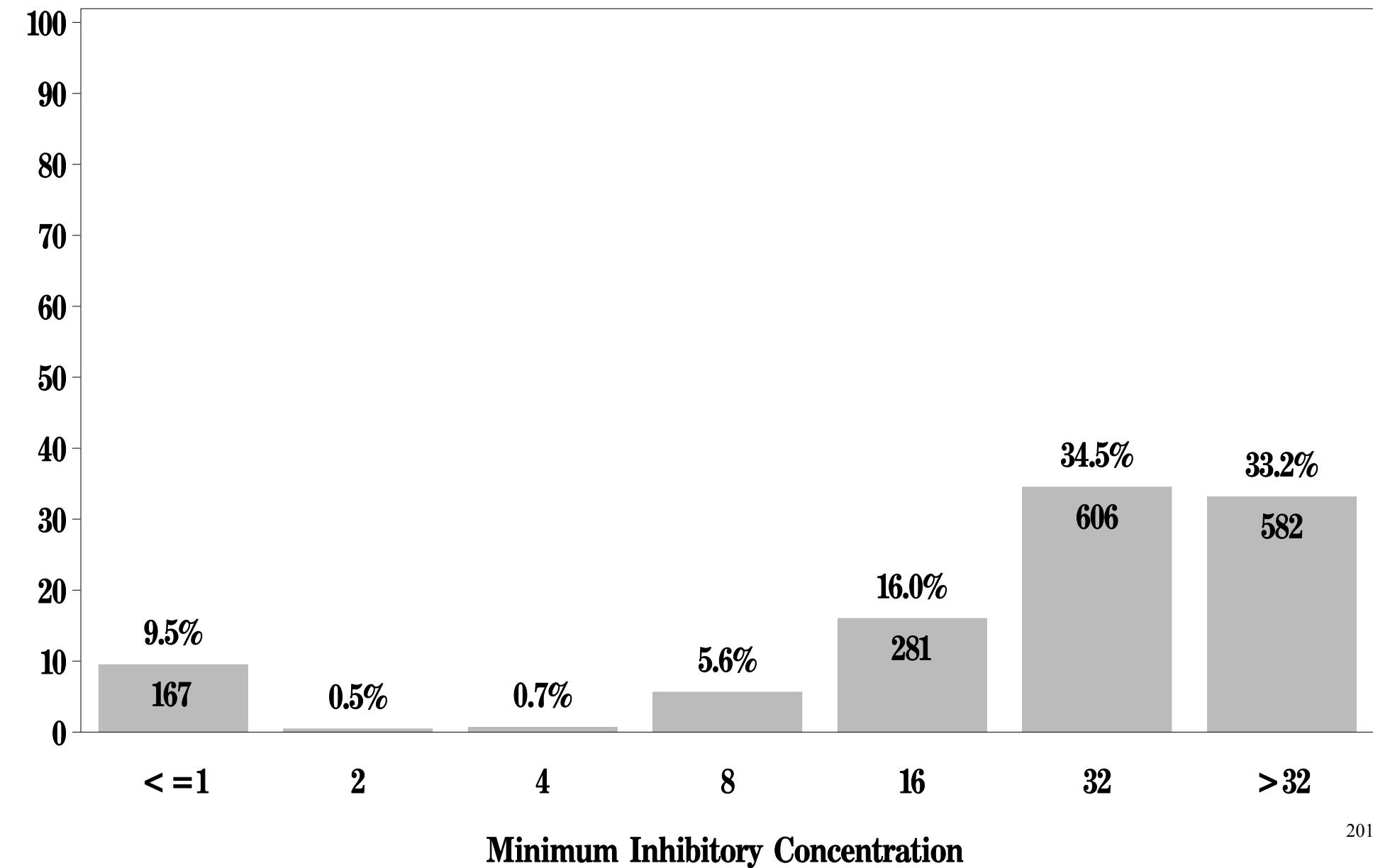
Breakpoints: Susceptible $\leq 128 \mu\text{g/mL}$ Resistant $\geq 512 \mu\text{g/mL}$



NARMS

Figure 13i: Minimum Inhibitory Concentration of Lincomycin
for *Enterococcus* (N=1755 Isolates)

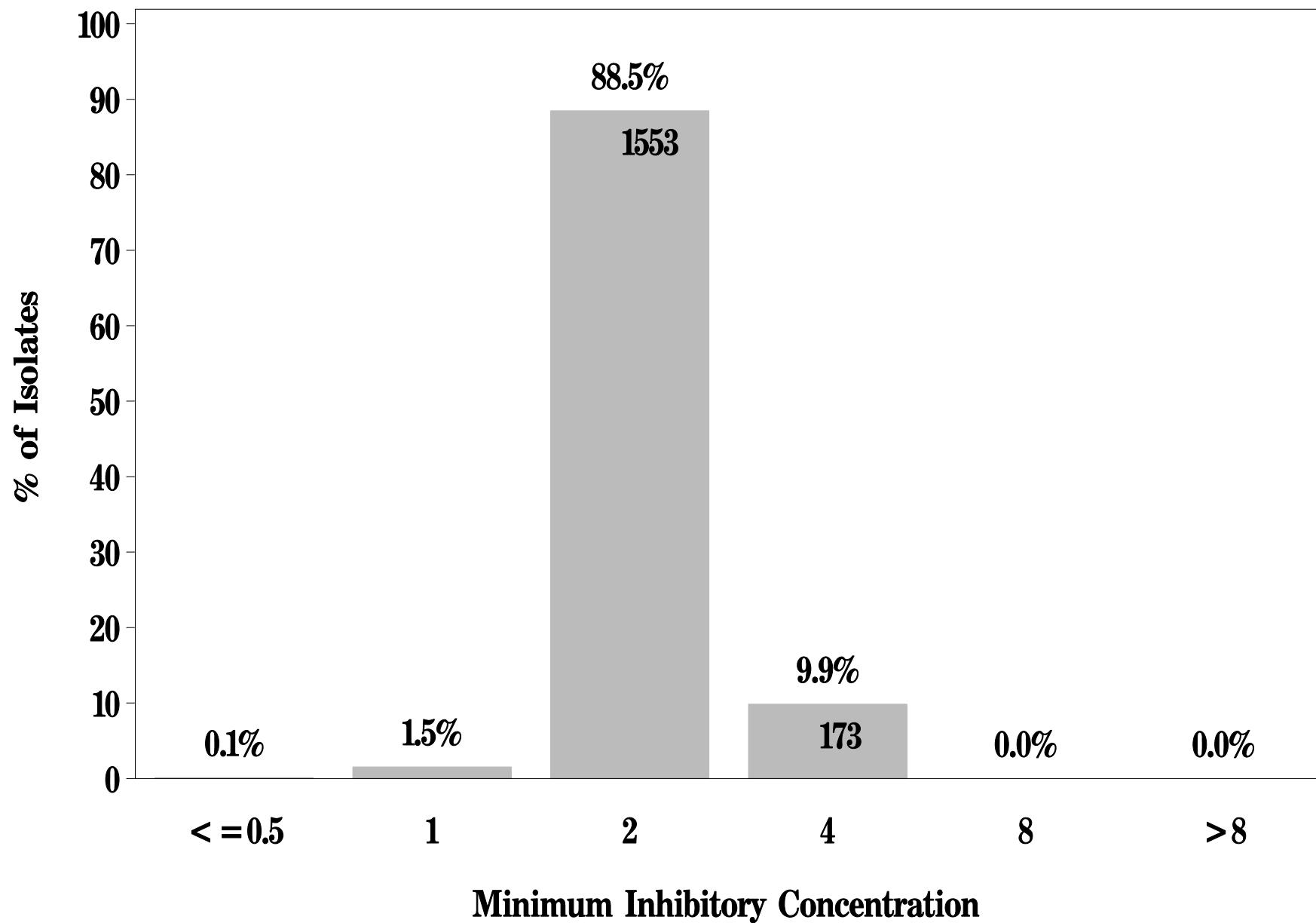
Breakpoints: Susceptible $\leq 8 \text{ } \mu\text{g/mL}$ Resistant $\geq 32 \text{ } \mu\text{g/mL}$



NARMS

**Figure 13j: Minimum Inhibitory Concentration of Linezolid
for *Enterococcus* (N=1755 Isolates)**

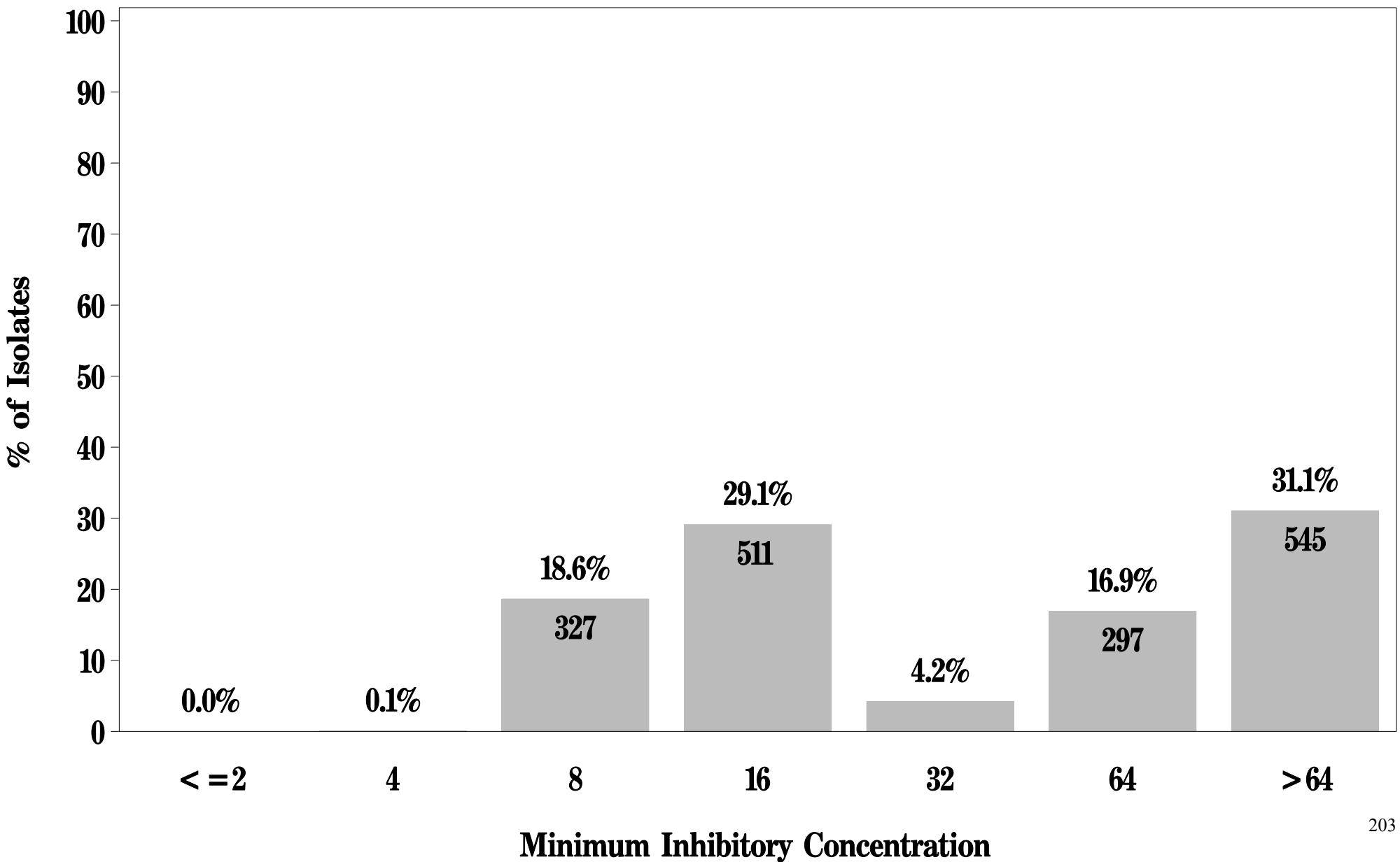
Breakpoints: Susceptible $\leq 2 \text{ } \mu\text{g/mL}$ Resistant $\geq 8 \text{ } \mu\text{g/mL}$



NARMS

**Figure 13k: Minimum Inhibitory Concentration of Nitrofurantoin
for *Enterococcus* (N=1755 Isolates)**

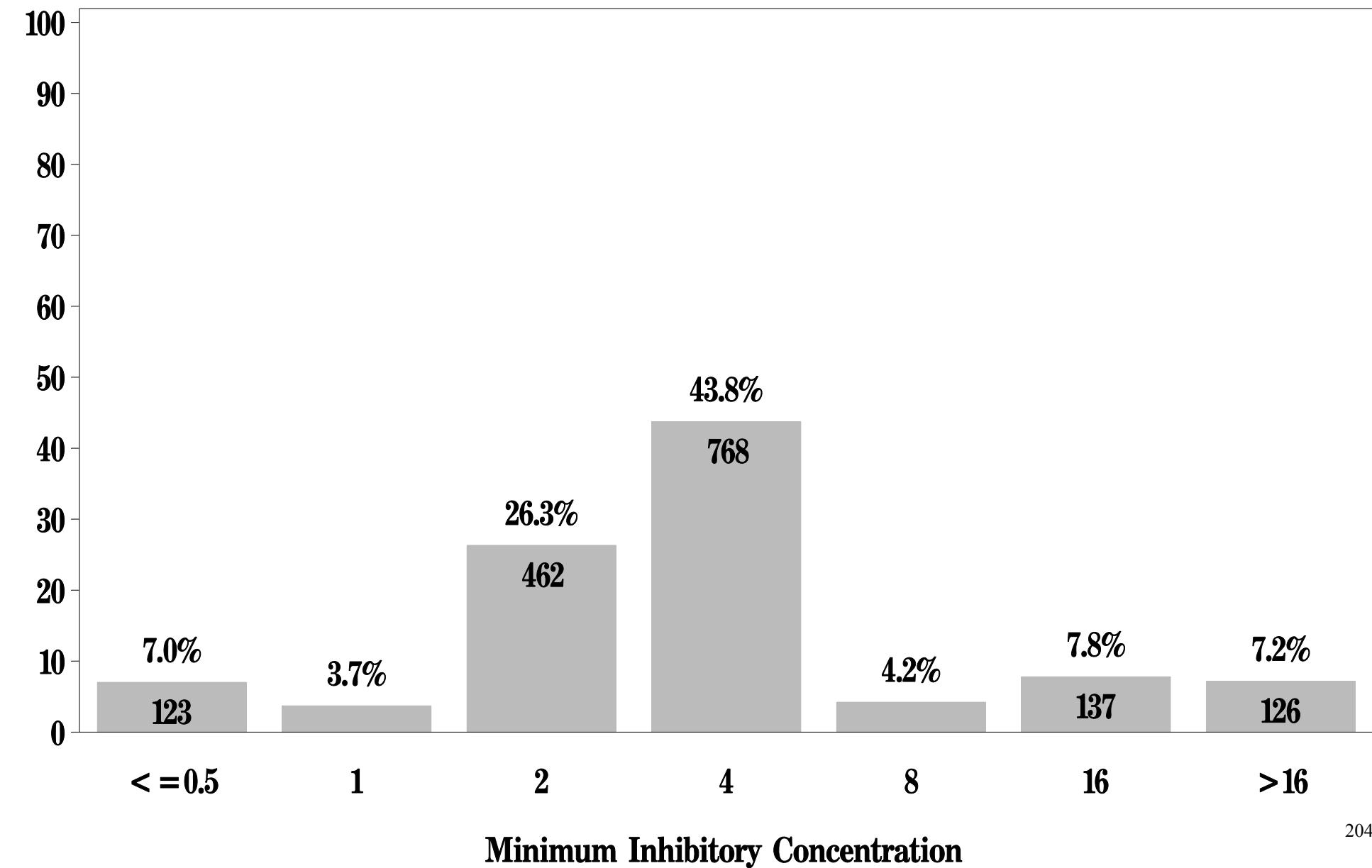
Breakpoints: Susceptible $\leq 32 \text{ } \mu\text{g/mL}$ Resistant $\geq 128 \text{ } \mu\text{g/mL}$



NARMS

**Figure 13l: Minimum Inhibitory Concentration of Penicillin
for *Enterococcus* (N=1755 Isolates)**

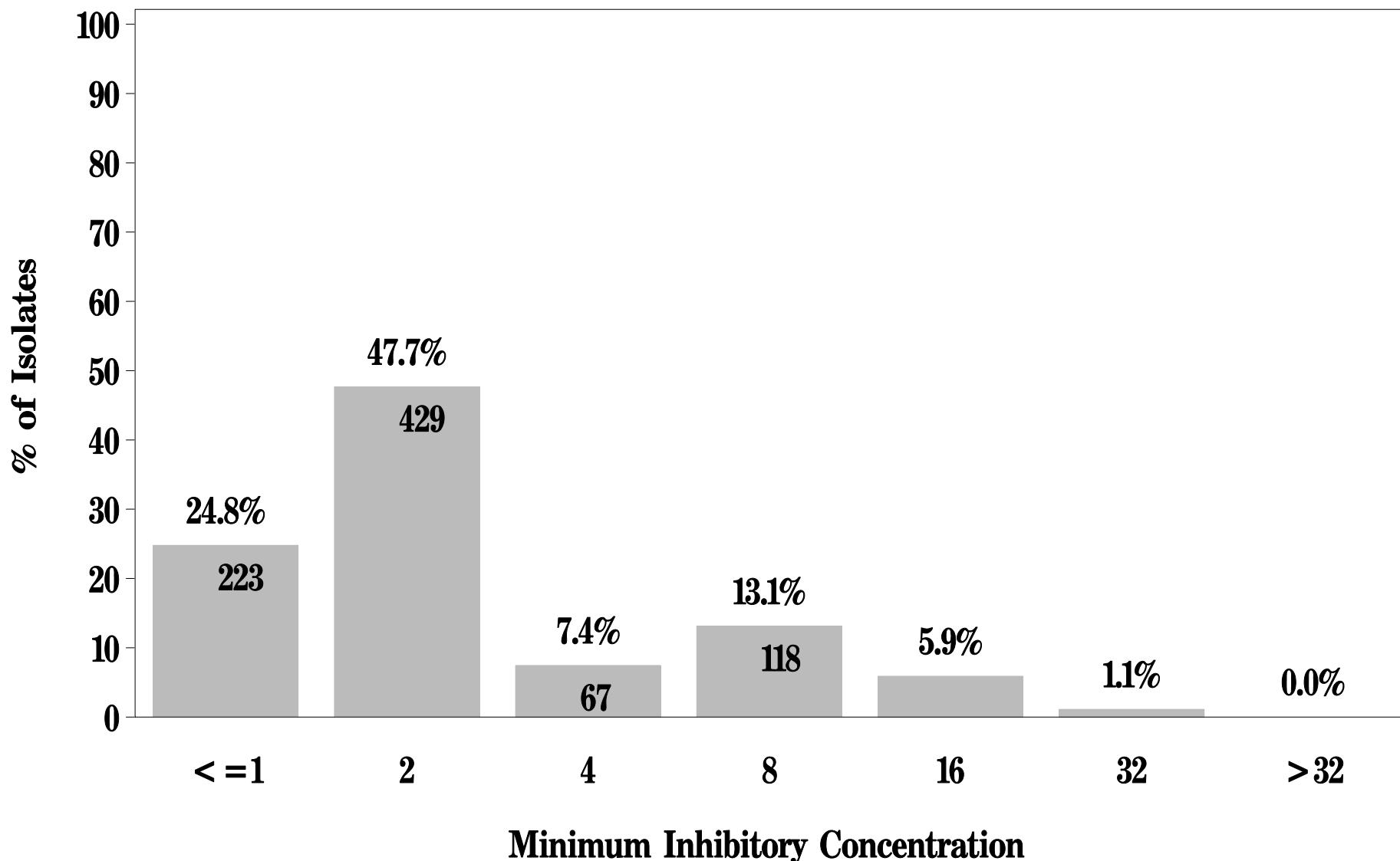
Breakpoints: Susceptible $\leq 8 \text{ } \mu\text{g/mL}$ Resistant $\geq 16 \text{ } \mu\text{g/mL}$



NARMS

Figure 13m: Minimum Inhibitory Concentration of Quinupristin – dalfopristin*
for *Enterococcus* (N=900 Isolates)

Breakpoints: Susceptible $\leq 1 \text{ } \mu\text{g/mL}$ Resistant $\geq 4 \text{ } \mu\text{g/mL}$

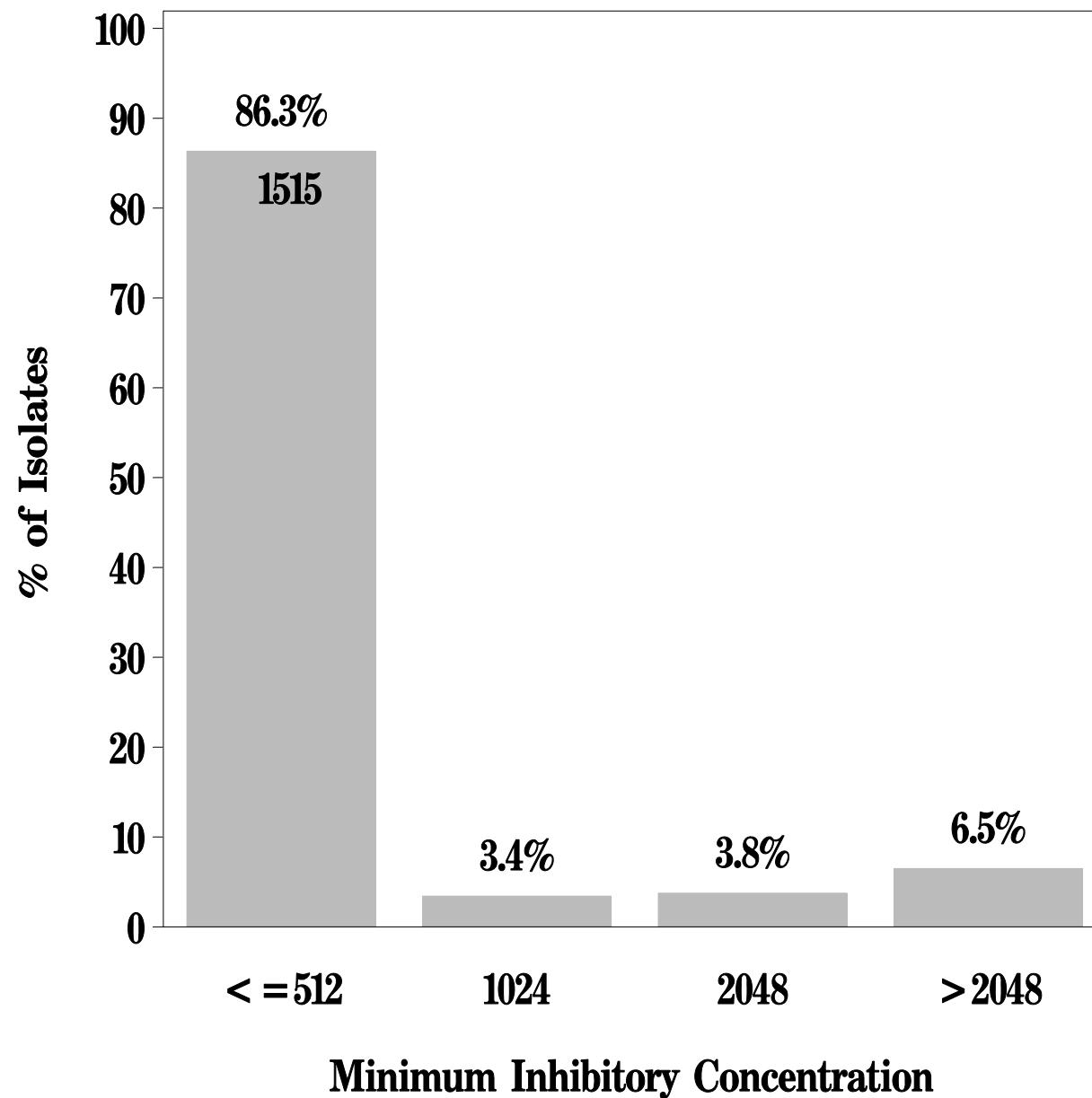


*Presented for all species except *E. faecalis* (N=1755 – 855 = 900)

NARMS

**Figure 13n: Minimum Inhibitory Concentration of Streptomycin
for *Enterococcus* (N=1755 Isolates)**

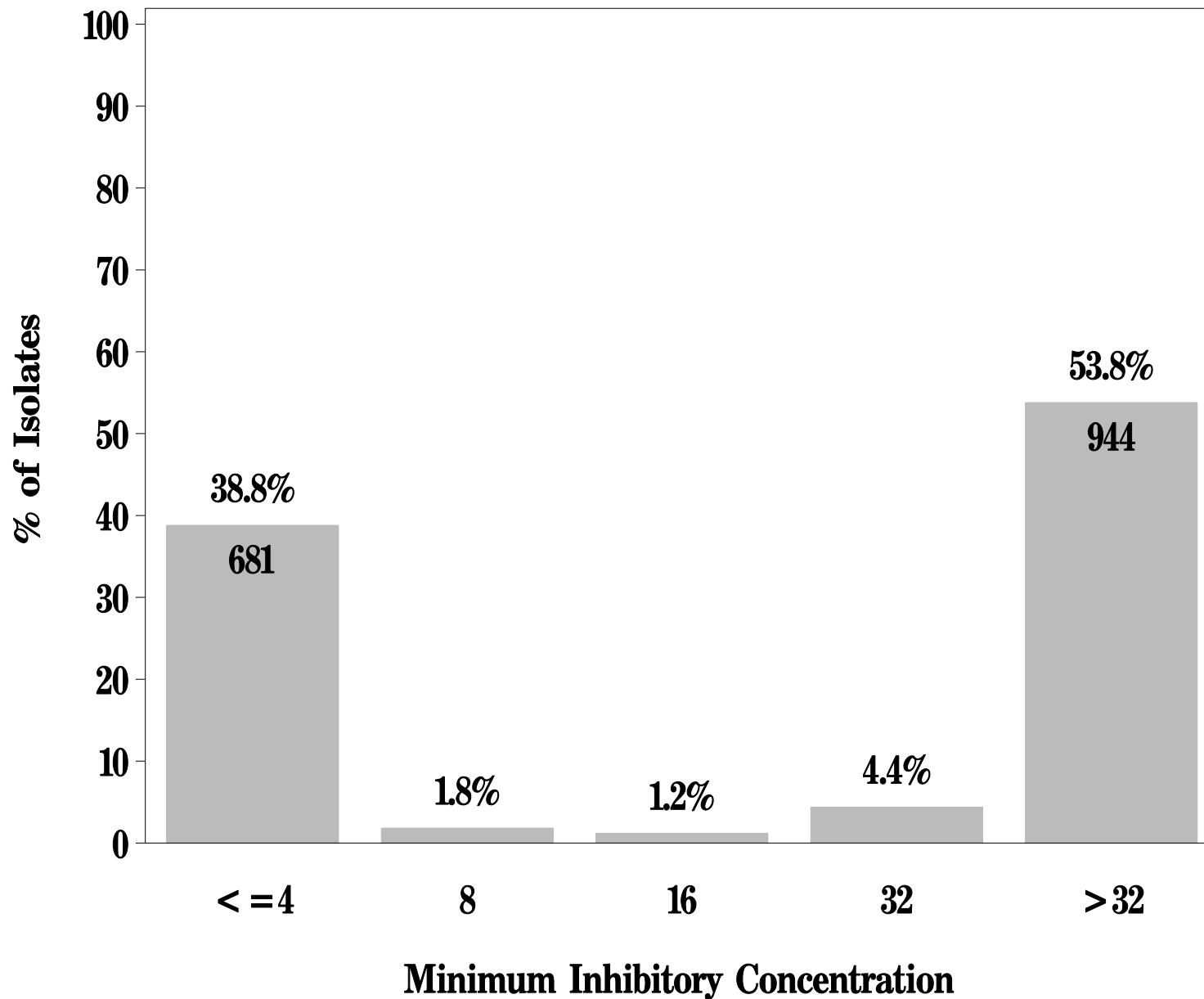
Breakpoints: Susceptible $\leq 1000 \mu\text{g/mL}$ Resistant $\geq 1000 \mu\text{g/mL}$



NARMS

**Figure 13o: Minimum Inhibitory Concentration of Tetracycline
for *Enterococcus* (N=1755 Isolates)**

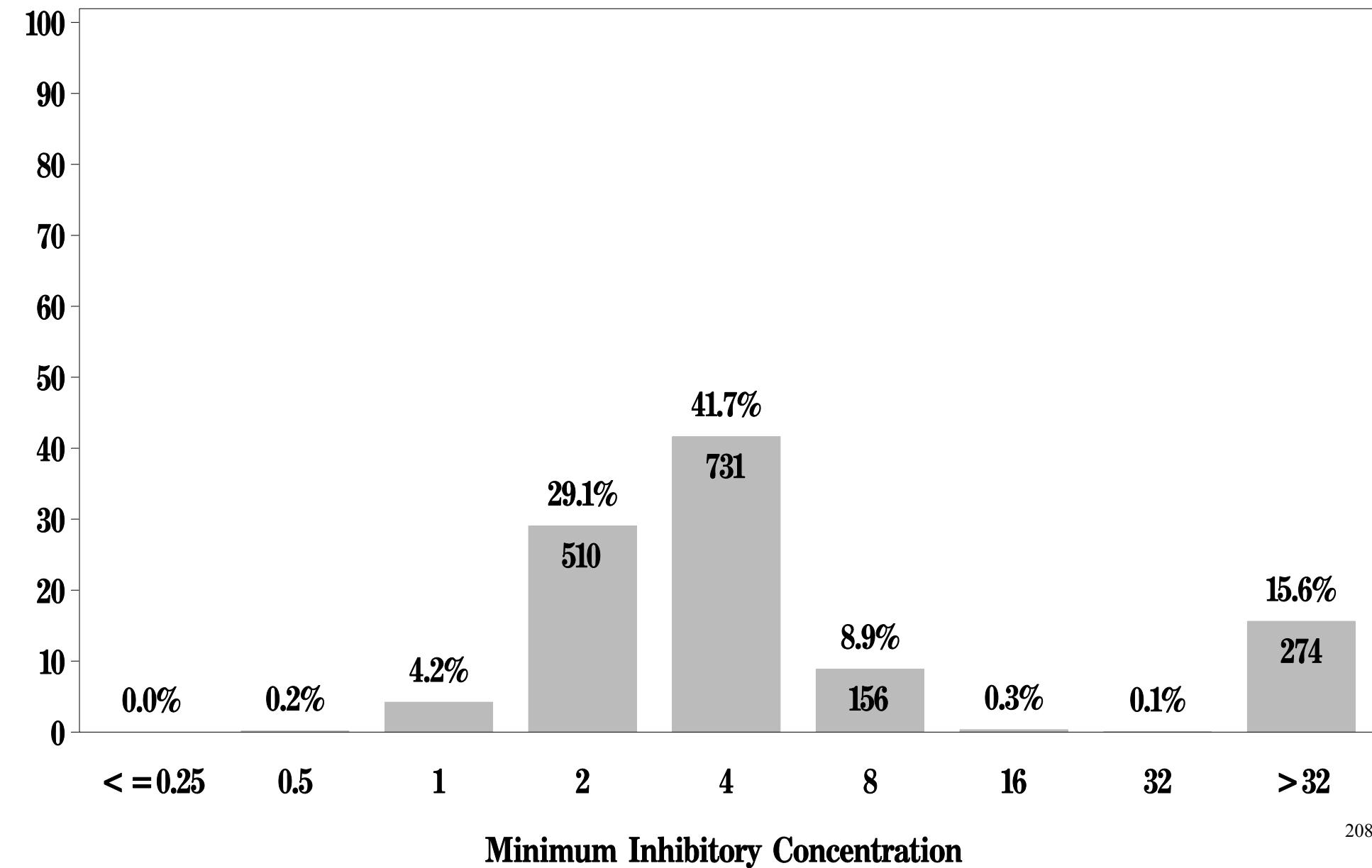
Breakpoints: Susceptible $\leq 4 \text{ } \mu\text{g/mL}$ Resistant $\geq 16 \text{ } \mu\text{g/mL}$



NARMS

Figure 13p: Minimum Inhibitory Concentration of Tylosin
for *Enterococcus* (N=1755 Isolates)

Breakpoints: Susceptible $\leq 8 \text{ } \mu\text{g/mL}$ Resistant $\geq 32 \text{ } \mu\text{g/mL}$



NARMS

**Figure 13q: Minimum Inhibitory Concentration of Vancomycin
for *Enterococcus* (N=1755 Isolates)**

Breakpoints: Susceptible $\leq 4 \text{ } \mu\text{g/mL}$ Resistant $\geq 32 \text{ } \mu\text{g/mL}$

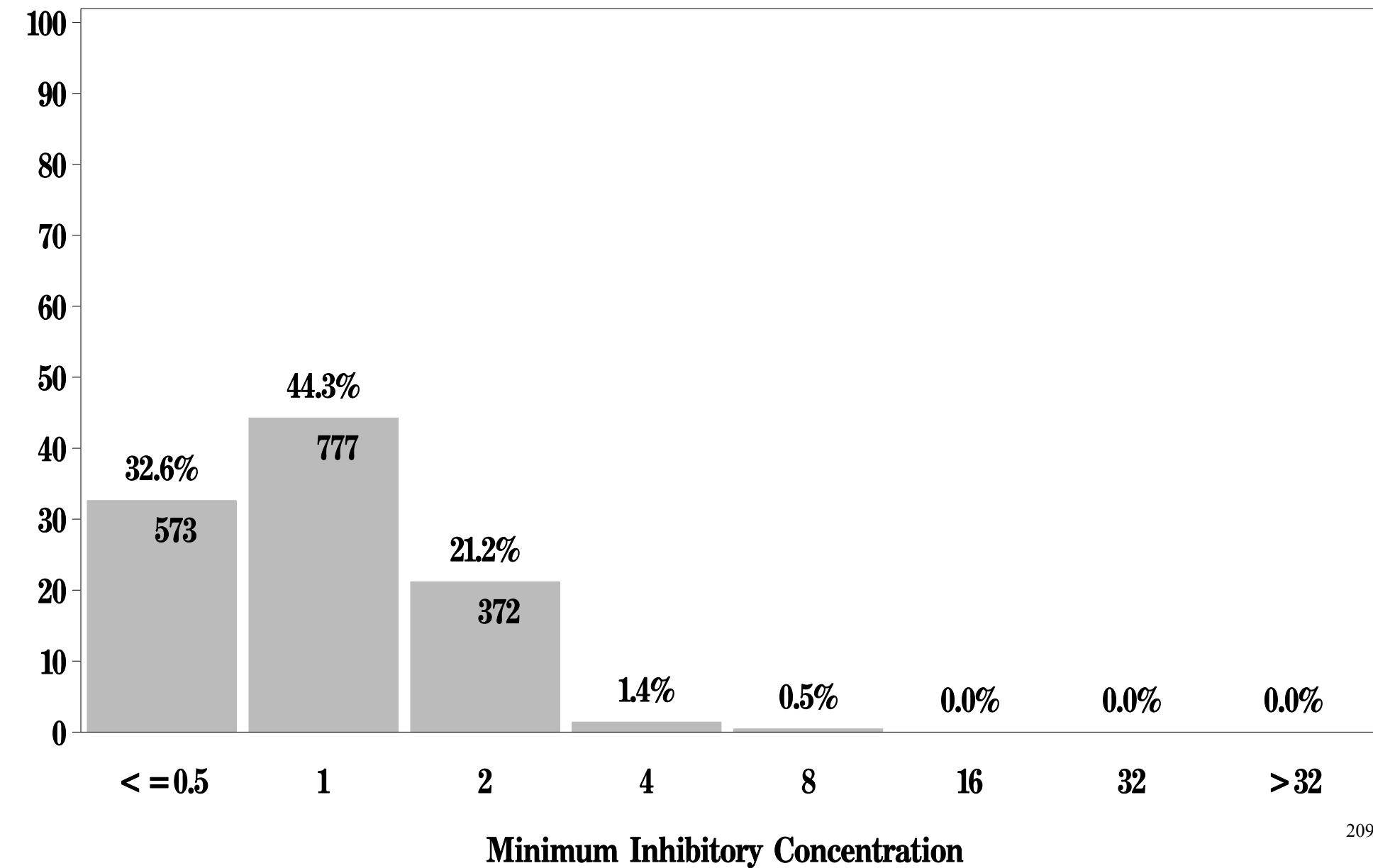


Figure 14a. MIC Distribution among *Enterococcus* from Chicken Breast

<i>Enterococcus</i> from Chicken Breast (N=466)		Distribution (%) of MICs (in µg/ml)																		
Antimicrobial Agent	%R	0.015	0.03	0.06	0.125	0.25	0.5	1	2	4	8	16	32	64	128	256	512	1024	2048	>2048
Bacitracin*	80.7									2.8	1.5	1.7	13.3	16.5	64.2					
Chloramphenicol	0.0									4.7	88.4	6.9								
Ciprofloxacin	40.8			0.2	0.4	4.9	13.1	40.6	32.6	8.2										
Daptomycin*	§						0.4	14.8	24.7	57.1	2.1	0.9								
Erythromycin	17.0						38.0	18.9	18.9	7.3	1.7	15.2								
Tylosin*	15.0							2.4	24.7	44.4	13.5				15.0					
Gentamicin	7.1										92.3	0.6	1.1	0.6	5.4					
Kanamycin*	34.8										41.0	24.2	23.0	2.6	9.2					
Streptomycin*	11.4											88.6	3.4	4.1	3.9					
Lincomycin*	67.2						13.1	0.2	1.9	17.6	12.9	54.3								
Linezolid	0.0						1.1	87.8	11.2											
Nitrofurantoin	65.5									4.7	13.1	2.8	13.9	65.5						
Flavomycin*	68.5						18.2	0.6	1.5	5.6	5.6	3.4	65.0							
Penicillin	30.9						1.1	3.0	20.4	35.6	9.0	16.5	14.4							
Tetracycline	49.1								45.3	5.6	2.1	3.2	43.8							
Quinupristin/Dalfopristin†	29.9						27.5	42.6	6.3	18.5	5.0									
Vancomycin	0.0						47.6	36.1	15.7	0.6										

Vertical bars show the CLSI Susceptible/Resistant breakpoints for each drug where appropriate.

*Currently no CLSI breakpoints have been established for this organism/antimicrobial combination. Indicated breakpoints were established by NARMS.

†Discrepancies between %R and sums of distribution %s are due to rounding.

Unshaded areas indicate the dilution ranges of the Sensititre plate used to test the 2004 isolates.

↑ Presented for all species except *E. faecalis* in QDA (n=466-88= 378 non *E. faecalis*)

§Absence of resistant strains precludes defining any results category other than "susceptible."

Figure 14b. MIC Distribution among *Enterococcus* from Ground Turkey

<i>Enterococcus</i> from Ground Turkey (N=437)		Distribution (%) of MICs (in µg/ml)																		
Antimicrobial Agent	%R	0.015	0.03	0.06	0.125	0.25	0.5	1	2	4	8	16	32	64	128	256	512	1024	2048	>2048
Bacitracin*	80.1										0.5	0.9	2.7	15.8	12.8	67.3				
Chloramphenicol	0.0									0.5	3.7	85.1	10.8							
Ciprofloxacin	24.7						0.2	3.7	25.4	46.0	19.5	5.3								
Daptomycin*	§								5.9	47.1	16.9	27.0	2.7	0.2						
Erythromycin	37.1							34.6	21.3	5.7	1.4	1.1	35.9							
Tylosin*	34.6							0.2	3.9	21.7	34.8	4.8		0.2	34.3					
Gentamicin	20.1										79.4	0.5	0.7	0.5		18.9				
Kanamycin*	41.0										49.4	9.6	9.2	0.9	30.9					
Streptomycin*	29.5											70.5	6.6	5.7	17.1					
Lincomycin*	86.0							4.3	0.5	0.5	0.7	8.0	30.0	56.1						
Linezolid	0.0							0.2	2.3	90.8	6.6		29.3	28.8	1.1	13.7	27.0			
Nitrofurantoin	27.0																			
Flavomycin*	35.7							55.8	2.1	1.1	2.7	2.5	0.9	34.8						
Penicillin	24.3							1.1	0.9	26.1	43.5	4.1	11.9	12.4						
Tetracycline	87.0									12.8	0.2	0.9	3.2	82.8						
Quinupristin/Dalfopristin†	62.7							14.7	22.6	11.3	26.6	19.2	5.6							
Vancomycin	0.0							22.7	46.0	28.6	1.8	0.9								

Vertical bars show the CLSI Susceptible/Resistant breakpoints for each drug where appropriate.

*Currently no CLSI breakpoints have been established for this organism/antimicrobial combination. Indicated breakpoints were established by NARMS.

†Discrepancies between %R and sums of distribution %s are due to rounding.

Unshaded areas indicate the dilution ranges of the Sensititre plate used to test the 2004 isolates.

↑ Presented for all species except *E. faecalis* in QDA (n=437-260= 177 non *E. faecalis*)

§Absence of resistant strains precludes defining any results category other than "susceptible."

Figure 14c. MIC Distribution among *Enterococcus* from Ground Beef

<i>Enterococcus</i> from Ground Beef (N=448)		Distribution (%) of MICs (in µg/ml)																		
Antimicrobial Agent	%R	0.015	0.03	0.06	0.125	0.25	0.5	1	2	4	8	16	32	64	128	256	512	1024	2048	>2048
Bacitracin*	33.3											20.1	4.2	13.4	29.0	21.2	12.1			
Chloramphenicol	0.4									6.5	88.6	4.5	0.4							
Ciprofloxacin	15.8				0.2	7.8	37.9	38.2	13.2	2.7										
Daptomycin*	§							2.9	34.6	33.3	24.6	3.3	1.3							
Erythromycin	6.5							46.7	23.0	13.2	10.7	0.9	5.6							
Tylosin*	5.1								6.7	39.1	38.6	9.4	1.1		5.1					
Gentamicin	0.4											99.1	0.4		0.2	0.2				
Kanamycin*	13.6											78.1	8.3	9.2	0.4	4.0				
Streptomycin*	5.4											94.6	2.0	1.1	2.2					
Lincomycin*	52.2							13.6	0.9	1.1	8.3	23.9	44.9	7.4						
Linezolid	0.0							0.2	0.4	87.7	11.6									
Nitrofurantoin	20.1									0.2	15.4	27.9	8.7	27.7	20.0					
Flavomycin*	53.3							39.5	3.6	1.1	0.4	2.0	1.1	52.2						
Penicillin	1.3							14.1	8.5	27.7	45.8	2.7	0.4	0.9						
Tetracycline	30.4									69.4	0.2	0.9	5.8	23.7						
Quinupristin/Dalfopristin†	7.5							29.1	63.4	7.5										
Vancomycin	0.0							43.1	38.8	16.1	1.3	0.7								

Vertical bars show the CLSI Susceptible/Resistant breakpoints for each drug where appropriate.

*Currently no CLSI breakpoints have been established for this organism/antimicrobial combination. Indicated breakpoints were established by NARMS.

†Discrepancies between %R and sums of distribution %s are due to rounding.

Unshaded areas indicate the dilution ranges of the Sensititre plate used to test the 2004 isolates.

↑ Presented for all species except *E. faecalis* in QDA (n=448-194= 254 non *E. faecalis*)

§Absence of resistant strains precludes defining any results category other than “susceptible.”

Figure 14d. MIC Distribution among *Enterococcus* from Pork Chops

<i>Enterococcus</i> from Pork Chops (N=404)		Distribution (%) of MICs (in µg/ml)																			
Antimicrobial Agent	%R	0.015	0.03	0.06	0.125	0.25	0.5	1	2	4	8	16	32	64	128	256	512	1024	2048	>2048	
Bacitracin*	29.2											5.4	5.9	12.9	46.5	19.3	9.9				
Chloramphenicol	0.5											3.2	91.3	5.0	0.2	0.2					
Ciprofloxacin	8.2						0.5	4.7	46.8	39.9	6.2	2.0									
Daptomycin*	§								7.7	57.7	21.5	13.1									
Erythromycin	8.7								41.6	32.7	9.1	7.9	1.0	7.7							
Tylosin*	7.7								0.5	4.0	30.9	49.2	7.4	0.2		7.7					
Gentamicin	1.5													98.0	0.5	0.5	0.5	0.5			
Kanamycin*	4.7													91.6	3.7	1.9	0.5	2.2			
Streptomycin*	8.4														91.5	1.5	4.2	2.7			
Lincomycin*	65.6								6.4	0.5	1.0	12.3	14.1	53.0	12.6						
Linezolid	0.0								2.5	87.6	9.9										
Nitrofurantoin	7.9											26.7	49.3	4.2	11.9	7.9					
Flavomycin*	21.5								72.3	5.4	0.2	0.2	0.2	0.2	0.2	21.2					
Penicillin	1.7								12.4	2.2	31.9	51.2	0.5	1.5	0.2						
Tetracycline	73.5										25.4	1.0	0.7	5.4	67.3						
Quinupristin/Dalfopristin†	5.5								20.9	73.6	4.4	1.1									
Vancomycin	0.0								14.6	57.9	25.2	2.0	0.2								

Vertical bars show the CLSI Susceptible/Resistant breakpoints for each drug where appropriate.

*Currently no CLSI breakpoints have been established for this organism/antimicrobial combination. Indicated breakpoints were established by NARMS.

†Discrepancies between %R and sums of distribution %s are due to rounding.

Unshaded areas indicate the dilution ranges of the Sensititre plate used to test the 2003 isolates.

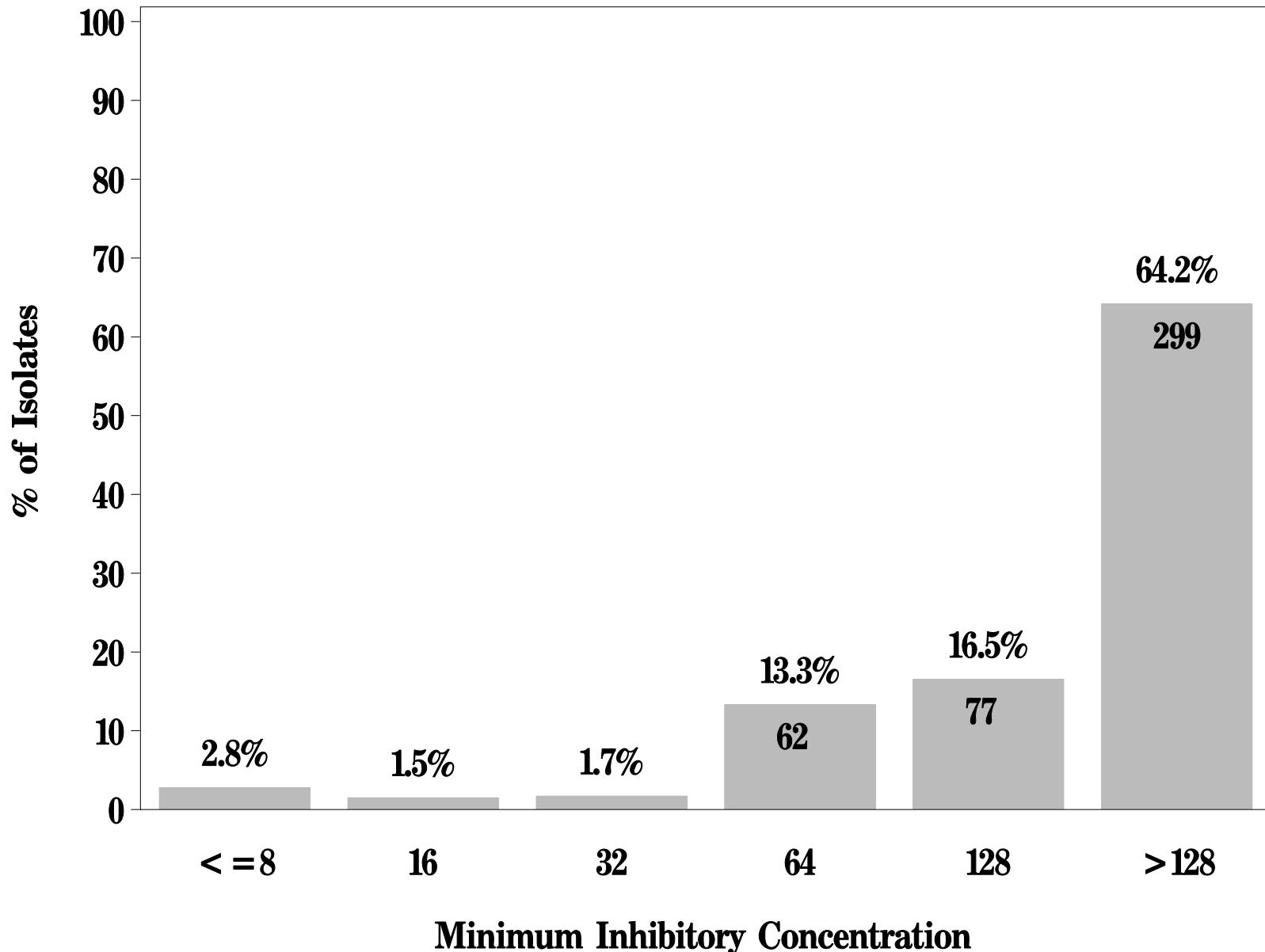
↑ Presented for all species except *E. faecalis* in QDA (n=404-313= 91 non *E. faecalis*)

§Absence of resistant strains precludes defining any results category other than "susceptible."

NARMS

**Figure 15a: Minimum Inhibitory Concentration of Bacitracin
for *Enterococcus* in Chicken Breast (N=466 Isolates)**

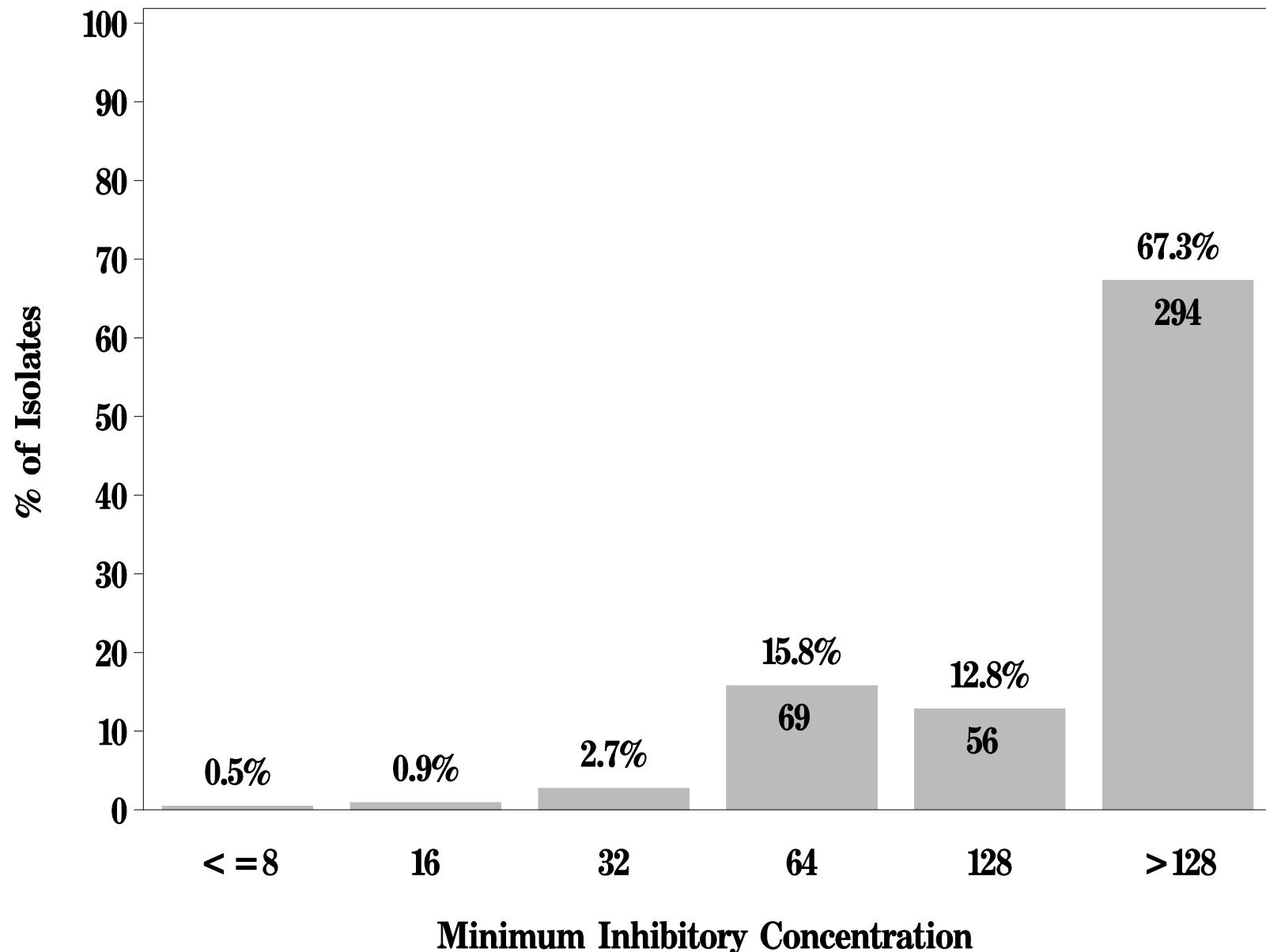
Breakpoints: Susceptible $\leq 32 \mu\text{g/mL}$ Resistant $\geq 128 \mu\text{g/mL}$



NARMS

**Figure 15a: Minimum Inhibitory Concentration of Bacitracin
for *Enterococcus* in Ground Turkey (N=437 Isolates)**

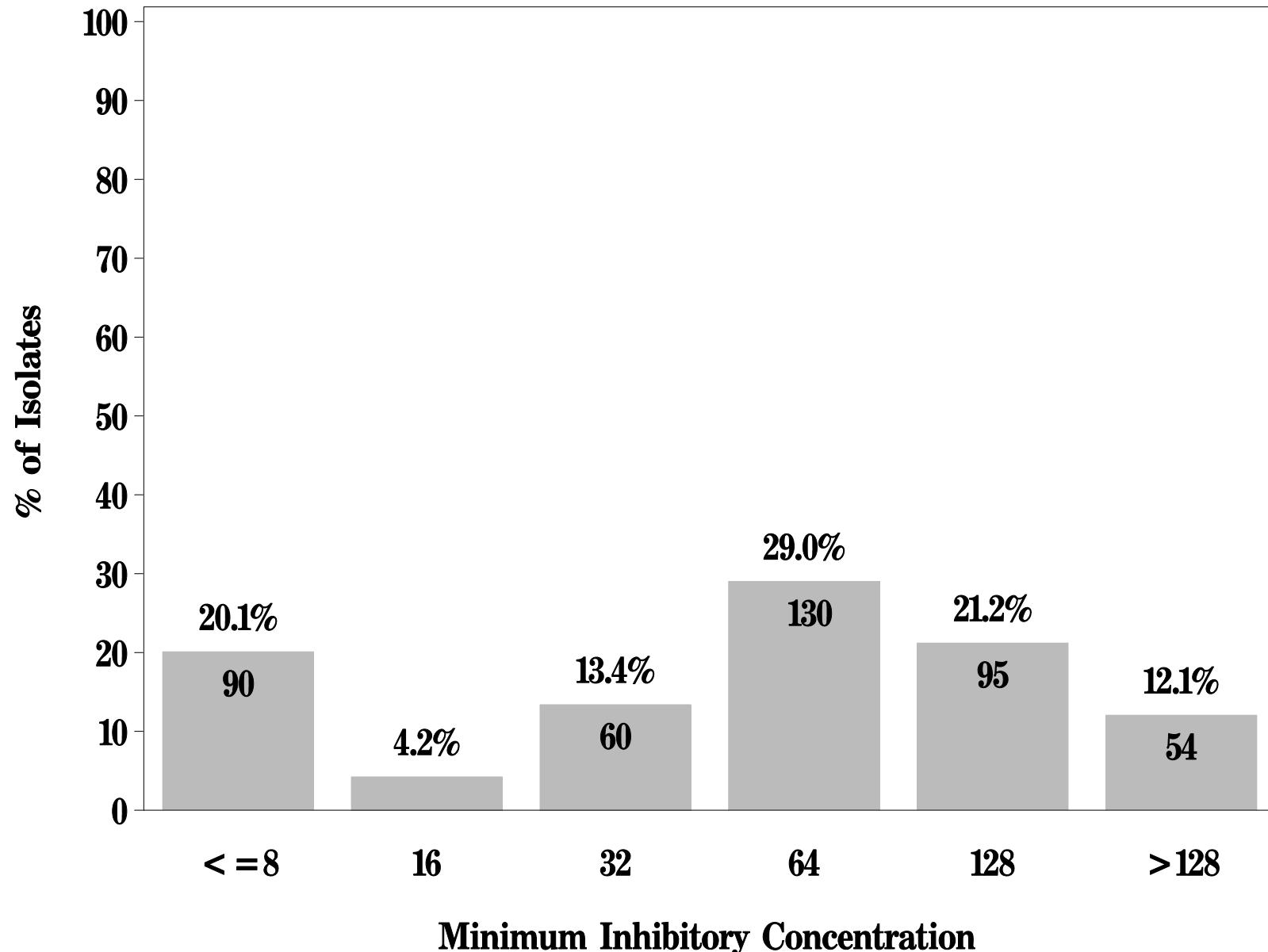
Breakpoints: Susceptible $\leq 32 \mu\text{g/mL}$ Resistant $\geq 128 \mu\text{g/mL}$



NARMS

**Figure 15a: Minimum Inhibitory Concentration of Bacitracin
for *Enterococcus* in Ground Beef (N=448 Isolates)**

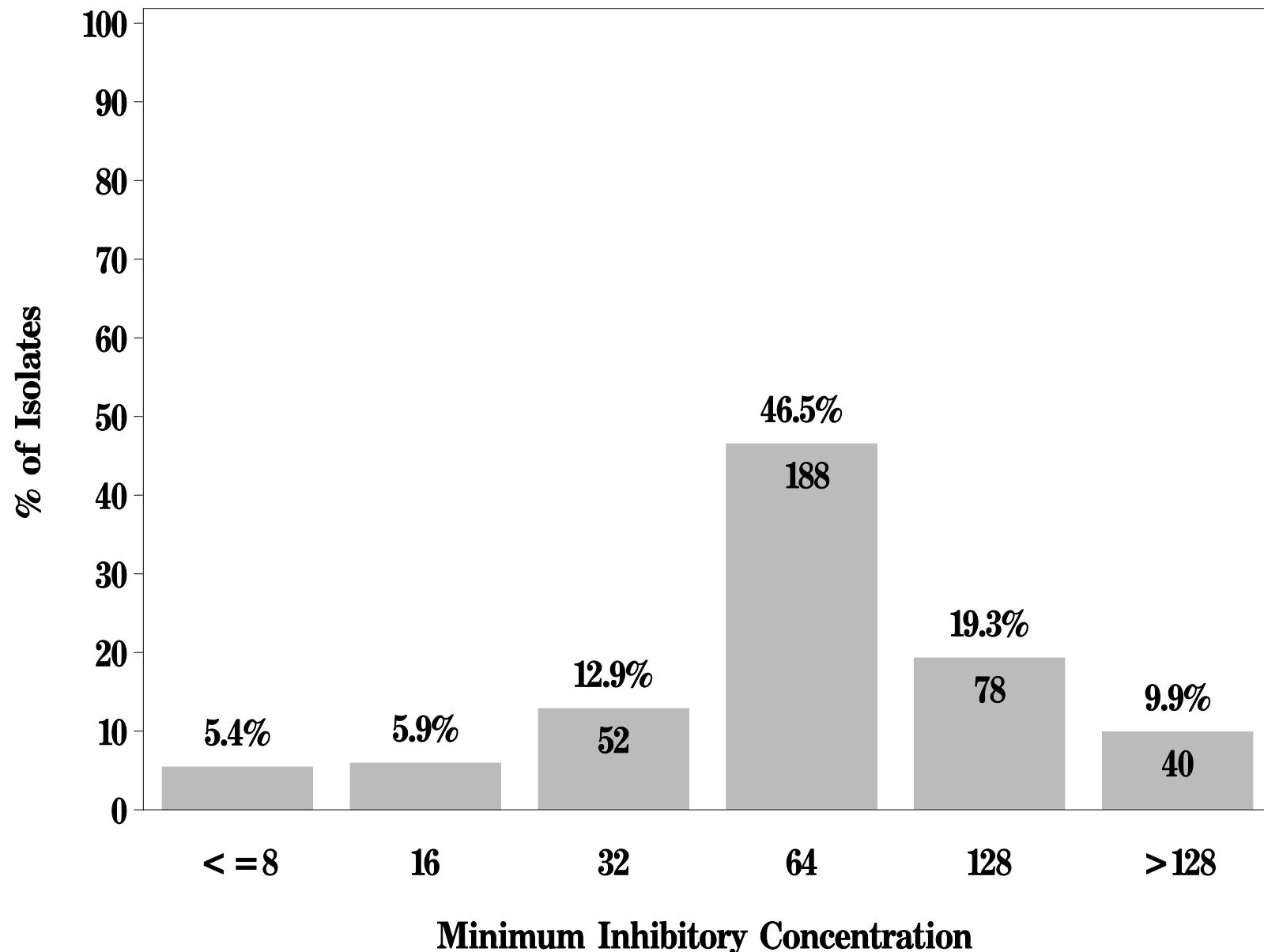
Breakpoints: Susceptible $\leq 32 \mu\text{g/mL}$ Resistant $\geq 128 \mu\text{g/mL}$



NARMS

**Figure 15a: Minimum Inhibitory Concentration of Bacitracin
for *Enterococcus* in Pork Chop (N=404 Isolates)**

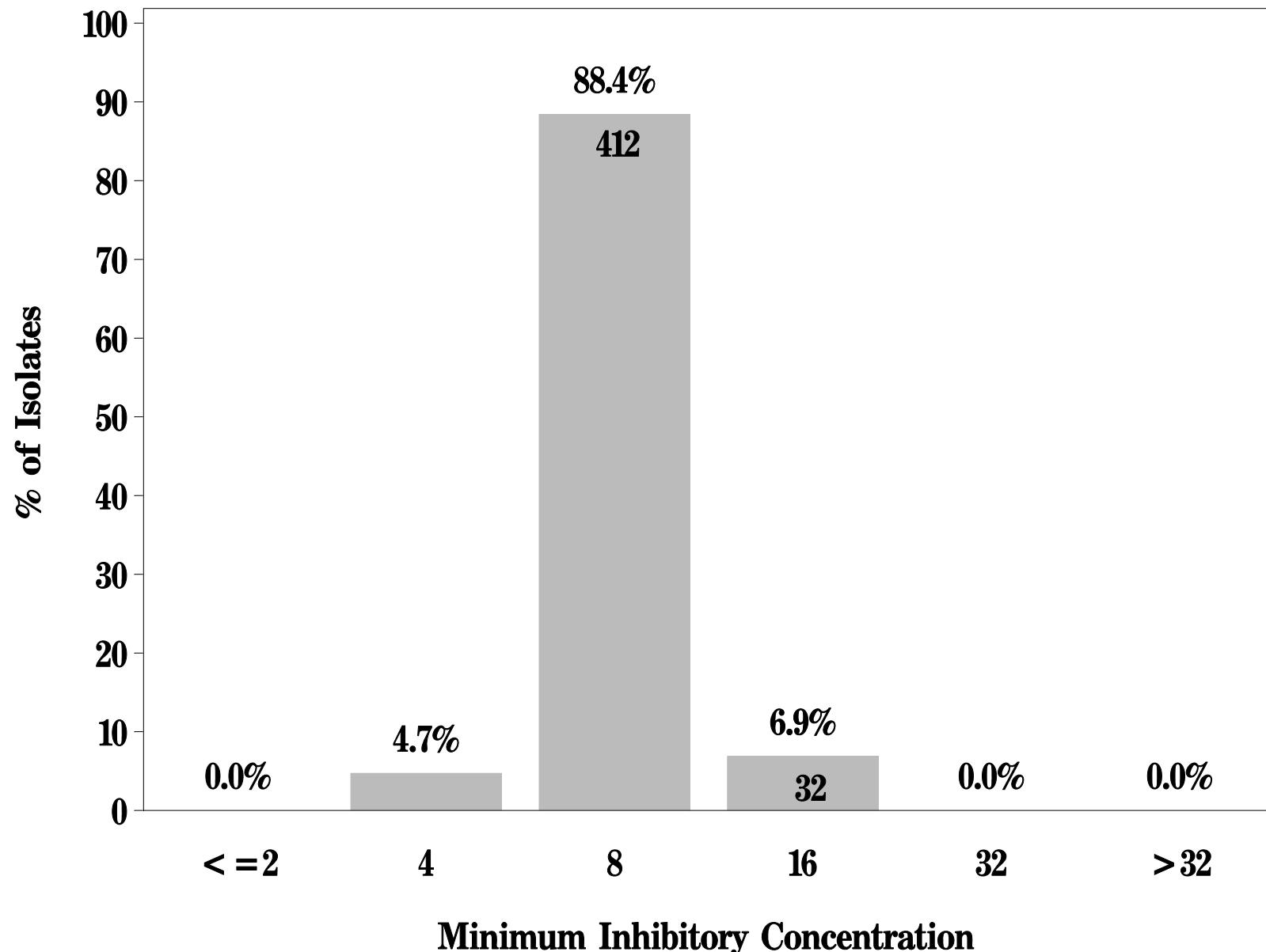
Breakpoints: Susceptible $\leq 32 \mu\text{g/mL}$ Resistant $\geq 128 \mu\text{g/mL}$



NARMS

**Figure 15b: Minimum Inhibitory Concentration of Chloramphenicol
for *Enterococcus* in Chicken Breast (N=466 Isolates)**

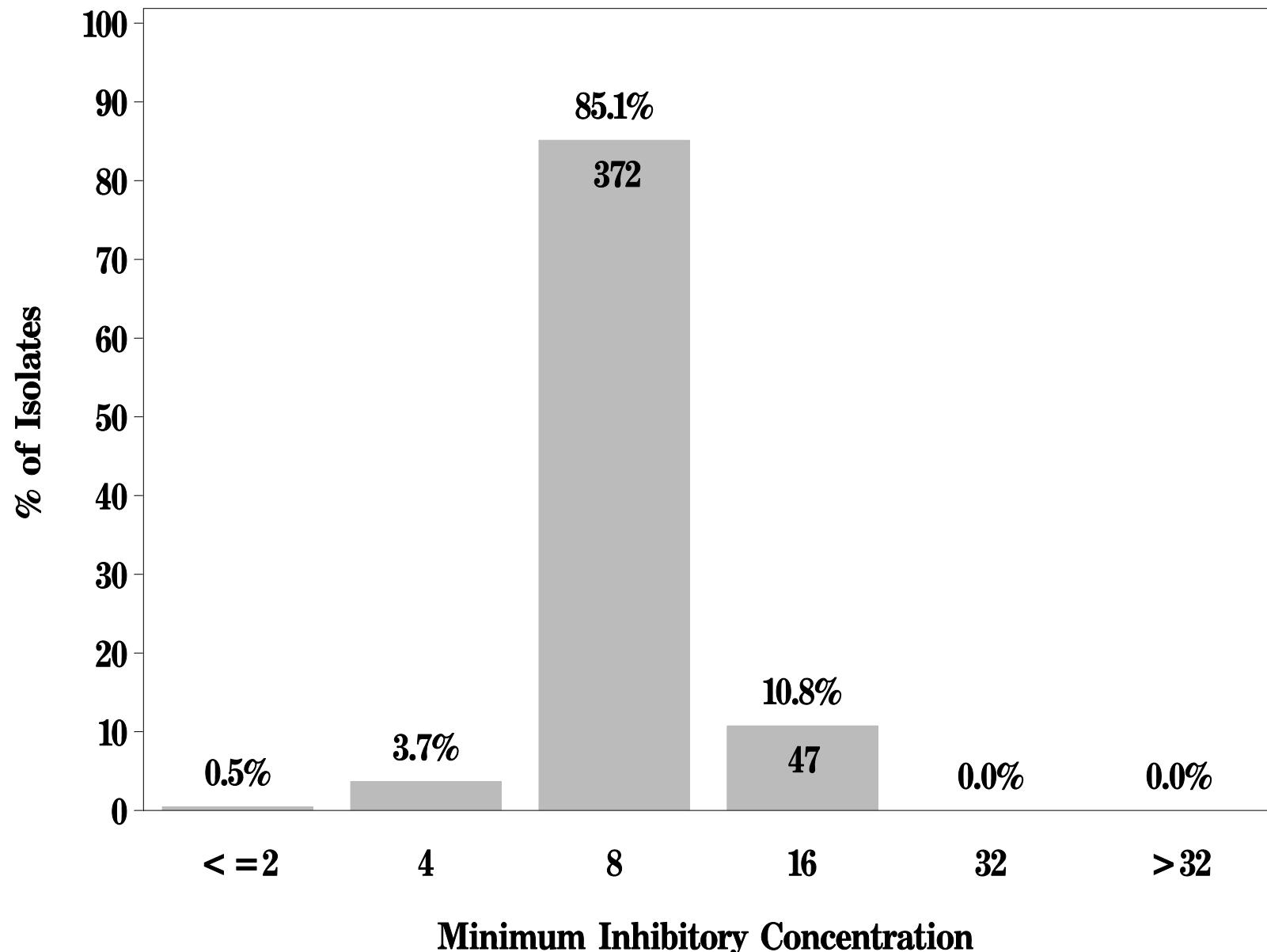
Breakpoints: Susceptible $\leq 8 \mu\text{g/mL}$ Resistant $\geq 32 \mu\text{g/mL}$



NARMS

**Figure 15b: Minimum Inhibitory Concentration of Chloramphenicol
for *Enterococcus* in Ground Turkey (N=437 Isolates)**

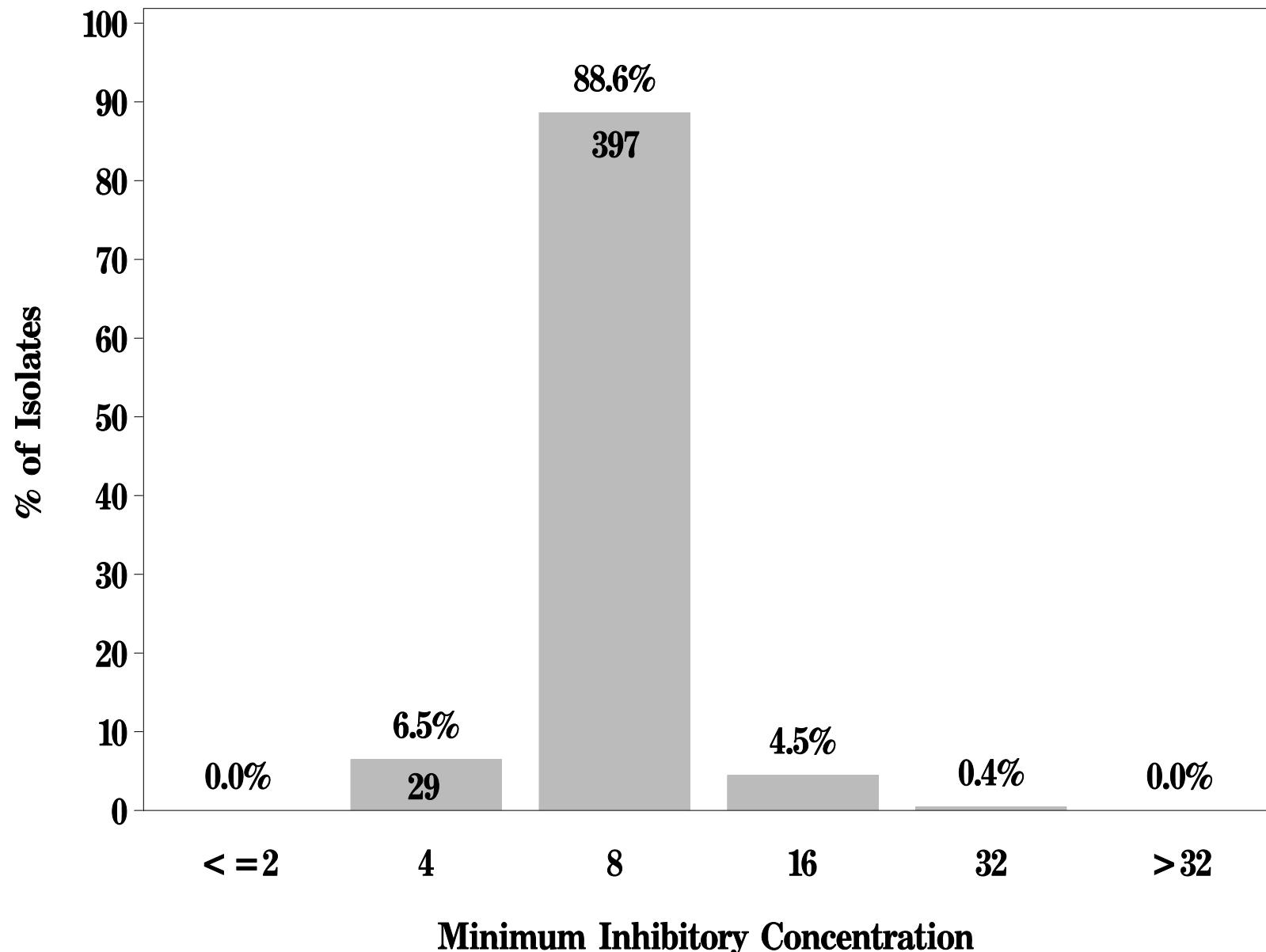
Breakpoints: Susceptible $\leq 8 \mu\text{g/mL}$ Resistant $\geq 32 \mu\text{g/mL}$



NARMS

**Figure 15b: Minimum Inhibitory Concentration of Chloramphenicol
for *Enterococcus* in Ground Beef (N=448 Isolates)**

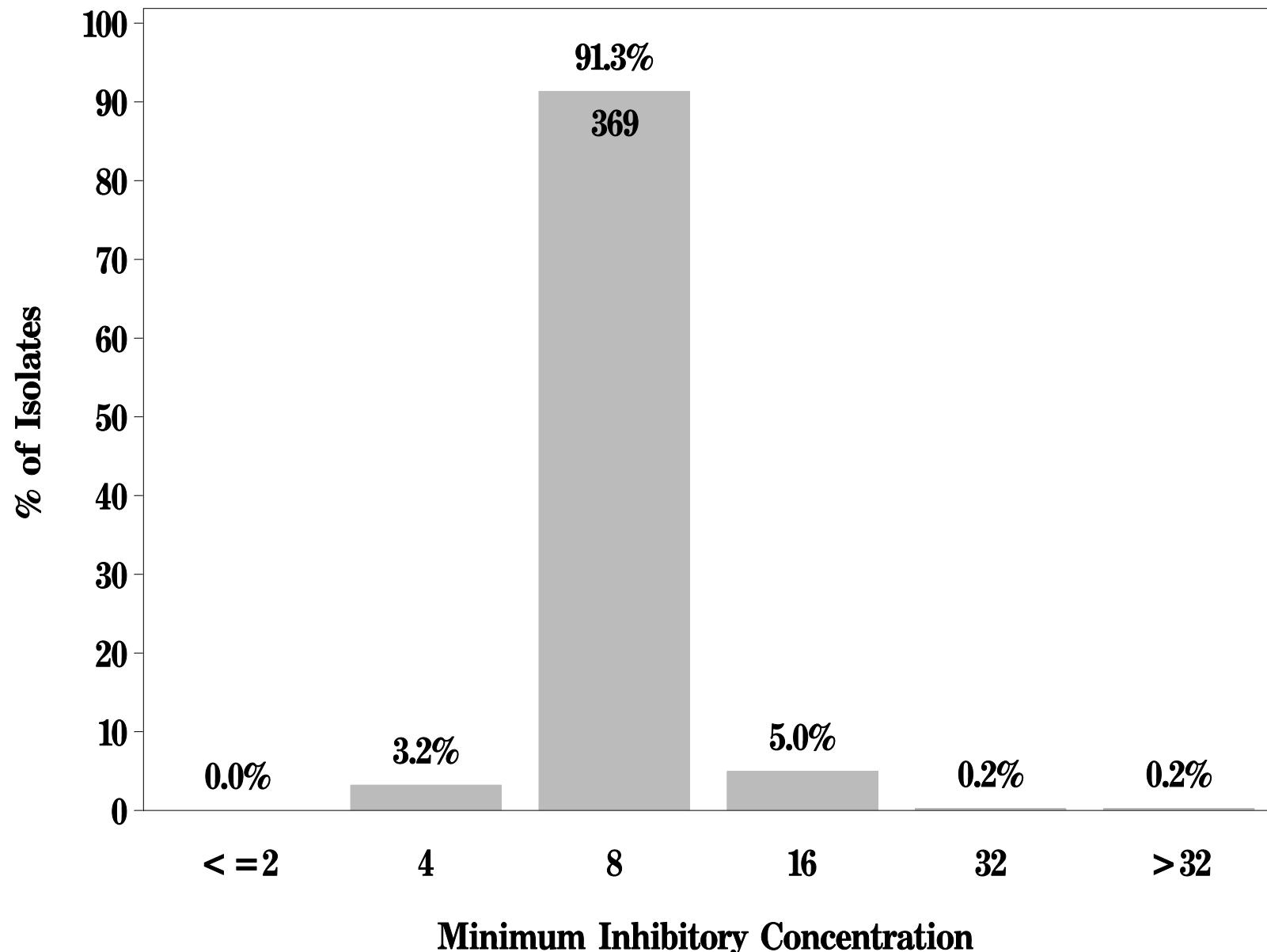
Breakpoints: Susceptible $\leq 8 \mu\text{g/mL}$ Resistant $\geq 32 \mu\text{g/mL}$



NARMS

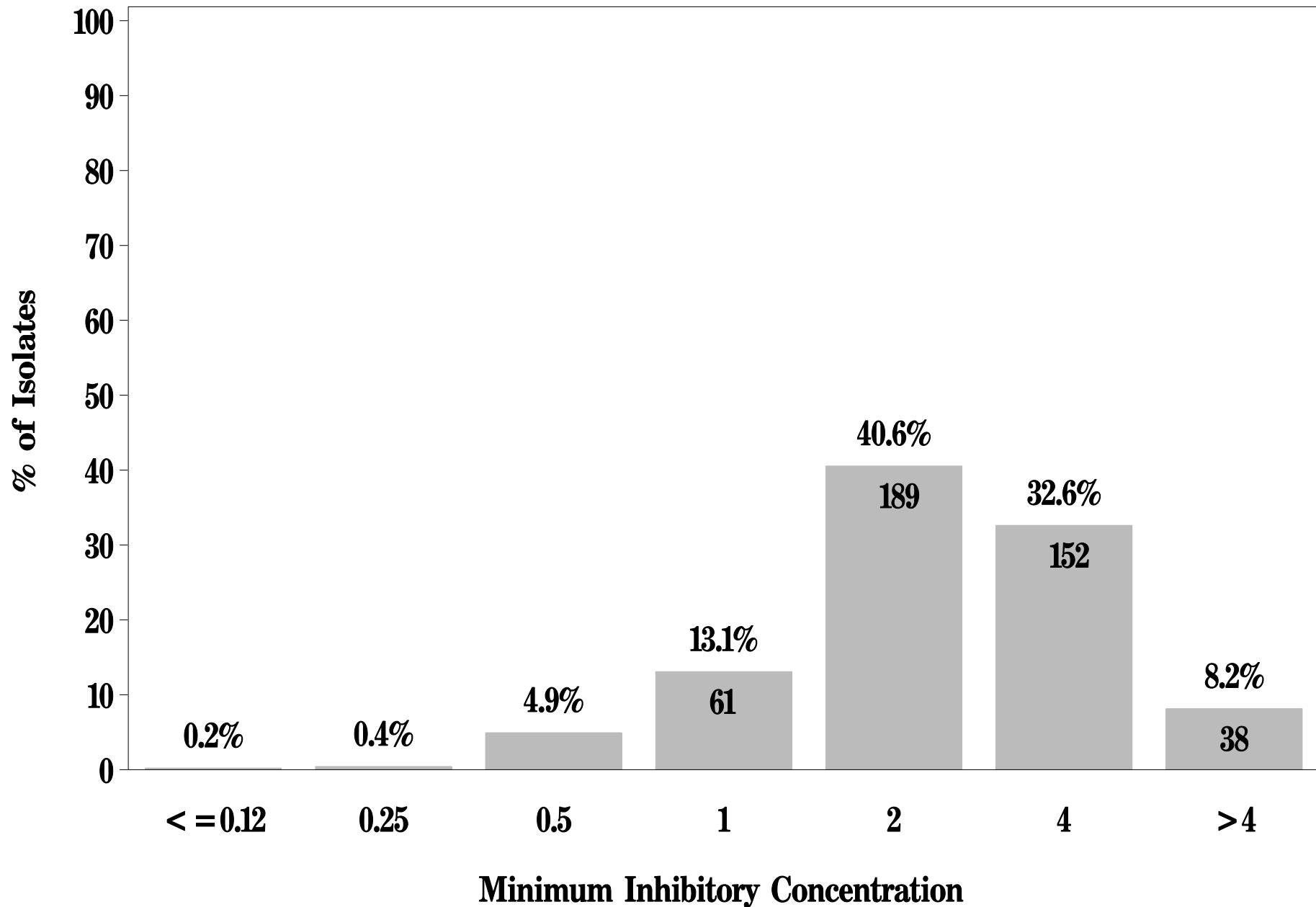
**Figure 15b: Minimum Inhibitory Concentration of Chloramphenicol
for *Enterococcus* in Pork Chop (N=404 Isolates)**

Breakpoints: Susceptible $\leq 8 \mu\text{g/mL}$ Resistant $\geq 32 \mu\text{g/mL}$



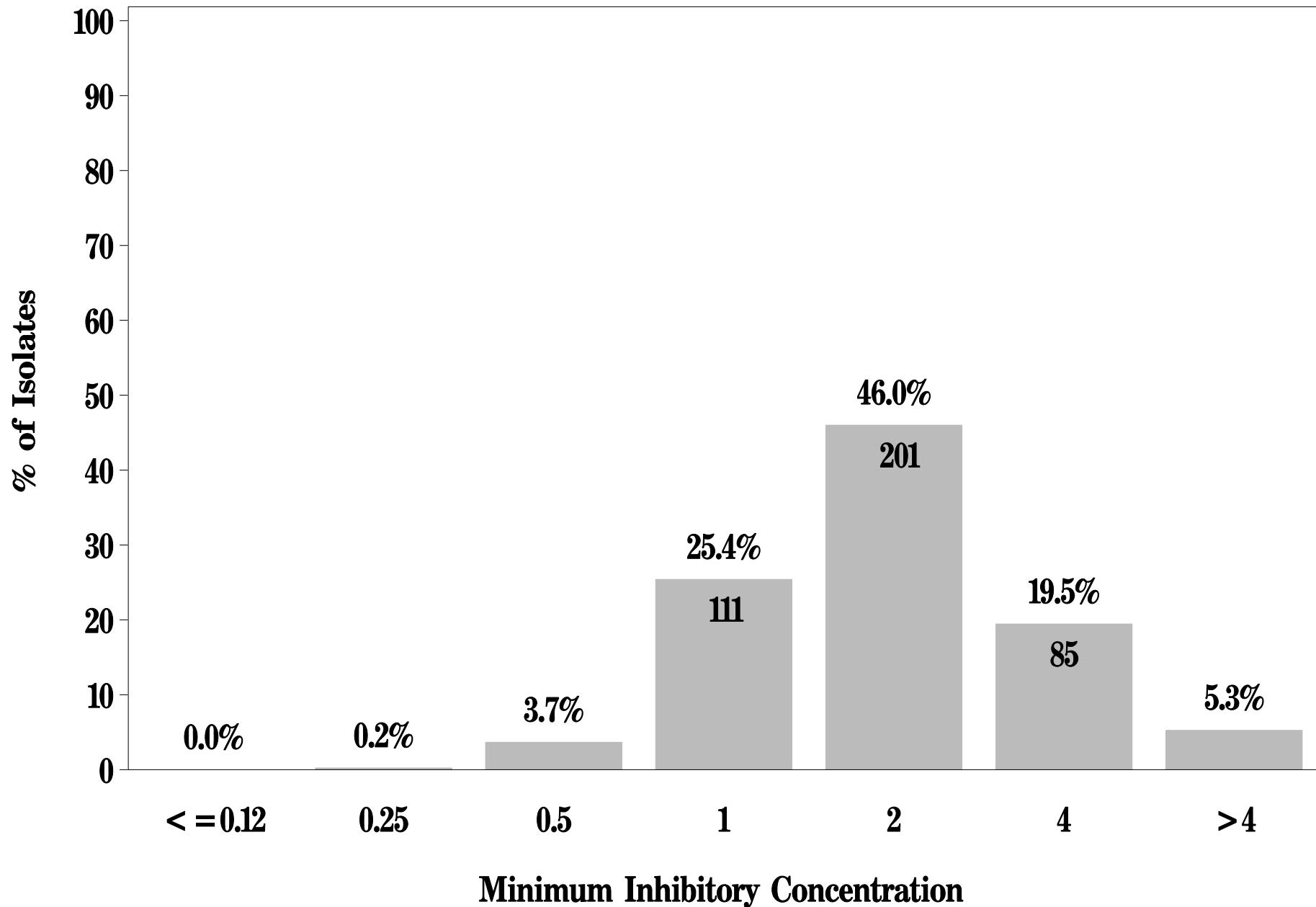
NARMS

**Figure 15c: Minimum Inhibitory Concentration of Ciprofloxacin
for *Enterococcus* in Chicken Breast (N=466 Isolates)**
Breakpoints: Susceptible $\leq 1 \mu\text{g/mL}$ Resistant $\geq 4 \mu\text{g/mL}$



NARMS

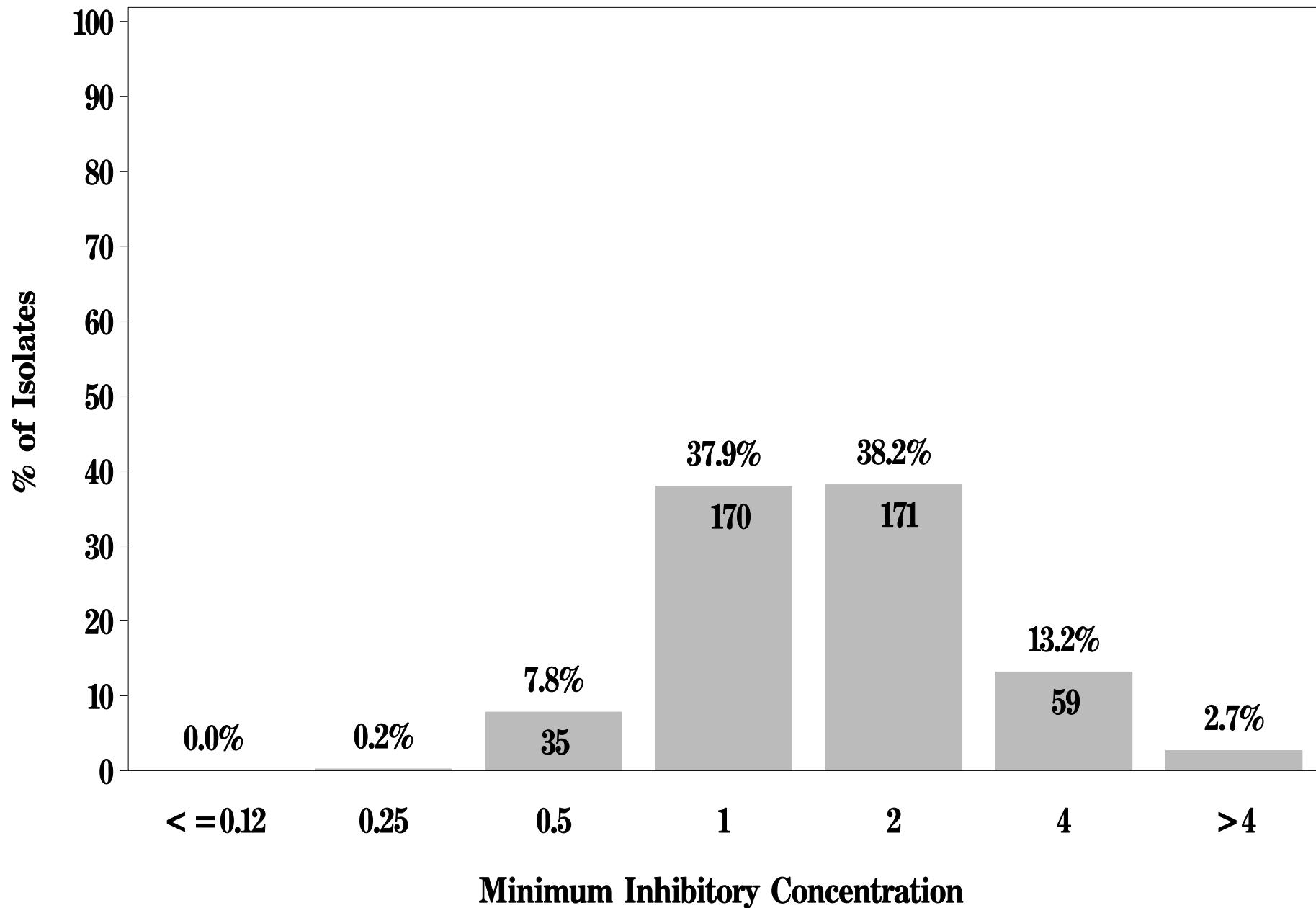
**Figure 15c: Minimum Inhibitory Concentration of Ciprofloxacin
for *Enterococcus* in Ground Turkey (N=437 Isolates)**
Breakpoints: Susceptible $\leq 1 \mu\text{g/mL}$ Resistant $\geq 4 \mu\text{g/mL}$



NARMS

**Figure 15c: Minimum Inhibitory Concentration of Ciprofloxacin
for *Enterococcus* in Ground Beef (N=448 Isolates)**

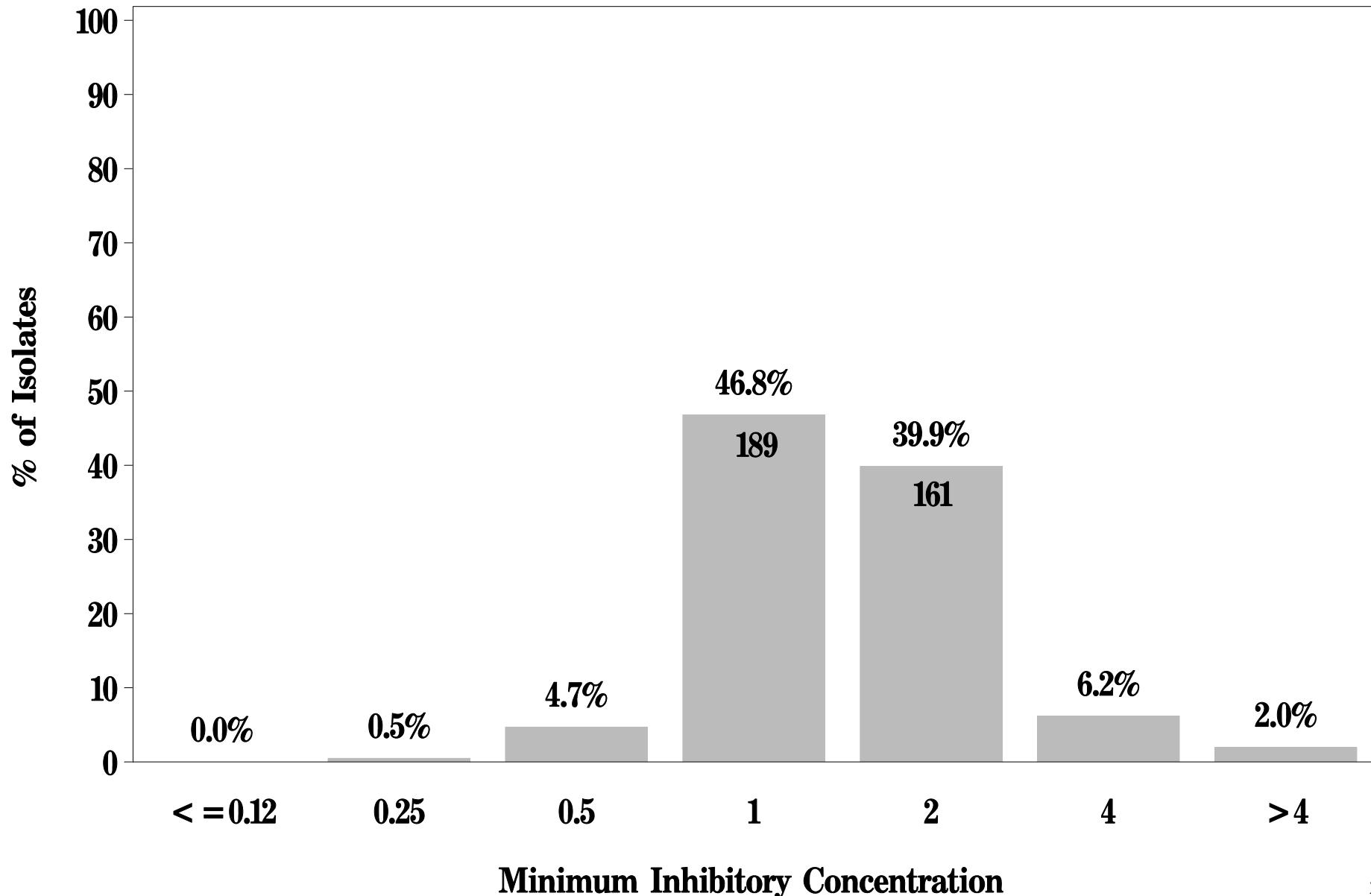
Breakpoints: Susceptible $\leq 1 \mu\text{g/mL}$ Resistant $\geq 4 \mu\text{g/mL}$



NARMS

**Figure 15c: Minimum Inhibitory Concentration of Ciprofloxacin
for *Enterococcus* in Pork Chop (N=404 Isolates)**

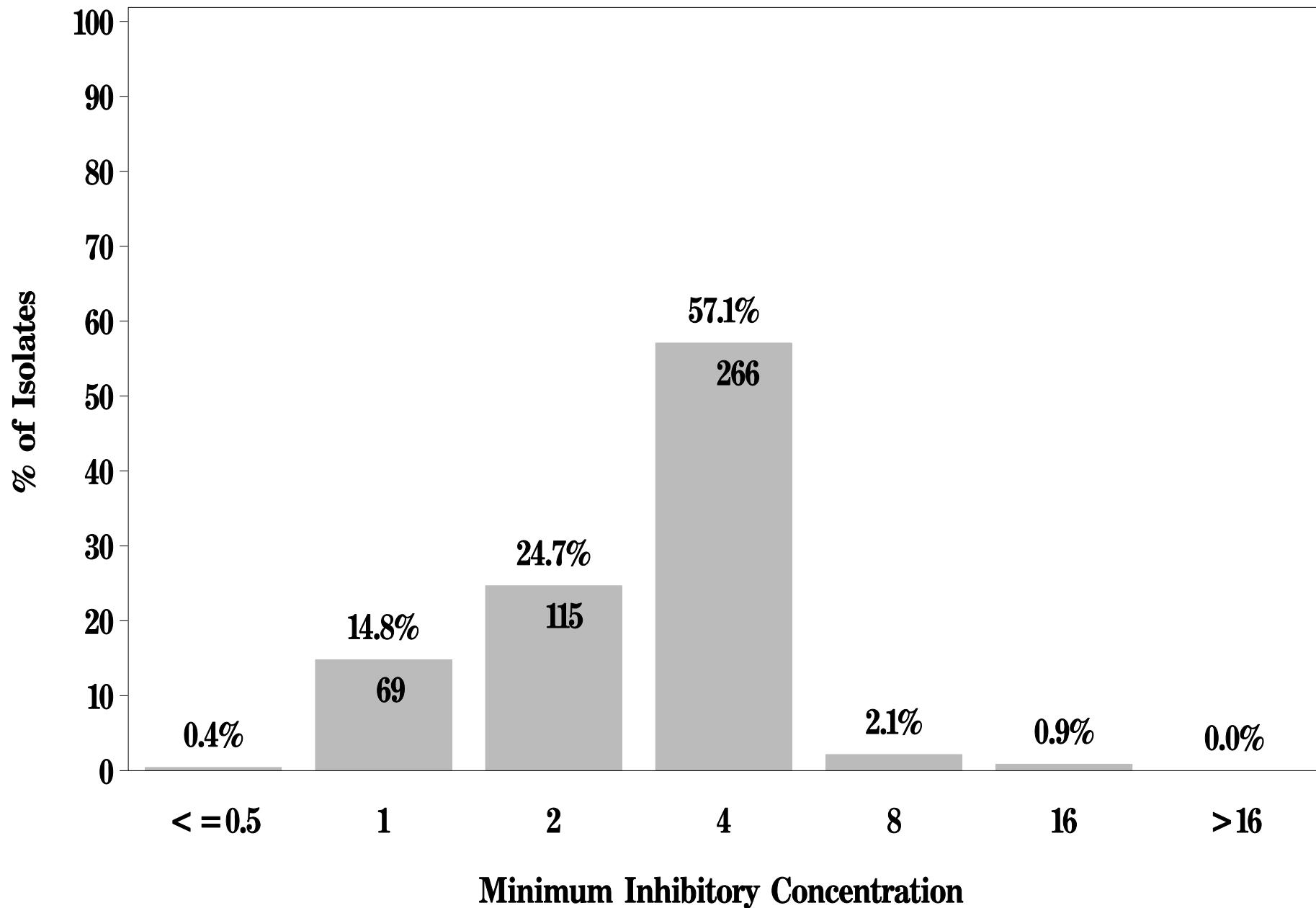
Breakpoints: Susceptible $\leq 1 \mu\text{g/mL}$ Resistant $\geq 4 \mu\text{g/mL}$



NARMS

Figure 15d: Minimum Inhibitory Concentration of Daptomycin
for *Enterococcus* in Chicken Breast (N=466 Isolates)

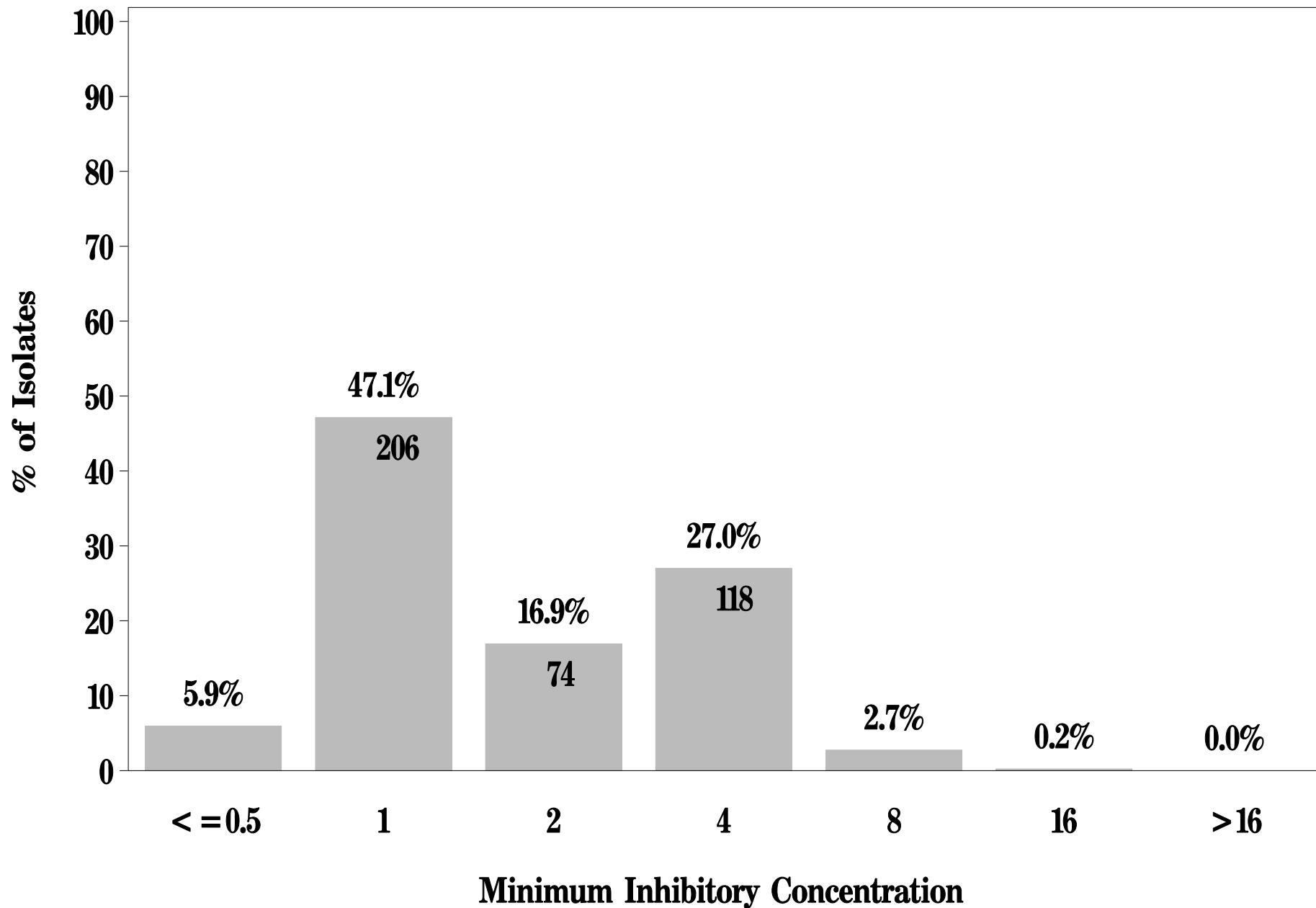
Breakpoint: Susceptible $\leq 4 \mu\text{g/mL}$



NARMS

Figure 15d: Minimum Inhibitory Concentration of Daptomycin
for *Enterococcus* in Ground Turkey (N=437 Isolates)

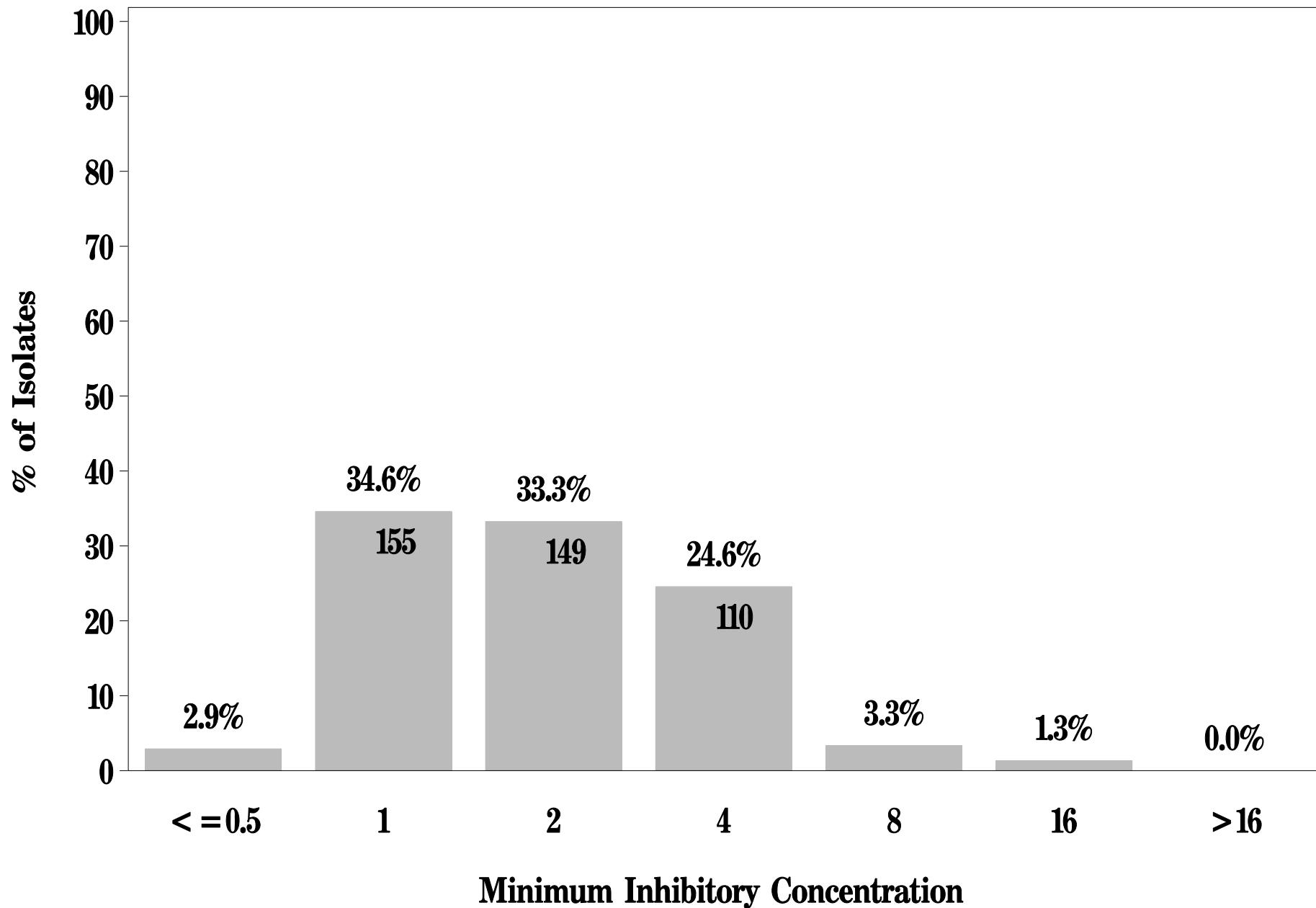
Breakpoint: Susceptible $\leq 4 \mu\text{g/mL}$



NARMS

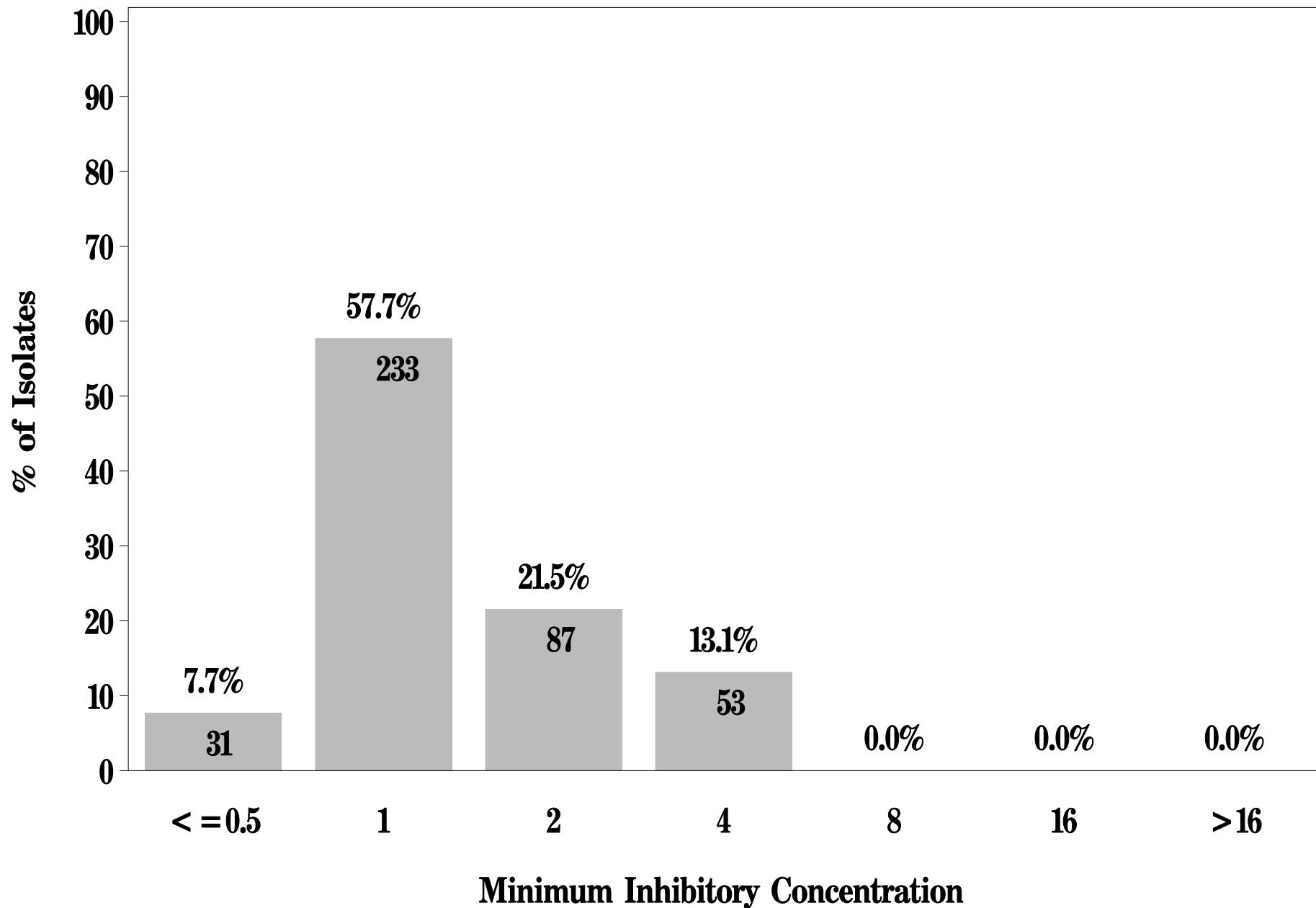
**Figure 15d: Minimum Inhibitory Concentration of Daptomycin
for *Enterococcus* in Ground Beef (N=448 Isolates)**

Breakpoint: Susceptible $\leq 4 \mu\text{g/mL}$



NARMS

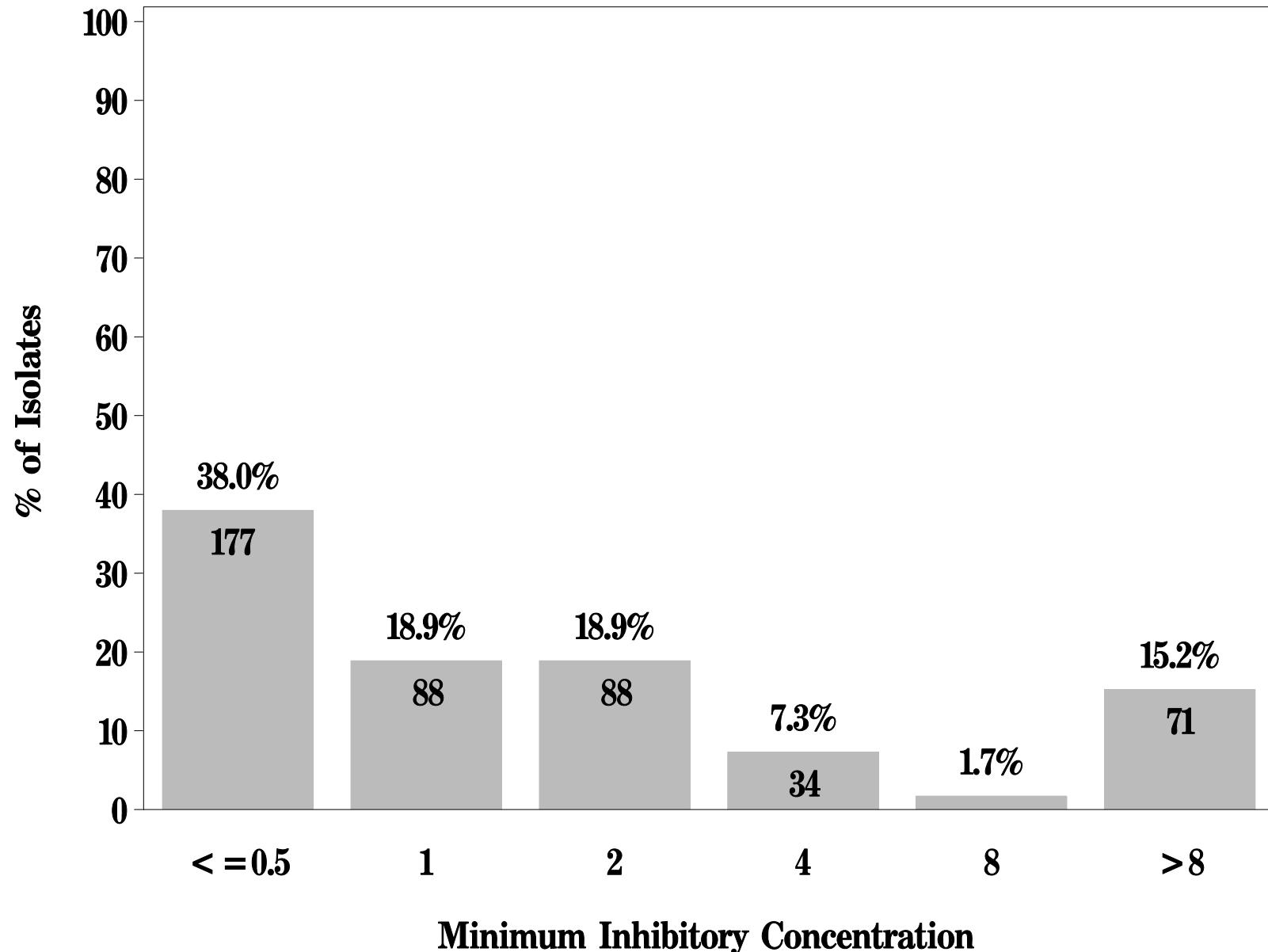
Figure 15d: Minimum Inhibitory Concentration of Daptomycin
for *Enterococcus* in Pork Chop (N=404 Isolates)
Breakpoint: Susceptible $\leq 4 \mu\text{g/mL}$



NARMS

**Figure 15e: Minimum Inhibitory Concentration of Erythromycin
for *Enterococcus* in Chicken Breast (N=466 Isolates)**

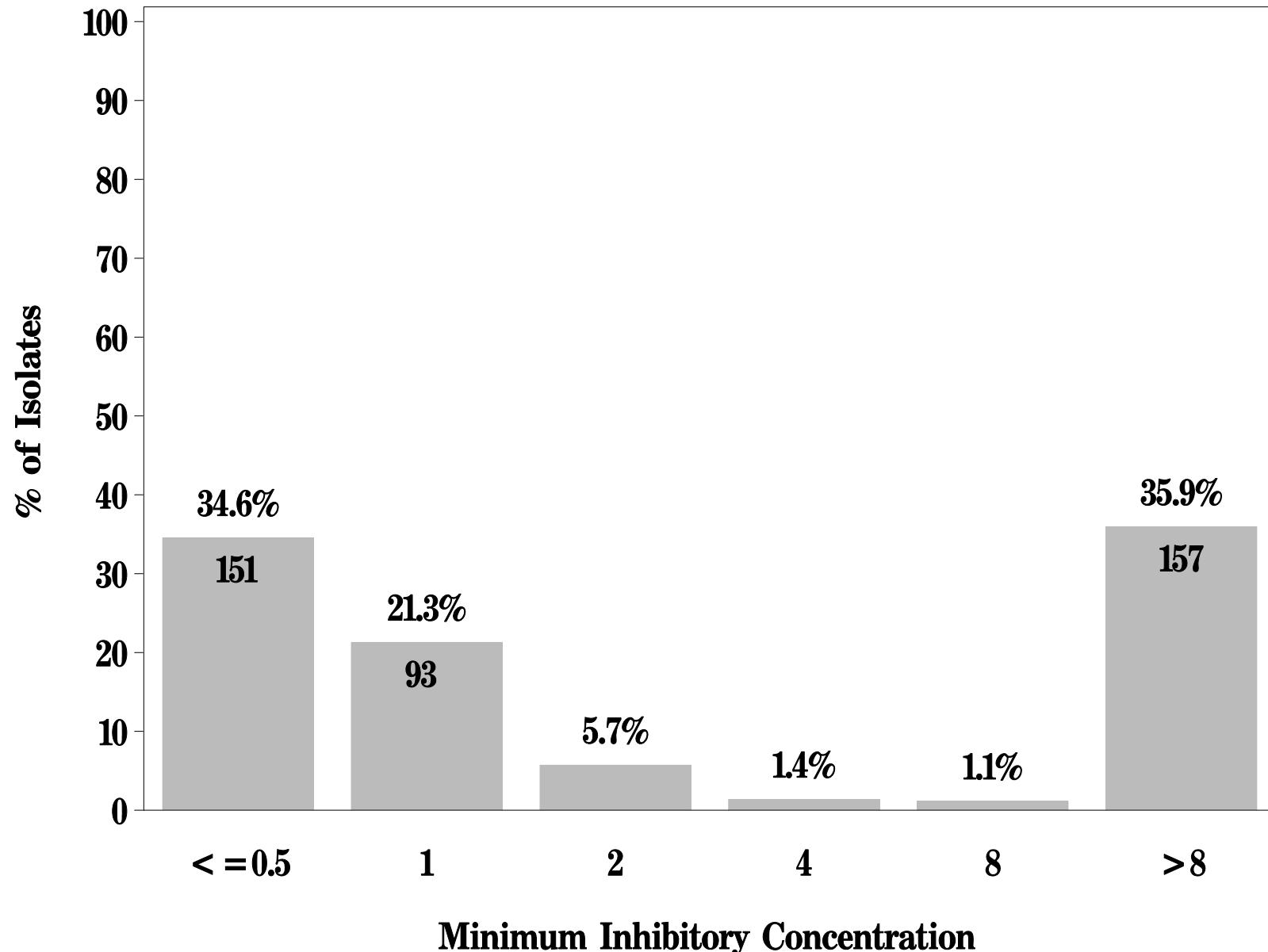
Breakpoints: Susceptible $\leq 0.5 \mu\text{g/mL}$ Resistant $\geq 8 \mu\text{g/mL}$



NARMS

**Figure 15e: Minimum Inhibitory Concentration of Erythromycin
for *Enterococcus* in Ground Turkey (N=437 Isolates)**

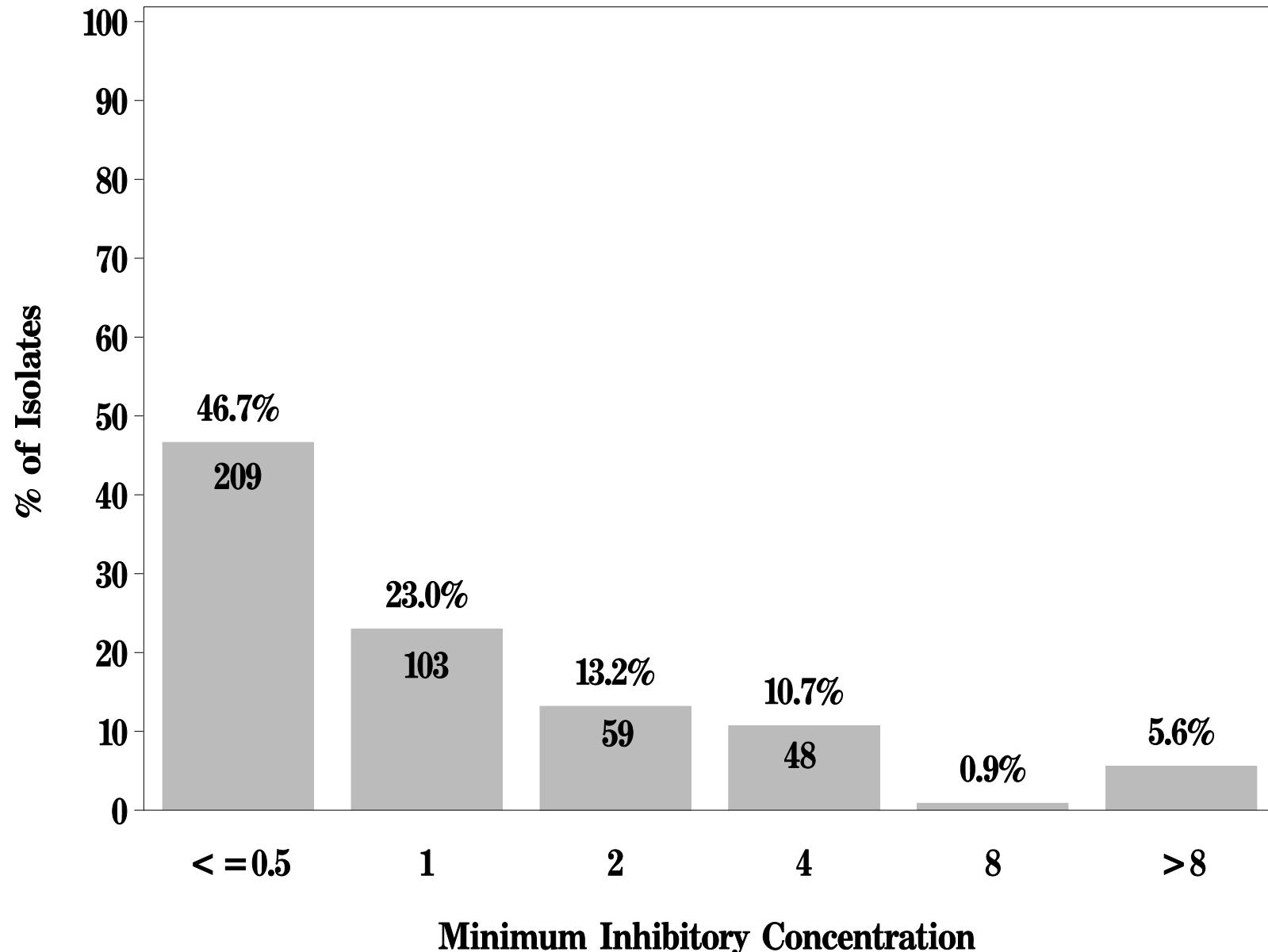
Breakpoints: Susceptible $\leq 0.5 \mu\text{g/mL}$ Resistant $\geq 8 \mu\text{g/mL}$



NARMS

**Figure 15e: Minimum Inhibitory Concentration of Erythromycin
for *Enterococcus* in Ground Beef (N=448 Isolates)**

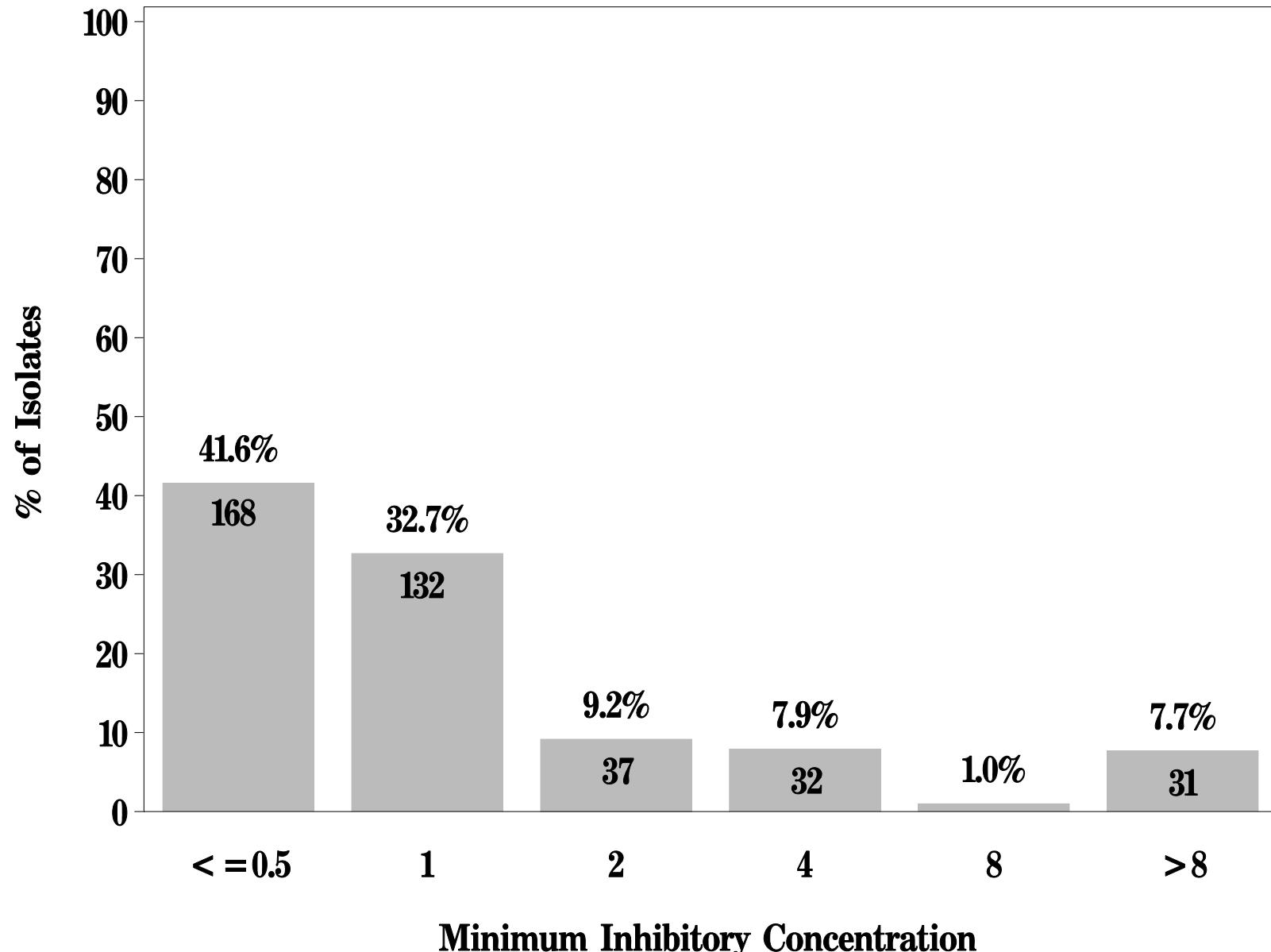
Breakpoints: Susceptible $\leq 0.5 \mu\text{g/mL}$ Resistant $\geq 8 \mu\text{g/mL}$



NARMS

**Figure 15e: Minimum Inhibitory Concentration of Erythromycin
for *Enterococcus* in Pork Chop (N=404 Isolates)**

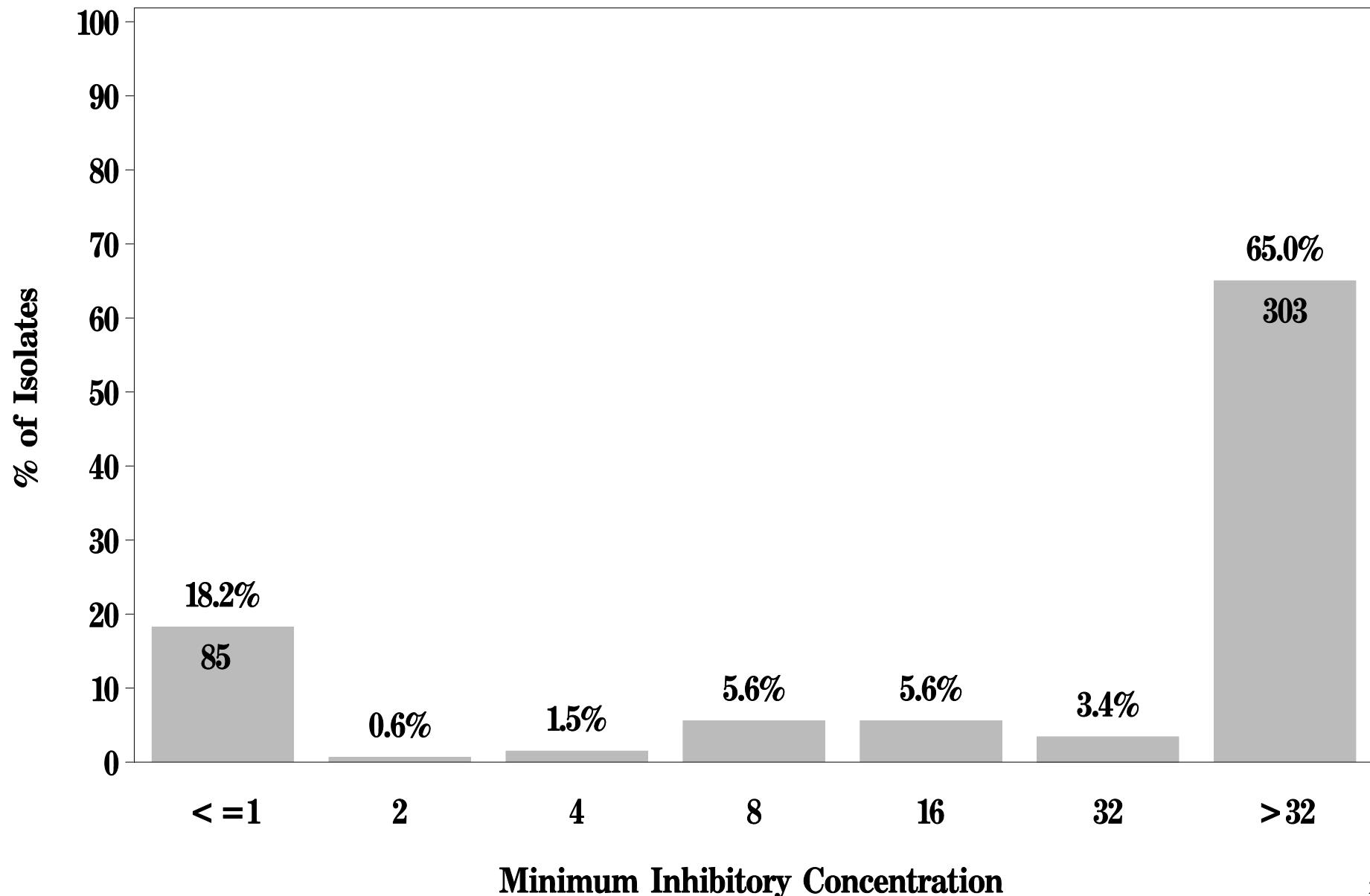
Breakpoints: Susceptible $\leq 0.5 \mu\text{g/mL}$ Resistant $\geq 8 \mu\text{g/mL}$



NARMS

Figure 15f: Minimum Inhibitory Concentration of Flavomycin for *Enterococcus* in Chicken Breast (N=466 Isolates)

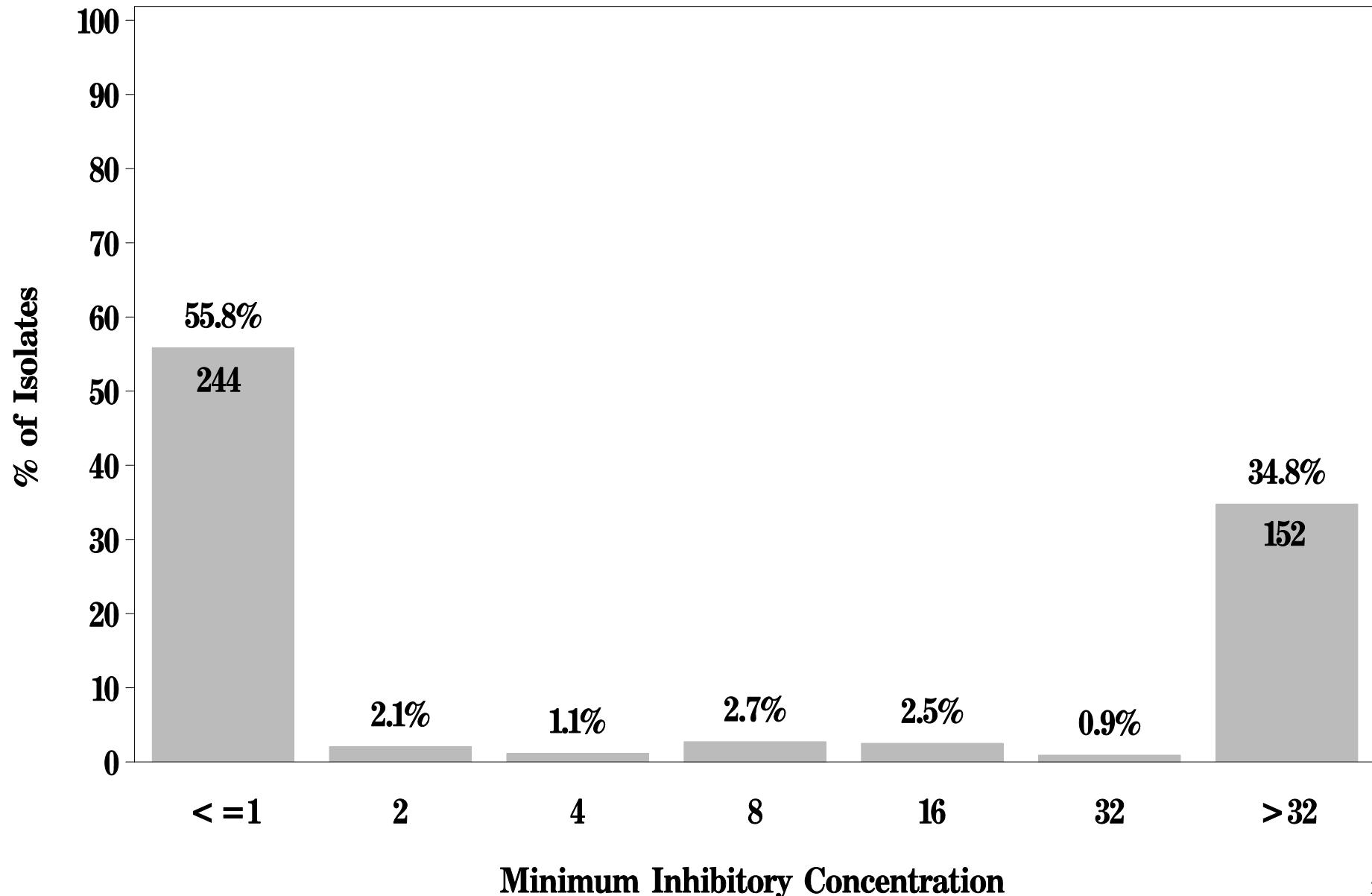
Breakpoints: Susceptible $\leq 8 \mu\text{g/mL}$ Resistant $\geq 32 \mu\text{g/mL}$



NARMS

Figure 15f: Minimum Inhibitory Concentration of Flavomycin for *Enterococcus* in Ground Turkey (N=437 Isolates)

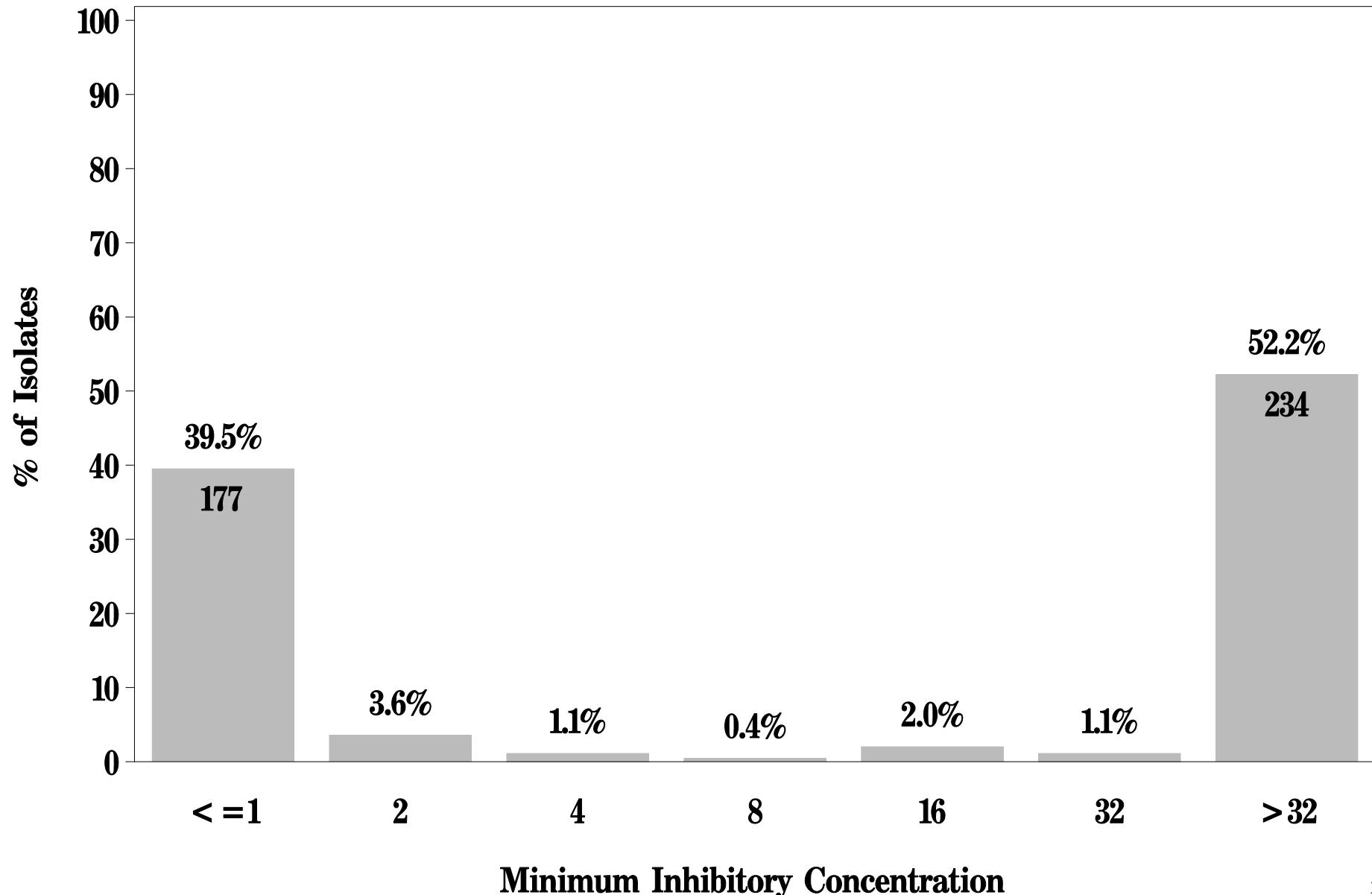
Breakpoints: Susceptible $\leq 8 \mu\text{g/mL}$ Resistant $\geq 32 \mu\text{g/mL}$



NARMS

**Figure 15f: Minimum Inhibitory Concentration of Flavomycin
for *Enterococcus* in Ground Beef (N=448 Isolates)**

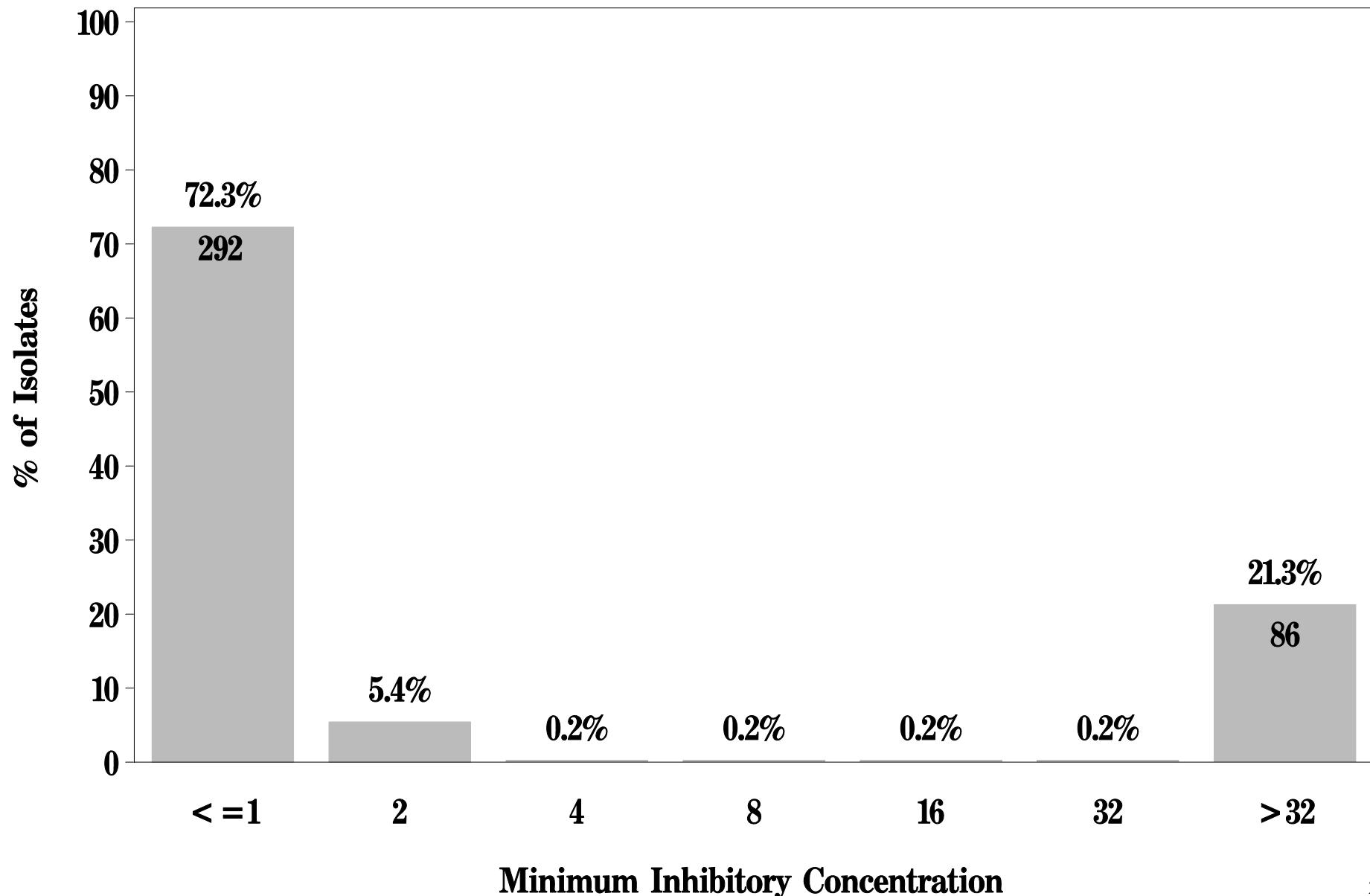
Breakpoints: Susceptible $\leq 8 \mu\text{g/mL}$ Resistant $\geq 32 \mu\text{g/mL}$



NARMS

**Figure 15f: Minimum Inhibitory Concentration of Flavomycin
for *Enterococcus* in Pork Chop (N=404 Isolates)**

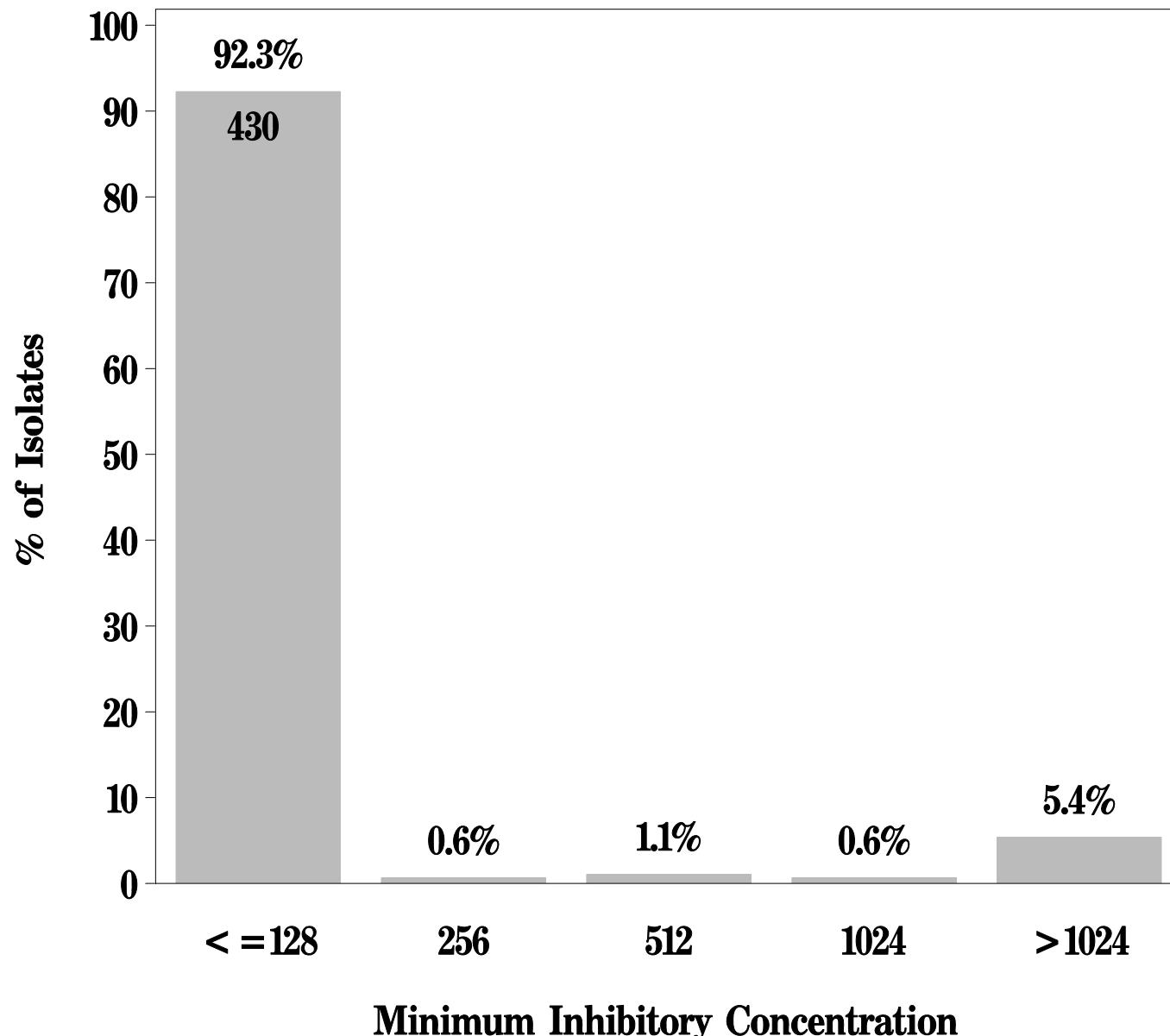
Breakpoints: Susceptible $\leq 8 \mu\text{g/mL}$ Resistant $\geq 32 \mu\text{g/mL}$



NARMS

**Figure 15g: Minimum Inhibitory Concentration of Gentamicin
for *Enterococcus* in Chicken Breast (N=466 Isolates)**

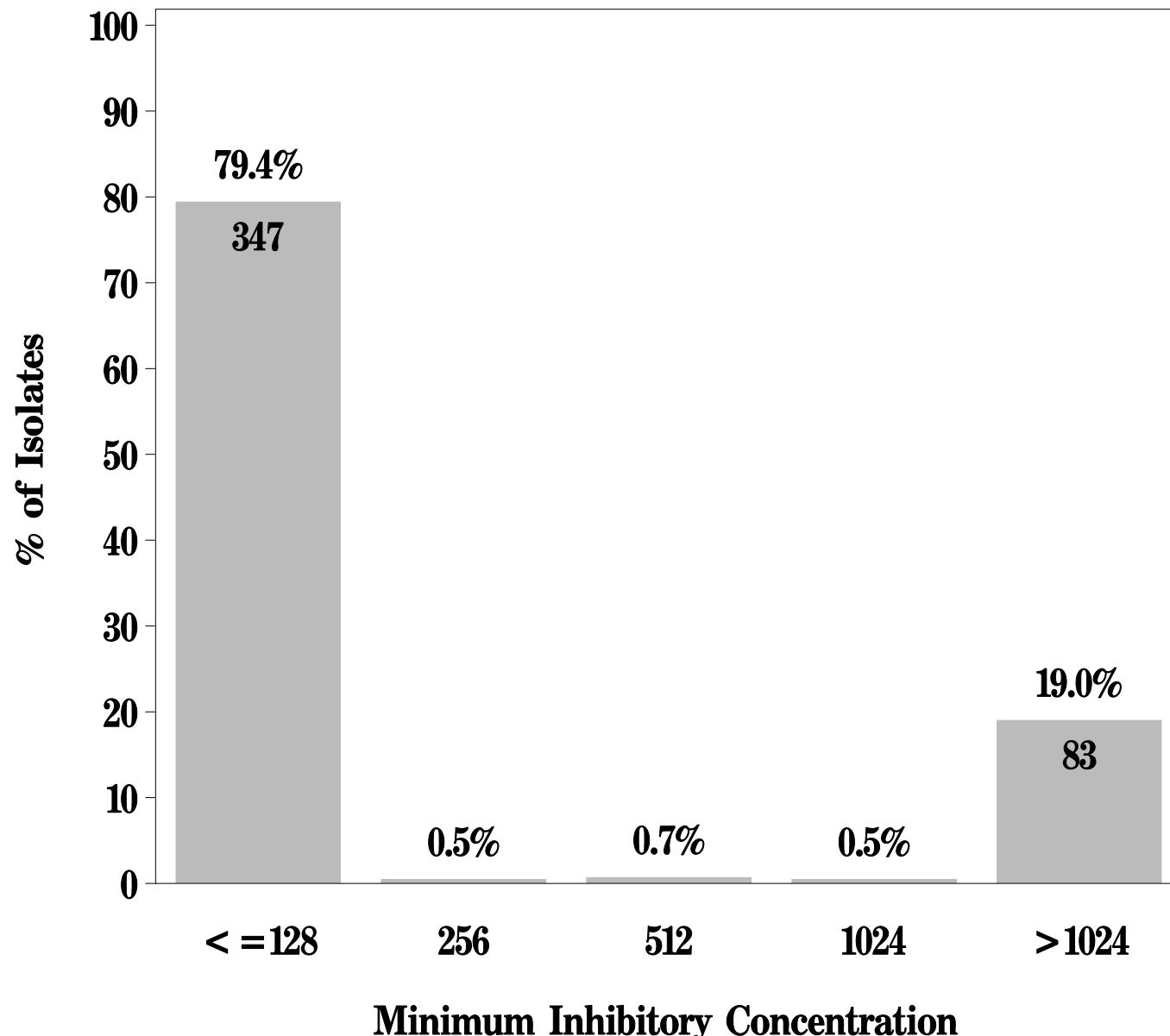
Breakpoints: Susceptible $\leq 500 \mu\text{g/mL}$ Resistant $\geq 500 \mu\text{g/mL}$



NARMS

**Figure 15g: Minimum Inhibitory Concentration of Gentamicin
for *Enterococcus* in Ground Turkey (N=437 Isolates)**

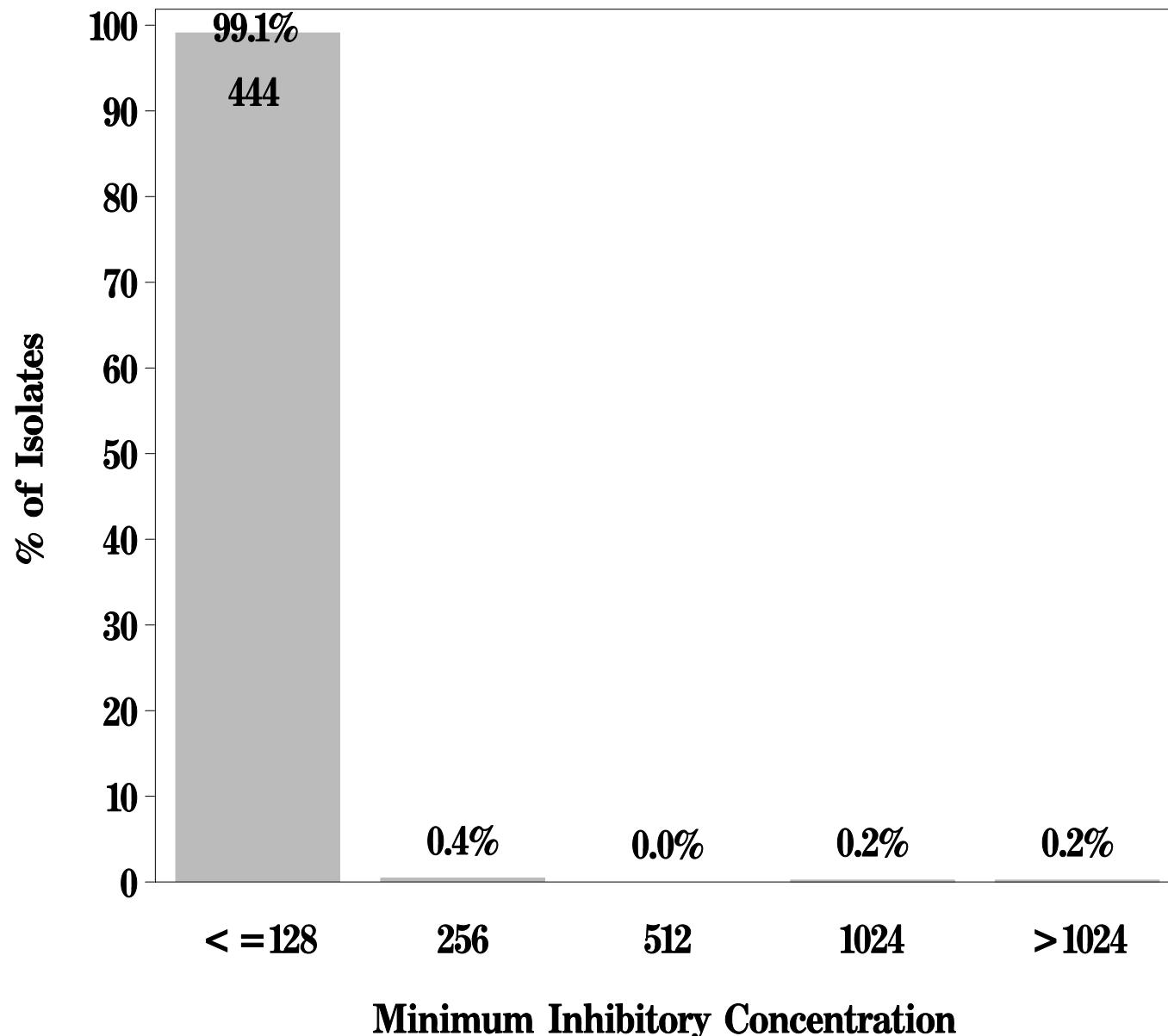
Breakpoints: Susceptible $\leq 500 \mu\text{g/mL}$ Resistant $\geq 500 \mu\text{g/mL}$



NARMS

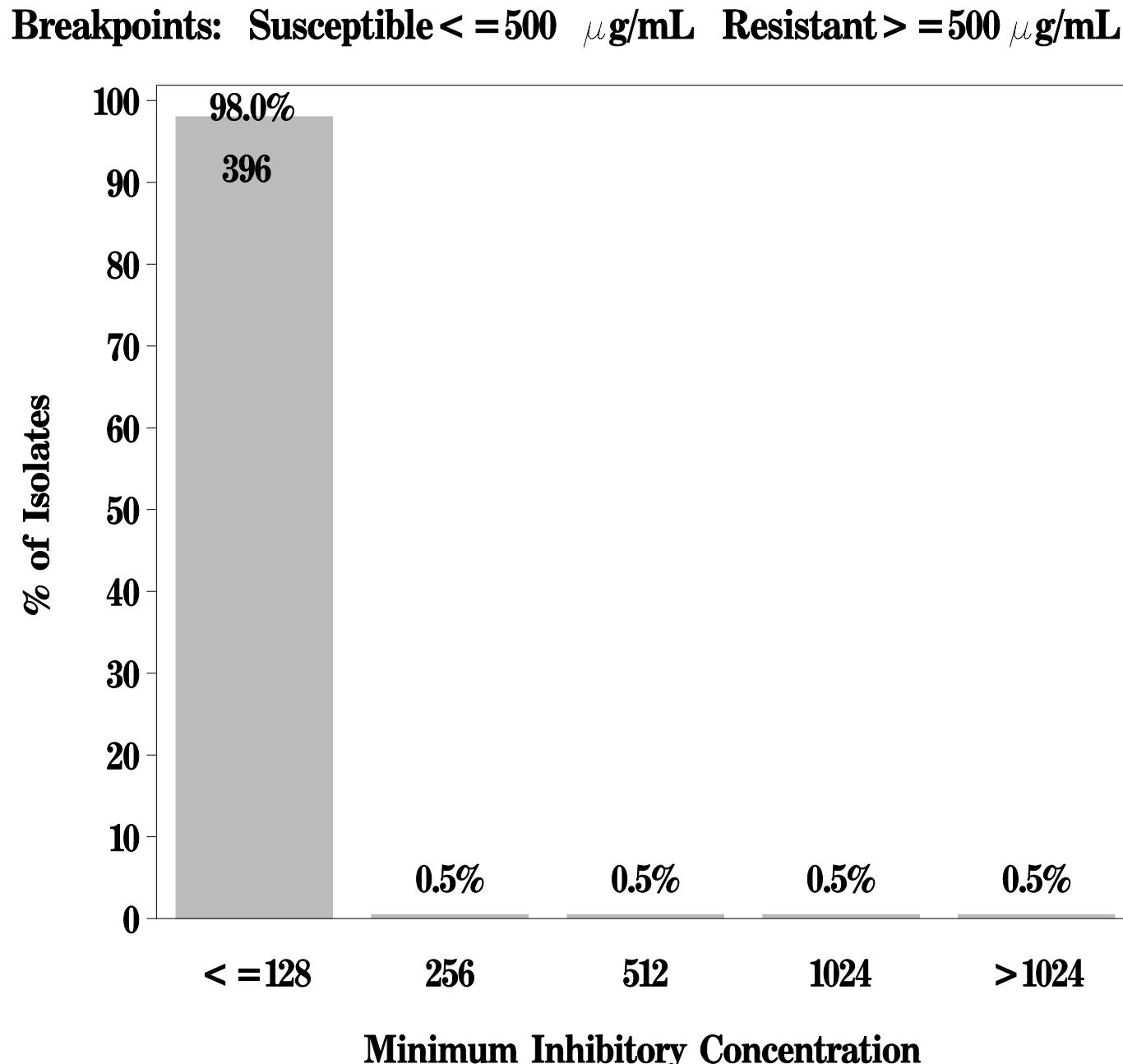
**Figure 15g: Minimum Inhibitory Concentration of Gentamicin
for *Enterococcus* in Ground Beef (N=448 Isolates)**

Breakpoints: Susceptible $\leq 500 \mu\text{g/mL}$ Resistant $\geq 500 \mu\text{g/mL}$



NARMS

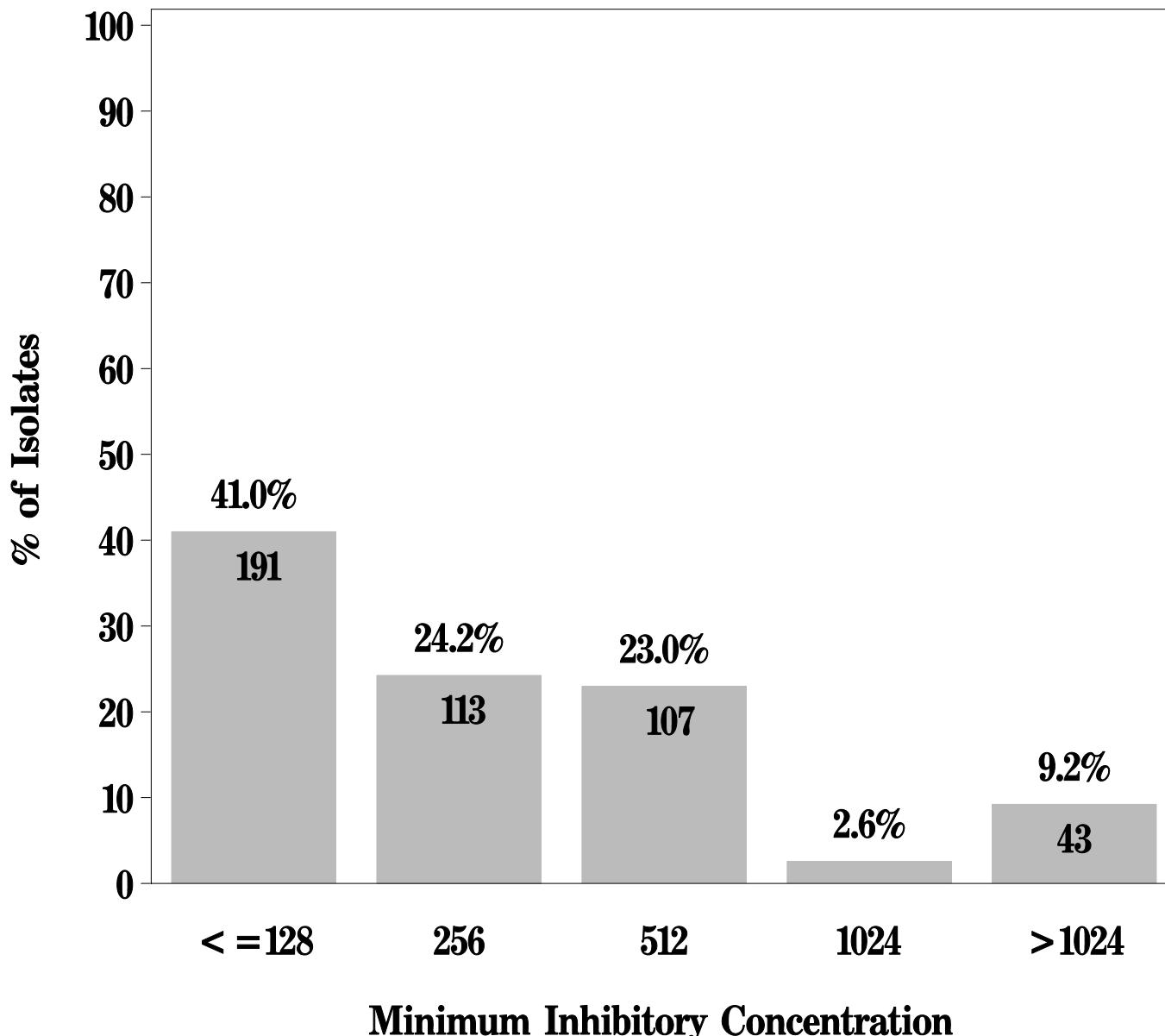
**Figure 15g: Minimum Inhibitory Concentration of Gentamicin
for *Enterococcus* in Pork Chop (N=404 Isolates)**



NARMS

**Figure 15h: Minimum Inhibitory Concentration of Kanamycin
for *Enterococcus* in Chicken Breast (N=466 Isolates)**

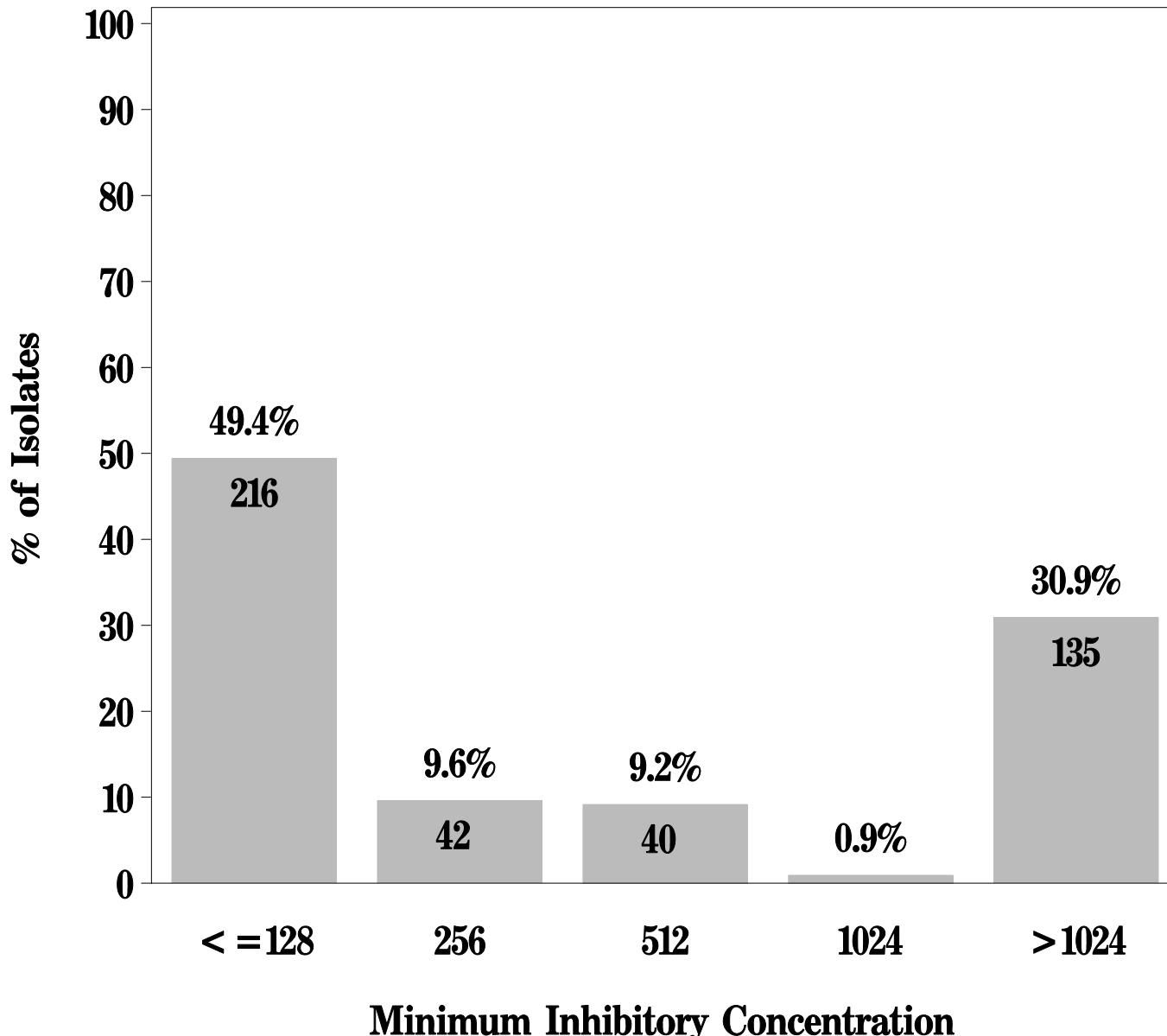
Breakpoints: Susceptible $\leq 128 \mu\text{g/mL}$ Resistant $\geq 512 \mu\text{g/mL}$



NARMS

**Figure 15h: Minimum Inhibitory Concentration of Kanamycin
for *Enterococcus* in Ground Turkey (N=437 Isolates)**

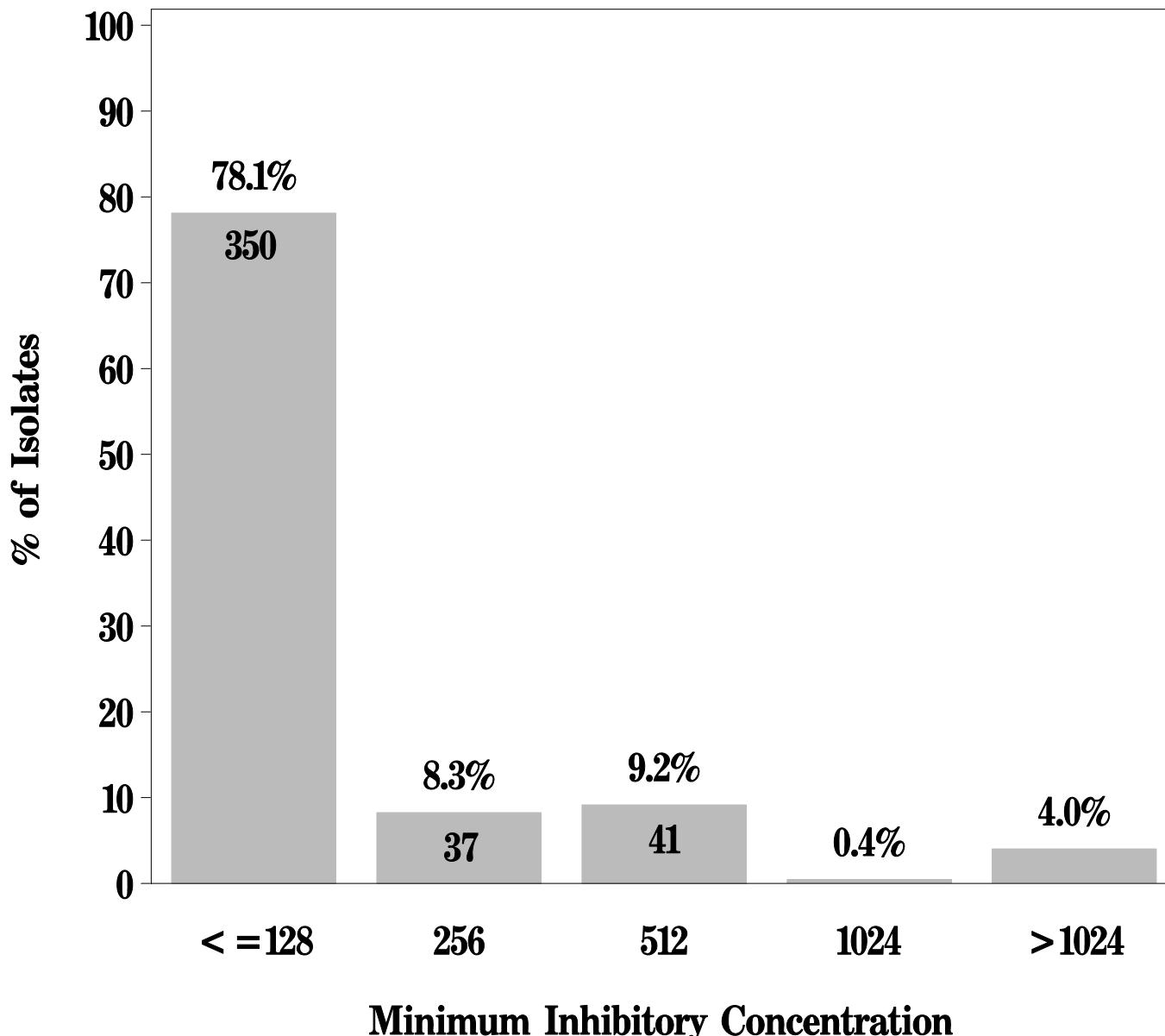
Breakpoints: Susceptible $\leq 128 \mu\text{g/mL}$ Resistant $\geq 512 \mu\text{g/mL}$



NARMS

**Figure 15h: Minimum Inhibitory Concentration of Kanamycin
for *Enterococcus* in Ground Beef (N=448 Isolates)**

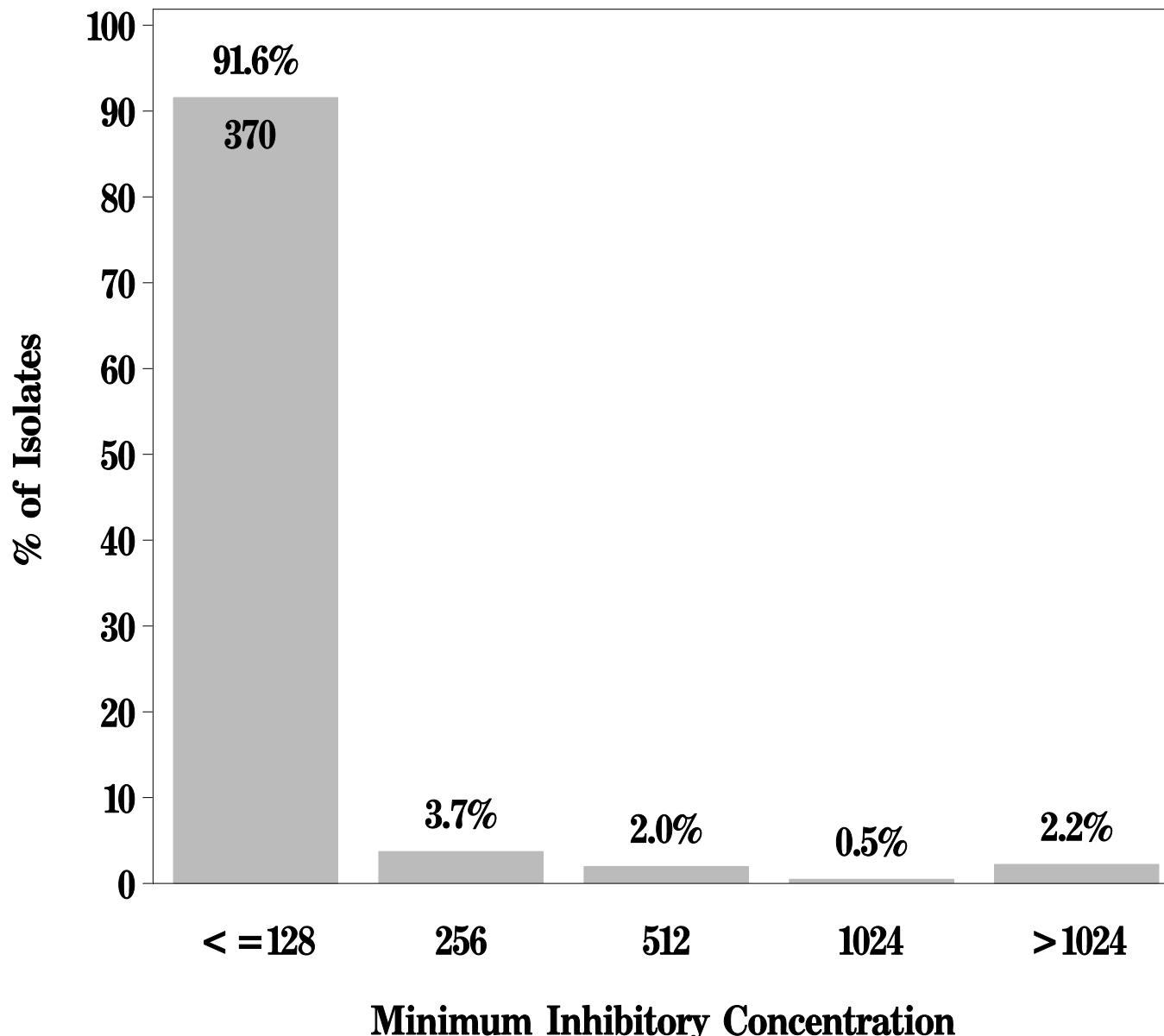
Breakpoints: Susceptible $\leq 128 \mu\text{g/mL}$ Resistant $\geq 512 \mu\text{g/mL}$



NARMS

**Figure 15h: Minimum Inhibitory Concentration of Kanamycin
for *Enterococcus* in Pork Chop (N=404 Isolates)**

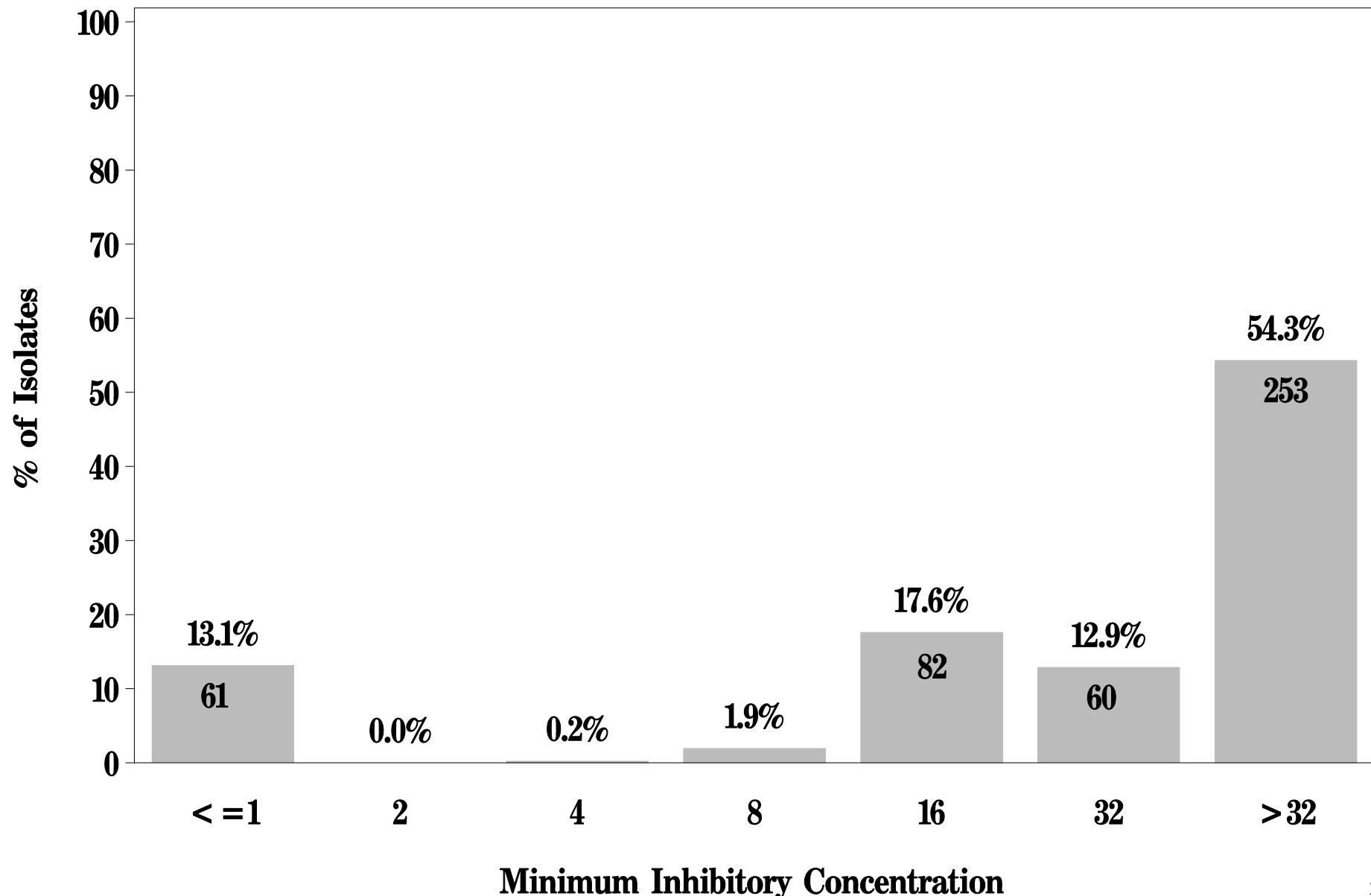
Breakpoints: Susceptible $\leq 128 \mu\text{g/mL}$ Resistant $\geq 512 \mu\text{g/mL}$



NARMS

**Figure 15i: Minimum Inhibitory Concentration of Lincomycin
for *Enterococcus* in Chicken Breast (N=466 Isolates)**

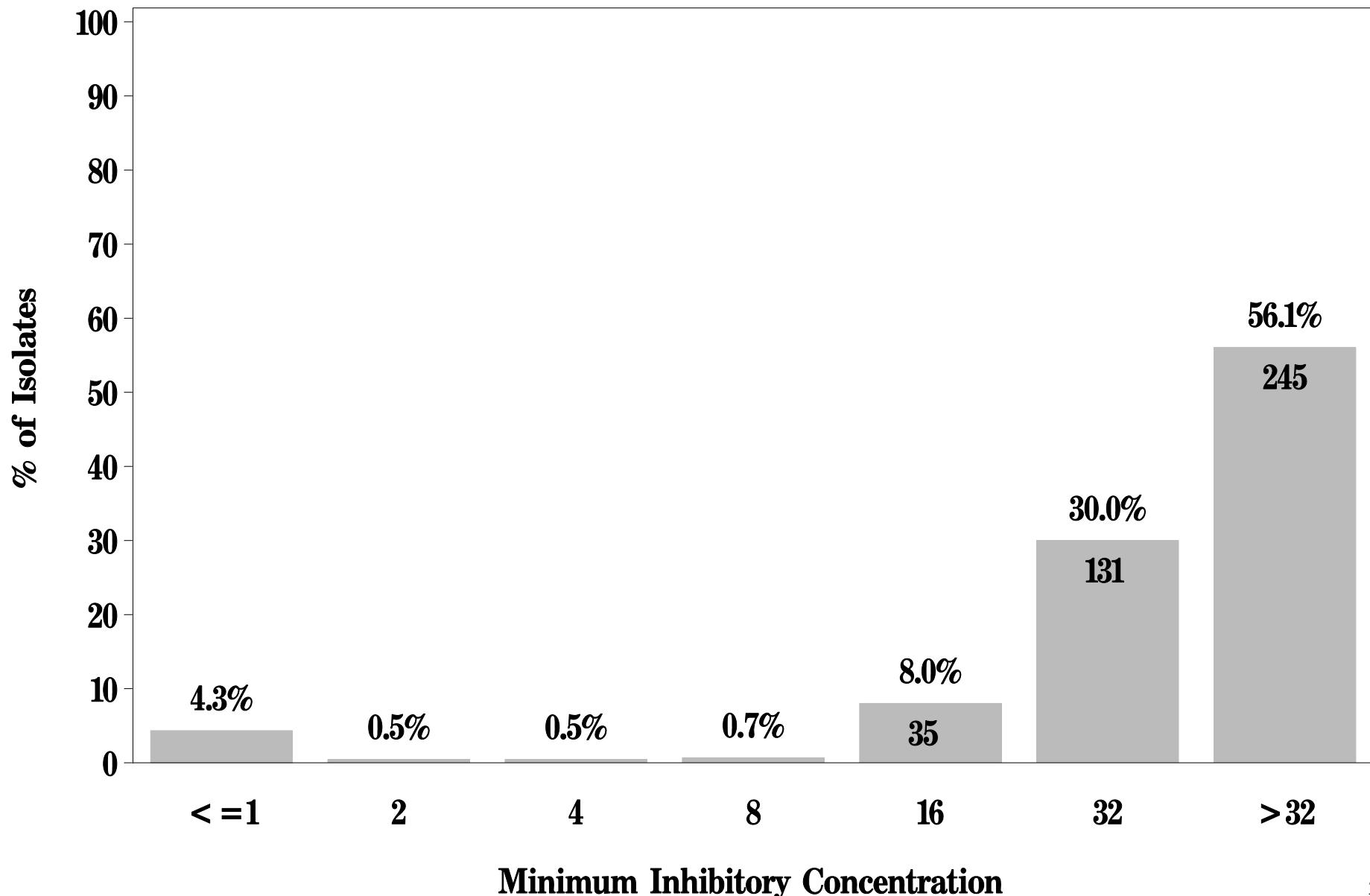
Breakpoints: Susceptible $\leq 8 \mu\text{g/mL}$ Resistant $\geq 32 \mu\text{g/mL}$



NARMS

**Figure 15i: Minimum Inhibitory Concentration of Lincomycin
for *Enterococcus* in Ground Turkey (N=437 Isolates)**

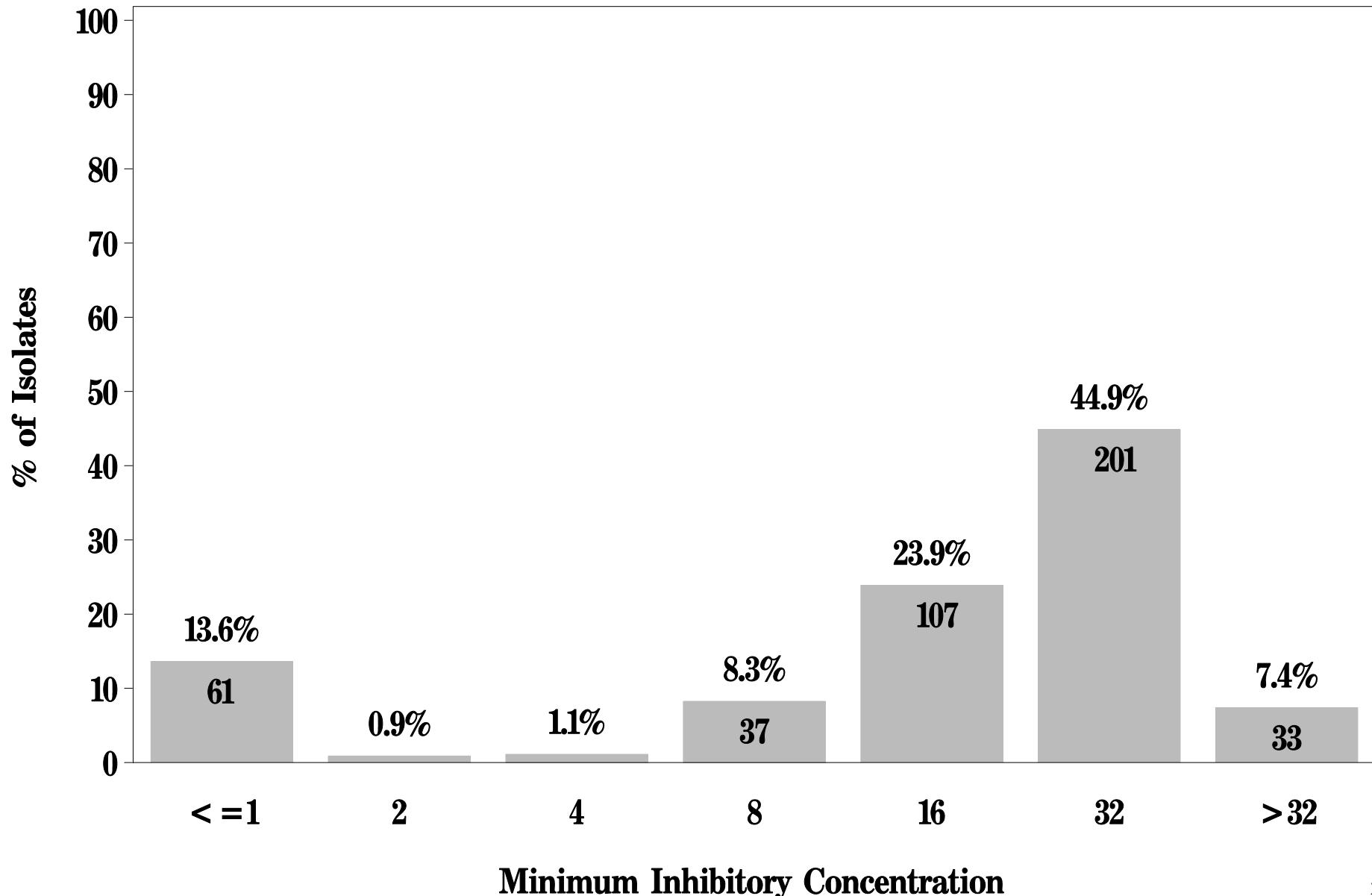
Breakpoints: Susceptible $\leq 8 \mu\text{g/mL}$ Resistant $\geq 32 \mu\text{g/mL}$



NARMS

**Figure 15i: Minimum Inhibitory Concentration of Lincomycin
for *Enterococcus* in Ground Beef (N=448 Isolates)**

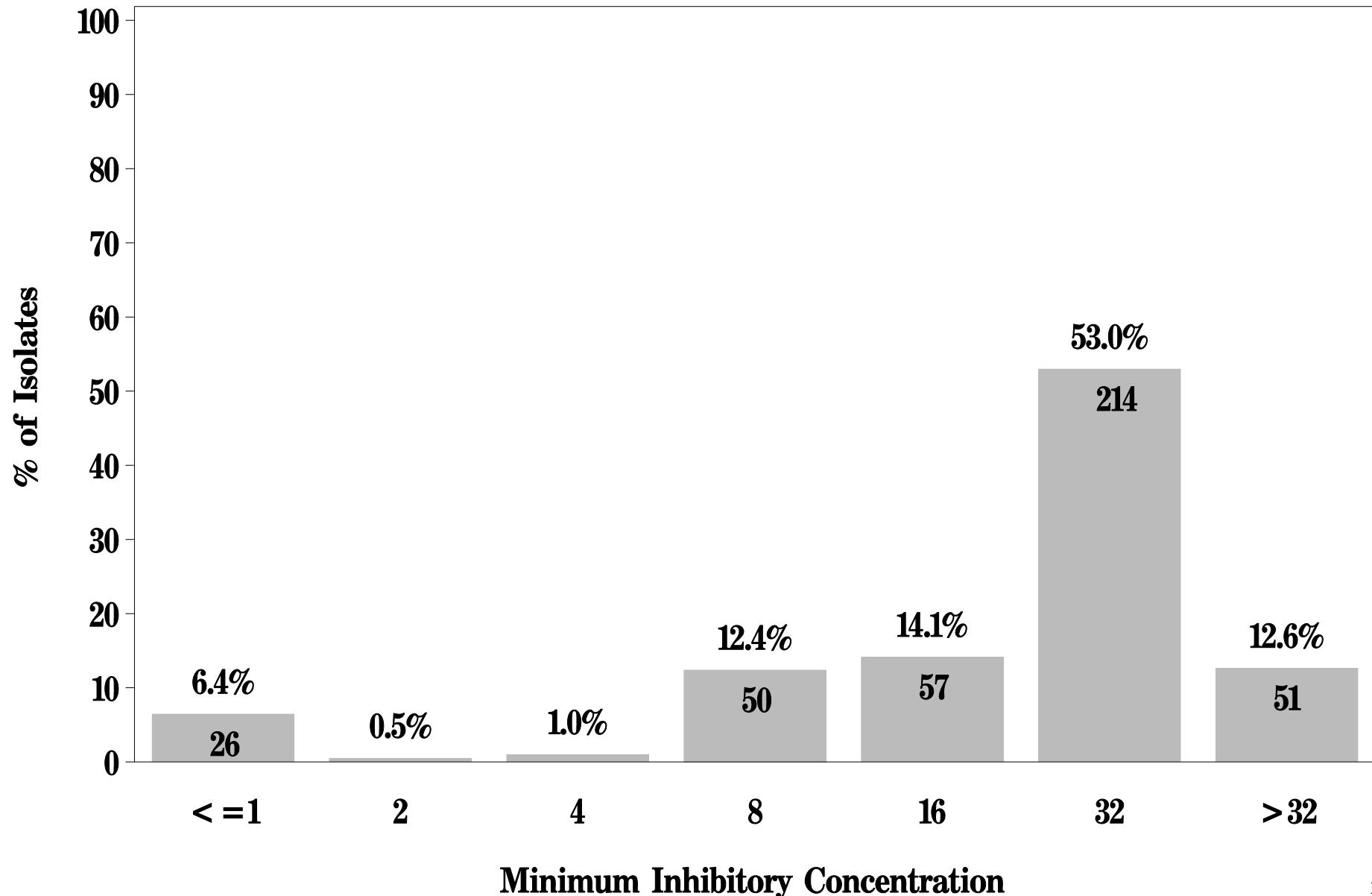
Breakpoints: Susceptible $\leq 8 \mu\text{g/mL}$ Resistant $\geq 32 \mu\text{g/mL}$



NARMS

**Figure 15i: Minimum Inhibitory Concentration of Lincomycin
for *Enterococcus* in Pork Chop (N=404 Isolates)**

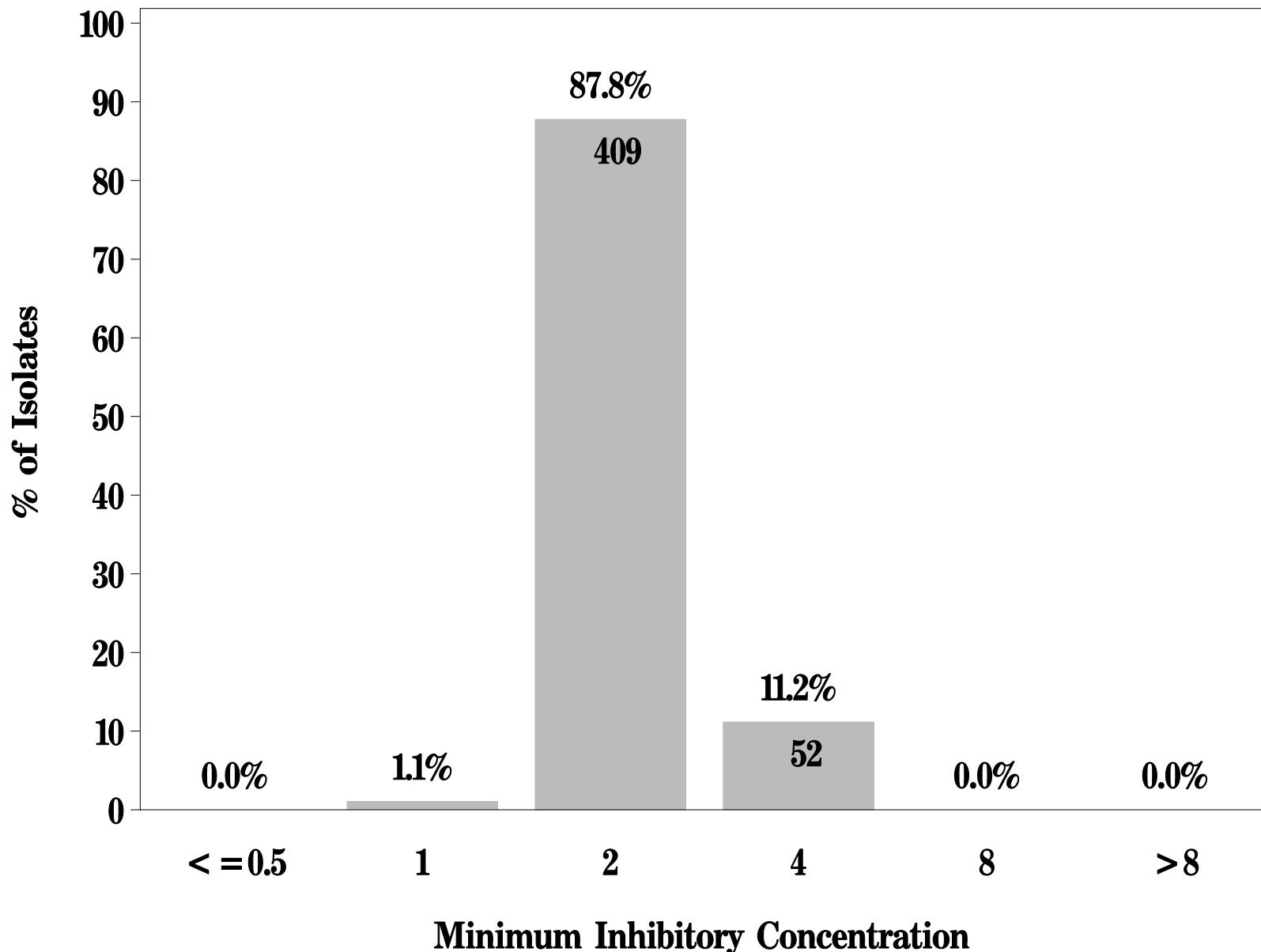
Breakpoints: Susceptible $\leq 8 \mu\text{g/mL}$ Resistant $\geq 32 \mu\text{g/mL}$



NARMS

**Figure 15j: Minimum Inhibitory Concentration of Linezolid
for *Enterococcus* in Chicken Breast (N=466 Isolates)**

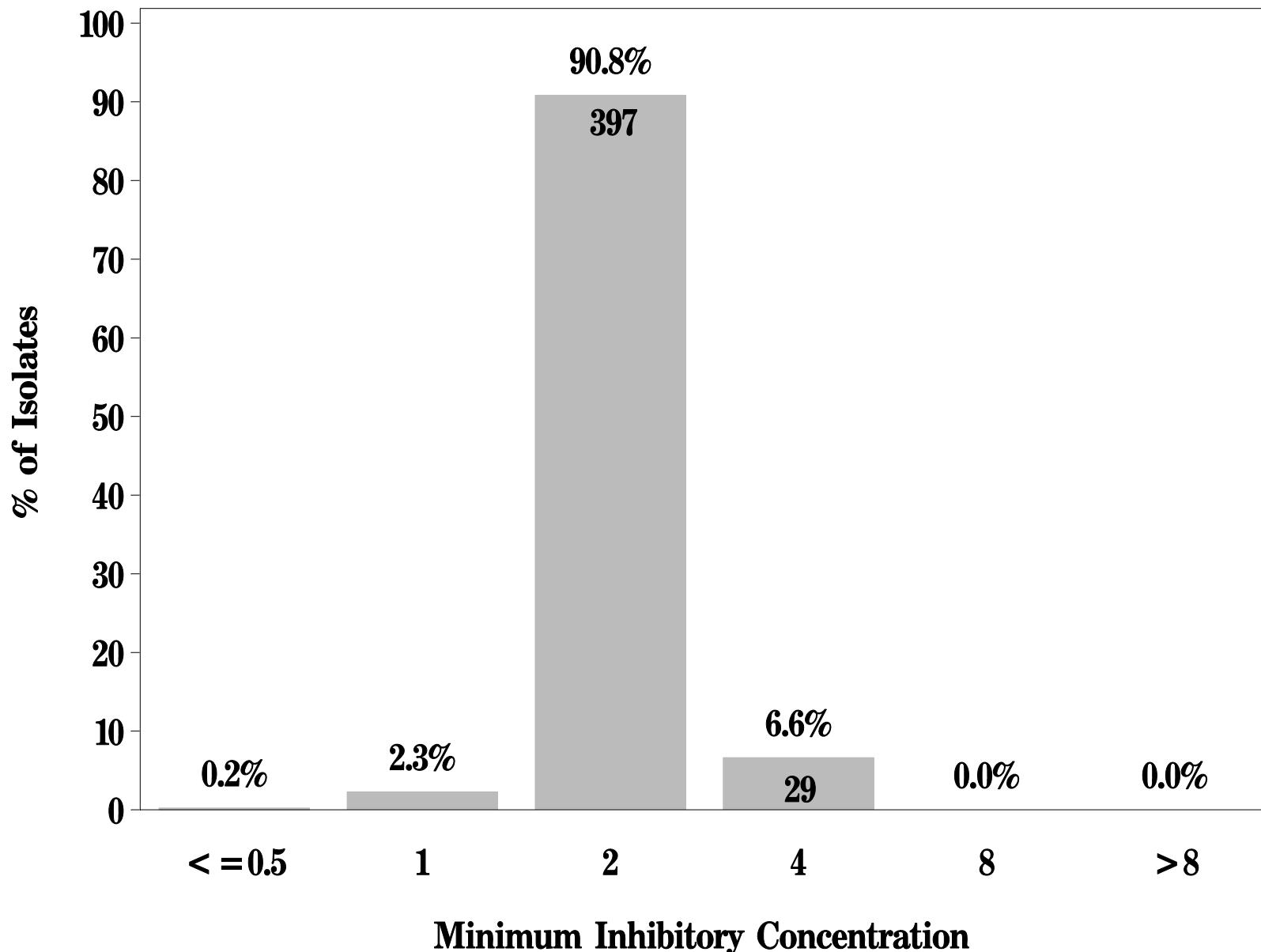
Breakpoints: Susceptible $\leq 2 \mu\text{g/mL}$ Resistant $\geq 8 \mu\text{g/mL}$



NARMS

**Figure 15j: Minimum Inhibitory Concentration of Linezolid
for *Enterococcus* in Ground Turkey (N=437 Isolates)**

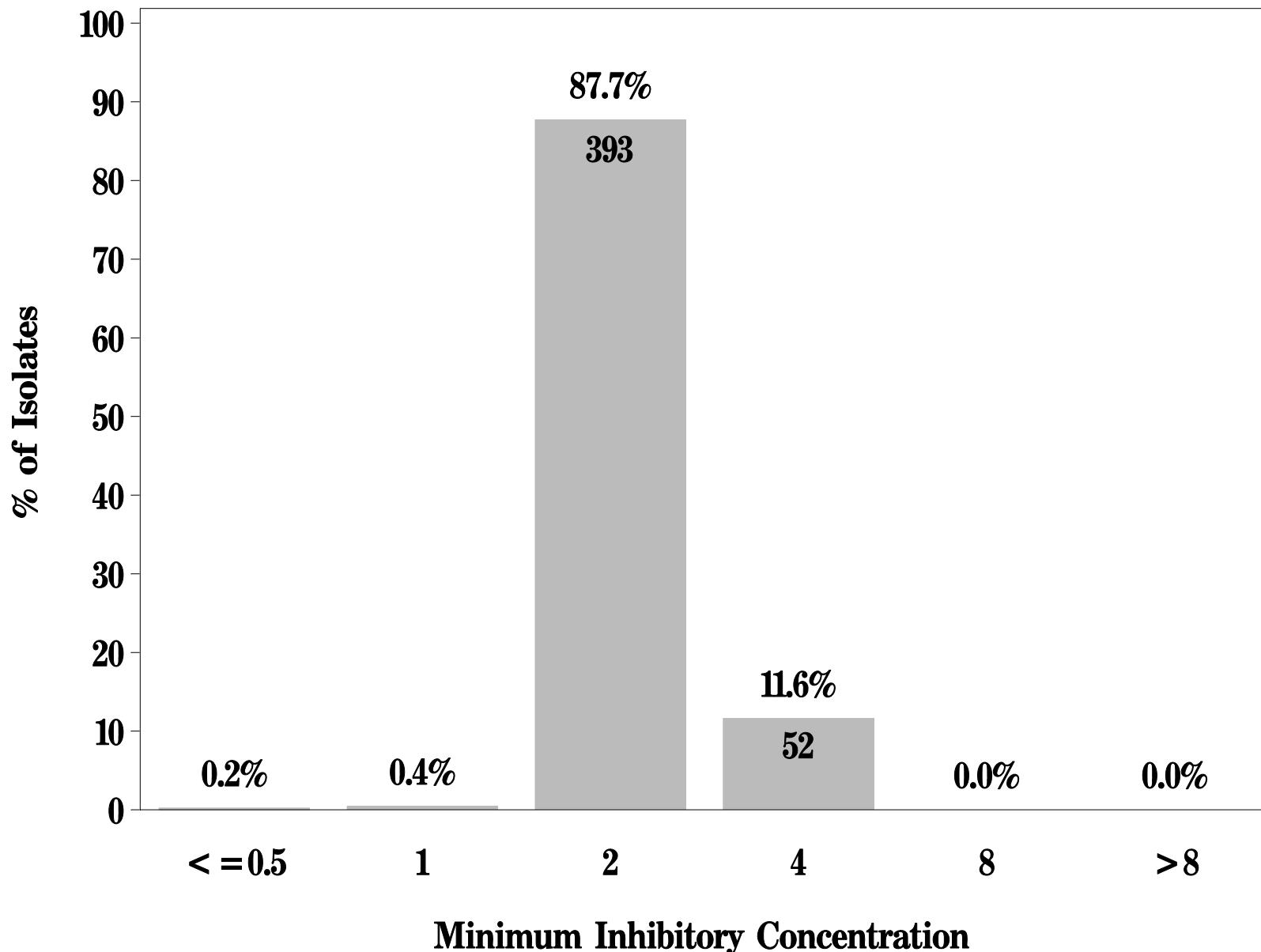
Breakpoints: Susceptible $\leq 2 \mu\text{g/mL}$ Resistant $\geq 8 \mu\text{g/mL}$



NARMS

**Figure 15j: Minimum Inhibitory Concentration of Linezolid
for *Enterococcus* in Ground Beef (N=448 Isolates)**

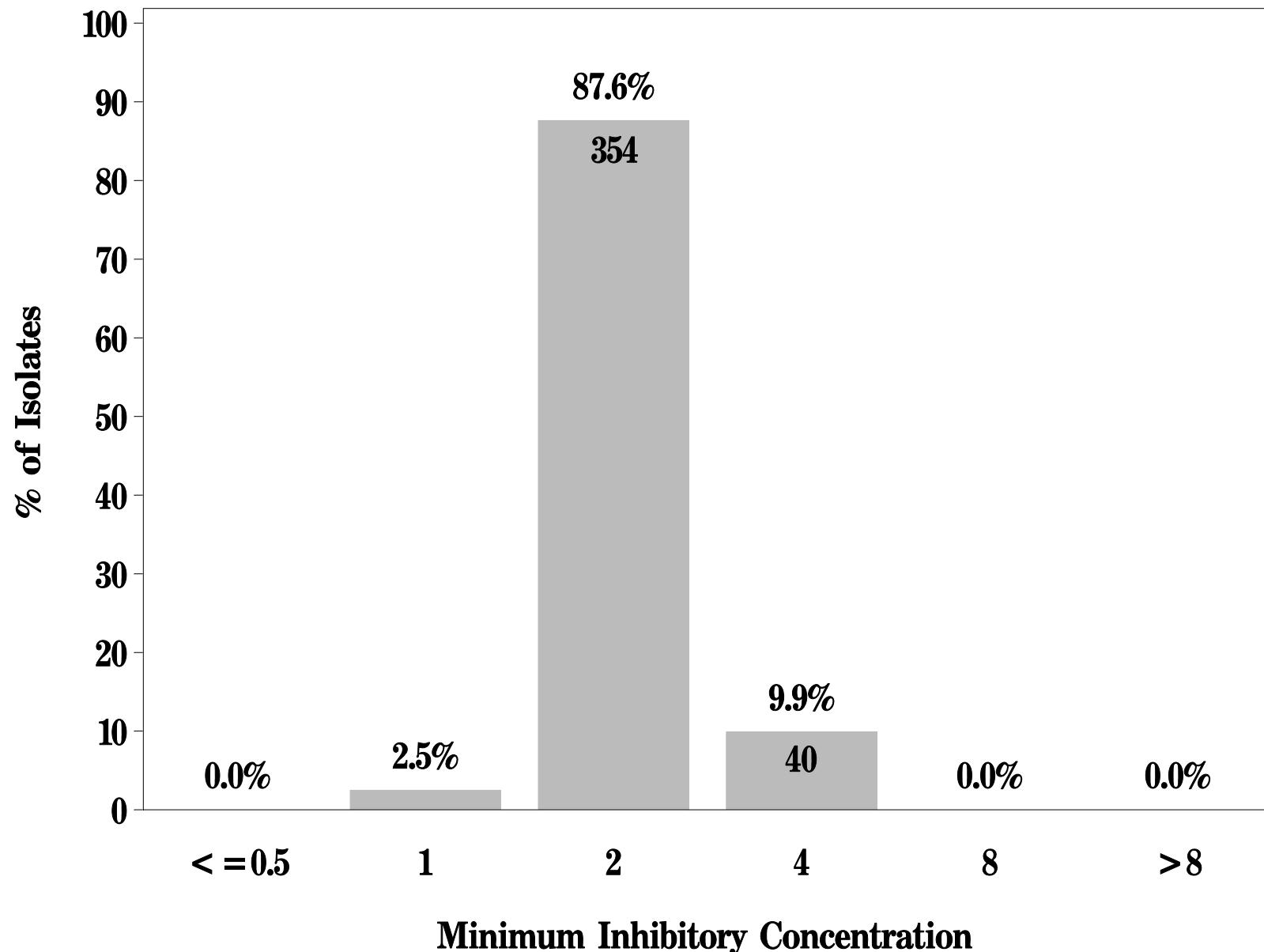
Breakpoints: Susceptible $\leq 2 \mu\text{g/mL}$ Resistant $\geq 8 \mu\text{g/mL}$



NARMS

**Figure 15j: Minimum Inhibitory Concentration of Linezolid
for *Enterococcus* in Pork Chop (N=404 Isolates)**

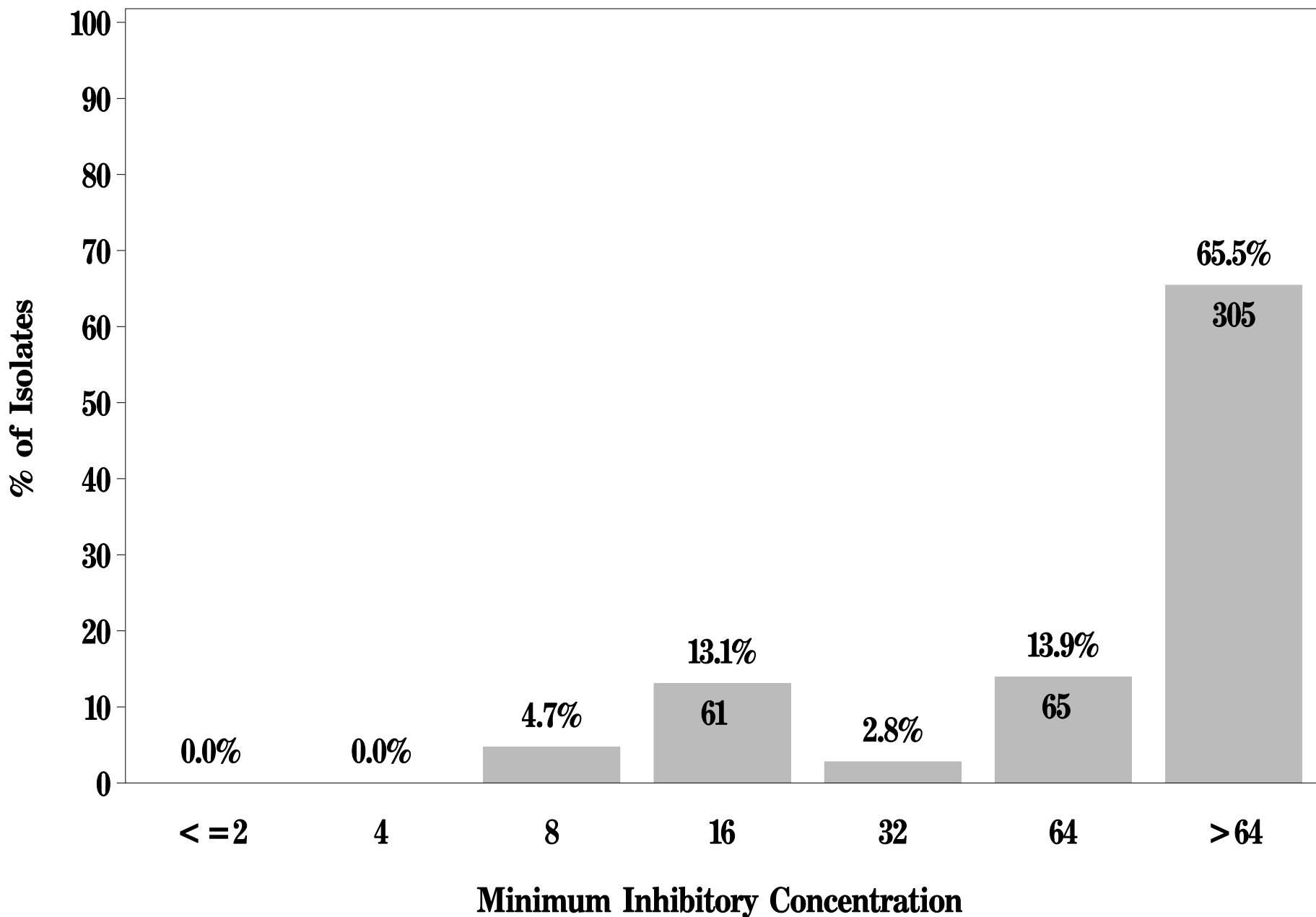
Breakpoints: Susceptible $\leq 2 \mu\text{g/mL}$ Resistant $\geq 8 \mu\text{g/mL}$



NARMS

**Figure 15k: Minimum Inhibitory Concentration of Nitrofurantoin
for *Enterococcus* in Chicken Breast (N=466 Isolates)**

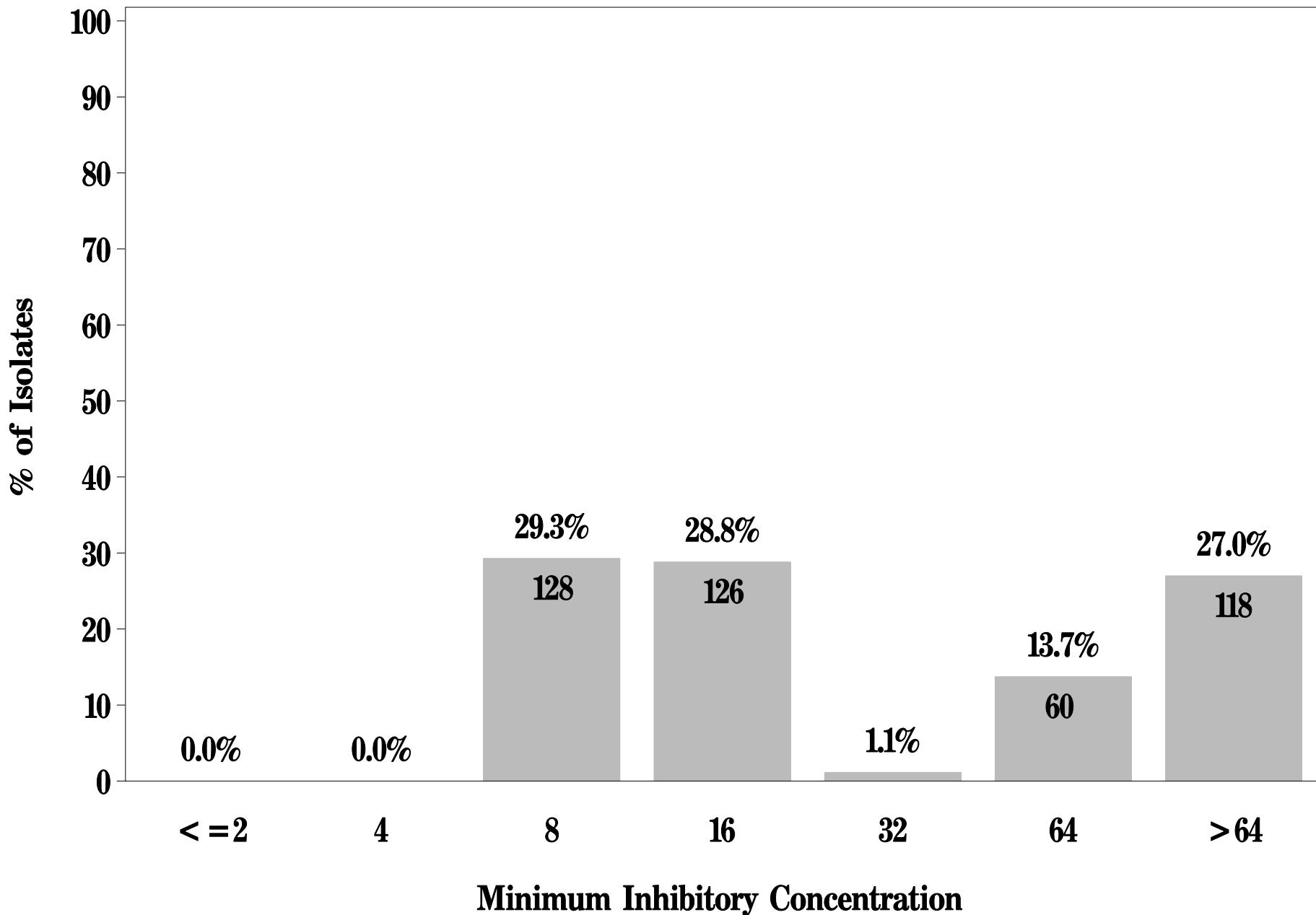
Breakpoints: Susceptible $\leq 32 \mu\text{g/mL}$ Resistant $\geq 128 \mu\text{g/mL}$



NARMS

**Figure 15k: Minimum Inhibitory Concentration of Nitrofurantoin
for *Enterococcus* in Ground Turkey (N=437 Isolates)**

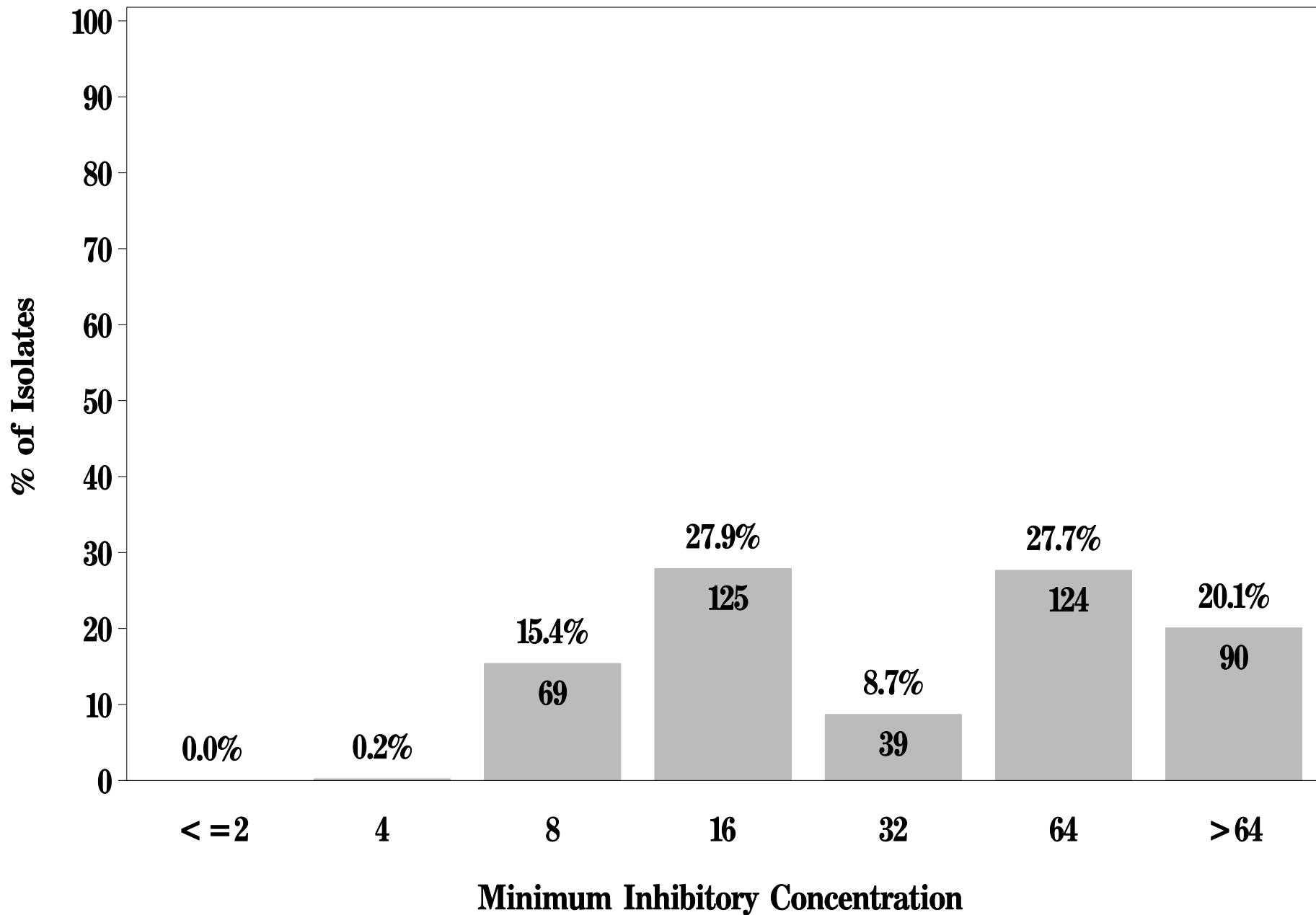
Breakpoints: Susceptible $\leq 32 \mu\text{g/mL}$ Resistant $\geq 128 \mu\text{g/mL}$



NARMS

**Figure 15k: Minimum Inhibitory Concentration of Nitrofurantoin
for *Enterococcus* in Ground Beef (N=448 Isolates)**

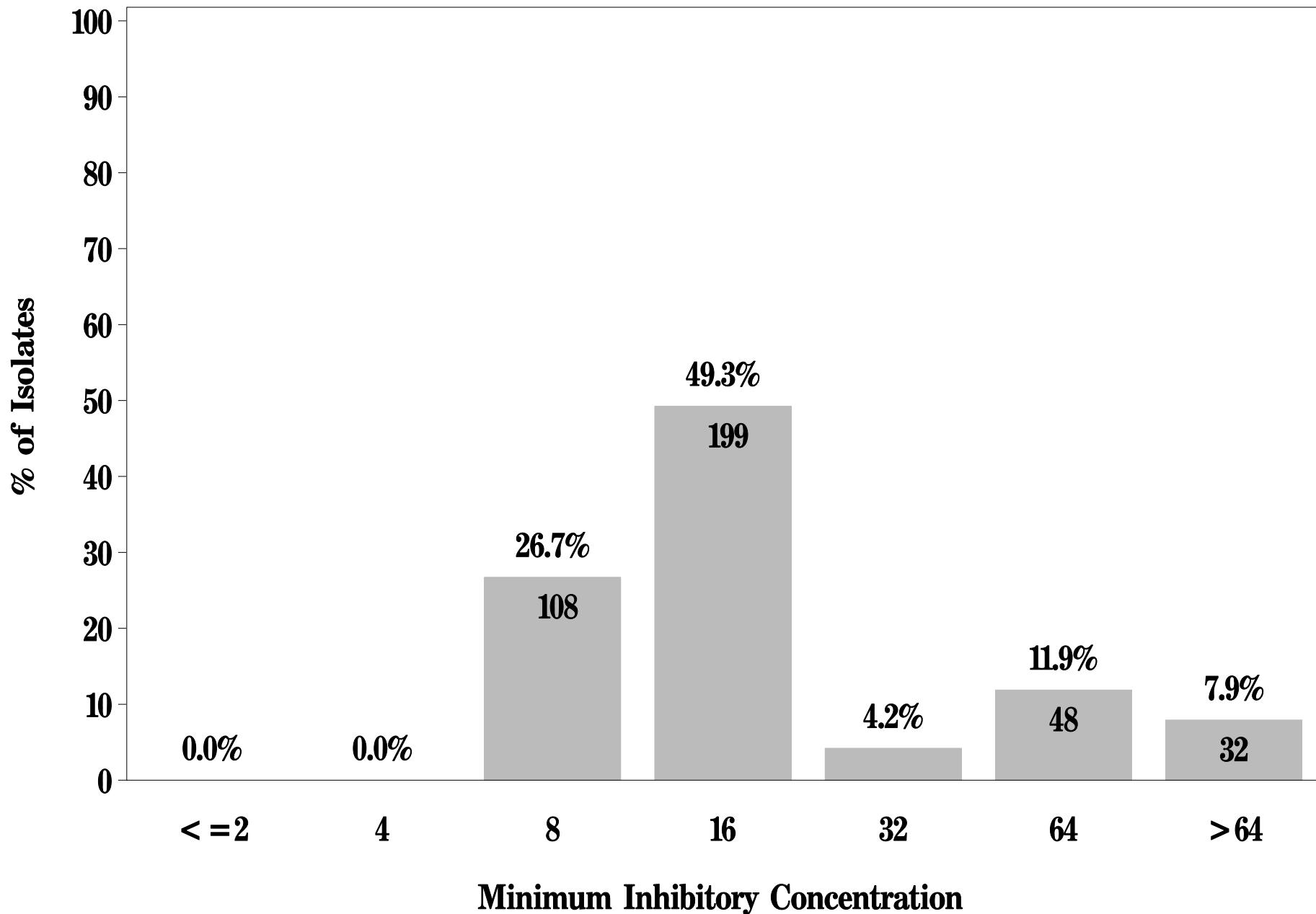
Breakpoints: Susceptible $\leq 32 \mu\text{g/mL}$ Resistant $\geq 128 \mu\text{g/mL}$



NARMS

**Figure 15k: Minimum Inhibitory Concentration of Nitrofurantoin
for *Enterococcus* in Pork Chop (N=404 Isolates)**

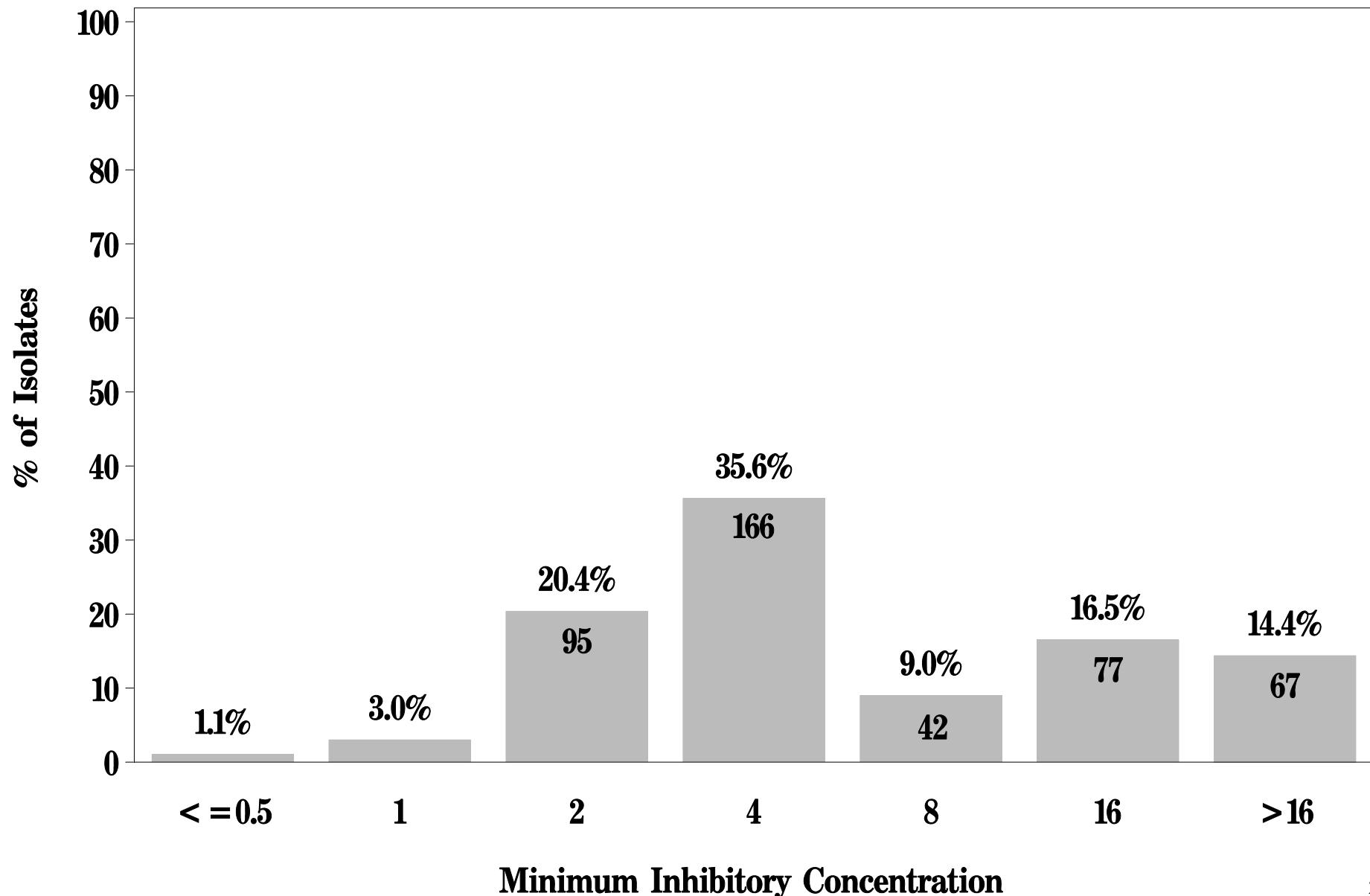
Breakpoints: Susceptible $\leq 32 \mu\text{g/mL}$ Resistant $\geq 128 \mu\text{g/mL}$



NARMS

**Figure 15l: Minimum Inhibitory Concentration of Penicillin
for *Enterococcus* in Chicken Breast (N=466 Isolates)**

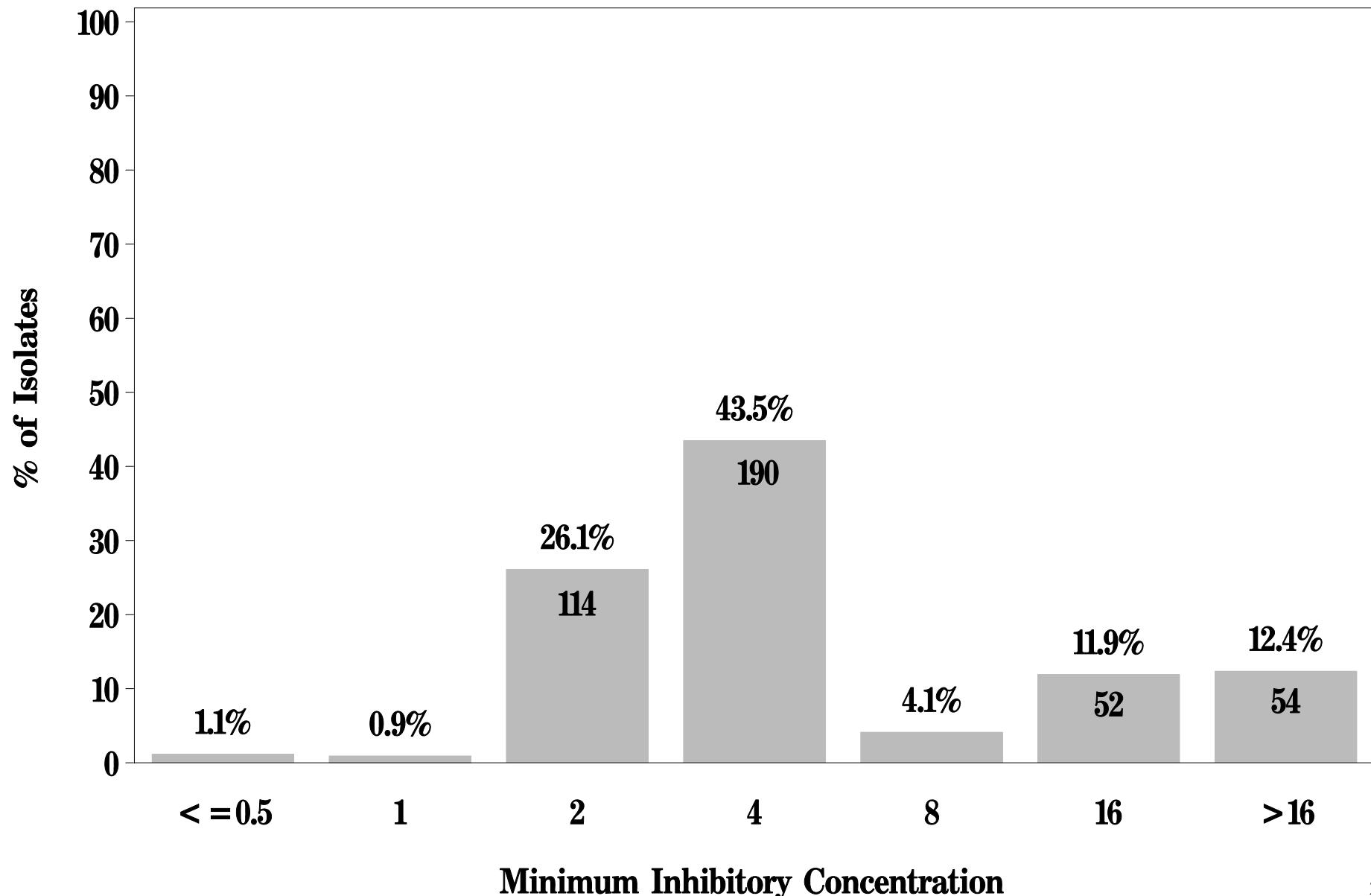
Breakpoints: Susceptible $\leq 8 \mu\text{g/mL}$ Resistant $\geq 16 \mu\text{g/mL}$



NARMS

**Figure 15l: Minimum Inhibitory Concentration of Penicillin
for *Enterococcus* in Ground Turkey (N=437 Isolates)**

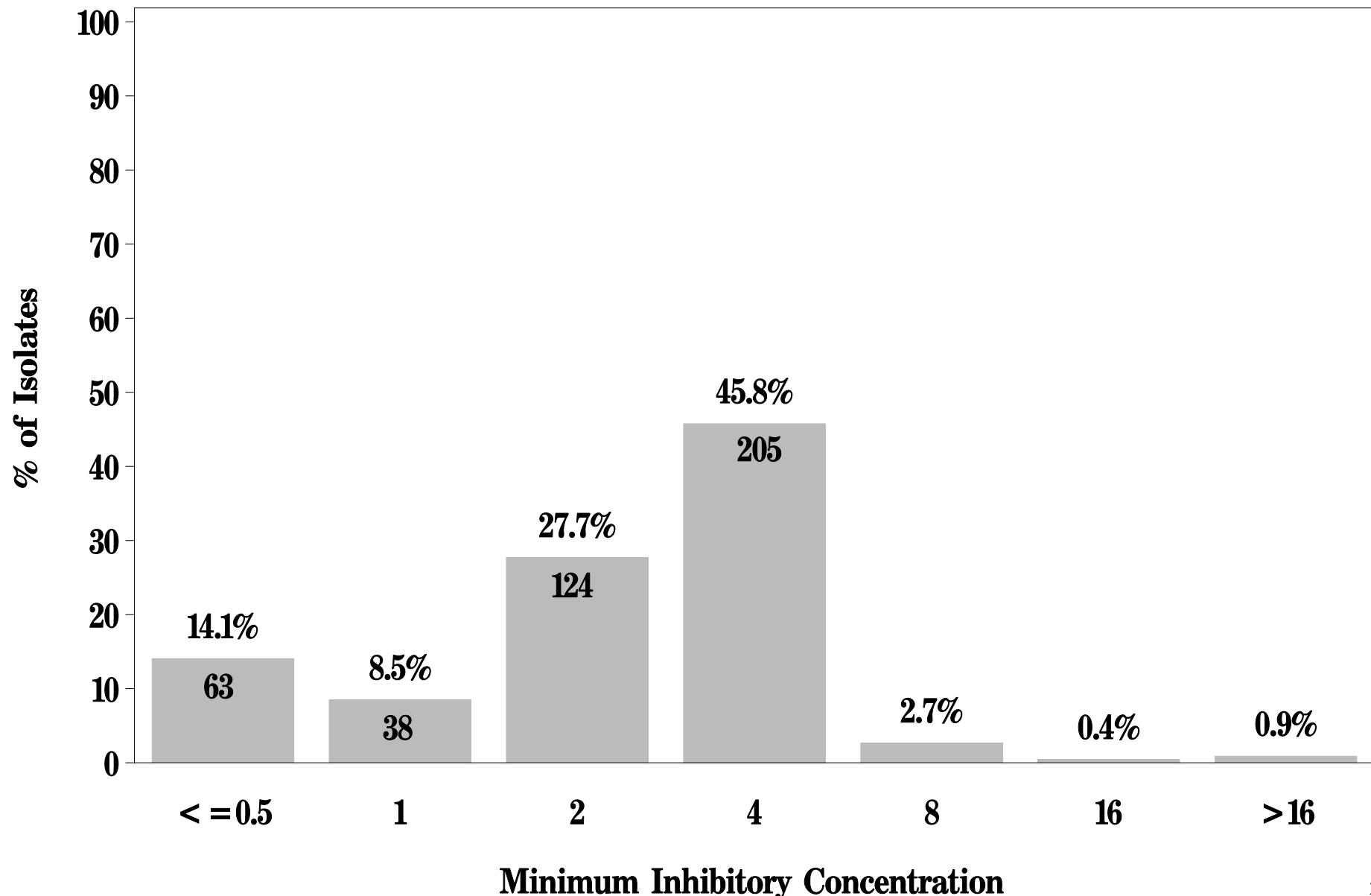
Breakpoints: Susceptible $\leq 8 \mu\text{g/mL}$ Resistant $\geq 16 \mu\text{g/mL}$



NARMS

**Figure 15l: Minimum Inhibitory Concentration of Penicillin
for *Enterococcus* in Ground Beef (N=448 Isolates)**

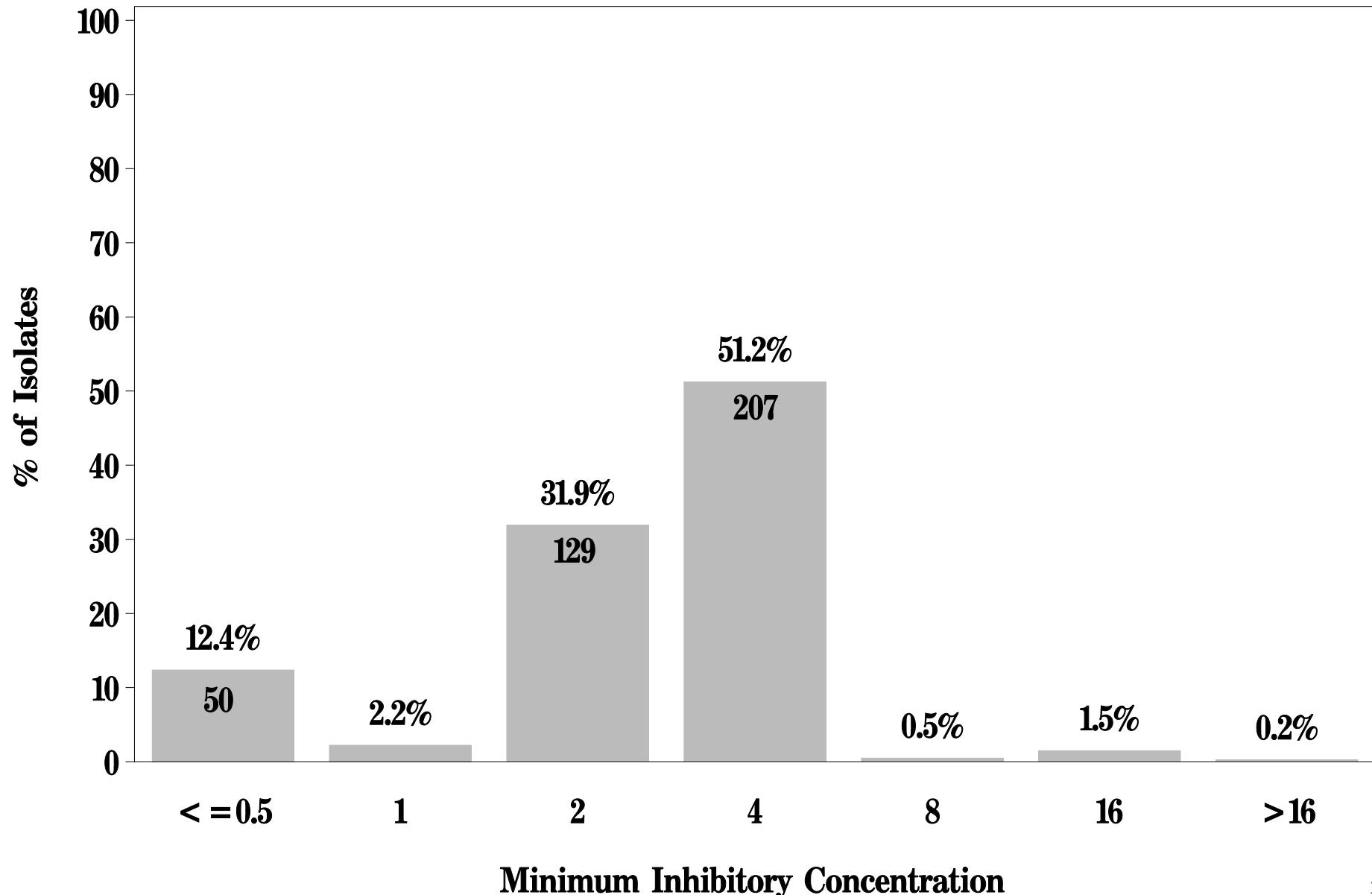
Breakpoints: Susceptible $\leq 8 \mu\text{g/mL}$ Resistant $\geq 16 \mu\text{g/mL}$



NARMS

**Figure 15l: Minimum Inhibitory Concentration of Penicillin
for *Enterococcus* in Pork Chop (N=404 Isolates)**

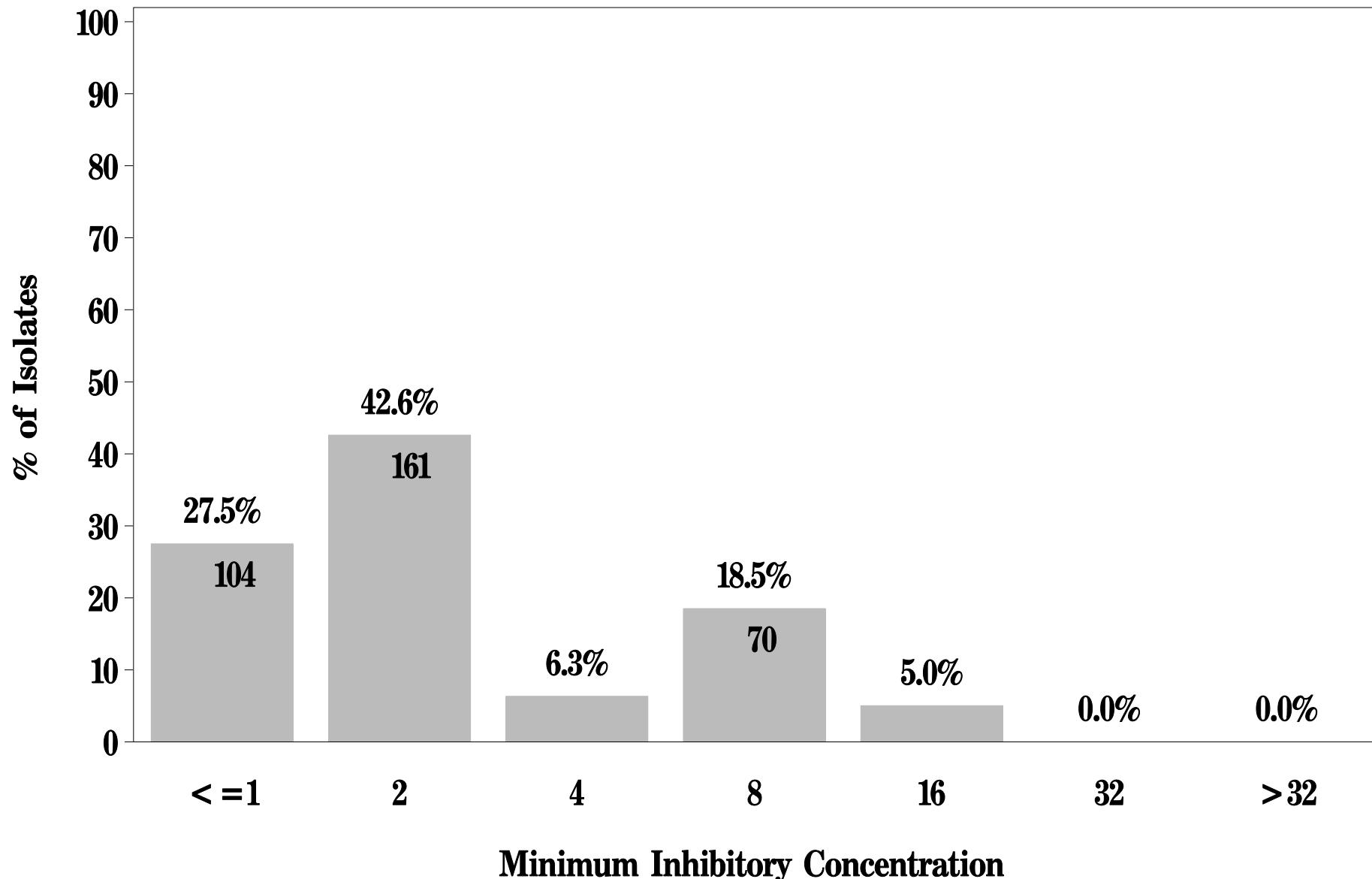
Breakpoints: Susceptible $\leq 8 \mu\text{g/mL}$ Resistant $\geq 16 \mu\text{g/mL}$



NARMS

**Figure 15m: Minimum Inhibitory Concentration of Quinupristin – dalfopristin
for *Enterococcus* in Chicken Breast (N=378 Isolates)**

Breakpoints: Susceptible < =1 $\mu\text{g/mL}$ Resistant > =4 $\mu\text{g/mL}$

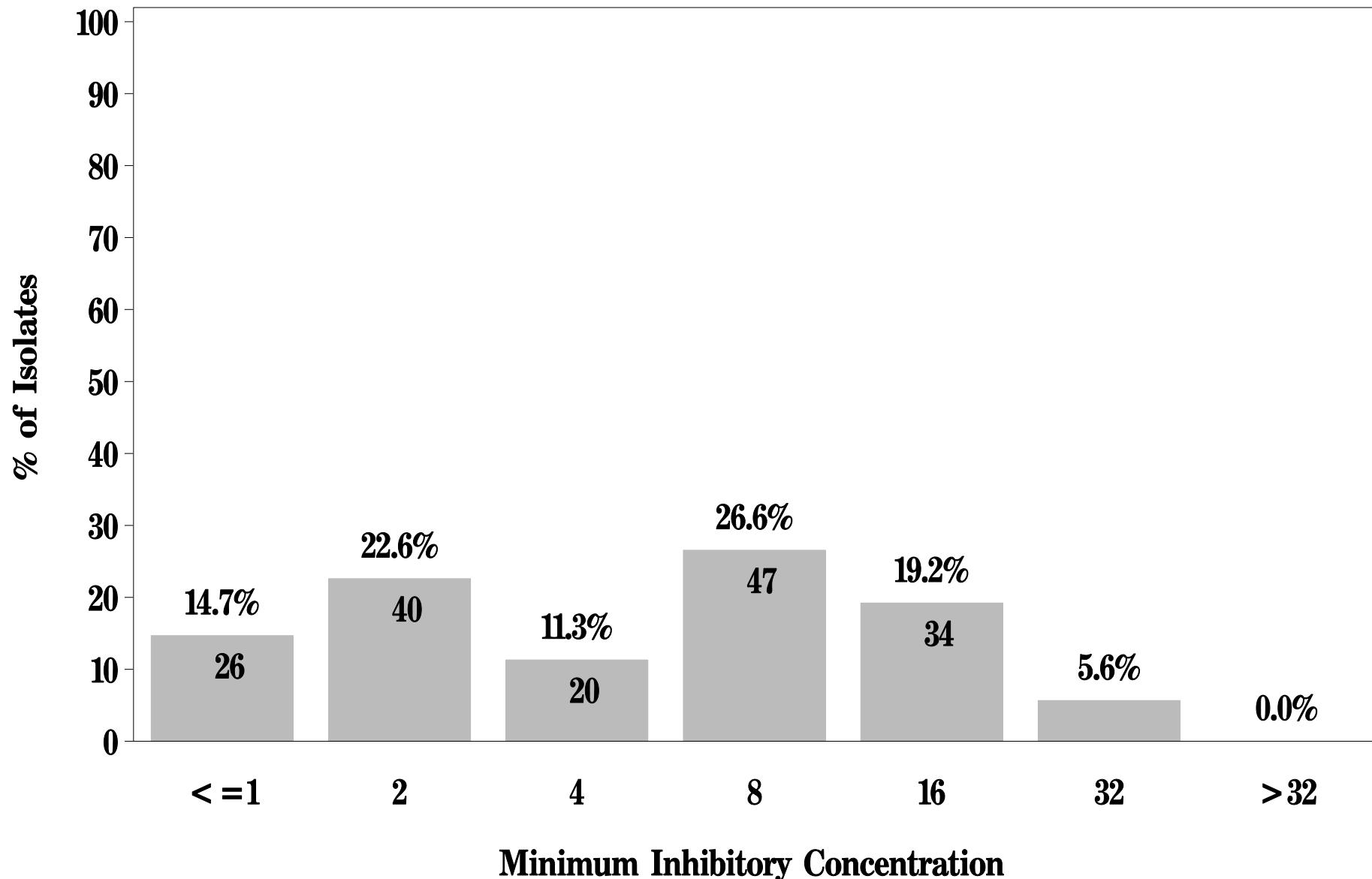


*Presented for all species except *E. faecalis* (N=466 – 88=378)

NARMS

**Figure 15m: Minimum Inhibitory Concentration of Quinupristin – dalfopristin
for *Enterococcus* in Ground Turkey (N=177 Isolates)**

Breakpoints: Susceptible $\leq 1 \mu\text{g/mL}$ Resistant $\geq 4 \mu\text{g/mL}$

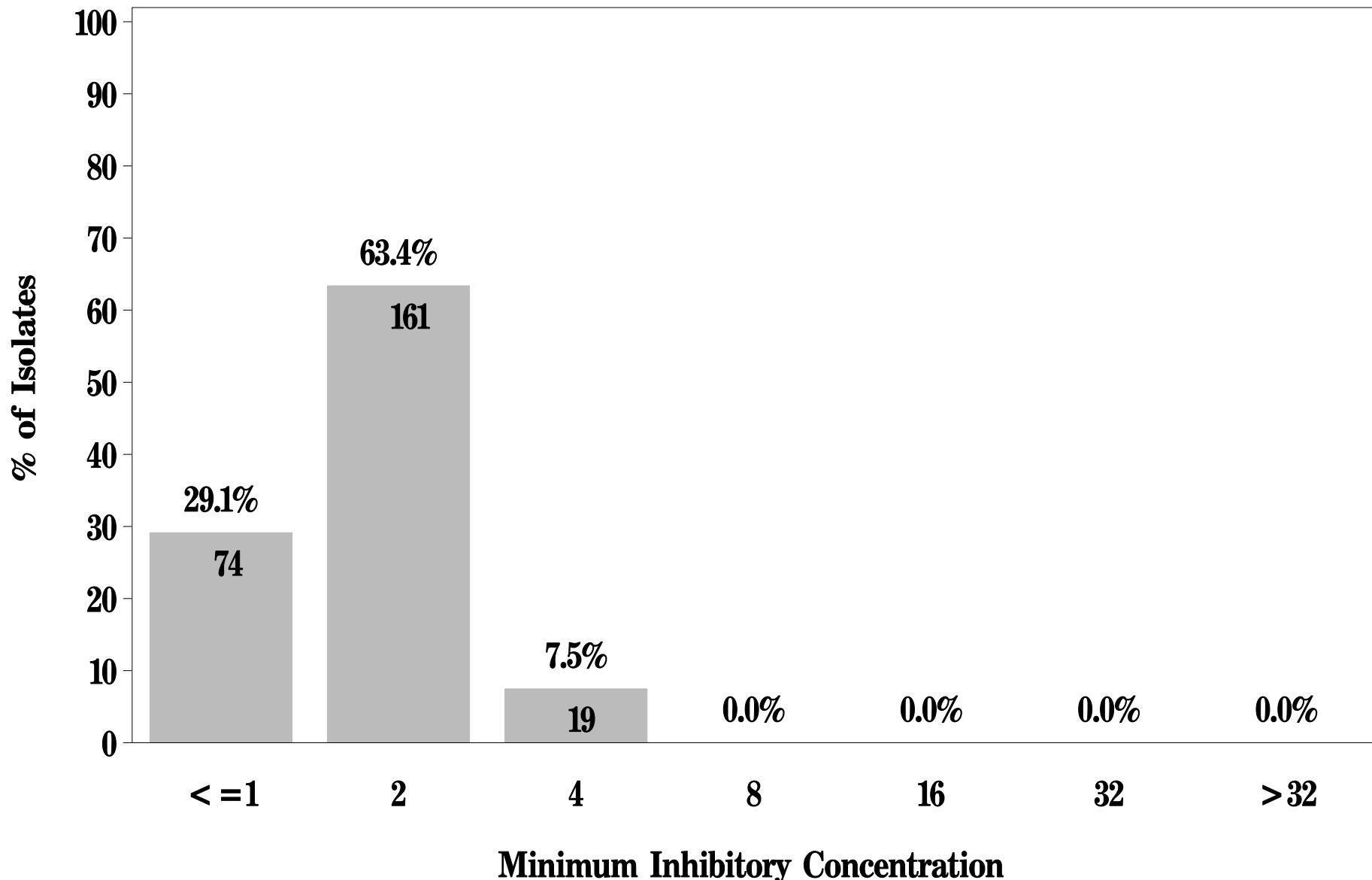


*Presented for all species except *E. faecalis* (N=437 - 260 = 177)

NARMS

Figure 15m: Minimum Inhibitory Concentration of Quinupristin – dalfopristin for *Enterococcus* in Ground Beef (N=254 Isolates)

Breakpoints: Susceptible $\leq 1 \mu\text{g/mL}$ Resistant $\geq 4 \mu\text{g/mL}$

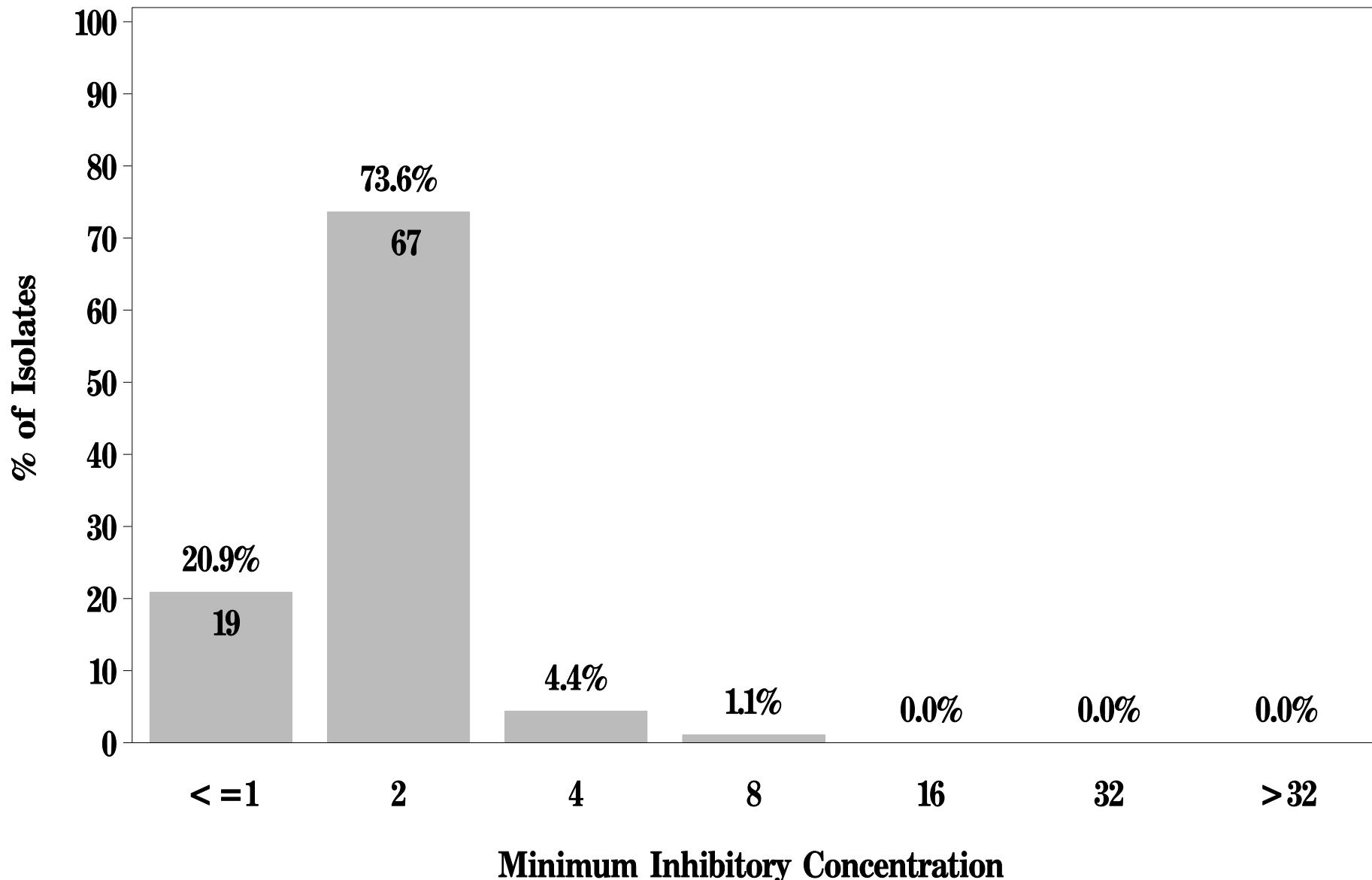


*Presented for all species except *E. faecalis* (N=488 – 194 = 254)

NARMS

**Figure 15m: Minimum Inhibitory Concentration of Quinupristin – dalfopristin
for *Enterococcus* in Pork Chop (N=91 Isolates)**

Breakpoints: Susceptible $\leq 1 \mu\text{g/mL}$ Resistant $\geq 4 \mu\text{g/mL}$

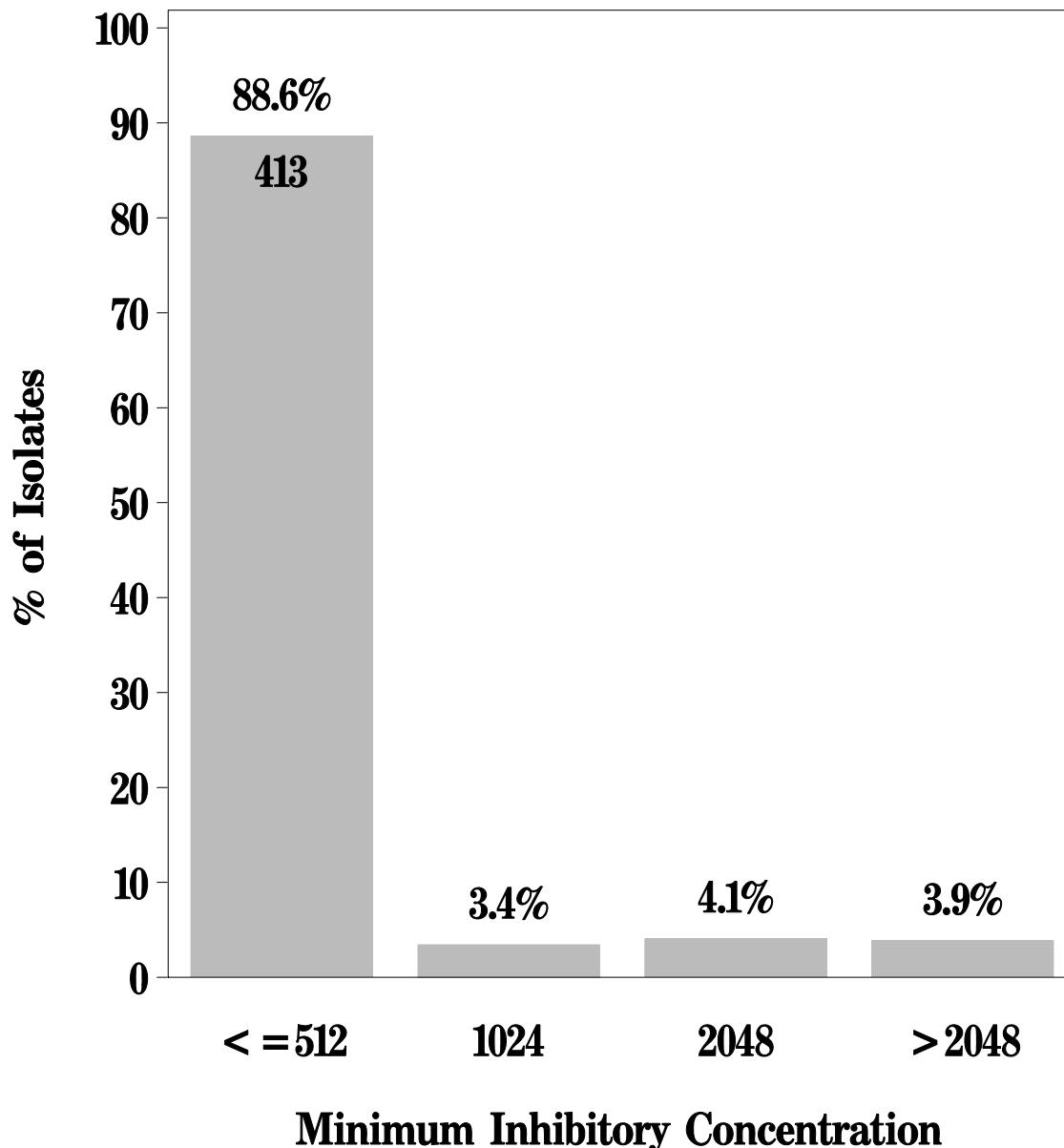


*Presented for all species except *E. faecalis* (N=404 – 313 = 91)

NARMS

**Figure 15n: Minimum Inhibitory Concentration of Streptomycin
for *Enterococcus* in Chicken Breast (N=466 Isolates)**

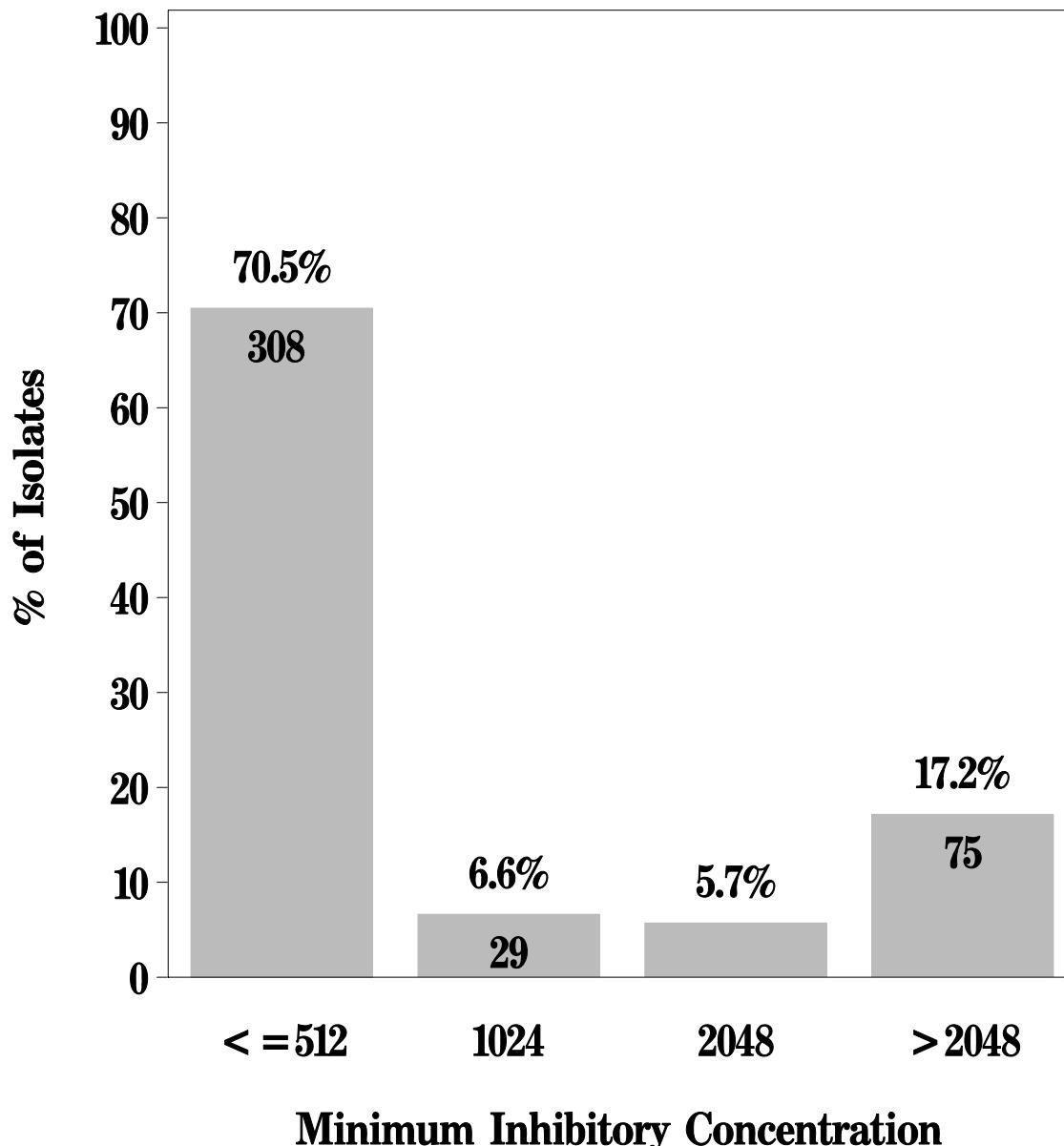
Breakpoints: Susceptible $\leq 1000 \mu\text{g/mL}$ Resistant $\geq 1000 \mu\text{g/mL}$



NARMS

**Figure 15n: Minimum Inhibitory Concentration of Streptomycin
for *Enterococcus* in Ground Turkey (N=437 Isolates)**

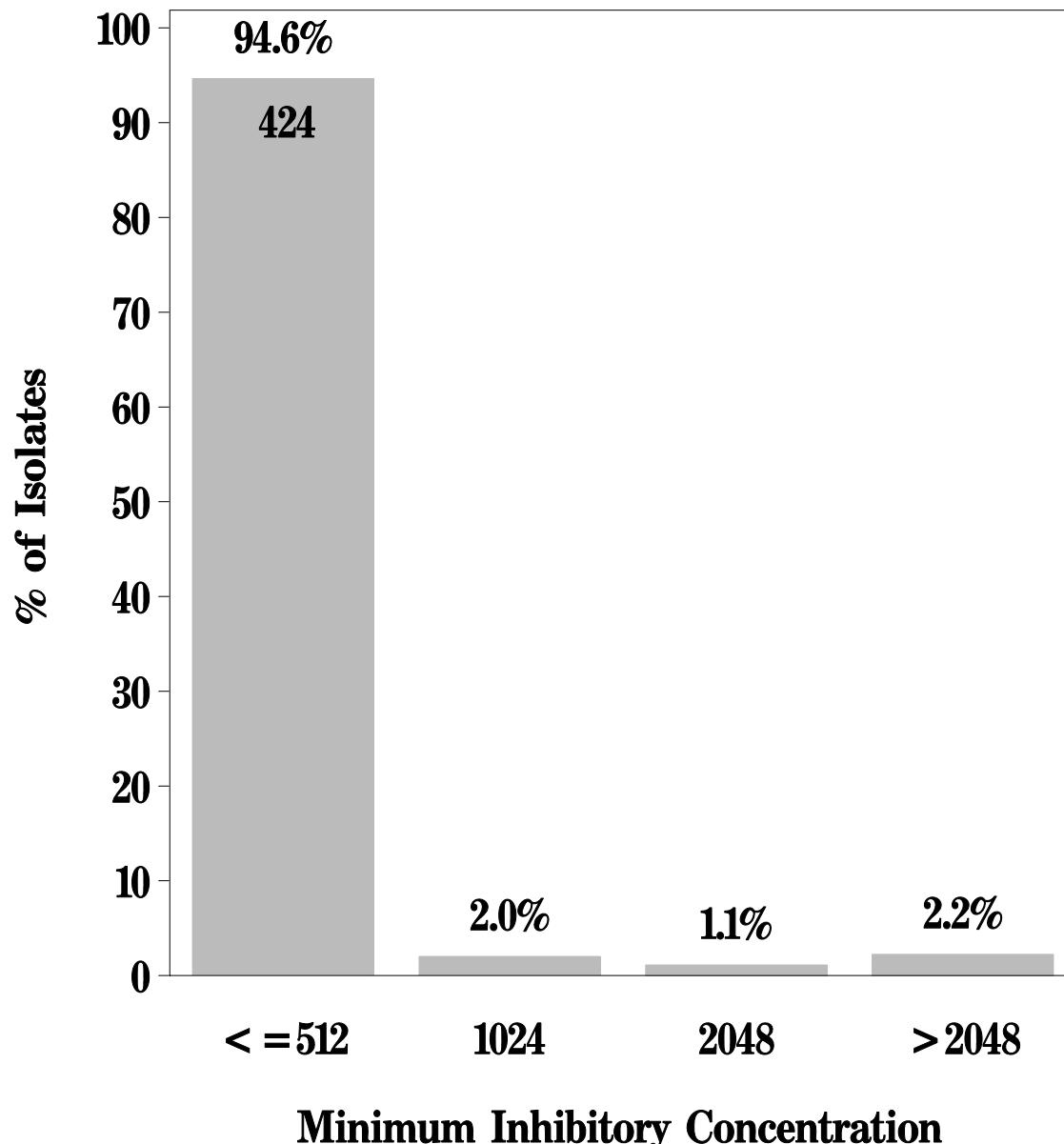
Breakpoints: Susceptible $\leq 1000 \mu\text{g/mL}$ Resistant $\geq 1000 \mu\text{g/mL}$



NARMS

**Figure 15n: Minimum Inhibitory Concentration of Streptomycin
for *Enterococcus* in Ground Beef (N=448 Isolates)**

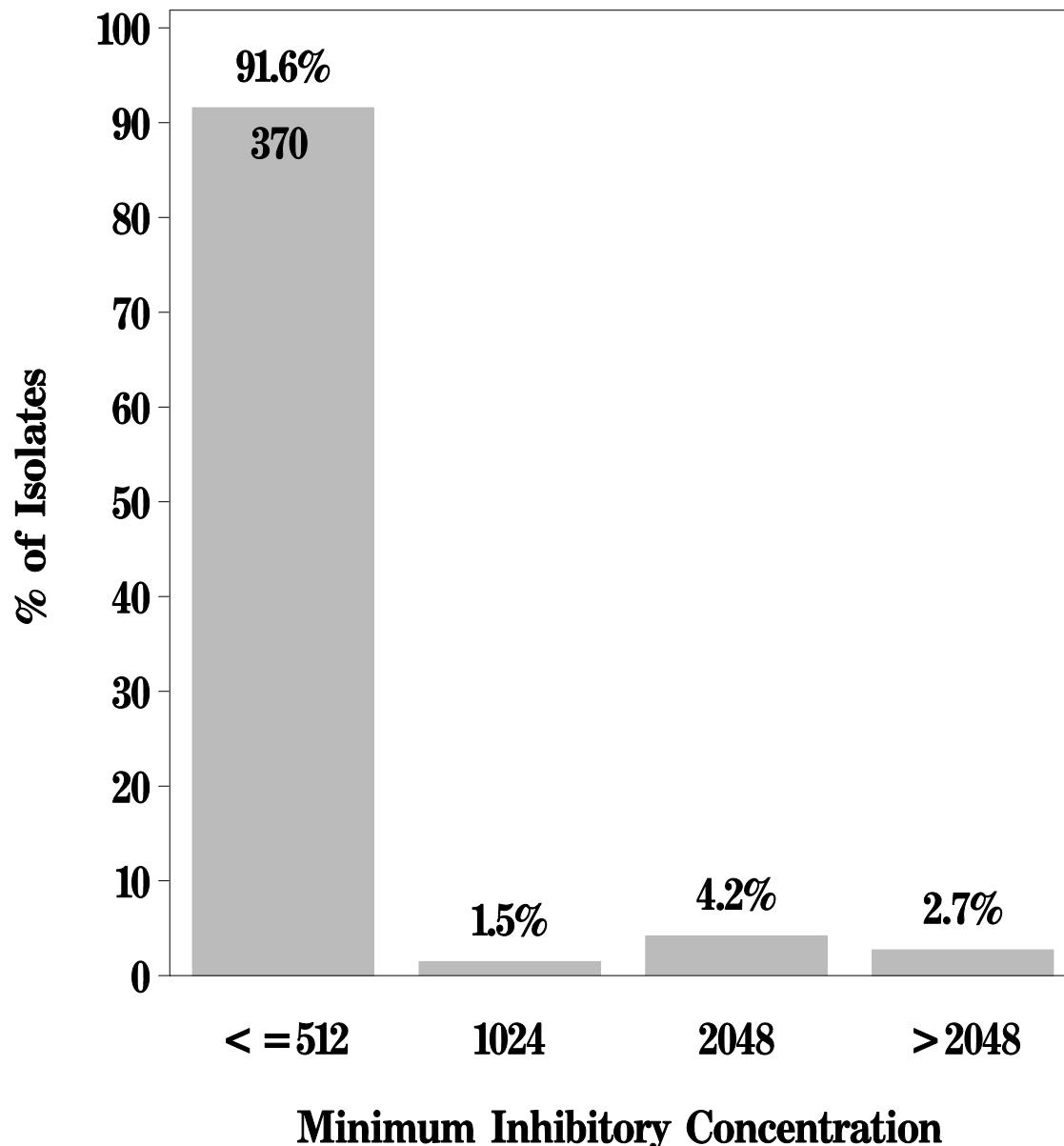
Breakpoints: Susceptible $\leq 1000 \mu\text{g/mL}$ Resistant $\geq 1000 \mu\text{g/mL}$



NARMS

**Figure 15n: Minimum Inhibitory Concentration of Streptomycin
for *Enterococcus* in Pork Chop (N=404 Isolates)**

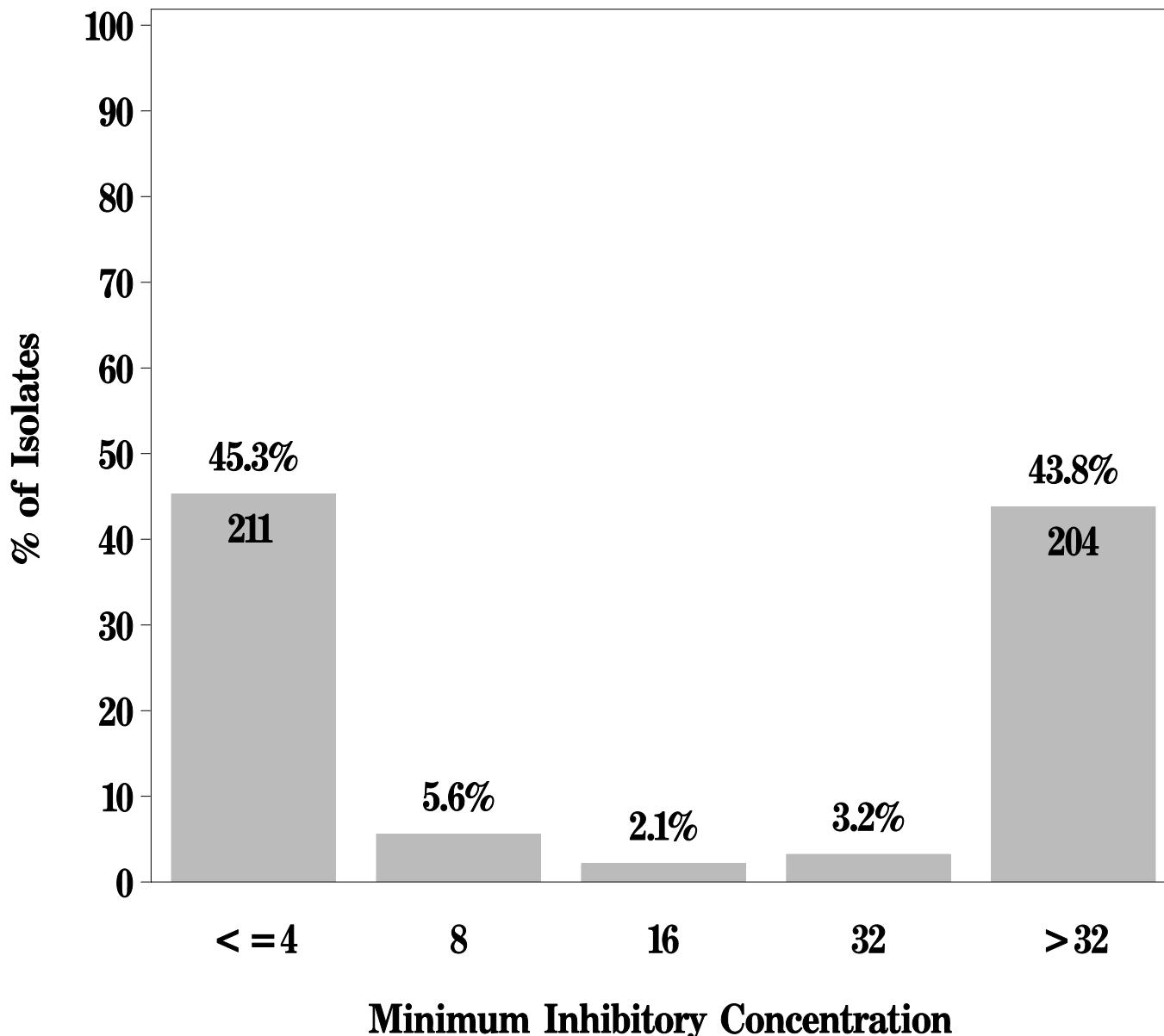
Breakpoints: Susceptible $\leq 1000 \mu\text{g/mL}$ Resistant $\geq 1000 \mu\text{g/mL}$



NARMS

**Figure 15o: Minimum Inhibitory Concentration of Tetracycline
for *Enterococcus* in Chicken Breast (N=466 Isolates)**

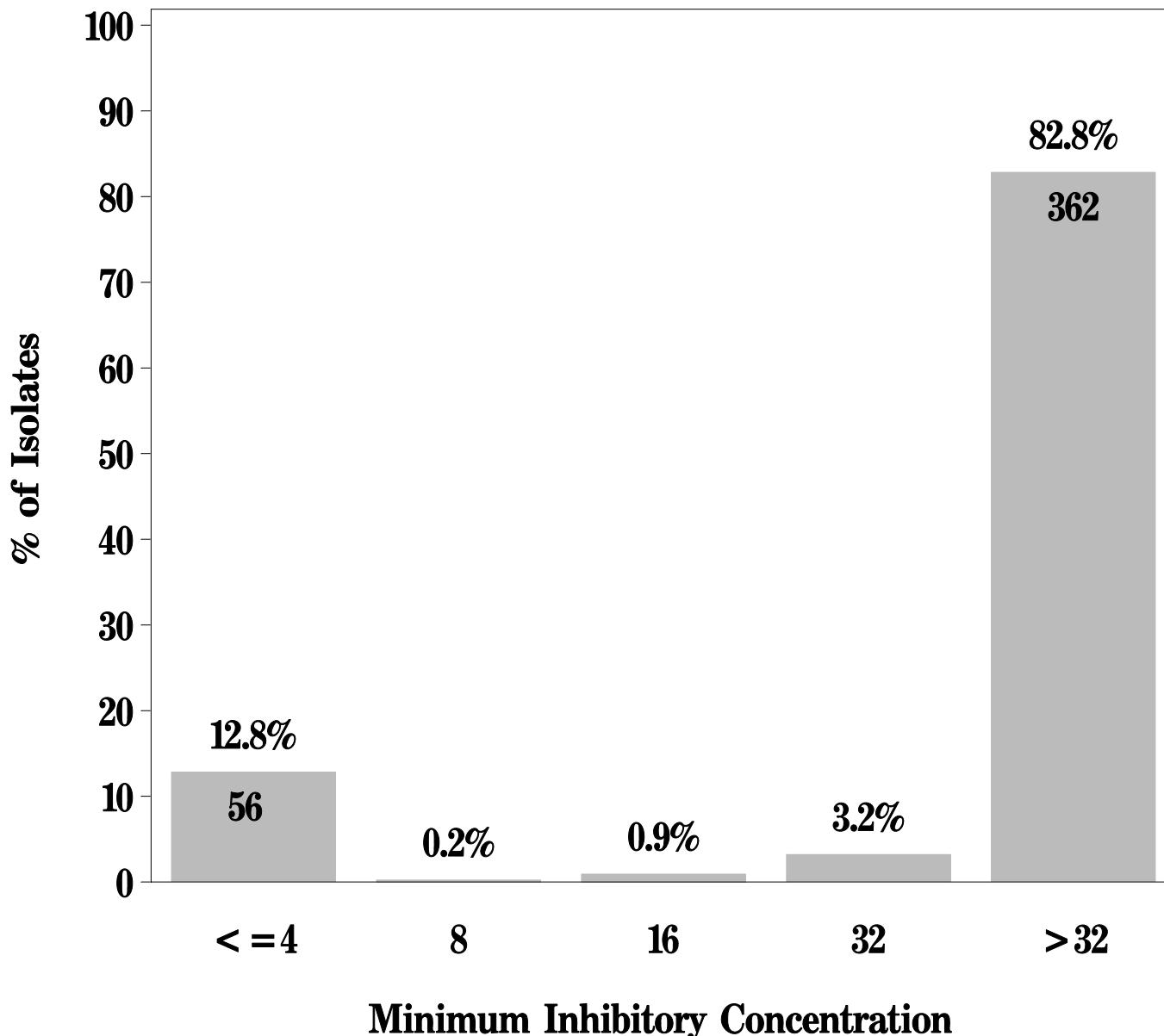
Breakpoints: Susceptible $\leq 4 \text{ } \mu\text{g/mL}$ Resistant $\geq 16 \text{ } \mu\text{g/mL}$



NARMS

**Figure 15o: Minimum Inhibitory Concentration of Tetracycline
for *Enterococcus* in Ground Turkey (N=437 Isolates)**

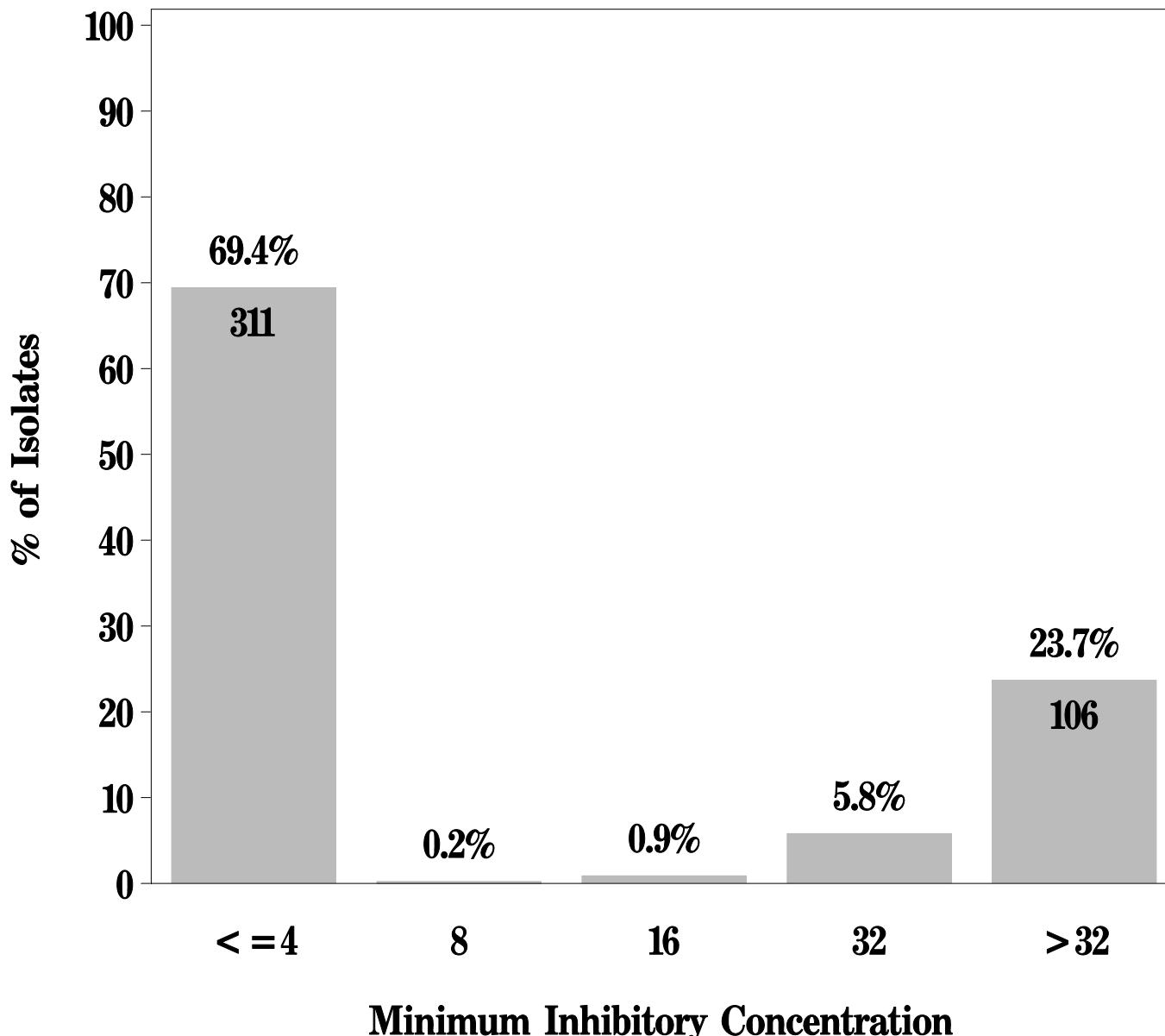
Breakpoints: Susceptible $\leq 4 \text{ } \mu\text{g/mL}$ Resistant $\geq 16 \text{ } \mu\text{g/mL}$



NARMS

**Figure 15o: Minimum Inhibitory Concentration of Tetracycline
for *Enterococcus* in Ground Beef (N=448 Isolates)**

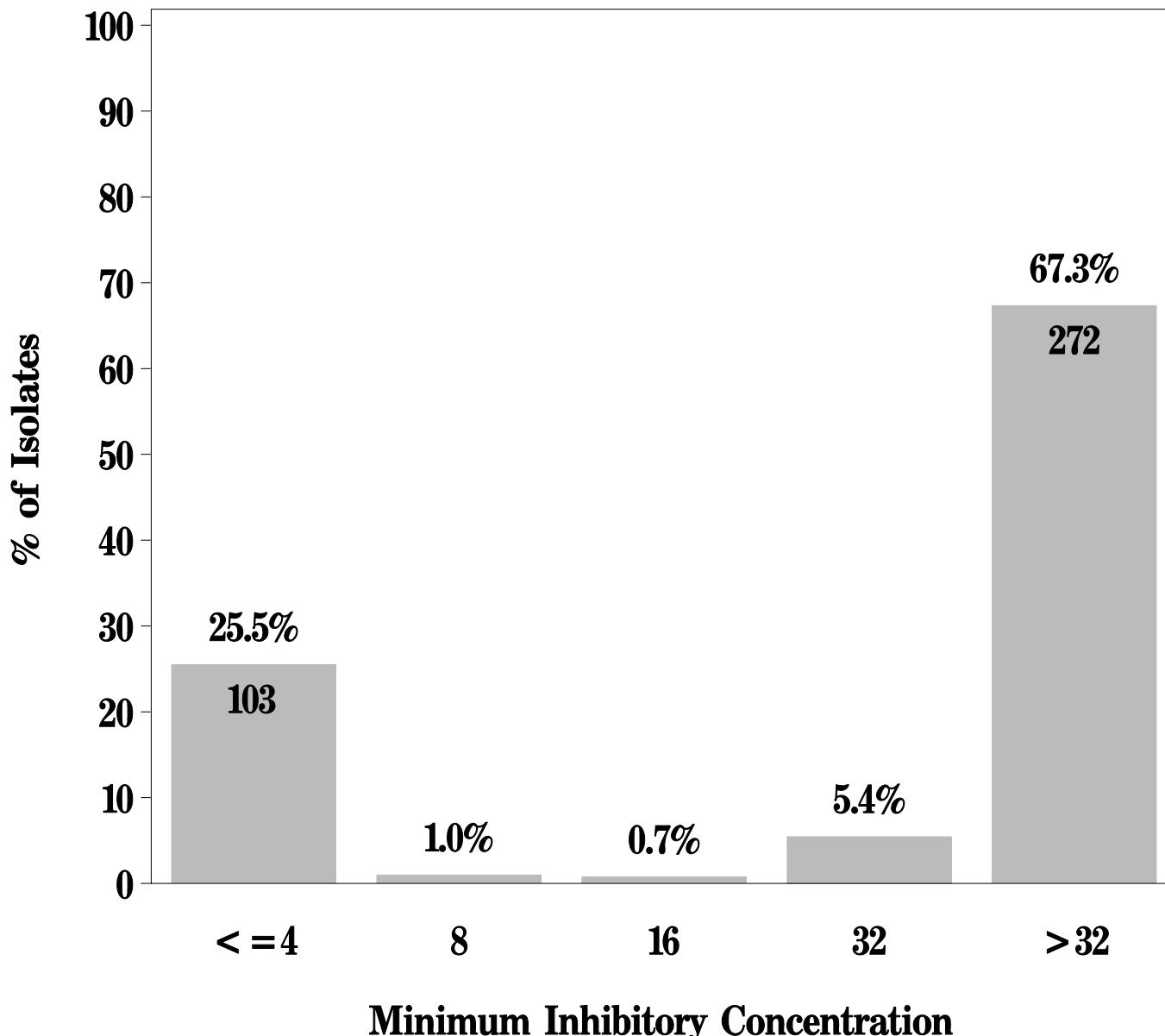
Breakpoints: Susceptible $\leq 4 \text{ } \mu\text{g/mL}$ Resistant $\geq 16 \text{ } \mu\text{g/mL}$



NARMS

**Figure 15o: Minimum Inhibitory Concentration of Tetracycline
for *Enterococcus* in Pork Chop (N=404 Isolates)**

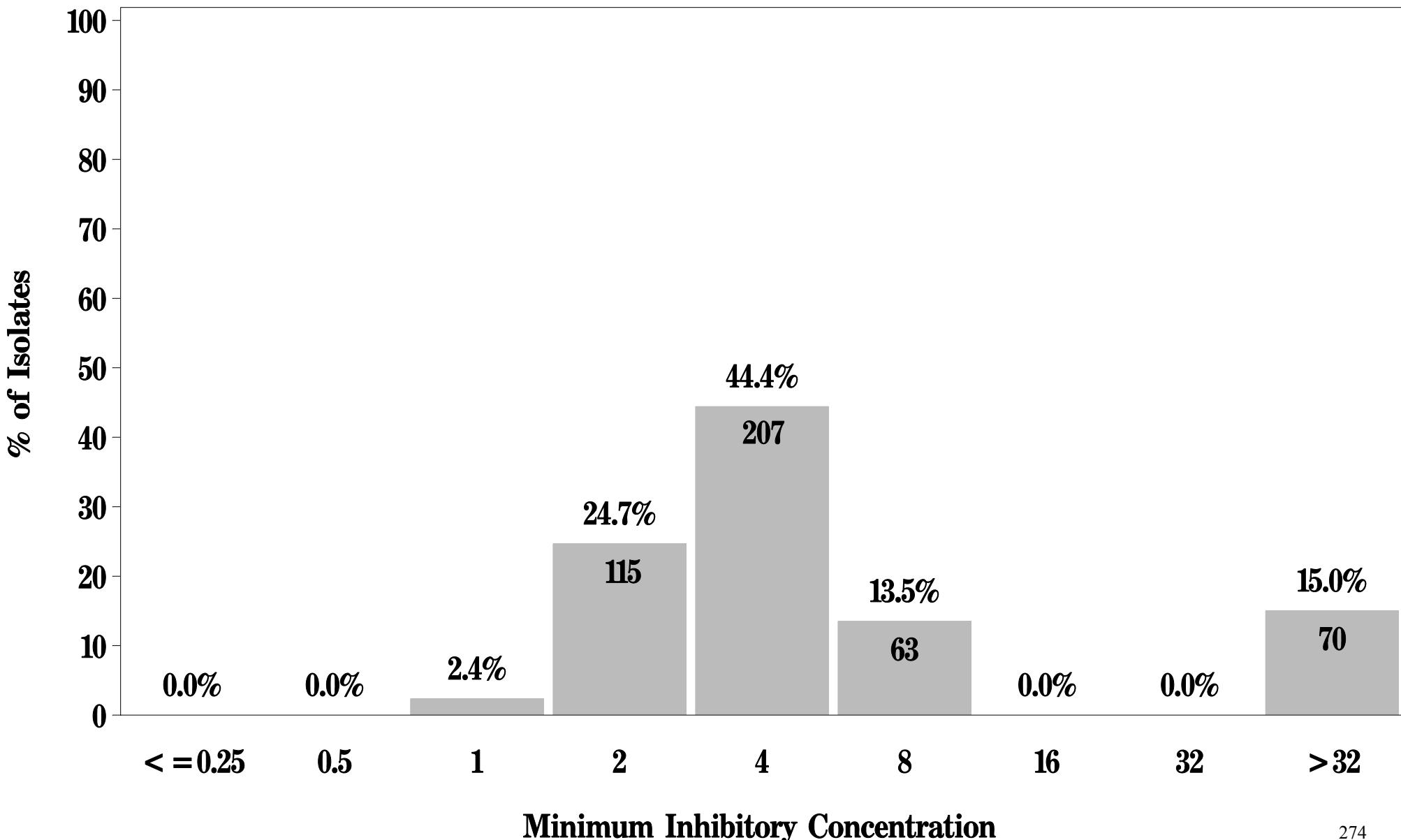
Breakpoints: Susceptible $\leq 4 \text{ } \mu\text{g/mL}$ Resistant $\geq 16 \text{ } \mu\text{g/mL}$



NARMS

**Figure 15p: Minimum Inhibitory Concentration of Tylosin
for *Enterococcus* in Chicken Breast (N=466 Isolates)**

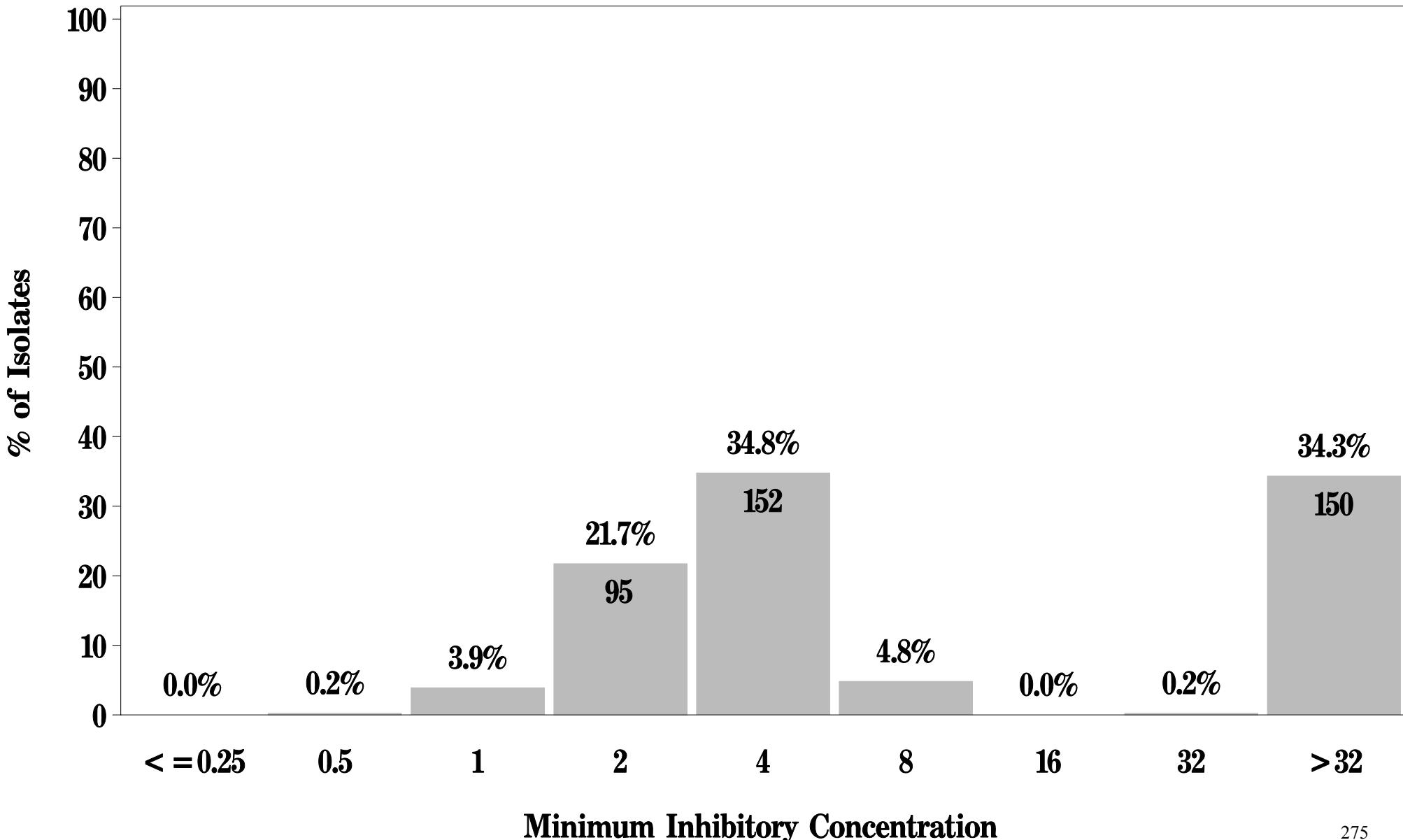
Breakpoints: Susceptible $\leq 8 \mu\text{g/mL}$ Resistant $\geq 32 \mu\text{g/mL}$



NARMS

**Figure 15p: Minimum Inhibitory Concentration of Tylosin
for *Enterococcus* in Ground Turkey (N=437 Isolates)**

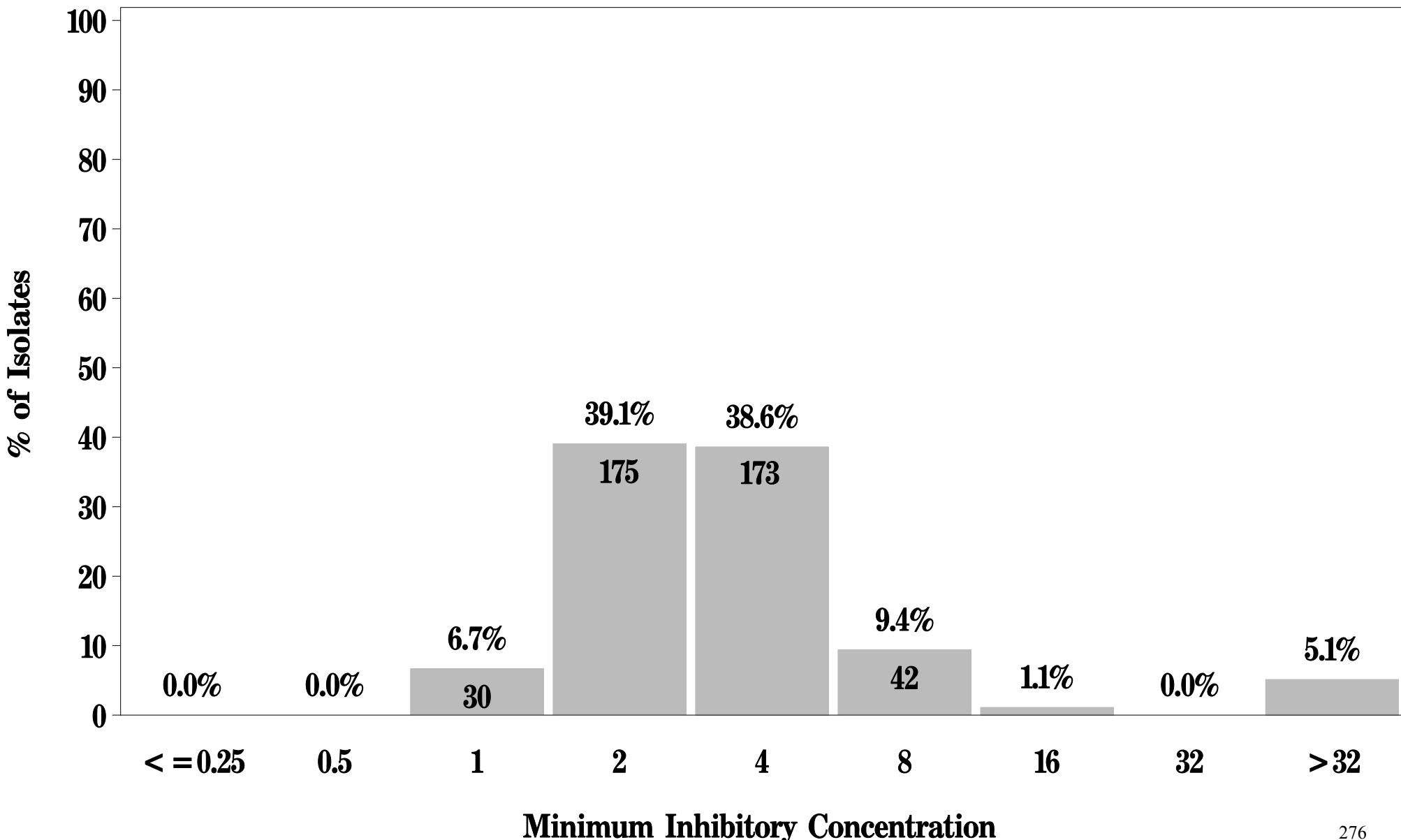
Breakpoints: Susceptible $\leq 8 \mu\text{g/mL}$ Resistant $\geq 32 \mu\text{g/mL}$



NARMS

**Figure 15p: Minimum Inhibitory Concentration of Tylosin
for *Enterococcus* in Ground Beef (N=448 Isolates)**

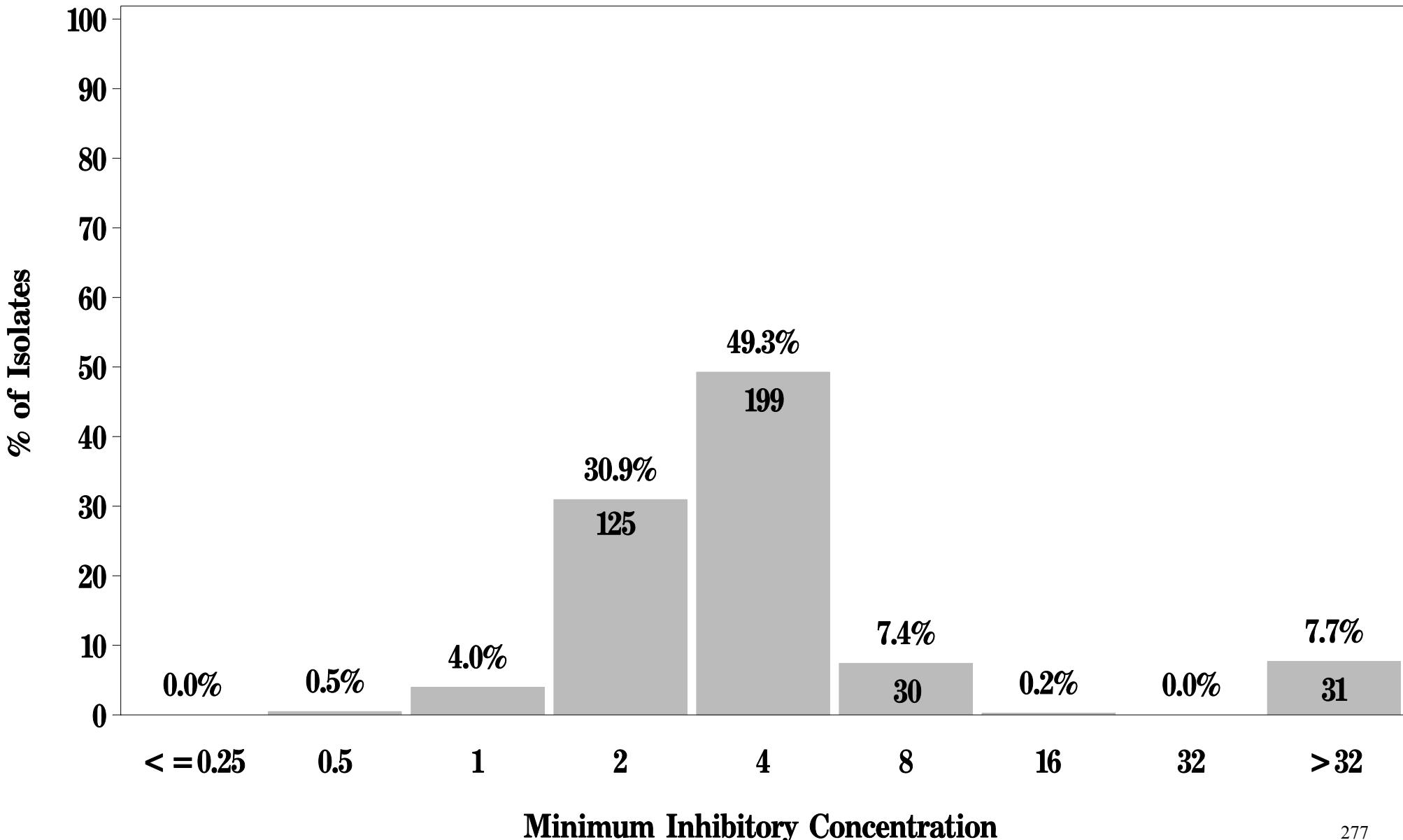
Breakpoints: Susceptible $\leq 8 \mu\text{g/mL}$ Resistant $\geq 32 \mu\text{g/mL}$



NARMS

**Figure 15p: Minimum Inhibitory Concentration of Tylosin
for *Enterococcus* in Pork Chop (N=404 Isolates)**

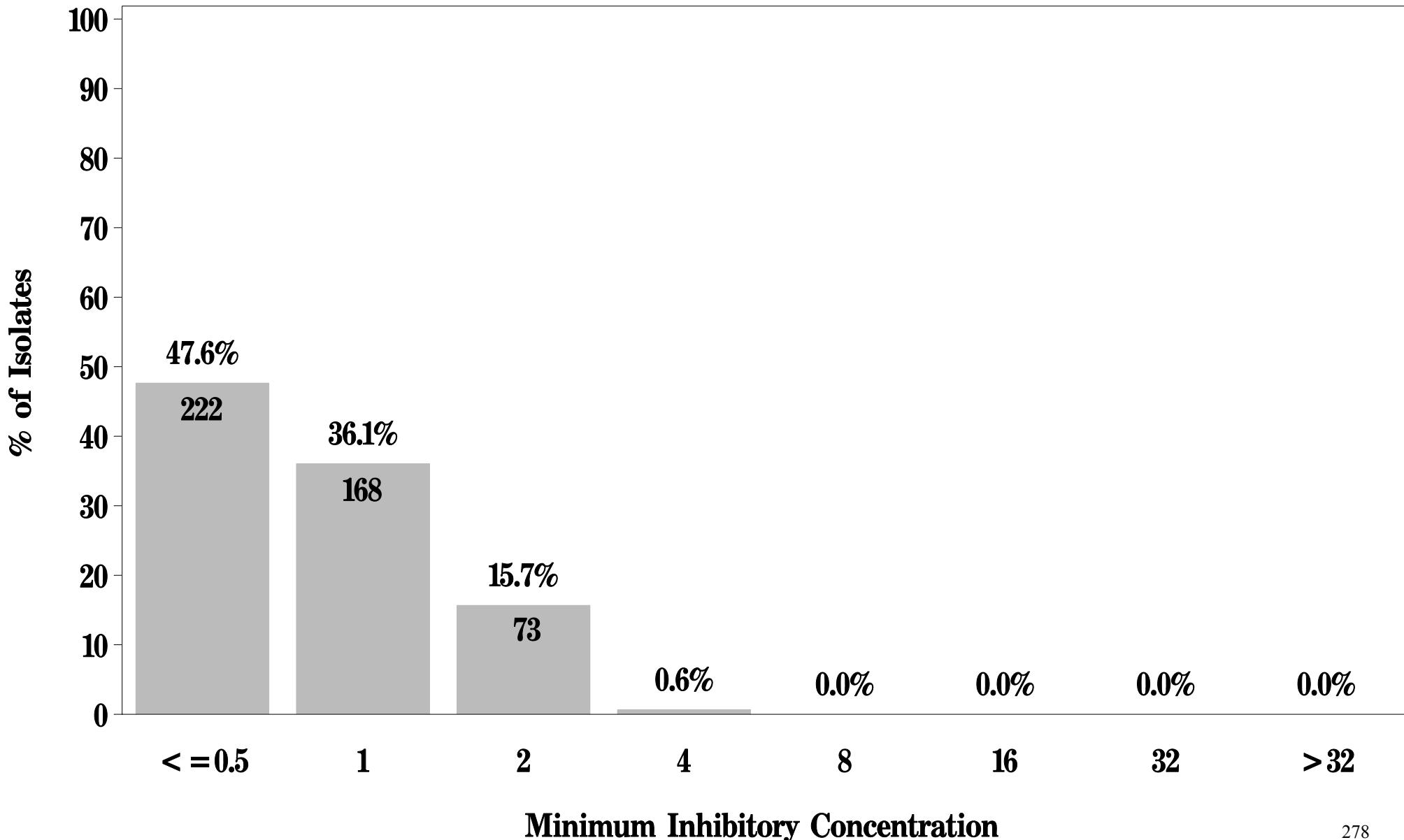
Breakpoints: Susceptible $\leq 8 \mu\text{g/mL}$ Resistant $\geq 32 \mu\text{g/mL}$



NARMS

**Figure 15q: Minimum Inhibitory Concentration of Vancomycin
for *Enterococcus* in Chicken Breast (N=466 Isolates)**

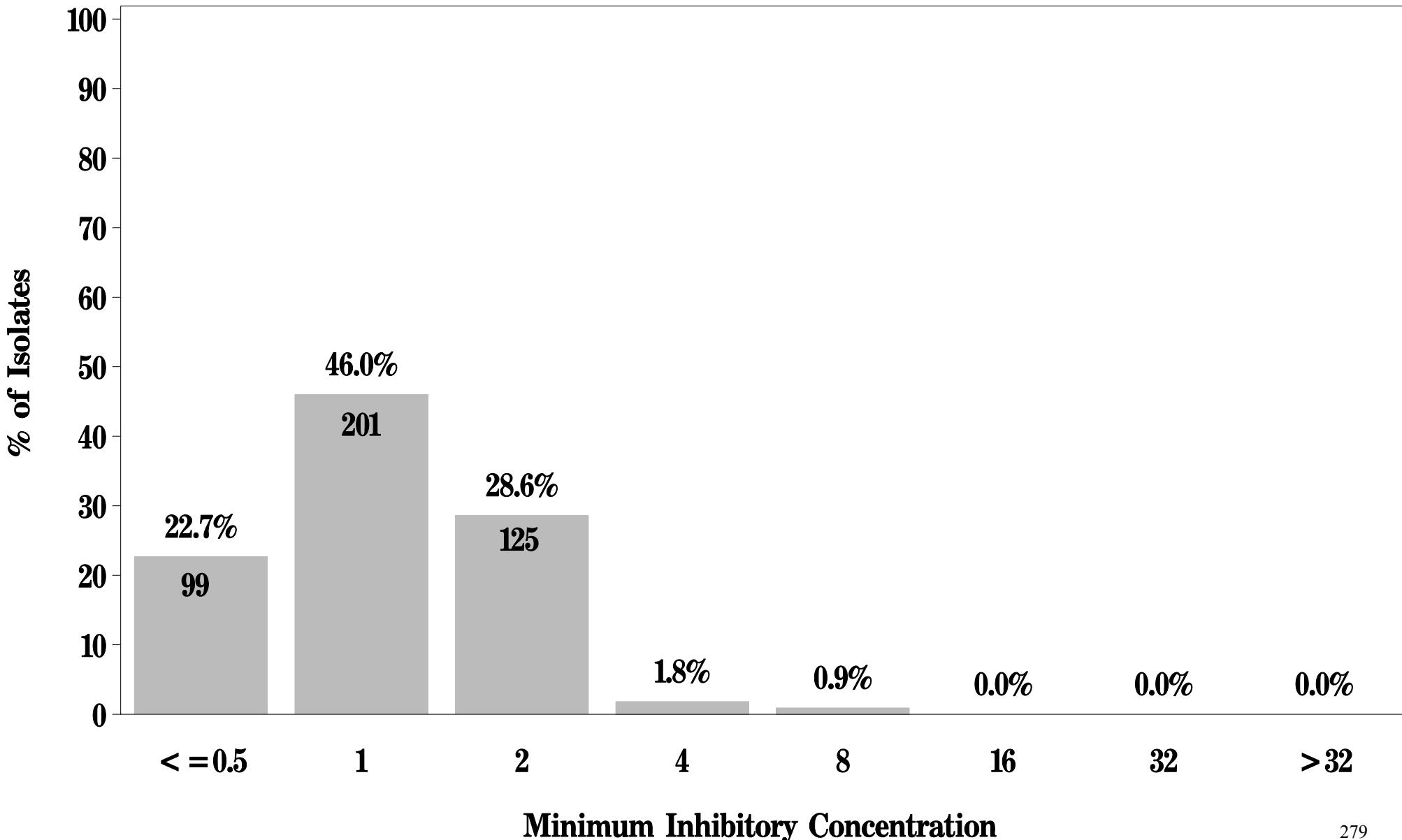
Breakpoints: Susceptible $\leq 4 \text{ } \mu\text{g/mL}$ Resistant $\geq 32 \text{ } \mu\text{g/mL}$



NARMS

**Figure 15q: Minimum Inhibitory Concentration of Vancomycin
for *Enterococcus* in Ground Turkey (N=437 Isolates)**

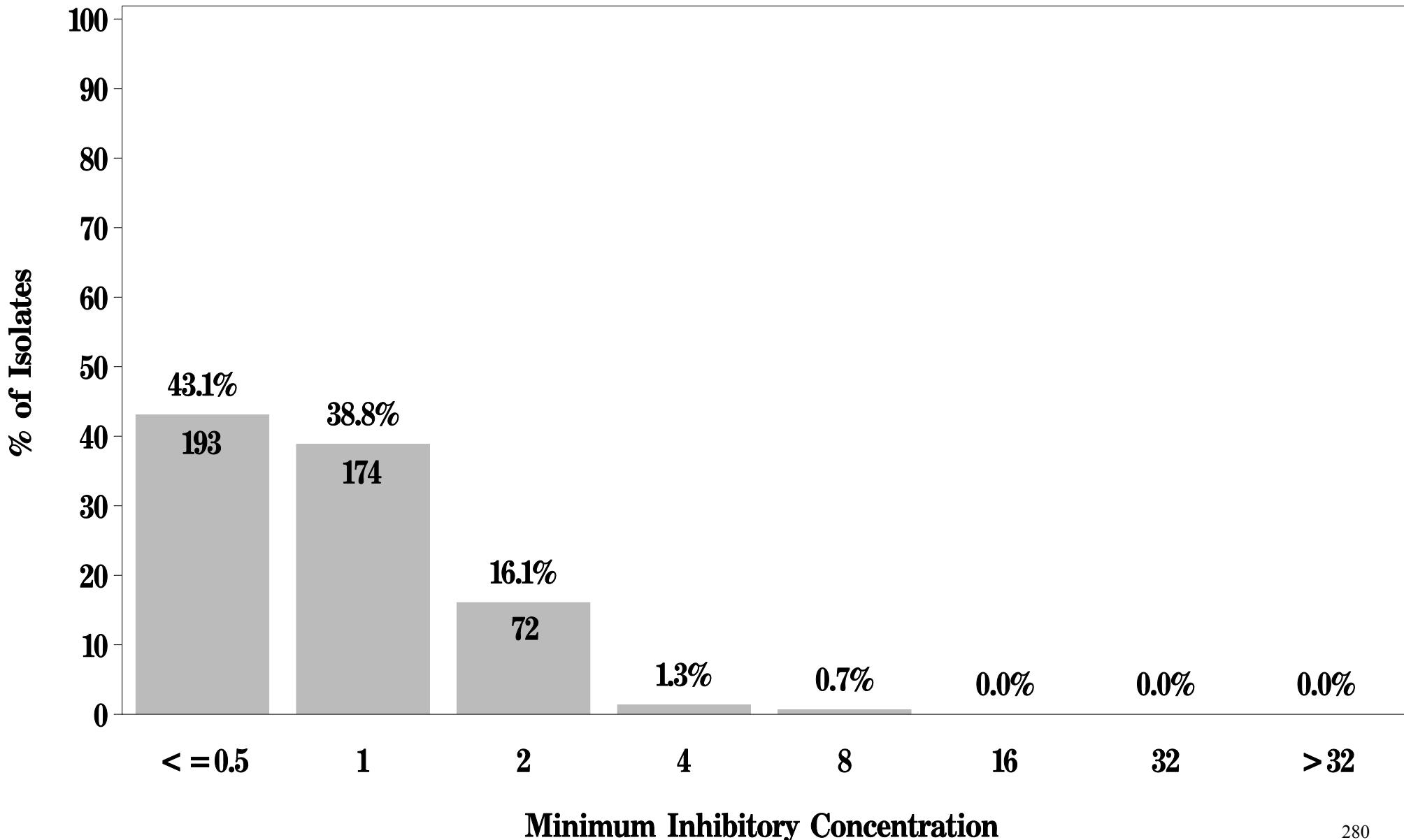
Breakpoints: Susceptible $\leq 4 \mu\text{g/mL}$ Resistant $\geq 32 \mu\text{g/mL}$



NARMS

**Figure 15q: Minimum Inhibitory Concentration of Vancomycin
for *Enterococcus* in Ground Beef (N=448 Isolates)**

Breakpoints: Susceptible $\leq 4 \text{ } \mu\text{g/mL}$ Resistant $\geq 32 \text{ } \mu\text{g/mL}$



NARMS

**Figure 15q: Minimum Inhibitory Concentration of Vancomycin
for *Enterococcus* in Pork Chop (N=404 Isolates)**

Breakpoints: Susceptible $\leq 4 \mu\text{g/mL}$ Resistant $\geq 32 \mu\text{g/mL}$

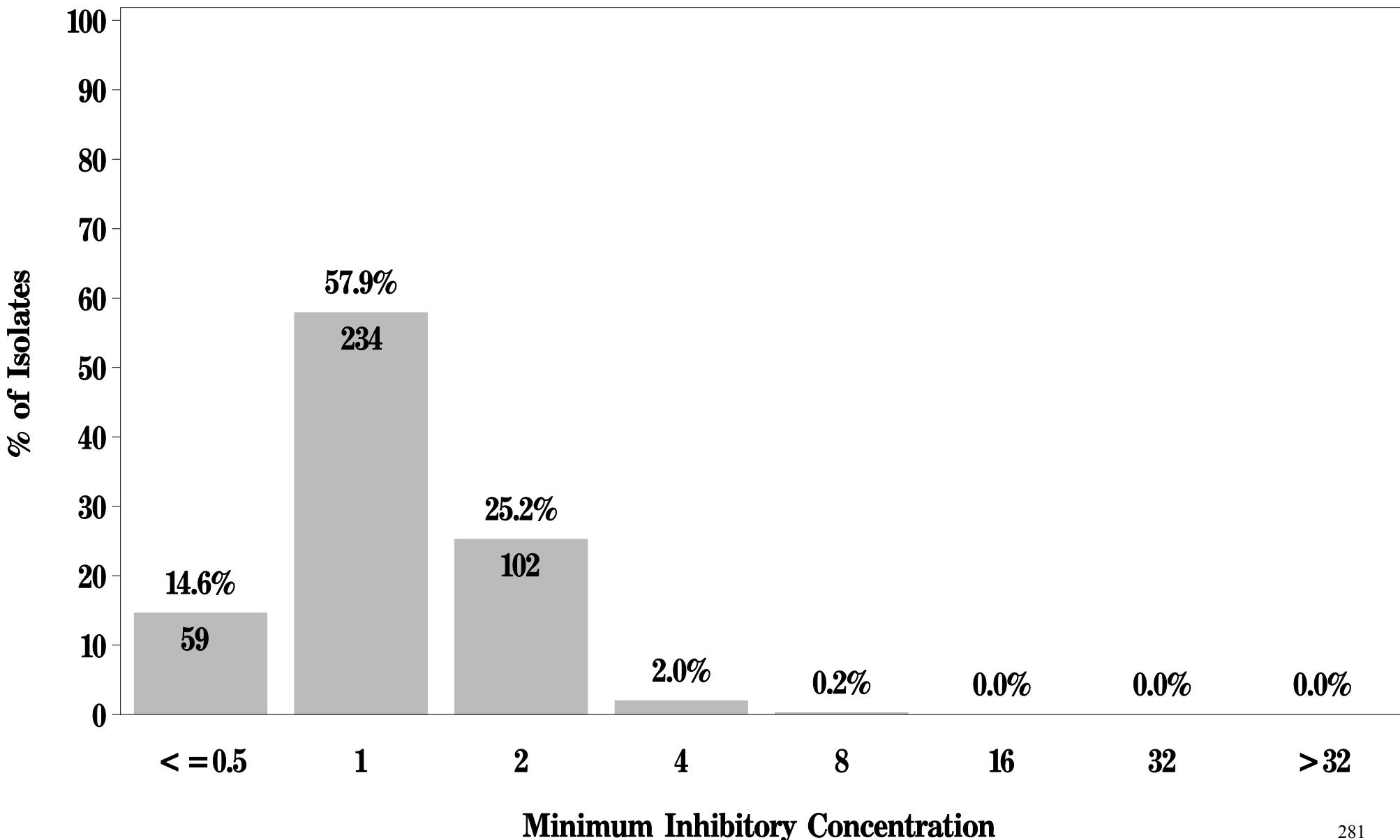


Table 31. Antimicrobial Resistance among *Enterococcus* by Meat Type for all Sites, 2004

Antimicrobial Agent	Chicken Breast (N=466)		Ground Turkey (N=418)		Ground Beef (N=432)		Pork Chop (N=426)	
	n	%	n	%	n	%	n	%
Lincomycin	313	67.2%*	376	86.0%	234	52.2%	265	65.6%
Quinupristin-Dalfopristin†‡	113	29.9%	111	62.7%	19	7.5%	5	5.5%
Tetracycline	229	49.1%	380	87.0%	136	30.4%	297	73.5%
Bacitracin	376	80.7%	350	80.1%	149	33.3%	118	29.2%
Flavomycin	319	68.5%	156	35.7%	239	53.3%	87	21.5%
Nitrofurantoin	305	65.5%	118	27.0%	90	20.1%	32	7.9%
Kanamycin	162	34.8%	179	41.0%	61	13.6%	19	4.7%
Ciprofloxacin	190	40.8%	108	24.7%	71	15.8%	33	8.2%
Erythromycin	79	17.0%	162	37.1%	29	6.5%	35	8.7%
Tylosin	70	15.0%	151	34.6%	23	5.1%	31	7.7%
Penicillin	144	30.9%	106	24.3%	6	1.3%	7	1.7%
Streptomycin	53	11.4%	129	29.5%	24	5.4%	34	8.4%
Gentamicin	33	7.1%	88	20.1%	2	0.4%	6	1.5%
Daptomycin	14	3.0%	13	3.0%	21	4.7%	0	0.0%
Chloramphenicol	0	-§	0	-	2	0.4%	2	0.5%
Linezolid,	0	-	0	-	0	-	0	-
Vancomycin	0	-	0	-	0	-	0	-

* Where % Resistance = (# isolates per meat type resistant to antimicrobial) / (total # isolates per meat type).

† Data presented for all species except *E. faecalis*, which is considered intrinsically resistant to Quinupristin-Dalfopristin.

‡ Number of *E. faecalis* in CB = 88, GT = 260, GB = 194, PC = 313.

§ Dashes indicate 0.0% resistance to antimicrobial.

Table 32. Antimicrobial Resistance among *Enterococcus* by Species, 2004

Species	Antimicrobial Agent																
	LIN	QDA	TET	BAC	FLA	DAP	NIT	KAN	CIP	ERY	TYL	PEN	STR	GEN	CHL	LZD	VAN
<i>E. casseliflavus</i> (n=3)	-	*	-	33.3% [†]	100.0%	-	-	-	-	-	-	-	-	-	-	-	-
<i>E. durans</i> (n=3)	66.7%	-	33.3%	66.7%	100.0%	-	66.7%	66.7%	-	-	-	33.3%	33.3%	-	-	-	-
<i>E. faecalis</i> (n=855)	86.3%	‡	66.8%	52.6%	-	-	0.6%	13.2%	7.7%	18.4%	18.5%	-	15.2%	10.4%	0.2%	-	-
<i>E. faecium</i> (n=757)	51.3%	31.2%	52.8%	69.0%	87.3%	3.7%	69.2%	39.1%	43.7%	18.1%	14.0%	33.4%	13.5%	5.0%	0.3%	-	-
<i>E. gallinarum</i> (n=7)	14.3%	-	42.9%	57.1%	100.0%	-	-	14.3%	28.6%	-	-	-	-	14.3%	-	-	-
<i>E. hirae</i> (n=129)	45.0%	8.5%	51.2%	10.1%	98.4%	15.5%	10.1%	6.2%	2.3%	7.8%	7.8%	7.0%	4.7%	-	-	-	-
<i>E. mundtii</i> (n=1)	100.0%	100.0%	100.0%	100.0%	-	-	100.0%	100.0%	-	100.0%	100.0%	-	100.0%	100.0%	-	-	-
Total (N=1755)	67.7%	27.6%	59.4%	56.6%	45.6%	2.7%	31.1%	24.0%	22.9%	17.4%	15.7%	15.0%	13.7%	7.4%	0.2%	-	-

* Dashes indicate 0.0% resistance to antimicrobial.

† Where % Resistance = (# isolates per species resistant to antimicrobial) / (total # isolates per species).

‡ QDA resistance is not presented for *E. faecalis*.

Table 33. Antimicrobial Resistance among *Enterococcus faecalis* & *E. faecium* by Meat Type, 2004

		Antimicrobial Agent																	
Meat Type	Species	LIN	QDA	BAC	TET	FLA	DAP	NIT	KAN	CIP	ERY	TYL	PEN	STR	GEN	CHL	LZD	VAN	
Chicken Breast	<i>E. faecalis</i> (n=88)	98.9%*	-†	78.4%	63.6%	-‡	-	1.1%	22.7%	8.0%	35.2%	34.1%	-	18.2%	19.3%	-	-	-	
	<i>E. faecium</i> (n=348)	60.3%	31.6%	84.8%	45.1%	83.6%	4.0%	85.3%	39.7%	52.3%	12.6%	10.3%	39.1%	8.3%	4.3%	-	-	-	
Ground Turkey	<i>E. faecalis</i> (n=260)	94.2%	-†	72.7%	88.1%	-	-	1.2%	30.0%	5.8%	33.8%	34.6%	-	26.9%	24.6%	-	-	-	
	<i>E. faecium</i> (n=172)	75.0%	64.5%	90.7%	86.6%	87.8%	7.6%	66.9%	57.6%	53.5%	43.0%	35.5%	61.6%	34.3%	13.4%	-	-	-	
Ground Beef	<i>E. faecalis</i> (n=194)	79.4%	-†	45.9%	25.3%	-	-	-	3.1%	12.9%	3.6%	3.6%	-	7.7%	1.0%	-	-	-	
	<i>E. faecium</i> (n=162)	24.7%	6.2%	35.2%	24.7%	91.4%	0.6%	51.9%	33.3%	27.2%	9.3%	5.6%	3.1%	5.6%	-	1.2%	-	-	
Pork Chop	<i>E. faecalis</i> (n=313)	80.5%	-†	32.9%	75.7%	-	-	0.3%	2.9%	6.1%	9.9%	9.9%	-	9.3%	1.9%	0.6%	-	-	
	<i>E. faecium</i> (n=75)	12.0%	6.7%	18.7%	72.0%	94.7%	-	37.3%	6.7%	17.3%	5.3%	-	8.0%	6.7%	-	-	-	-	
Total (N=1612)		69.9%	31.2%	60.3%	60.2%	41.0%	8.3%	32.8%	25.4%	24.6%	18.2%	16.4%	15.7%	14.4%	7.9%	0.2%	-	-	

* Where % Resistance = (# isolates resistant to antimicrobial per meat type per site) / (total # isolates per meat type per site).

† QDA resistance is not presented for *E. faecalis*.

‡ Dashes indicate 0.0% resistance to antimicrobial.

Table 34. Antimicrobial Resistance among *Enterococcus* by Site, Meat Type, and Antimicrobial Agent, 2004

Site	Meat Type	Antimicrobial Agent																
		LIN	QDA*	TET	BAC	FLA	DAP	NIT	KAN	CIP	ERY	TYL	PEN	STR	GEN	CHL	LZD	VAN
GA	CB (n=120)	77.5% [†]	40.9%	58.3%	79.2%	48.3%	4.2%	40.8%	30.0%	29.2%	29.2%	25.8%	14.2%	14.2%	13.3%	-‡	-	-
	GT (n=120)	96.7%	58.3%	80.8%	73.3%	9.2%	0.8%	9.2%	25.8%	8.3%	29.2%	30.0%	4.2%	18.3%	16.7%	-	-	-
	GB (n=117)	71.8%	10.3%	29.1%	40.2%	33.3%	1.7%	13.7%	9.4%	10.3%	8.5%	6.8%	-	8.5%	0.9%	-	-	-
	PC (n=116)	79.3%	10.0%	74.1%	37.1%	7.8%	-	6.9%	1.7%	3.4%	5.2%	5.2%	0.9%	9.5%	0.9%	0.9%	-	-
	Total (N=473)	81.4%	30.7%	60.7%	57.7%	24.7%	1.7%	17.8%	16.9%	12.9%	18.2%	17.1%	4.9%	12.7%	8.0%	0.2%	-	-
MD	CB (n=114)	71.1%	36.0%	53.5%	80.7%	77.2%	5.3%	83.3%	50.9%	52.6%	18.4%	14.9%	54.4%	7.9%	3.5%	-	-	-
	GT (n=106)	87.7%	69.7%	91.5%	78.3%	62.3%	6.6%	49.1%	60.4%	40.6%	46.2%	37.7%	52.8%	35.8%	25.5%	-	-	-
	GB (n=100)	29.0%	7.6%	37.0%	21.0%	75.0%	6.0%	29.0%	23.0%	21.0%	7.0%	5.0%	5.0%	9.0%	-	1.0%	-	-
	PC (n=77)	48.1%	-	68.8%	27.3%	37.7%	-	19.5%	7.8%	11.7%	3.9%	-	6.5%	5.2%	-	-	-	-
	Total (N=397)	60.5%	33.6%	62.5%	54.7%	65.0%	4.8%	48.1%	38.0%	33.5%	20.2%	15.6%	32.2%	15.1%	7.8%	0.3%	-	-
OR	CB (n=118)	74.6%	17.0%	46.6%	90.7%	71.2%	0.8%	66.9%	34.7%	41.5%	14.4%	13.6%	38.1%	10.2%	5.9%	-	-	-
	GT (n=105)	81.9%	60.5%	87.6%	88.6%	35.2%	1.0%	15.2%	39.0%	17.1%	41.9%	41.0%	15.2%	37.1%	25.7%	-	-	-
	GB (n=115)	47.8%	3.8%	28.7%	35.7%	67.0%	9.6%	13.0%	11.3%	9.6%	5.2%	4.3%	-	0.9%	-	-	-	-
	PC (n=108)	60.2%	13.8%	69.4%	29.6%	25.0%	-	2.8%	2.8%	4.6%	7.4%	7.4%	0.9%	6.5%	1.9%	0.9%	-	-
	Total (N=446)	65.9%	19.2%	57.2%	61.2%	50.4%	2.9%	25.3%	22.0%	18.6%	16.8%	16.1%	13.9%	13.2%	8.1%	0.2%	-	-
TN	CB (n=114)	44.7%	28.7%	37.7%	71.9%	78.1%	1.8%	71.9%	23.7%	40.4%	5.3%	5.3%	17.5%	13.2%	5.3%	-	-	-
	GT (n=106)	76.4%	54.9%	88.7%	81.1%	39.6%	3.8%	36.8%	40.6%	34.9%	32.1%	30.2%	27.4%	28.3%	13.2%	-	-	-
	GB (n=116)	56.9%	10.3%	27.6%	34.5%	41.4%	1.7%	25.9%	12.1%	23.3%	5.2%	4.3%	0.9%	3.4%	0.9%	0.9%	-	-
	PC (n=103)	68.9%	-	80.6%	21.4%	21.4%	-	5.8%	7.8%	14.6%	17.5%	16.5%	-	11.7%	2.9%	-	-	-
	Total (N=439)	61.3%	27.0%	57.4%	52.4%	45.8%	1.8%	35.8%	21.0%	28.5%	14.6%	13.7%	11.4%	13.9%	5.5%	0.2%	-	-
Total (N=1755)		67.7%	27.6%	59.4%	56.6%	45.6%	2.7%	31.1%	24.0%	22.9%	17.4%	15.7%	15.0%	13.7%	7.4%	0.2%	-	-

* Data does not include *E. faecalis* in QDA, as it is considered intrinsically resistant.

† Where % Resistance = (# isolates resistant to antimicrobial per meat type per site) / (total # isolates per meat type per site).

‡ Dashes indicate 0.0% resistance to antimicrobial.

Table 35. Number of *Enterococcus faecalis* (N=855) Resistant to Multiple Antimicrobial Agents, * 2004

Meat Type	Number of Antimicrobials				
	0	1	2-4	5-7	≥8
Chicken Breast	0	9	50	28	1
Ground Turkey	3	13	153	91	0
Ground Beef	25	56	108	5	0
Pork Chop	19	72	209	11	2
Total	47	150	520	135	3

* Data does not include QDA, as *E. faecalis* is considered intrinsically resistant.

Table 36. Number of *Enterococcus faecium* (N=757) Resistant to Multiple Antimicrobial Agents, 2004

Meat Type	Number of Antimicrobials				
	0	1	2-4	5-7	≥ 8
Chicken Breast	1	12	152	152	31
Ground Turkey	0	1	29	70	72
Ground Beef	6	53	84	13	6
Pork Chop	0	10	58	7	0
Total	7	76	323	242	109

Table 37. *Escherichia coli* by Meat Type, 2004

Meat Type	N[*]	n[†]	% [‡]
Chicken Breast	476	400	84.0%
Ground Turkey	466	376	80.7%
Ground Beef	480	338	70.4%
Pork Chop	478	232	48.5%
Total	1900	1346	70.8%

^{*} Where N = Number of retail meat samples.

[†] Where n = number of *E. coli* positive samples.

[‡] Where % = (n / N).

Table 38. *Escherichia coli* by Site and Meat Type, 2004

Site	Chicken Breast		Ground Turkey		Ground Beef		Pork Chop	
	n	%*	n	%	n	%	n	%
Georgia (n=389)	115	29.6%	119	30.6%	91	23.4%	64	16.5%
Maryland (n=364)	110	30.2%	109	29.9%	83	22.8%	62	17.0%
Oregon (n=276)	73	26.4%	53	19.2%	99	35.9%	51	18.5%
Tennessee (n=317)	102	32.1%	95	29.9%	65	20.4%	55	17.3%
Total (N=1346)	400	29.7%	376	27.9%	338	25.1%	232	17.2%

* Where % Positive = (# isolates per meat type per site) / (total # isolates for that site).

Table 39. *Escherichia coli* Isolates by Month for All Sites, 2004

Month	n	%*
January	117	8.7%
February	106	7.9%
March	107	7.9%
April	115	8.5%
May	127	9.4%
June	96	7.1%
July	107	7.9%
August	117	8.7%
September	111	8.2%
October	118	8.8%
November	113	8.4%
December	112	8.3%
Total	1346	100%

* Where % Positive = (# isolates per month) / (total # isolates).

Table 40. Antimicrobial Resistance among *E. coli* Isolates (N=1346), 2004

Antimicrobial Agent	n	%R*
Tetracycline	678	50.4%
Streptomycin	501	37.2%
Sulfisoxazole	436	32.4%
Ampicillin	246	18.3%
Gentamicin	235	17.5%
Kanamycin	114	8.5%
Amoxicillin/Clavulanic Acid	86	6.4%
Nalidixic Acid	73	5.4%
Cefoxitin	59	4.4%
Trimethoprim/Sulfamethoxazole	42	3.1%
Chloramphenicol	32	2.4%
Ceftiofur	31	2.3%
Ciprofloxacin	3	0.2%
Amikacin	0	0.0%
Ceftriaxone	0	0.0%

*

* Where % R = (n / N).

Figure 16. Antimicrobial Resistance among *E. coli* isolates (n =1346), 2004

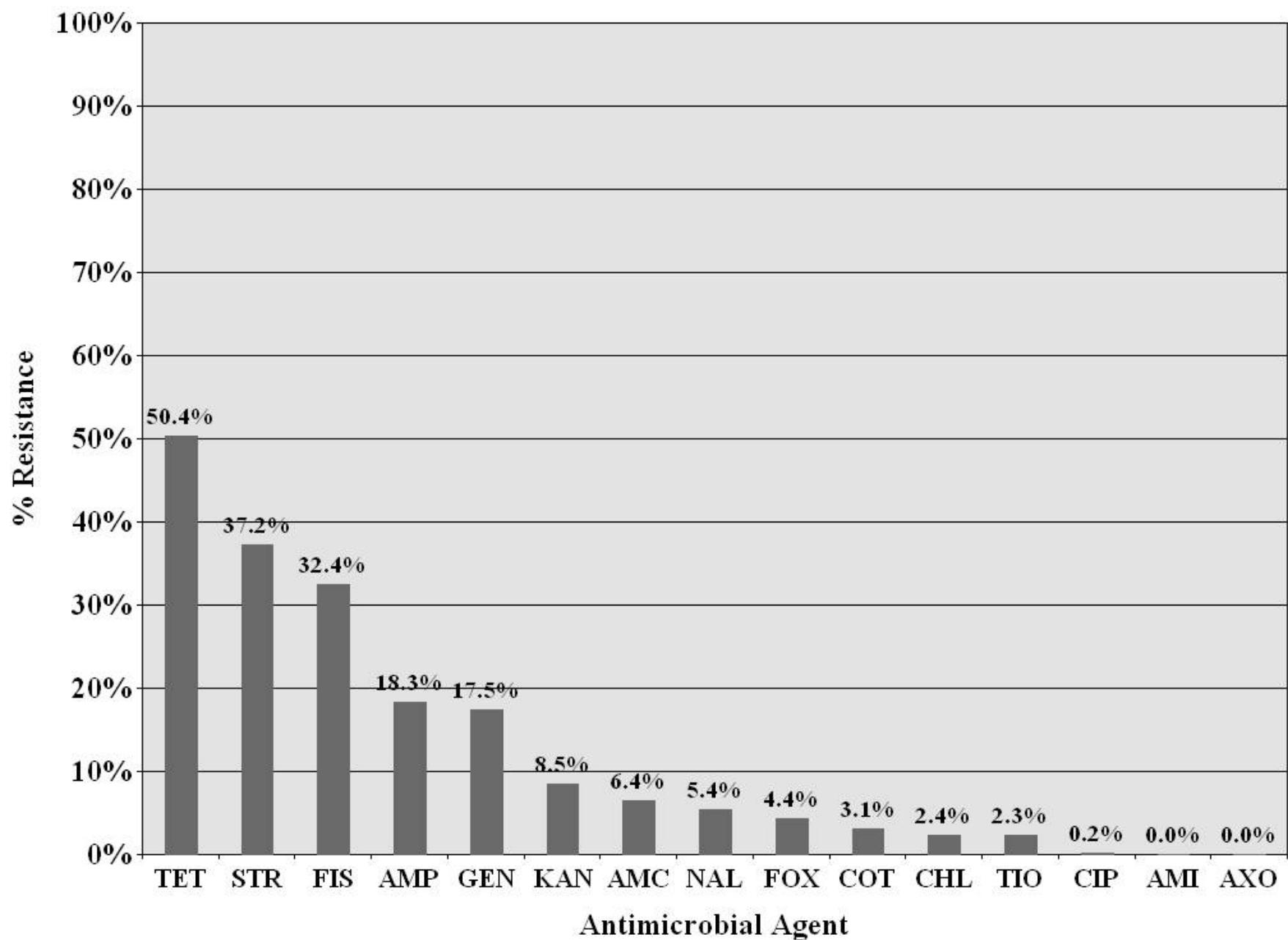


Figure 17. MIC Distribution among all Antimicrobial Agents

<i>E. coli</i> from All Meats Types (N=1346)		Distribution (%) of MICs (in µg/ml)																
Antimicrobial Agent	%R	0.015	0.03	0.06	0.125	0.25	0.5	1	2	4	8	16	32	64	128	256	512	>512
Ampicillin	18.3							8.2	40.5	31.4	1.0	0.5	0.5	17.8				
Amoxicillin/Clavulanic Acid	6.4							2.7	22.7	50.2	16.7	1.3	5.1	1.3				
Cefoxitin	4.4						0.1	1.8	23.0	55.2	14.1	1.4	2.2	2.2				
Ceftiofur	2.3				4.5	49.7	40.4	2.2	0.4	0.5	1.5	0.8						
Ceftriaxone	0.0					94.2	1.5	1.4	0.1		1.3	0.9	0.5					
Nalidixic Acid	5.4						5.4	64.8	23.0	1.0	0.3	0.4	5.0					
Ciprofloxacin	0.2	90.9	2.7	0.4	1.6	3.6	0.5				0.2							
Sulfisoxazole	32.4									60.1	3.3	4.1	0.1	0.1	32.4			
Trimethoprim/Sulfamethoxazole	3.1				89.2	5.6	1.6	0.4	0.1		3.1							
Amikacin	0.0						0.1	15.9	65.1	16.6	2.1	0.3						
Gentamicin	17.5					7.1	51.7	19.5	2.4	0.3	1.5	6.5	11.0					
Kanamycin	8.5									84.6	6.0	0.9	0.1	8.4				
Streptomycin*	37.2										62.8	11.7	25.5					
Chloramphenicol	2.4							1.6	33.1	61.6	1.3	0.4	2.0					
Tetracycline	50.4								47.3	2.4	0.8	4.2	45.3					

Vertical bars show the CLSI/NCCLS Susceptible/Resistant breakpoints for each drug where appropriate.

*Currently no CLSI/NCCLS breakpoints have been established for this organism/antimicrobial combination. Indicated breakpoints were established by NARMS.

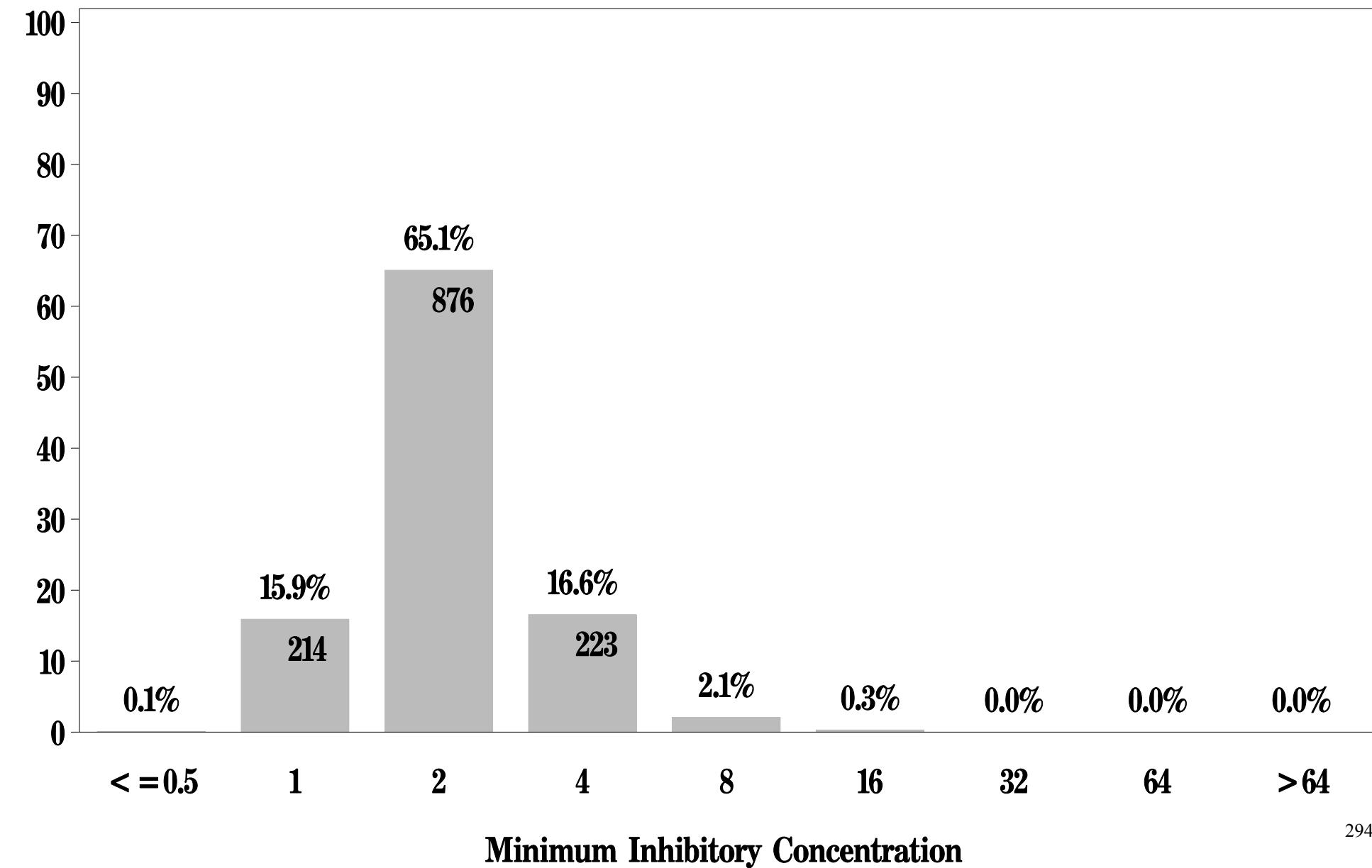
†Discrepancies between %R and sums of distribution %s are due to rounding.

Unshaded areas indicate the dilution ranges of the Sensititre plate used to test the 2004 isolates.

NARMS

**Figure 17a: Minimum Inhibitory Concentration of Amikacin
for *Escherichia coli* (N=1346 Isolates)**

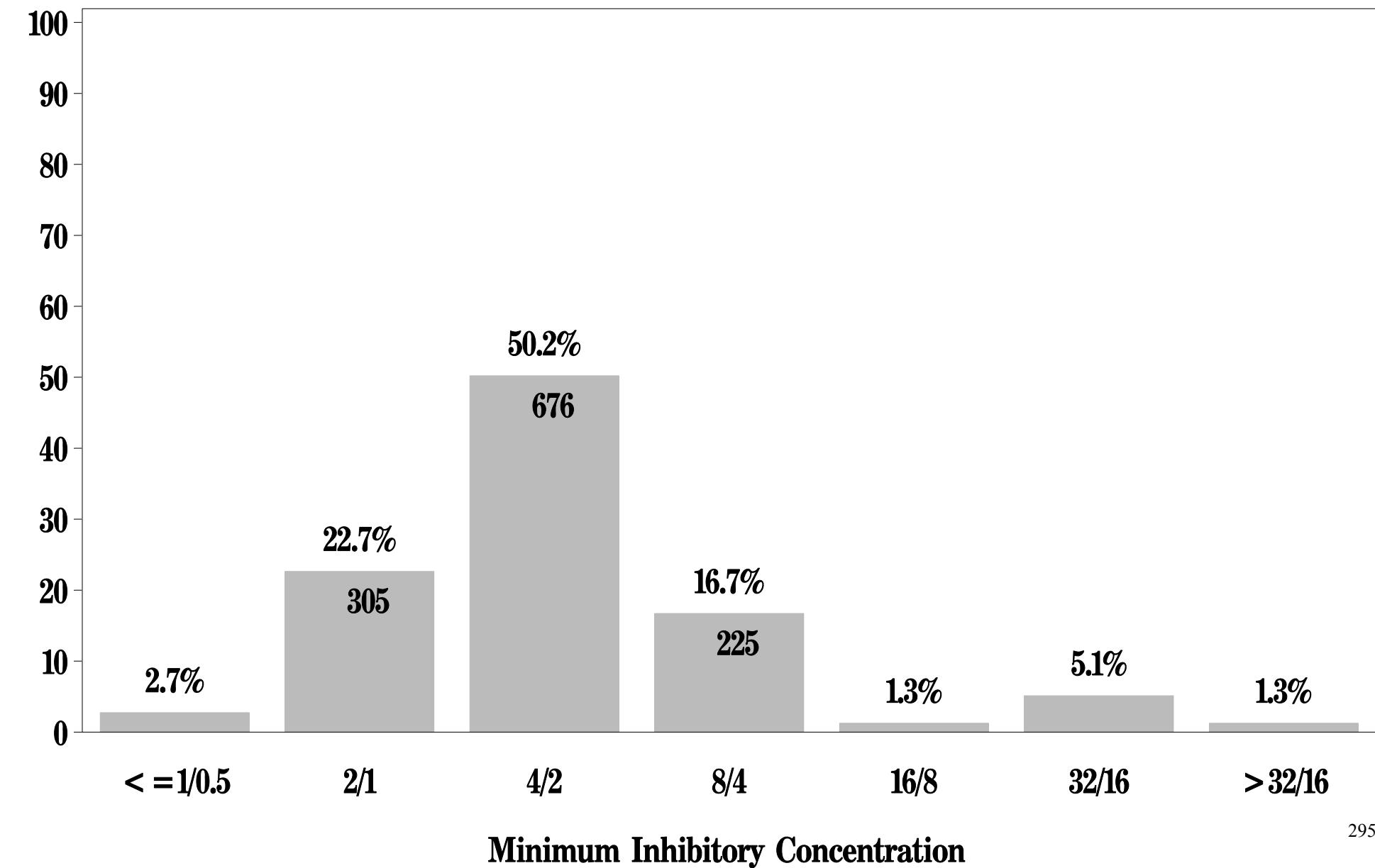
Breakpoints: Susceptible $\leq 16 \text{ } \mu\text{g/mL}$ Resistant $\geq 64 \text{ } \mu\text{g/mL}$



NARMS

Figure 17b: Minimum Inhibitory Concentration of Amoxicillin/Clavulanic acid
for *Escherichia coli* (N=1346 Isolates)

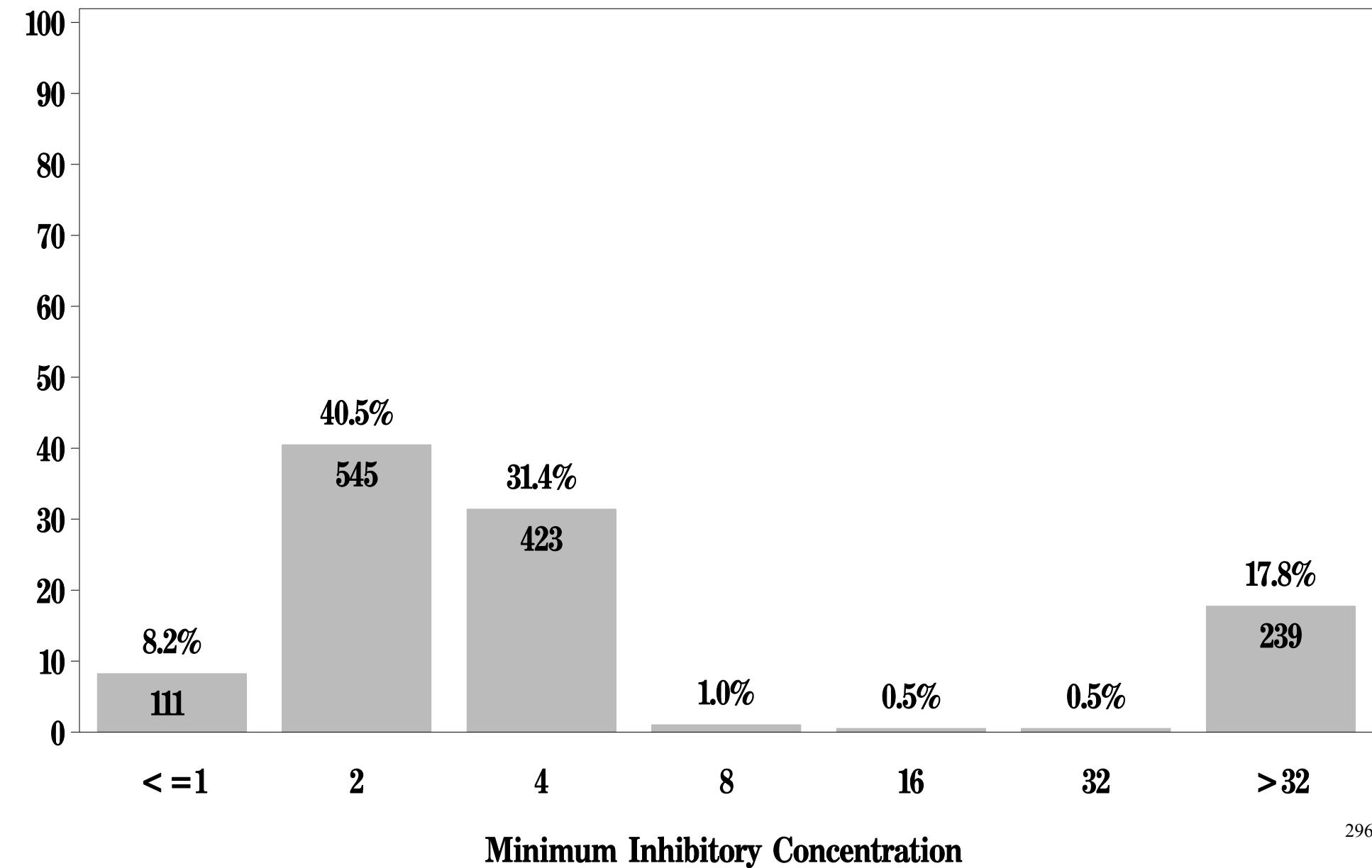
Breakpoints: Susceptible $\leq 8 \text{ } \mu\text{g/mL}$ Resistant $\geq 32 \text{ } \mu\text{g/mL}$



NARMS

**Figure 17c: Minimum Inhibitory Concentration of Ampicillin
for *Escherichia coli* (N=1346 Isolates)**

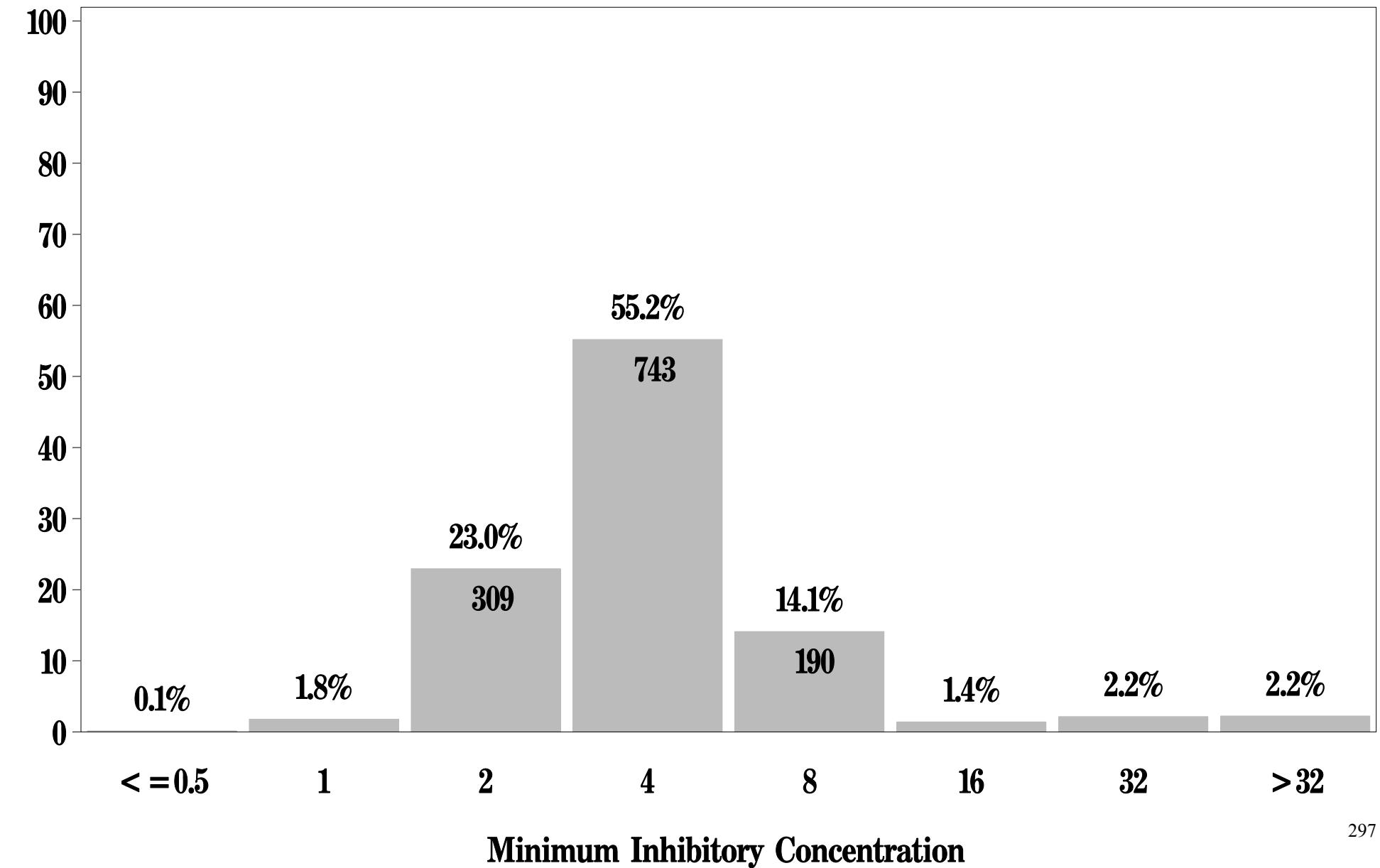
Breakpoints: Susceptible $\leq 8 \text{ } \mu\text{g/mL}$ Resistant $\geq 32 \text{ } \mu\text{g/mL}$



NARMS

Figure 17d: Minimum Inhibitory Concentration of Cefoxitin
for *Escherichia* (N=1346 Isolates)

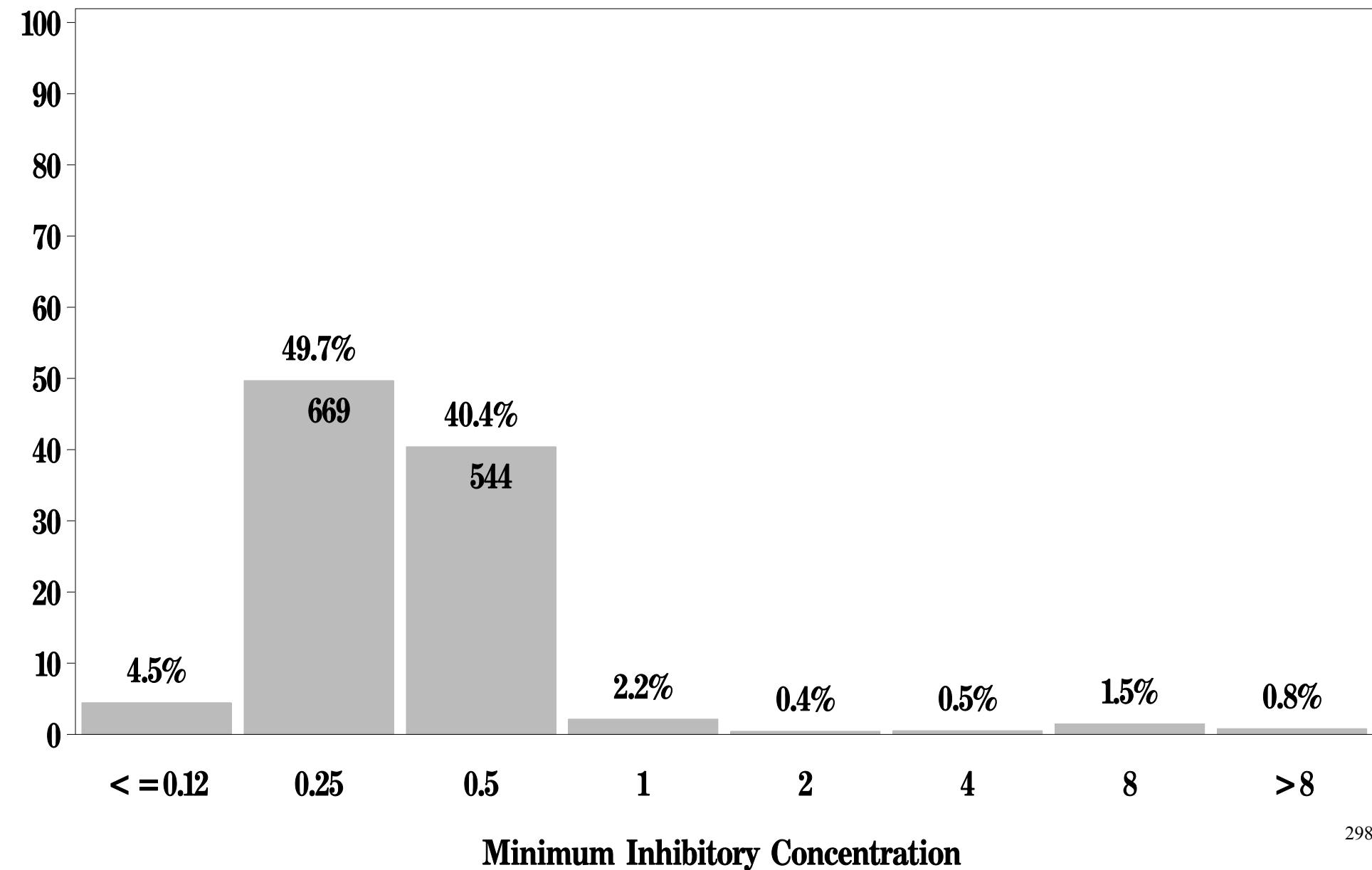
Breakpoints: Susceptible $\leq 8 \text{ } \mu\text{g/mL}$ Resistant $\geq 32 \text{ } \mu\text{g/mL}$



NARMS

**Figure 17e: Minimum Inhibitory Concentration of Ceftiofur
for *Escherichia coli* (N=1346 Isolates)**

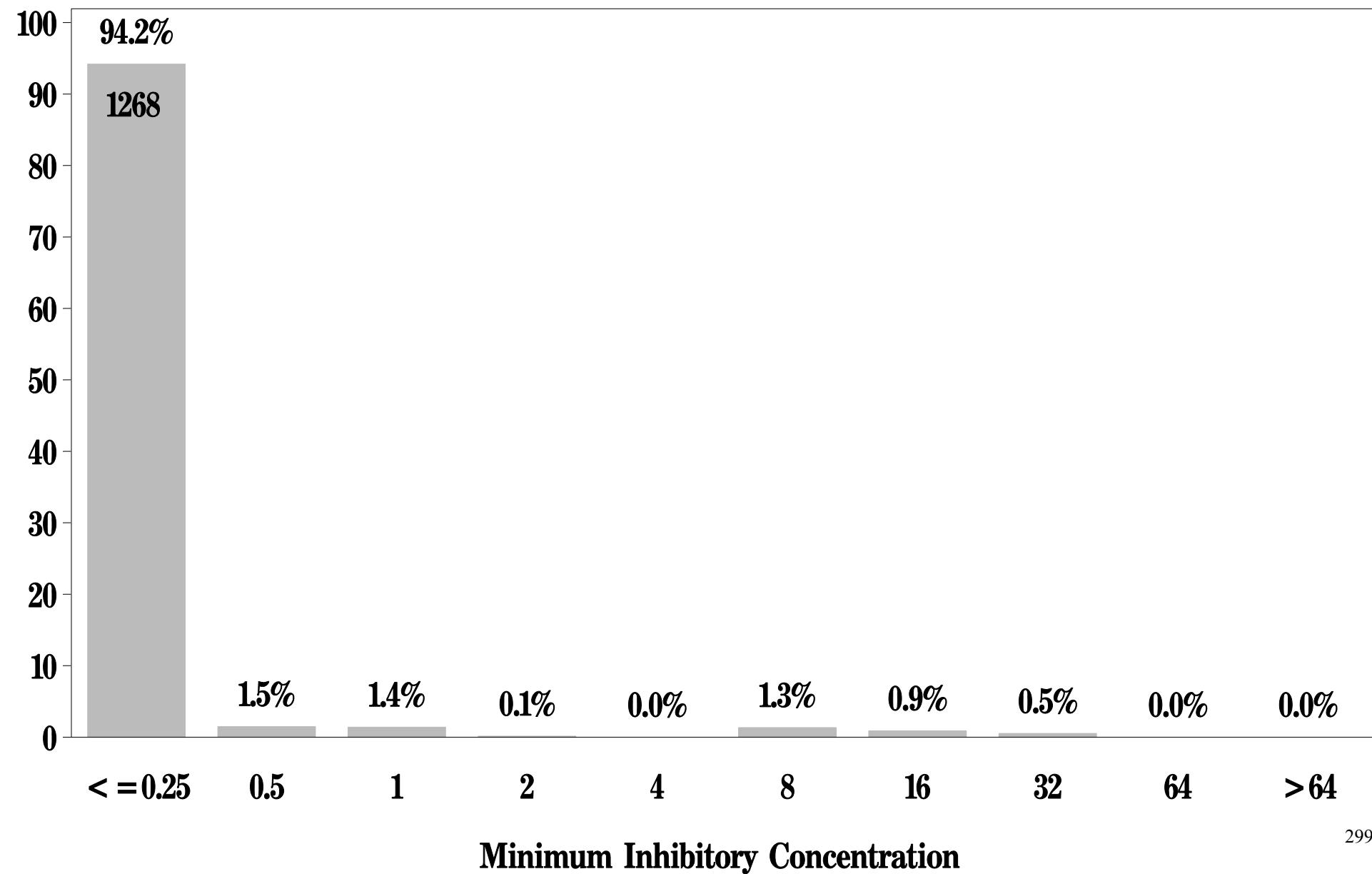
Breakpoints: Susceptible $\leq 2 \text{ } \mu\text{g/mL}$ Resistant $\geq 8 \text{ } \mu\text{g/mL}$



NARMS

Figure 17f: Minimum Inhibitory Concentration of Ceftriaxone
for *Escherichia* (N=1346 Isolates)

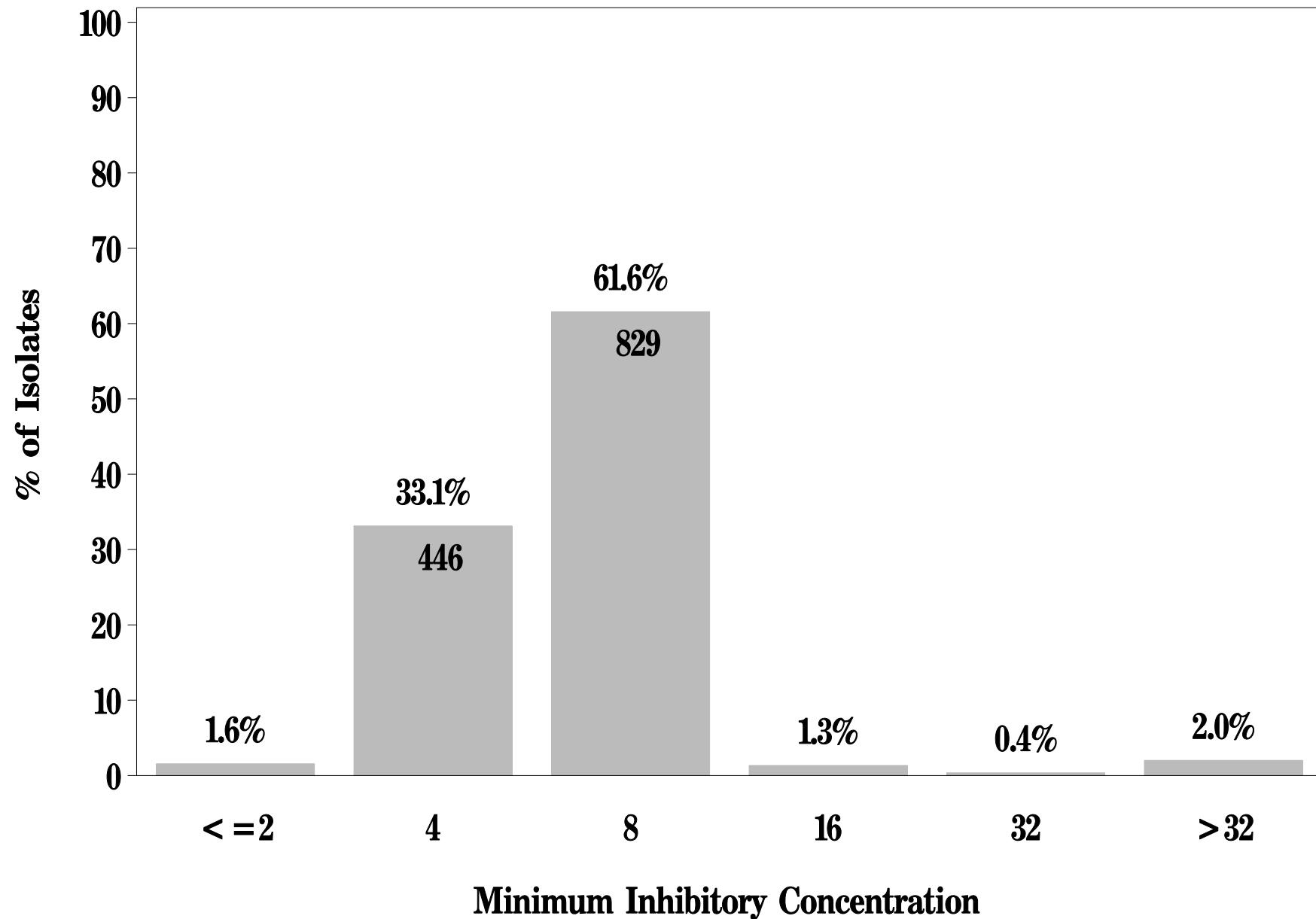
Breakpoints: Susceptible $\leq 8 \mu\text{g/mL}$ Resistant $\geq 64 \mu\text{g/mL}$



NARMS

**Figure 17g: Minimum Inhibitory Concentration of Chloramphenicol
for *Escherichia coli* (N=1346 Isolates)**

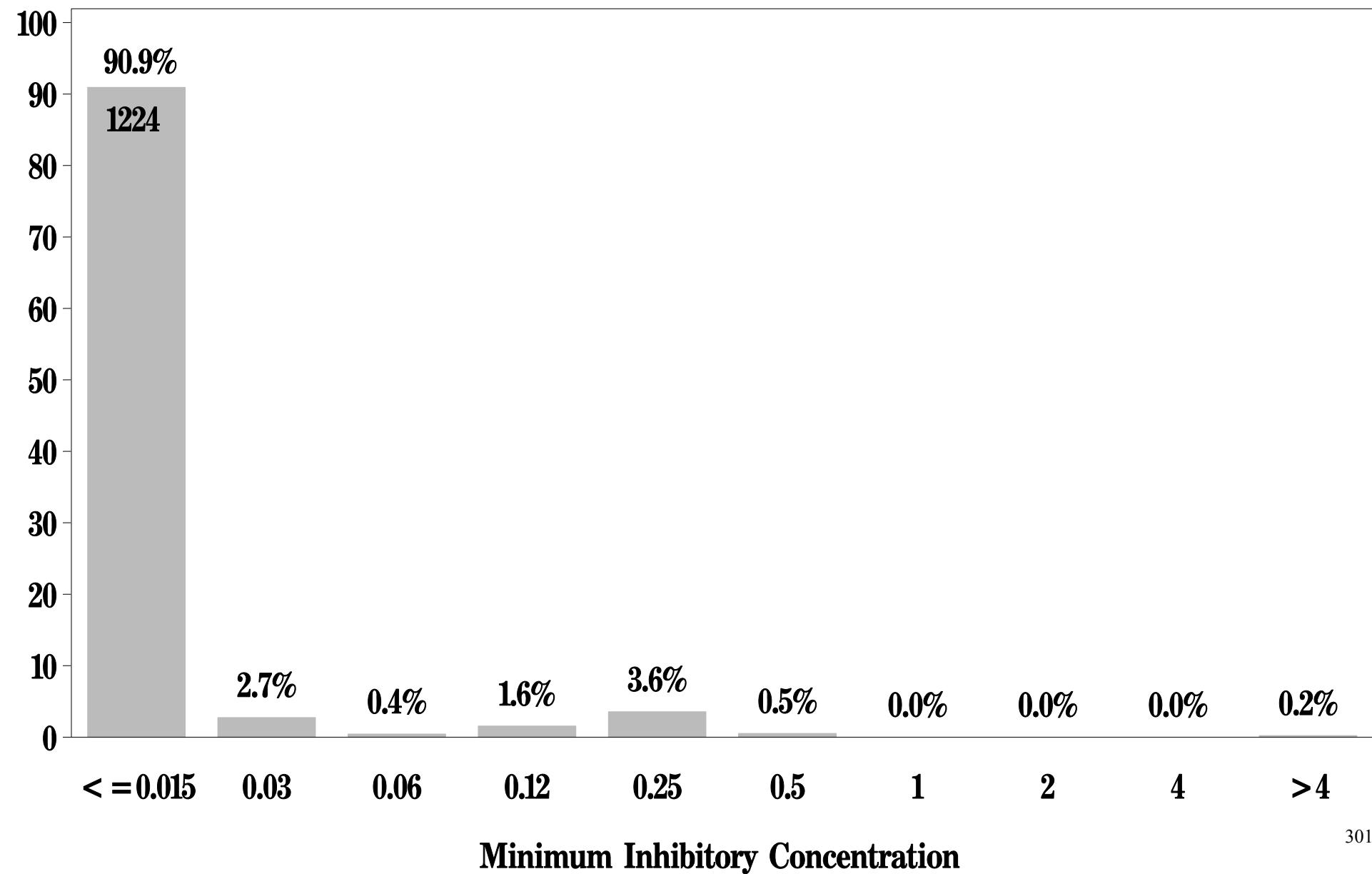
Breakpoints: Susceptible $\leq 8 \text{ } \mu\text{g/mL}$ Resistant $\geq 32 \text{ } \mu\text{g/mL}$



NARMS

Figure 17h: Minimum Inhibitory Concentration of Ciprofloxacin
for *Escherichia coli* (N=1346 Isolates)

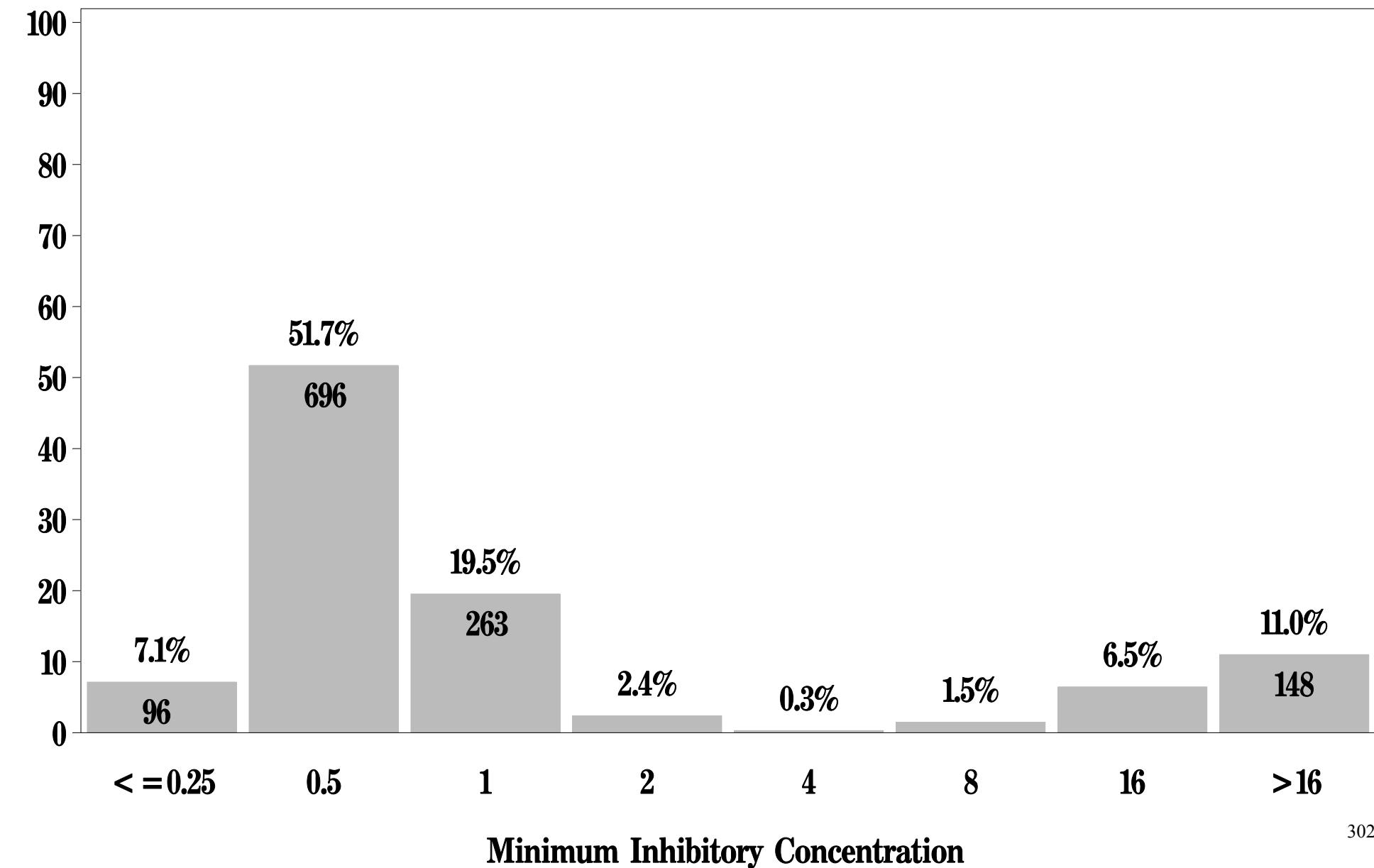
Breakpoints: Susceptible $\leq 1 \text{ } \mu\text{g/mL}$ Resistant $\geq 4 \text{ } \mu\text{g/mL}$



NARMS

**Figure 17i: Minimum Inhibitory Concentration of Gentamicin
for *Escherichia coli* (N=1346 Isolates)**

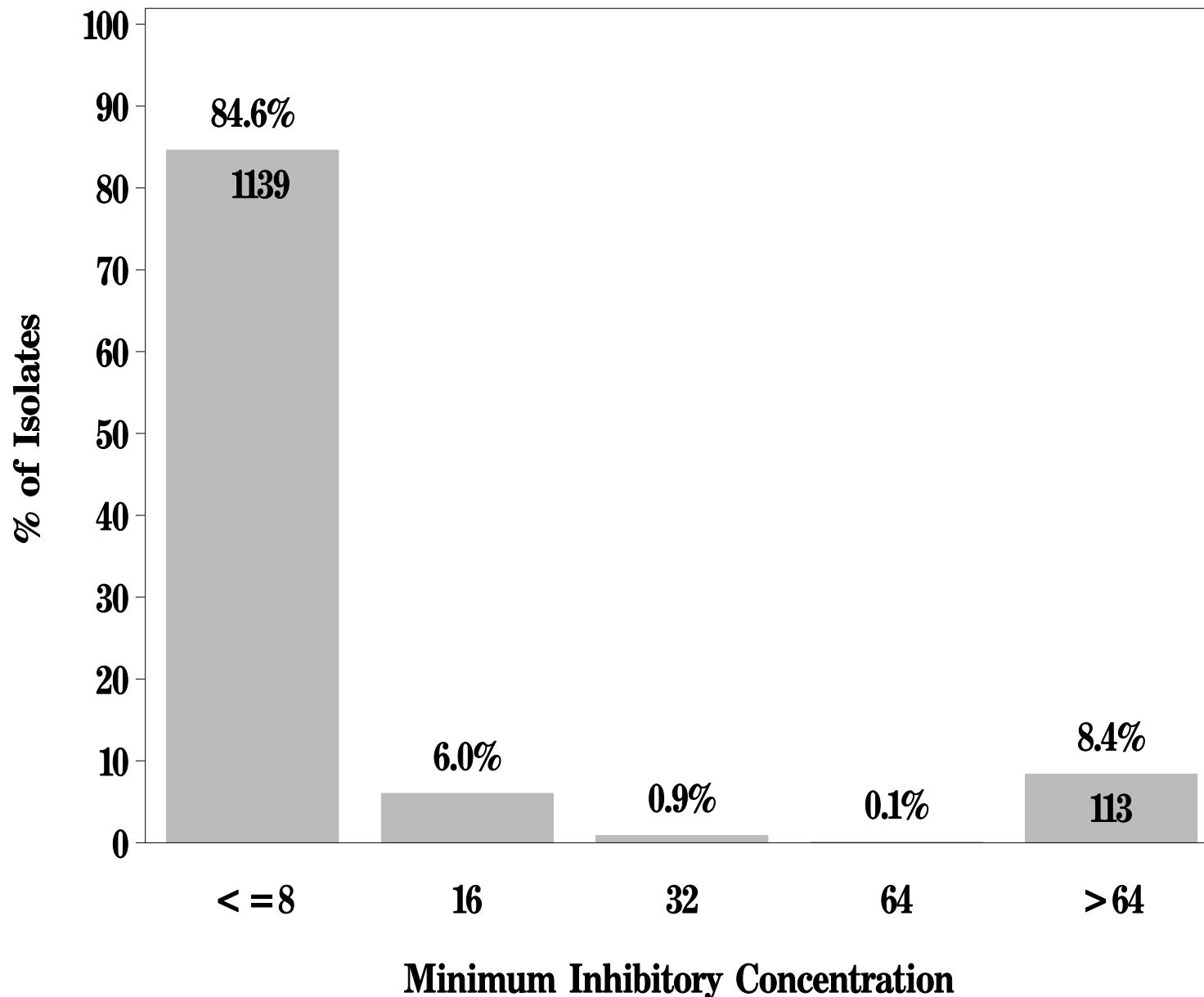
Breakpoints: Susceptible $\leq 4 \text{ } \mu\text{g/mL}$ Resistant $\geq 16 \text{ } \mu\text{g/mL}$



NARMS

**Figure 17j: Minimum Inhibitory Concentration of Kanamycin
for *Escherichia coli* (N=1346 Isolates)**

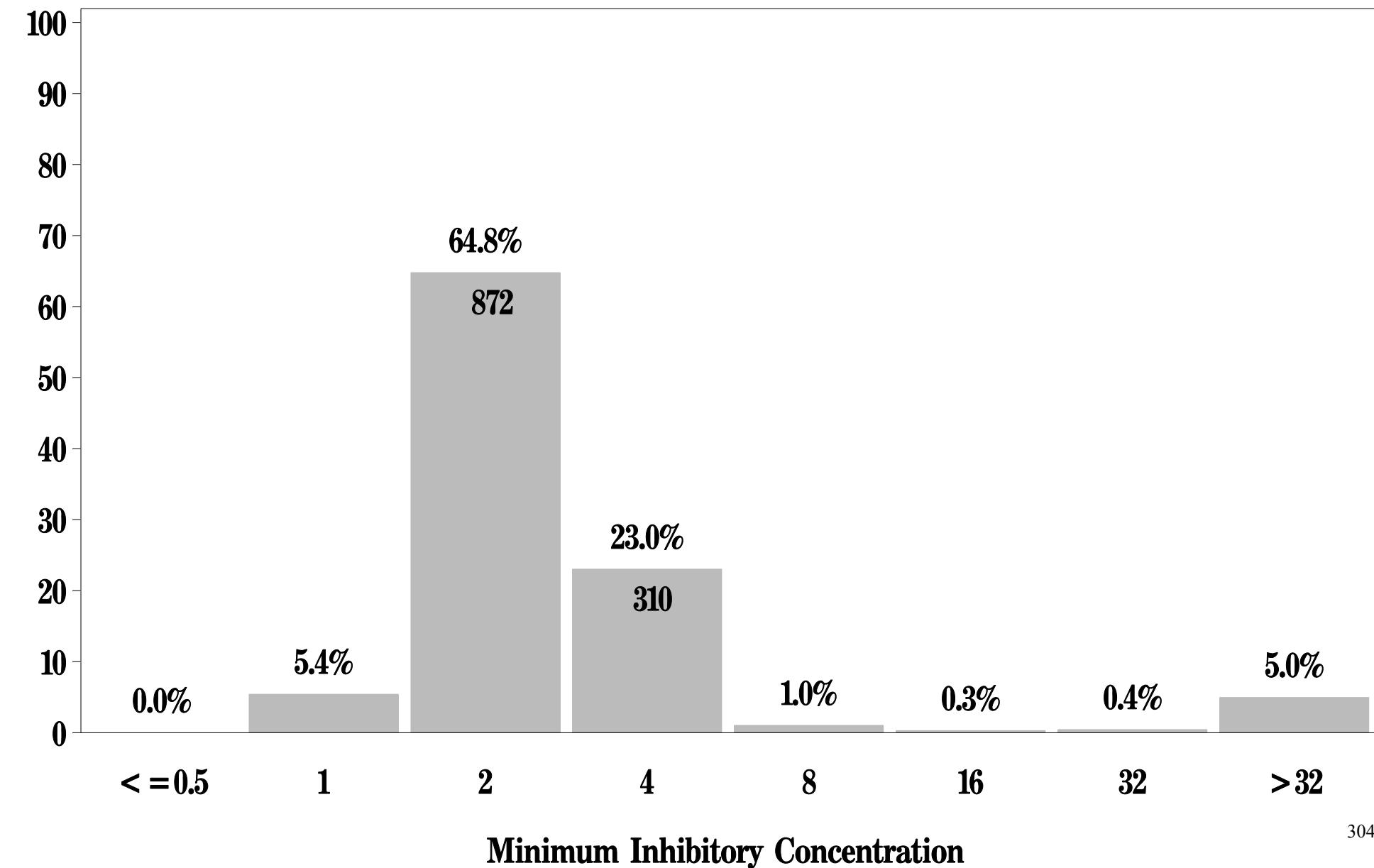
Breakpoints: Susceptible $\leq 16 \text{ } \mu\text{g/mL}$ Resistant $\geq 64 \text{ } \mu\text{g/mL}$



NARMS

Figure 17k: Minimum Inhibitory Concentration of Nalidixic acid
for *Escherichia coli* (N=1346 Isolates)

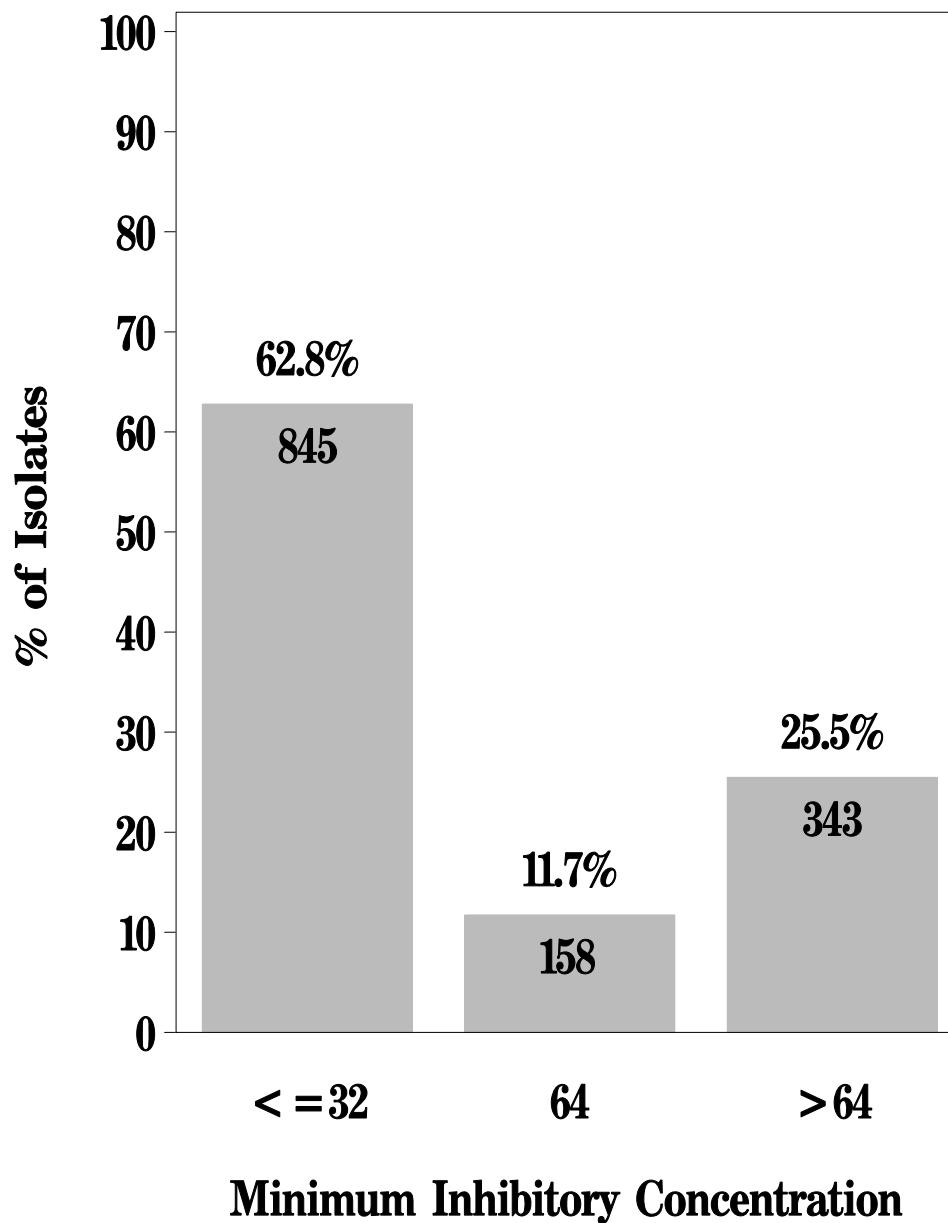
Breakpoints: Susceptible $\leq 16 \text{ } \mu\text{g/mL}$ Resistant $\geq 32 \text{ } \mu\text{g/mL}$



NARMS

**Figure 17l: Minimum Inhibitory Concentration of Streptomycin
for *Escherichia coli* (N=1346 Isolates)**

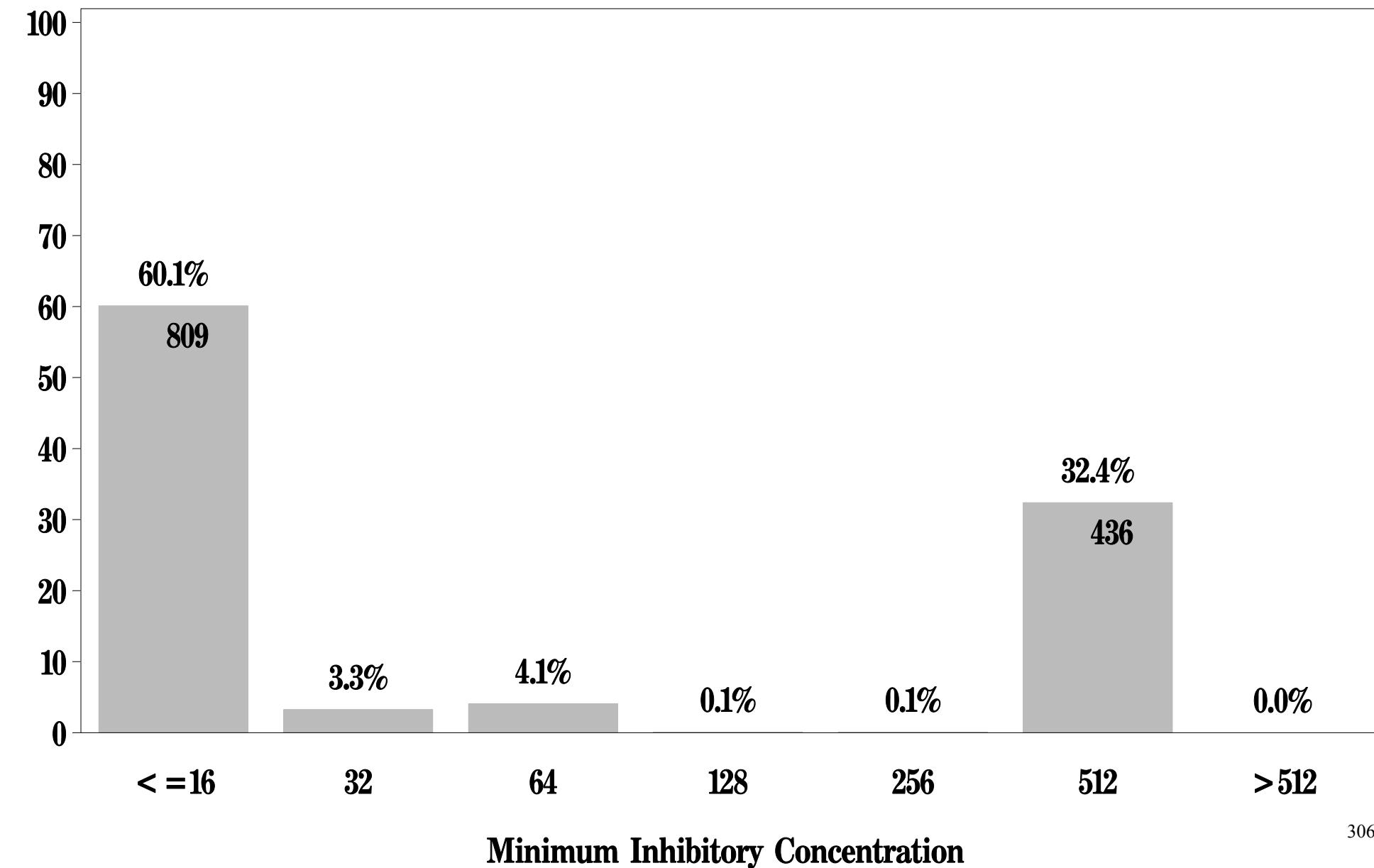
Breakpoints: Susceptible $\leq 32 \text{ } \mu\text{g/mL}$ Resistant $> 64 \text{ } \mu\text{g/mL}$



NARMS

Figure 17m: Minimum Inhibitory Concentration of Sulfisoxazole
for *Escherichia coli* (N=1346 Isolates)

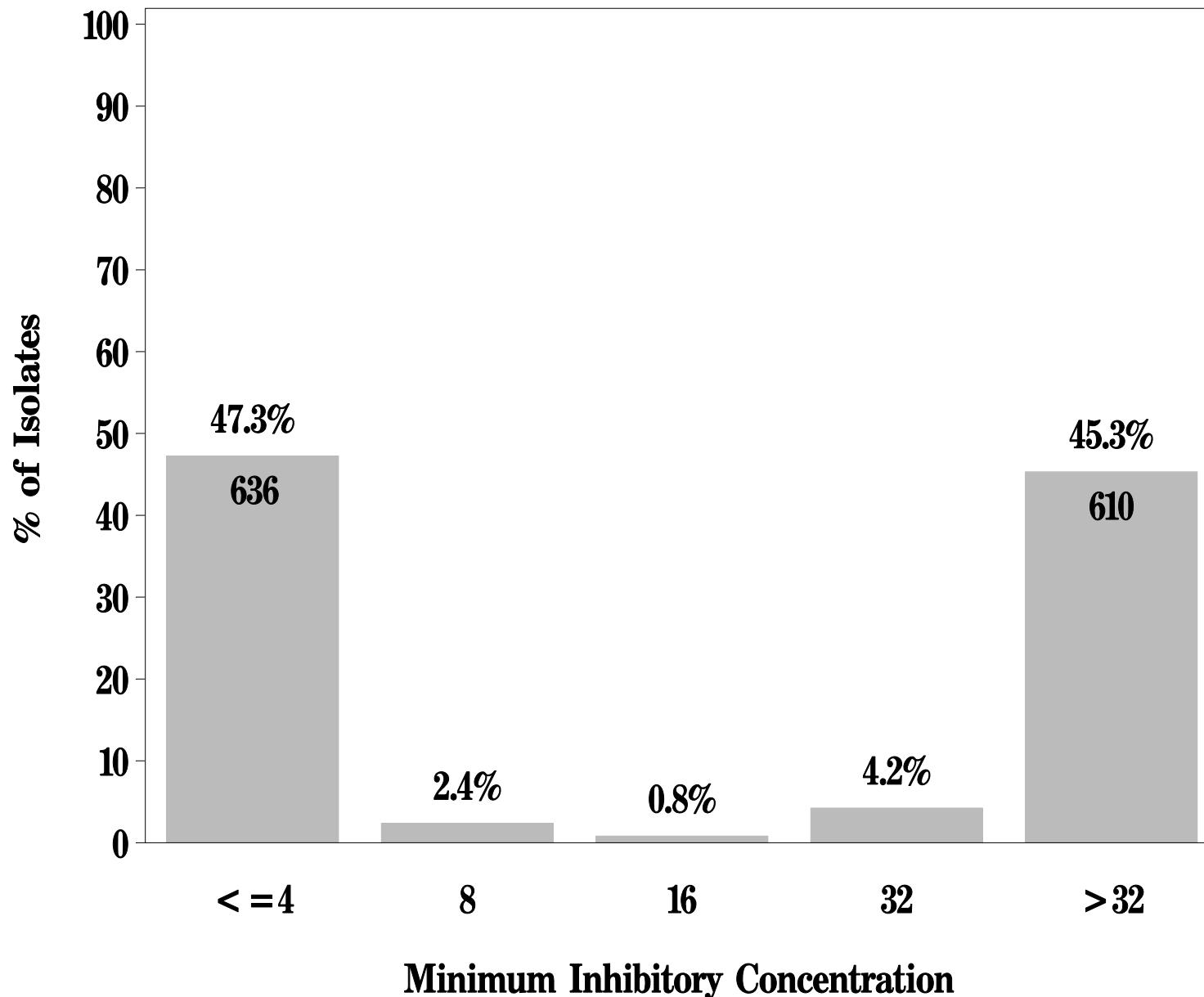
Breakpoints: Susceptible $\leq 256 \text{ } \mu\text{g/mL}$ Resistant $\geq 512 \text{ } \mu\text{g/mL}$



NARMS

**Figure 17n: Minimum Inhibitory Concentration of Tetracycline
for *Escherichia coli* (N=1346 Isolates)**

Breakpoints: Susceptible $\leq 4 \text{ } \mu\text{g/mL}$ Resistant $\geq 16 \text{ } \mu\text{g/mL}$



NARMS

**Figure 17o: Minimum Inhibitory Concentration of Trimethoprim/sulfamethoxazole
for *Escherichia coli* (N=1346 Isolates)**

Breakpoints: Susceptible $\leq 2 \text{ } \mu\text{g/mL}$ Resistant $\geq 4 \text{ } \mu\text{g/mL}$

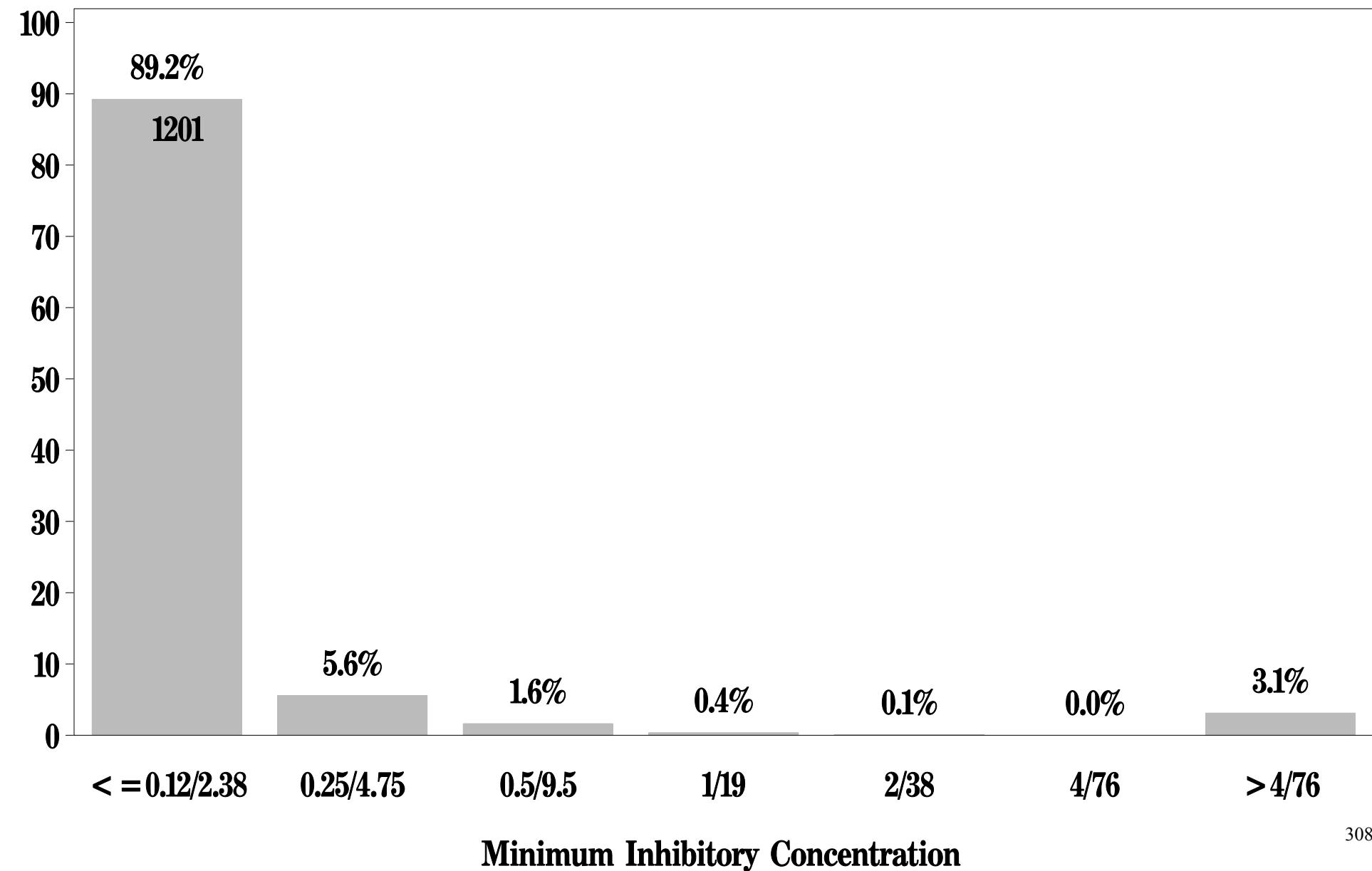


Figure 18a. MIC Distribution among *E. coli* from Chicken Breast

<i>E. coli</i> from Chicken Breast (N=400)		Distribution (%) of MICs (in µg/ml)																
Antimicrobial Agent	%R	0.015	0.03	0.06	0.125	0.25	0.5	1	2	4	8	16	32	64	128	256	512	>512
Ampicillin	17.0							6.8	40.3	34.0	1.8	0.3	0.3	16.8				
Amoxicillin/Clavulanic Acid	10.0							1.8	21.8	51.3	14.8	0.5	7.3	2.8				
Cefoxitin	8.3							0.3	15.5	53.0	20.8	2.3	3.8	4.5				
Ceftiofur	5.8					4.8	50.5	35.3	2.8		1.0	4.3	1.5					
Ceftriaxone	0.0						90.0	1.3	2.0	0.3		3.5	2.0	1.0				
Nalidixic Acid	7.0							6.5	63.0	23.3	0.3		0.3	6.8				
Ciprofloxacin	0.0	90.3	2.3	0.5	1.8	4.0	1.3											
Sulfisoxazole	41.3											48.5	6.3	4.0			41.3	
Trimethoprim/Sulfamethoxazole	4.3				85.5	7.0	2.5	0.5	0.3			4.3						
Amikacin	0.0							15.0	65.0	17.0	2.5	0.5						
Gentamicin	30.0					5.8	43.3	14.8	2.5	1.0	2.8	10.0	20.0					
Kanamycin	6.8										81.8	10.5	1.0			6.8		
Streptomycin*	56.8											43.3	13.0	43.8				
Chloramphenicol	1.8									3.3	34.5	58.0	2.5	0.3	1.5			
Tetracycline	48.0									51.3	0.8	0.5	3.3	44.3				

Vertical bars show the CLSI Susceptible/Resistant breakpoints for each drug where appropriate.

*Currently no CLSI breakpoints have been established for this organism/antimicrobial combination. Indicated breakpoints were established by NARMS.

†Discrepancies between %R and sums of distribution %s are due to rounding.

Unshaded areas indicate the dilution ranges of the Sensititre plate used to test the 2004 isolates.

Figure 18b. MIC Distribution among *E. coli* from Ground Turkey

<i>E. coli</i> from Ground Turkey (N=376)		Distribution (%) of MICs (in µg/ml)																
Antimicrobial Agent	%R	0.015	0.03	0.06	0.125	0.25	0.5	1	2	4	8	16	32	64	128	256	512	>512
Ampicillin	33.2							6.4	33.2	26.9		0.3	0.8	32.4				
Amoxicillin/Clavulanic Acid	5.3							1.3	19.9	41.8	28.2	3.5	4.5	0.8				
Cefoxitin	4.5							0.8	22.1	55.9	16.0	0.8	2.7	1.9				
Ceftiofur	1.1				1.9	47.9	45.2	2.4	1.3	0.3	0.5							
Ceftriaxone	0.0					95.5	1.3	1.9			0.8	0.3	0.3					
Nalidixic Acid	10.6							3.7	62.0	21.5	1.6	0.5	0.5	10.1				
Ciprofloxacin	0.8	84.3	3.5	0.8	2.9	7.4	0.3				0.8							
Sulfisoxazole	48.4										44.4	3.2	4.0			48.4		
Trimethoprim/Sulfamethoxazole	3.7				83.8	9.3	2.7	0.5			3.7							
Amikacin	0.0							17.3	66.5	13.8	2.4							
Gentamicin	29.3						4.8	42.6	19.1	2.1	2.1	12.5	16.8					
Kanamycin	16.0										75.0	6.9	2.1	0.3	15.7			
Streptomycin*	49.2										50.8	18.6	30.6					
Chloramphenicol	0.8							1.3	36.7	60.4	0.8			0.8				
Tetracycline	74.2								25.3	0.5		6.9	67.3					

Vertical bars show the CLSI/ Susceptible/Resistant breakpoints for each drug where appropriate.

*Currently no CLSI breakpoints have been established for this organism/antimicrobial combination. Indicated breakpoints were established by NARMS.

†Discrepancies between %R and sums of distribution %s are due to rounding.

Unshaded areas indicate the dilution ranges of the Sensititre plate used to test the 2004 isolates.

Figure 18c. MIC Distribution among *E. coli* from Ground Beef

<i>E. coli</i> from Ground Beef (N=338)		Distribution (%) of MICs (in µg/ml)																
Antimicrobial Agent	%R	0.015	0.03	0.06	0.125	0.25	0.5	1	2	4	8	16	32	64	128	256	512	>512
Ampicillin	5.3							8.9	46.2	37.9	0.9	0.9	0.3	5.0				
Amoxicillin/Clavulanic Acid	3.8							4.4	23.4	60.9	7.1	0.3	3.6	0.3				
Cefoxitin	1.2							4.1	30.2	53.8	8.9	1.8	0.3	0.9				
Ceftiofur	0.9					5.0	49.4	41.7	2.1	0.3	0.6		0.9					
Ceftriaxone	0.0						95.9	1.8	0.6	0.3		0.3	0.6	0.6				
Nalidixic Acid	1.5							3.0	67.5	26.9	1.2		0.9	0.6				
Ciprofloxacin	0.0	94.4	3.8		0.6	0.9	0.3											
Sulfisoxazole	13.0										84.6		2.4		13.0			
Trimethoprim/Sulfamethoxazole	0.6				97.0	2.1		0.3				0.6						
Amikacin	0.0							15.7	69.8	12.4	1.8	0.3						
Gentamicin	0.6						9.2	67.8	20.7	1.8			0.6					
Kanamycin	2.4										95.6	2.1			2.4			
Streptomycin*	11.8										88.2		4.7	7.1				
Chloramphenicol	3.6							0.3	26.9	68.3	0.9	0.3	3.3					
Tetracycline	22.8								70.7	6.5	2.7	1.2	18.9					

Vertical bars show the CLSI Susceptible/Resistant breakpoints for each drug where appropriate.

*Currently no CLSI/ breakpoints have been established for this organism/antimicrobial combination. Indicated breakpoints were established by NARMS.

†Discrepancies between %R and sums of distribution %s are due to rounding.

Unshaded areas indicate the dilution ranges of the Sensititre plate used to test the 2004 isolates.

Figure 18d. MIC Distribution among *E. coli* from Pork Chop

<i>E. coli</i> from Pork Chop (N=232)		Distribution (%) of MICs (in µg/ml)																
Antimicrobial Agent	%R	0.015	0.03	0.06	0.125	0.25	0.5	1	2	4	8	16	32	64	128	256	512	>512
Ampicillin	15.1							12.9	44.4	25.0	1.7	0.9	0.9	14.2				
Amoxicillin/Clavulanic Acid	5.6							4.3	27.6	46.6	15.5	0.4	4.7	0.9				
Cefoxitin	2.2						0.9	2.6	26.7	59.9	7.3	0.4	1.3	0.9				
Ceftiofur	0.4					7.3	51.7	39.7	0.9		0.4							
Ceftriaxone	0.0						97.0	1.7	0.9		0.4							
Nalidixic Acid	0.0							9.9	68.5	19.4	1.3	0.9						
Ciprofloxacin	0.0	97.8	0.9	0.4	0.4	0.4						69.8	3.0	6.9	0.4	0.4	19.4	
Sulfisoxazole	19.4																	
Trimethoprim/Sulfamethoxazole	3.9				93.1	2.2	0.9					3.9						
Amikacin	0.0						0.4	15.5	56.0	26.3	1.3	0.4						
Gentamicin	1.3						10.3	57.8	26.7	3.4		0.4		1.3				
Kanamycin	8.2									89.2	2.6					8.2		
Streptomycin*	21.1											78.9	8.6	12.5				
Chloramphenicol	4.3							0.9	34.1	59.9	0.9	1.3	3.0					
Tetracycline	56.0								41.8	2.2		6.0	50.0					

Vertical bars show the CLSI Susceptible/Resistant breakpoints for each drug where appropriate.

*Currently no CLSI breakpoints have been established for this organism/antimicrobial combination. Indicated breakpoints were established by NARMS.

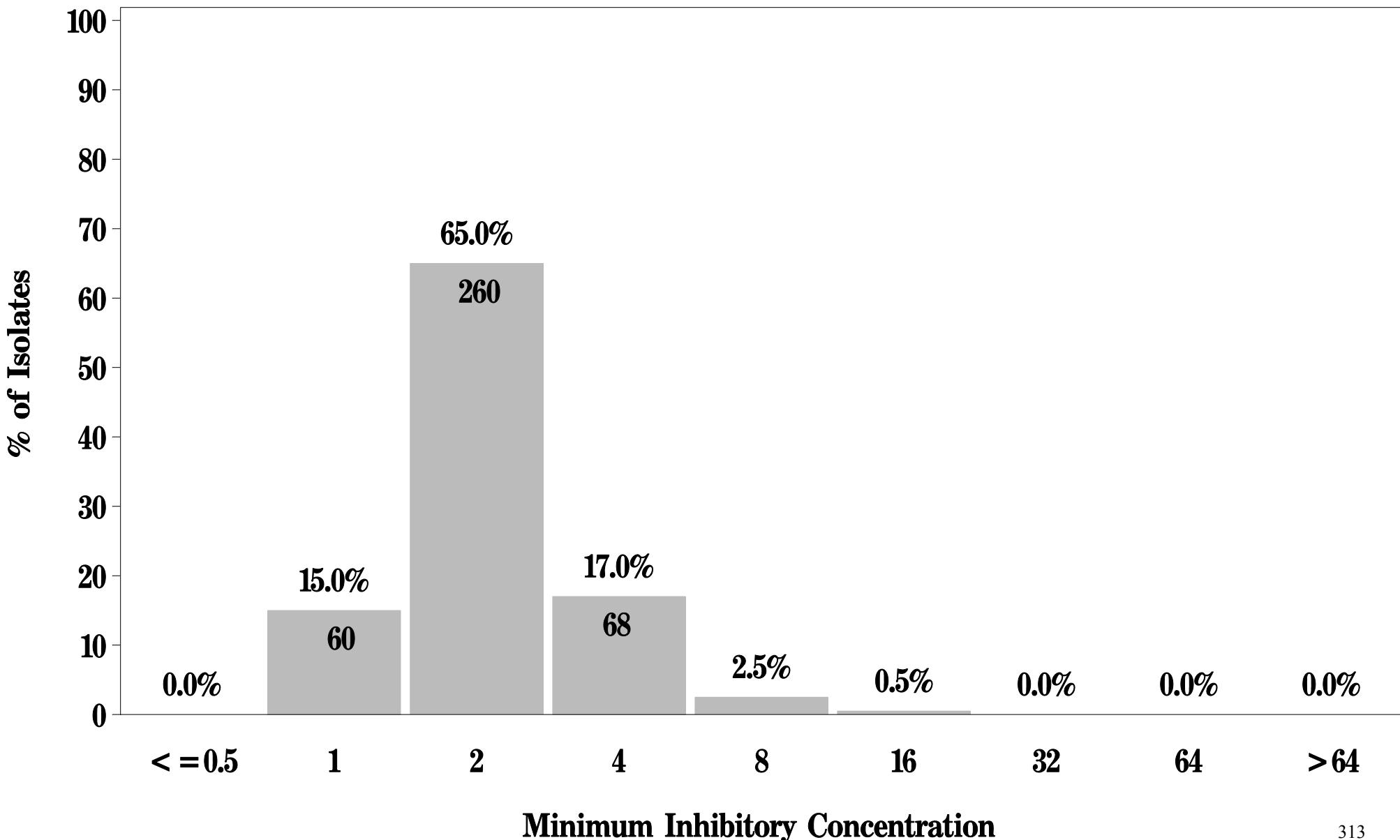
†Discrepancies between %R and sums of distribution %s are due to rounding.

Unshaded areas indicate the dilution ranges of the Sensititre plate used to test the 2004 isolates.

NARMS

**Figure 19a: Minimum Inhibitory Concentration of Amikacin
for *Escherichia coli* in Chicken Breast (N=400 Isolates)**

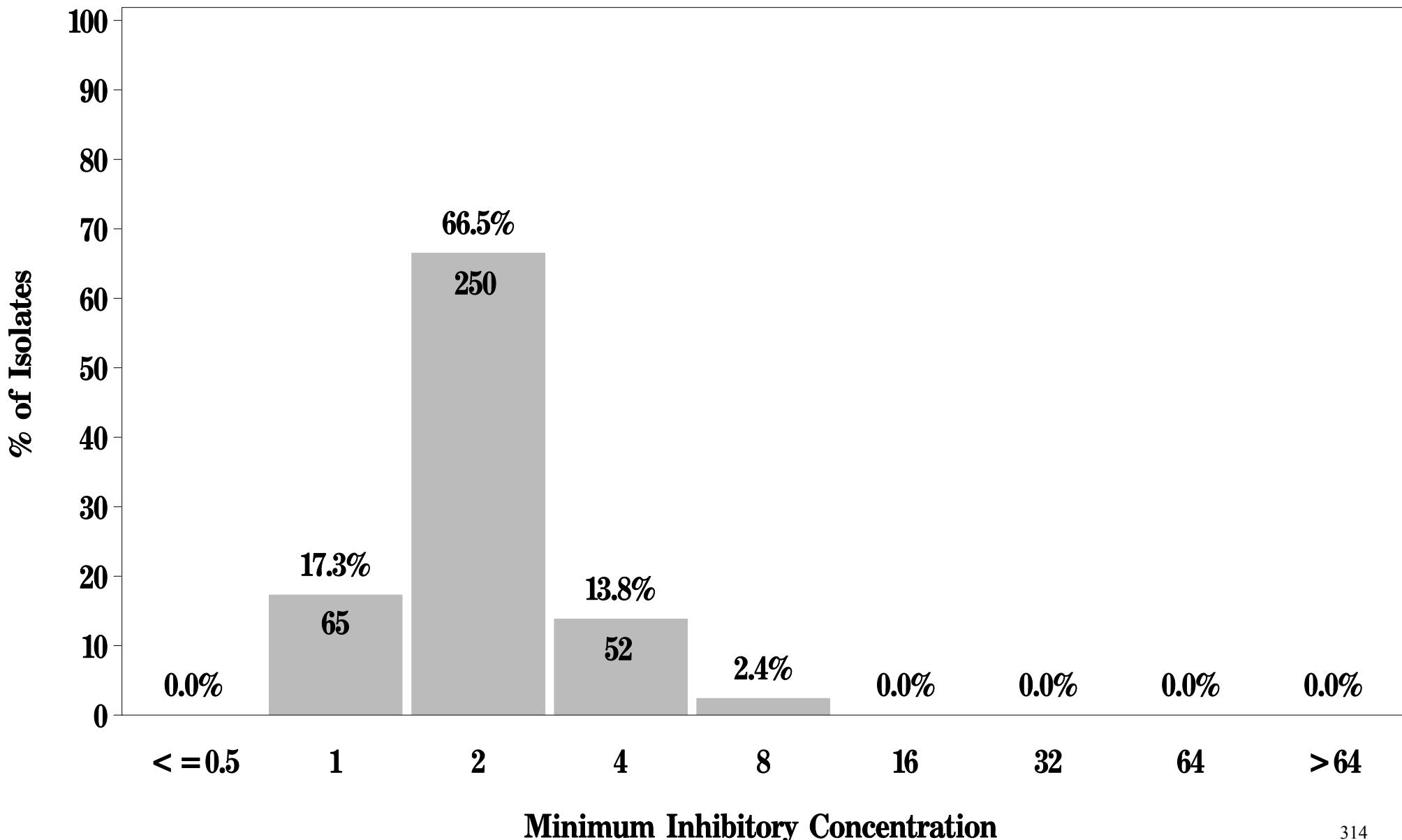
Breakpoints: Susceptible $\leq 16 \mu\text{g/mL}$ Resistant $\geq 64 \mu\text{g/mL}$



NARMS

**Figure 19a: Minimum Inhibitory Concentration of Amikacin
for *Escherichia coli* in Ground Turkey (N=376 Isolates)**

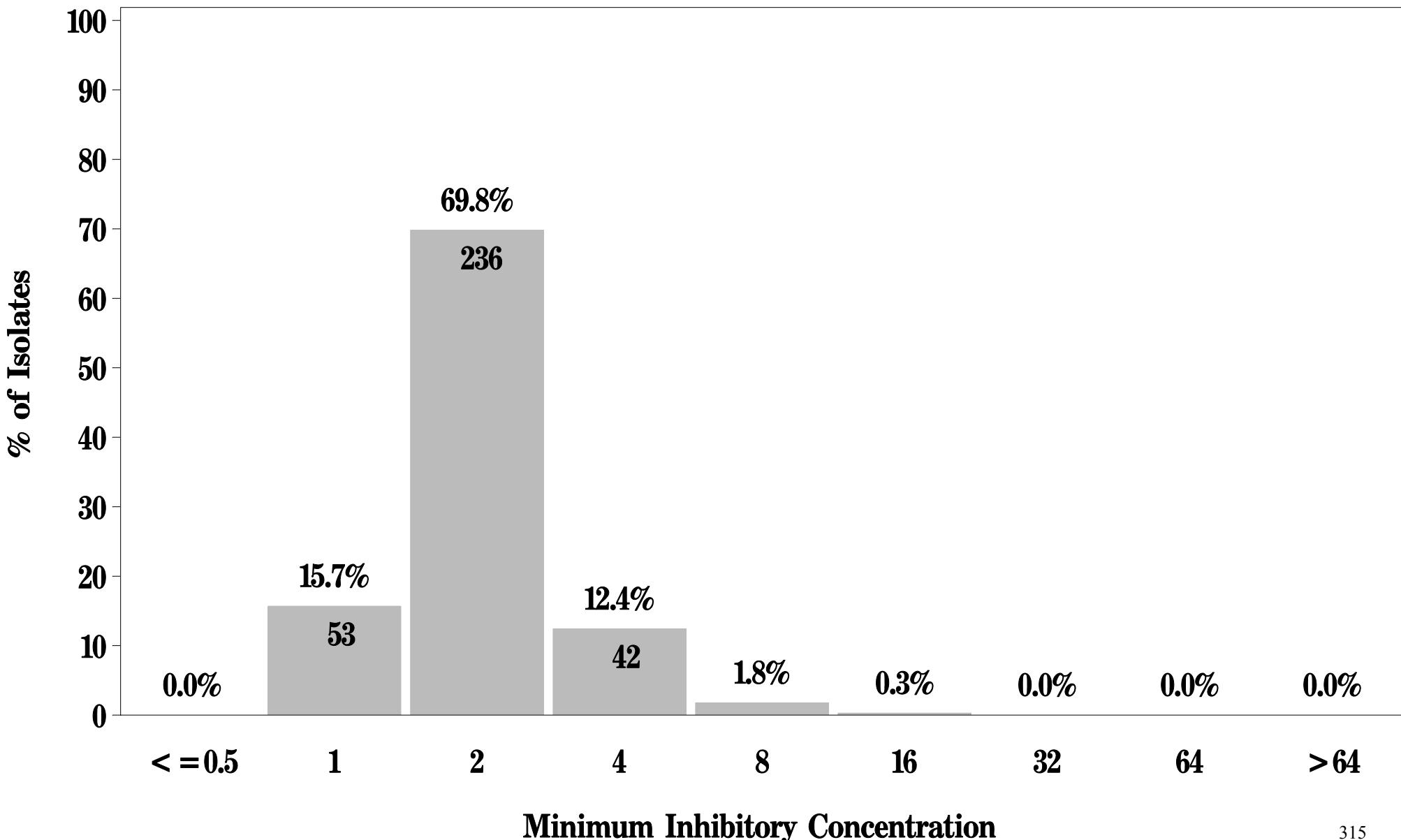
Breakpoints: Susceptible $\leq 16 \mu\text{g/mL}$ Resistant $\geq 64 \mu\text{g/mL}$



NARMS

**Figure 19a: Minimum Inhibitory Concentration of Amikacin
for *Escherichia coli* in Ground Beef (N=338 Isolates)**

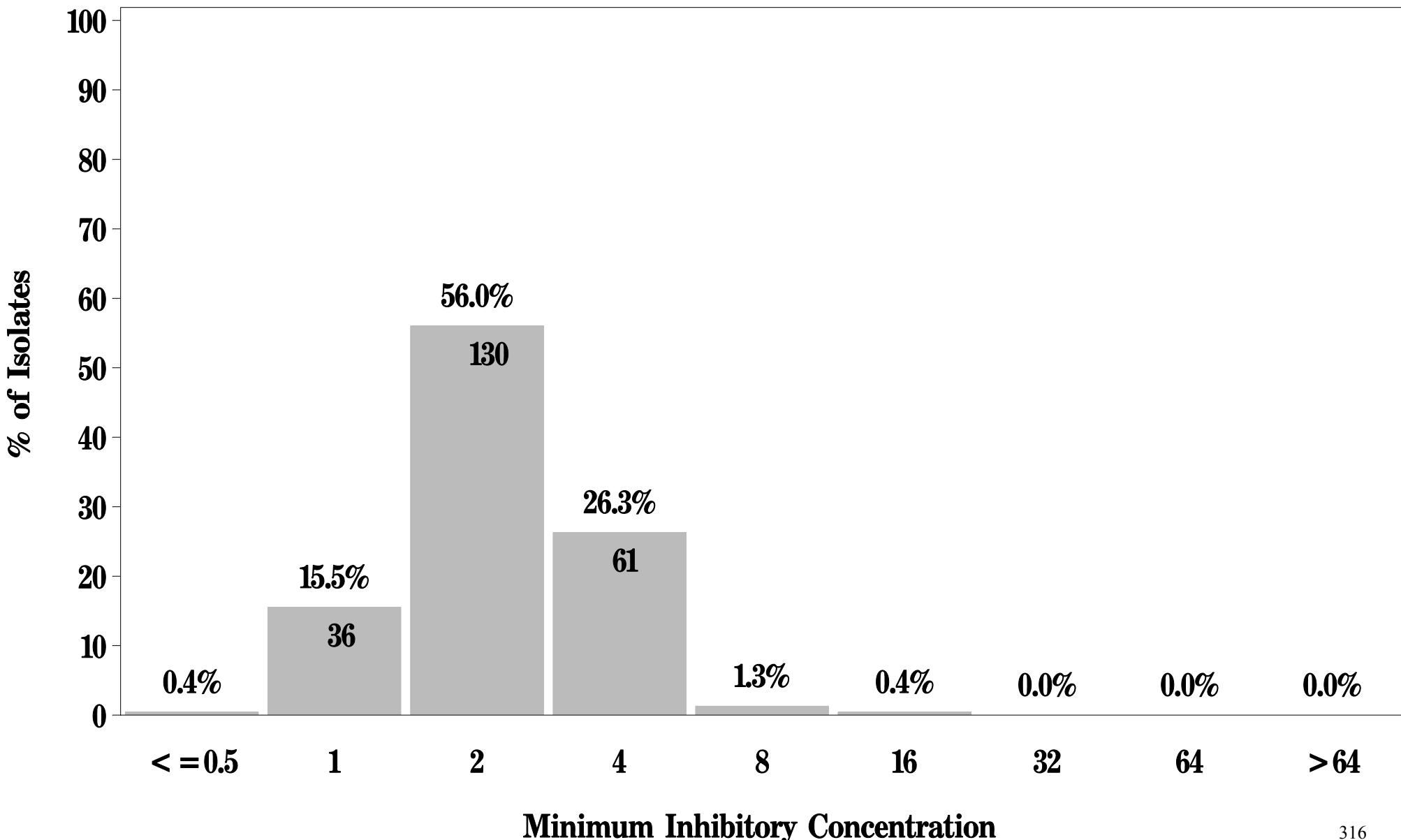
Breakpoints: Susceptible $\leq 16 \mu\text{g/mL}$ Resistant $\geq 64 \mu\text{g/mL}$



NARMS

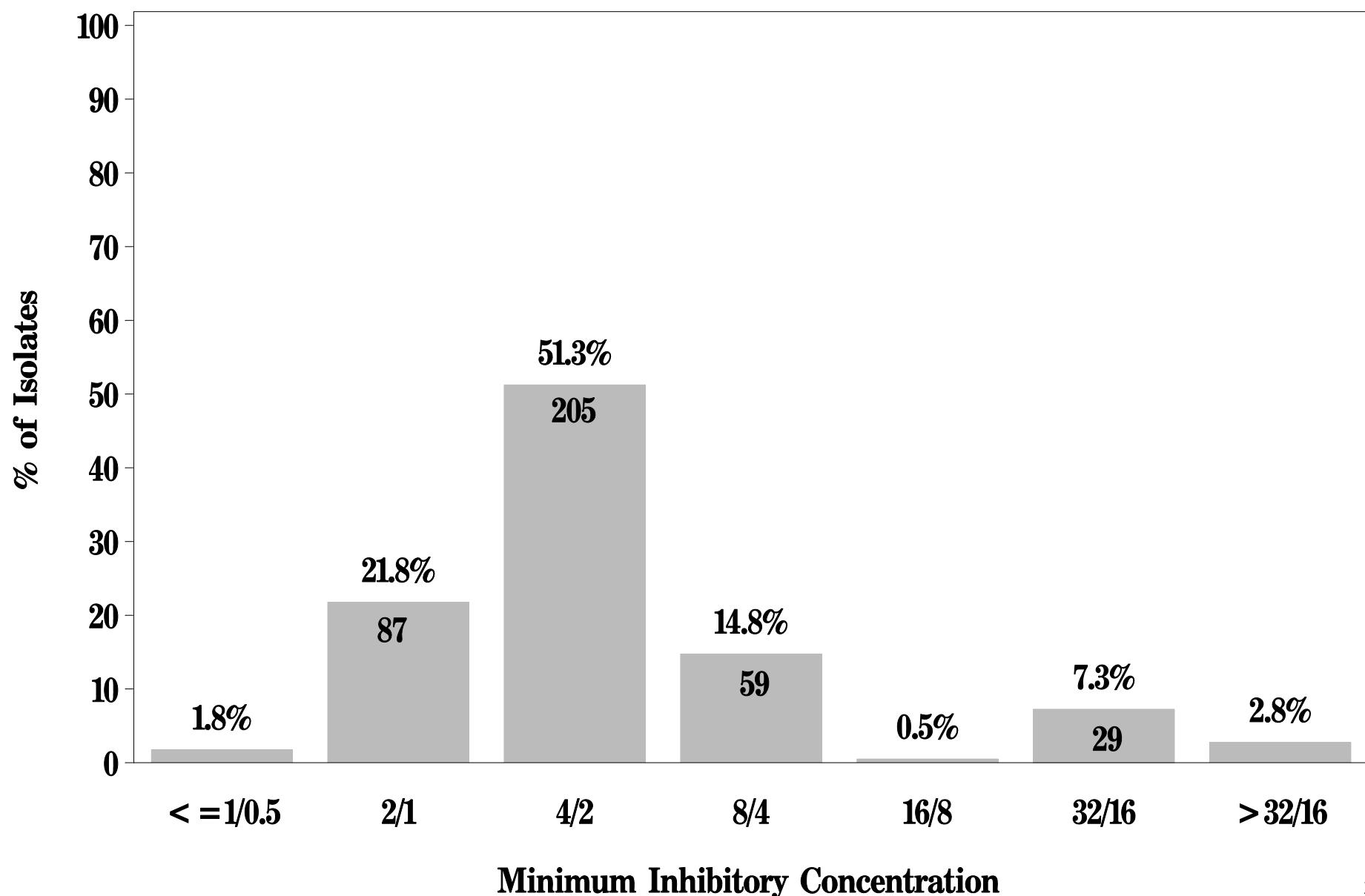
**Figure 19a: Minimum Inhibitory Concentration of Amikacin
for *Escherichia coli* in Pork Chop (N=232 Isolates)**

Breakpoints: Susceptible $\leq 16 \mu\text{g/mL}$ Resistant $\geq 64 \mu\text{g/mL}$



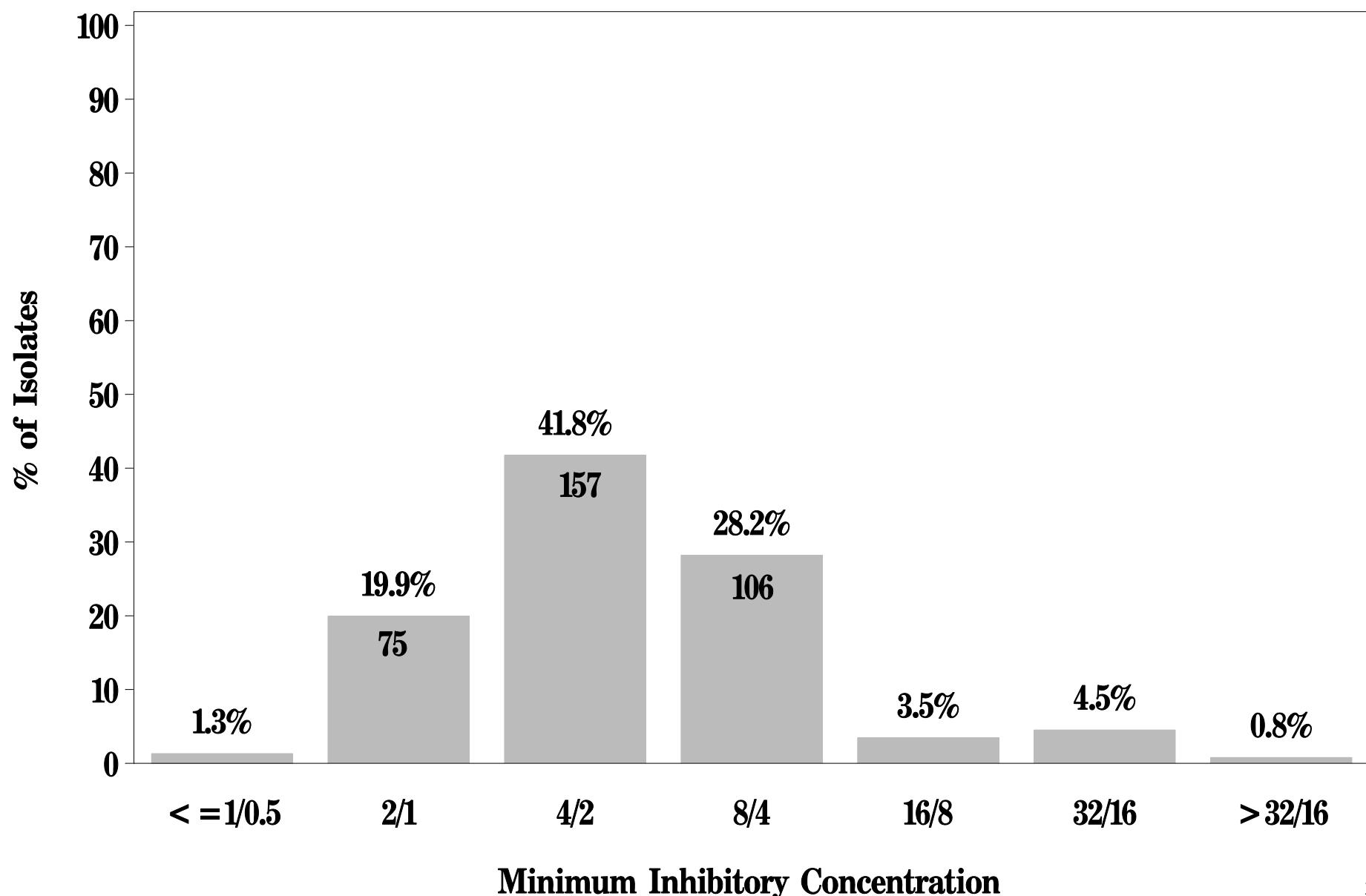
NARMS

Figure 19b: Minimum Inhibitory Concentration of Amoxicillin/Clavulanic acid for *Escherichia coli* in Chicken Breast (N=400 Isolates)
Breakpoints: Susceptible $\leq 8 \text{ } \mu\text{g/mL}$ Resistant $\geq 32 \text{ } \mu\text{g/mL}$



NARMS

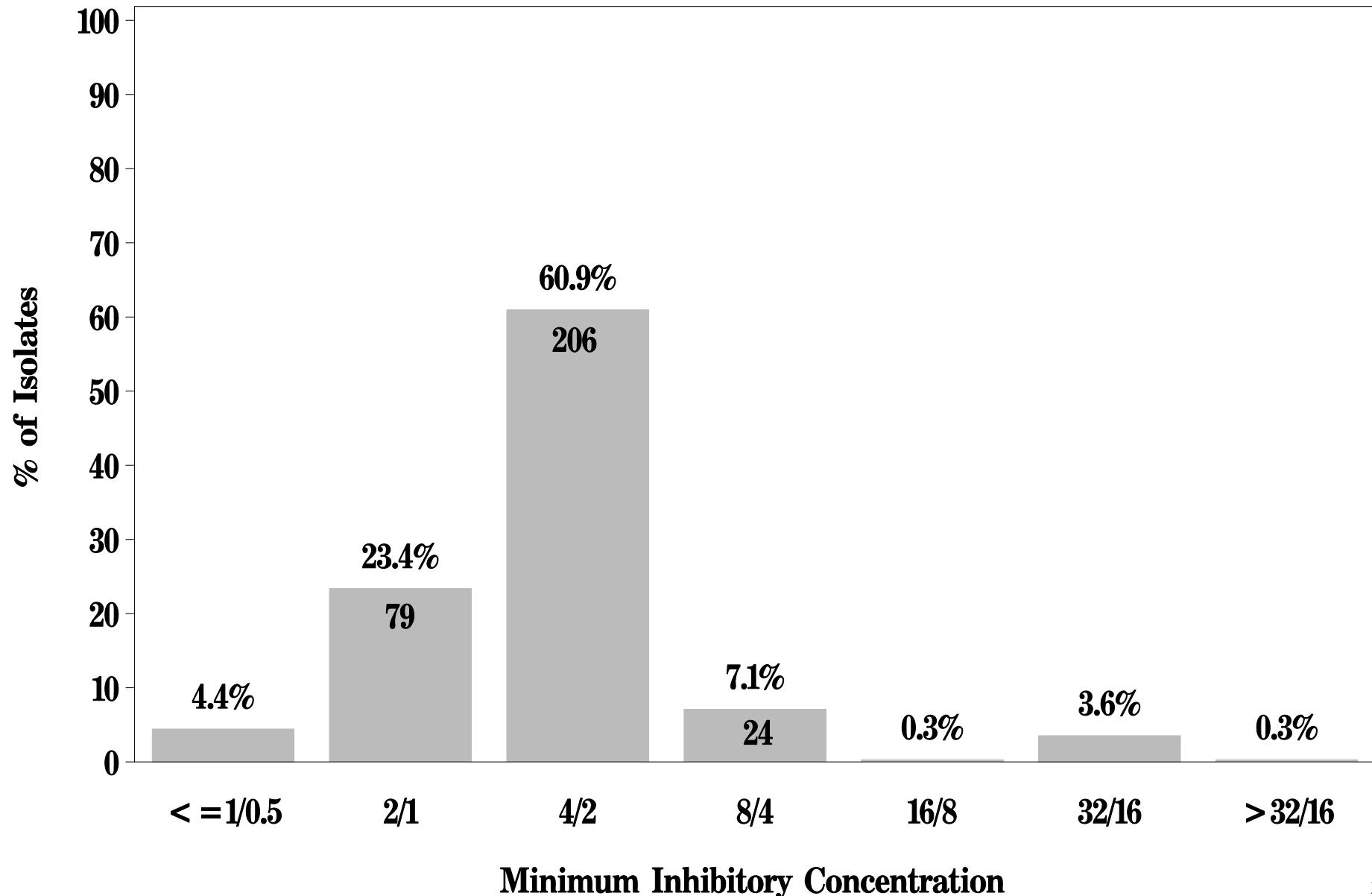
Figure 19b: Minimum Inhibitory Concentration of Amoxicillin/Clavulanic acid for *Escherichia coli* in Ground Turkey (N=376 Isolates)
Breakpoints: Susceptible $\leq 8 \text{ } \mu\text{g/mL}$ Resistant $\geq 32 \text{ } \mu\text{g/mL}$



NARMS

Figure 19b: Minimum Inhibitory Concentration of Amoxicillin/Clavulanic acid for *Escherichia coli* in Ground Beef (N=338 Isolates)

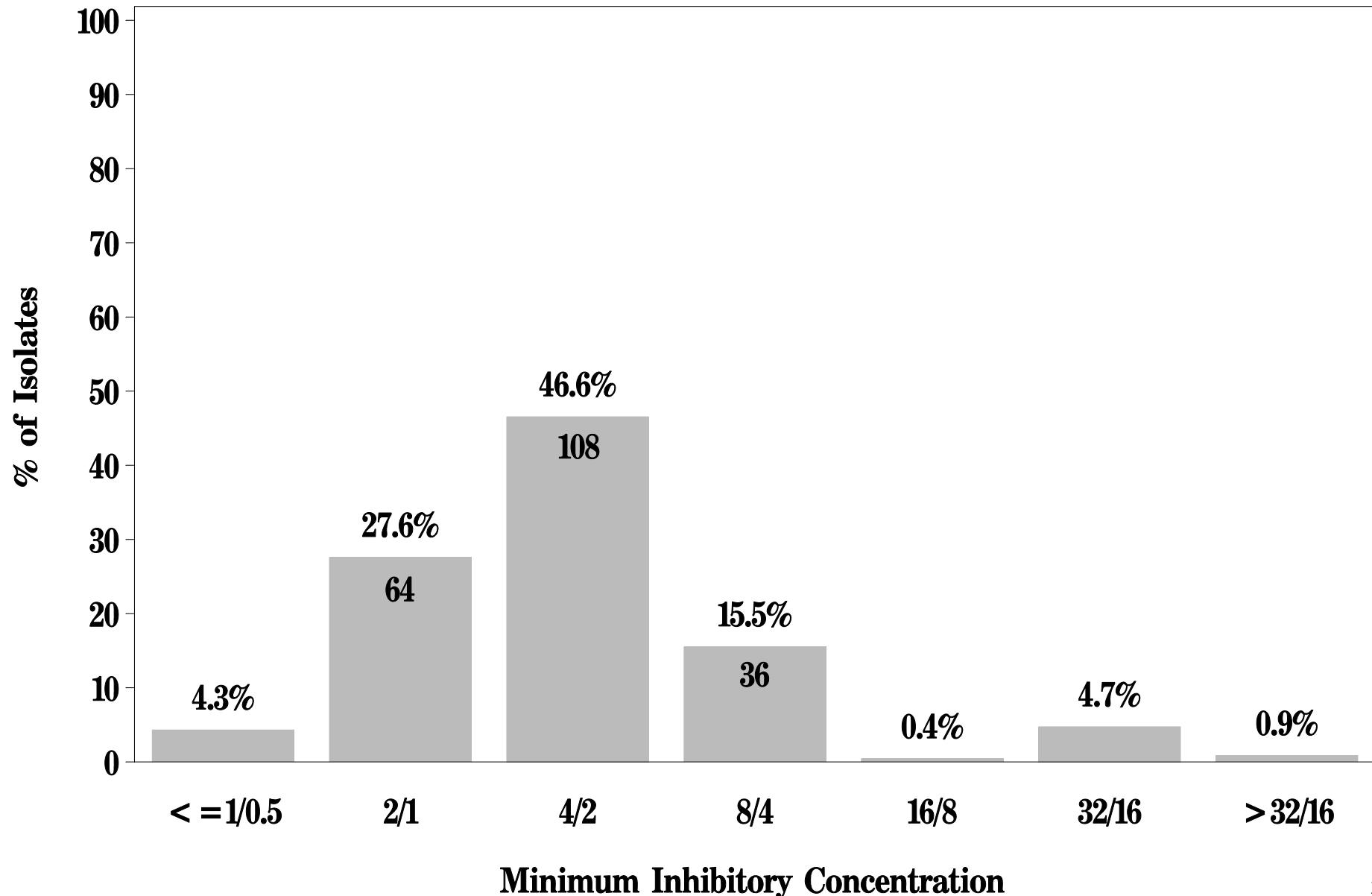
Breakpoints: Susceptible $\leq 8 \mu\text{g/mL}$ Resistant $\geq 32 \mu\text{g/mL}$



NARMS

**Figure 19b: Minimum Inhibitory Concentration of Amoxicillin/Clavulanic acid
for *Escherichia coli* in Pork Chop (N=232 Isolates)**

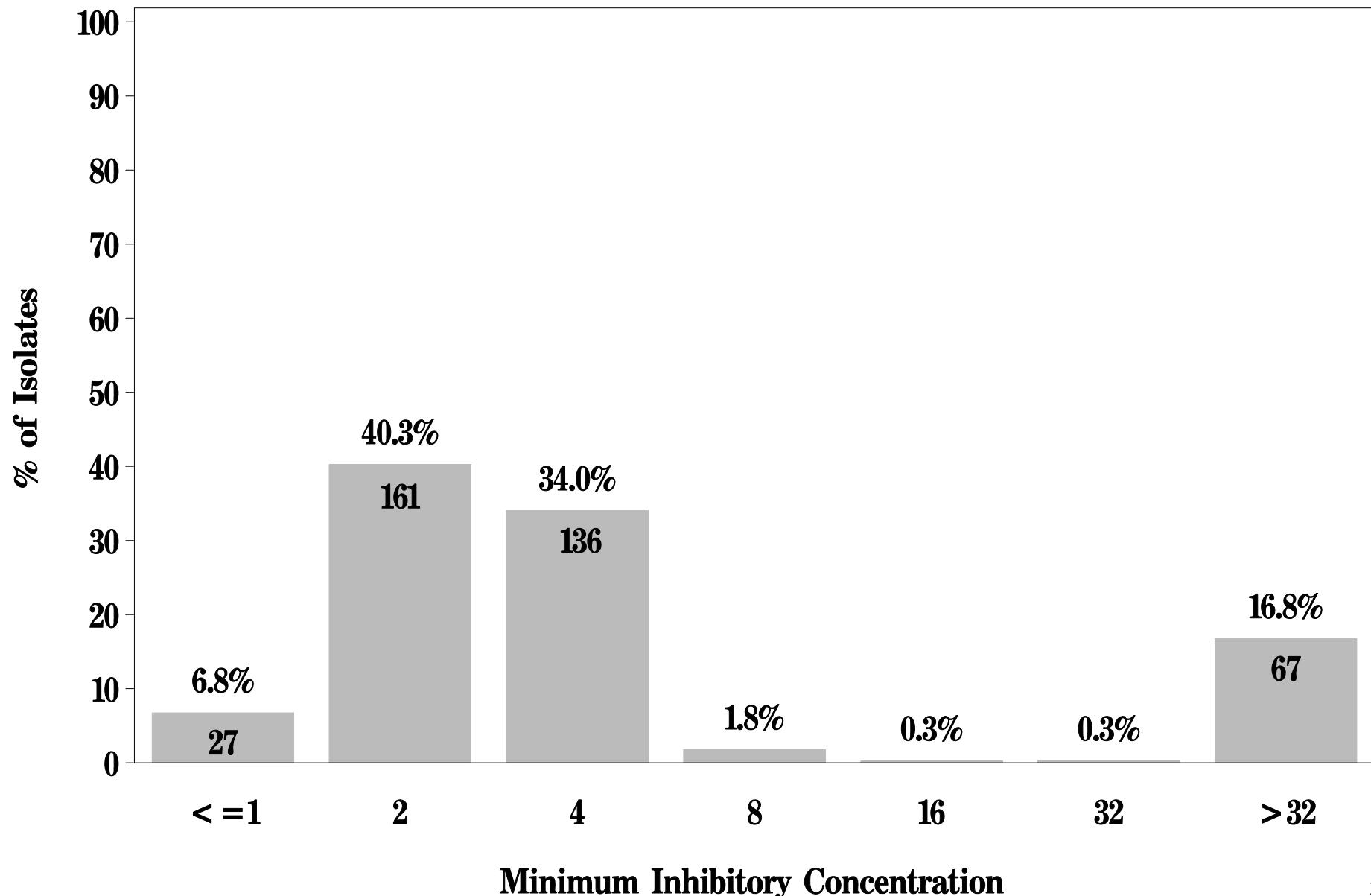
Breakpoints: Susceptible $\leq 8 \mu\text{g/mL}$ Resistant $\geq 32 \mu\text{g/mL}$



NARMS

**Figure 19c: Minimum Inhibitory Concentration of Ampicillin
for *Escherichia coli* in Chicken Breast (N=400 Isolates)**

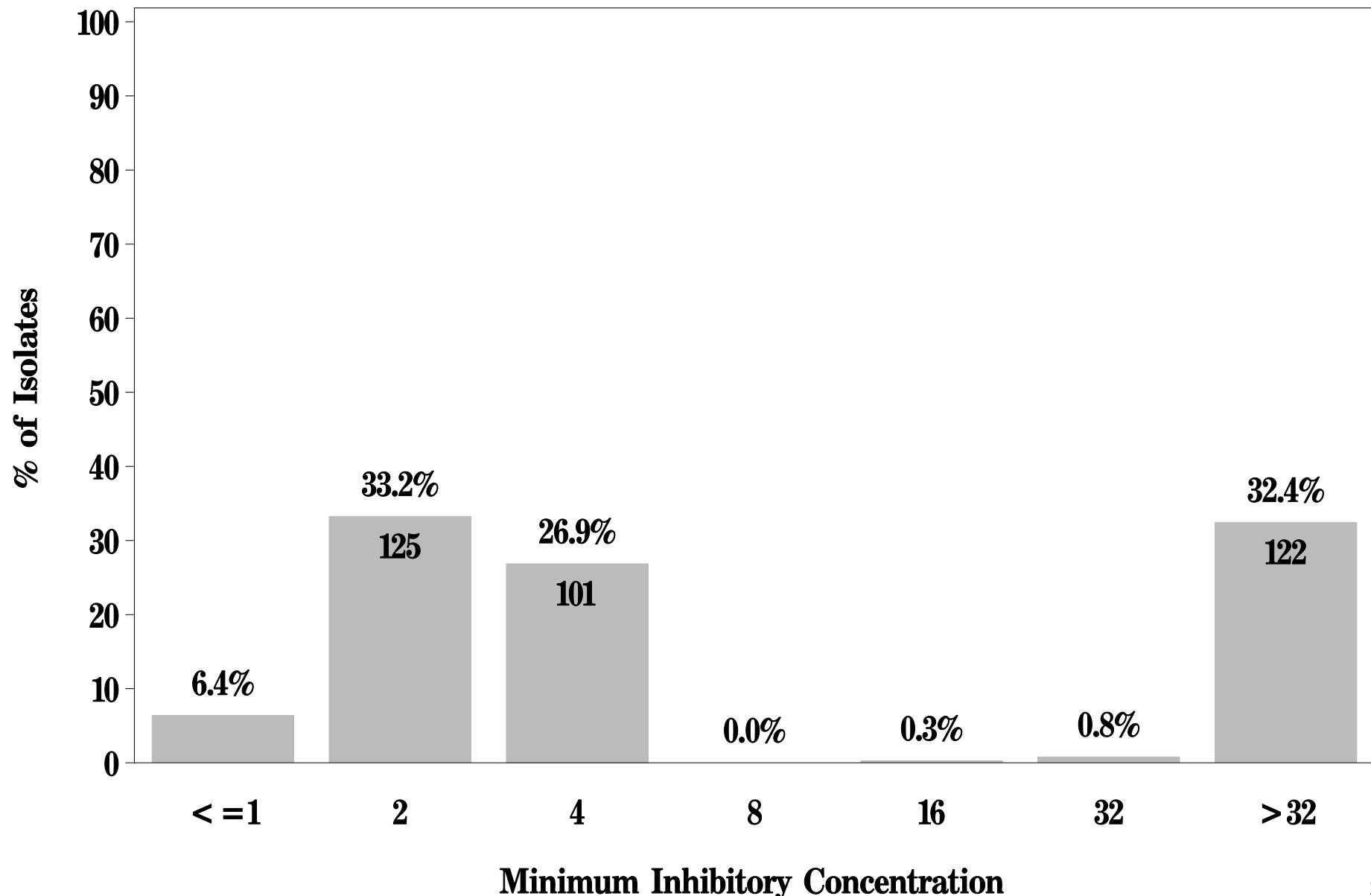
Breakpoints: Susceptible $\leq 8 \mu\text{g/mL}$ Resistant $\geq 32 \mu\text{g/mL}$



NARMS

**Figure 19c: Minimum Inhibitory Concentration of Ampicillin
for *Escherichia coli* in Ground Turkey (N=376 Isolates)**

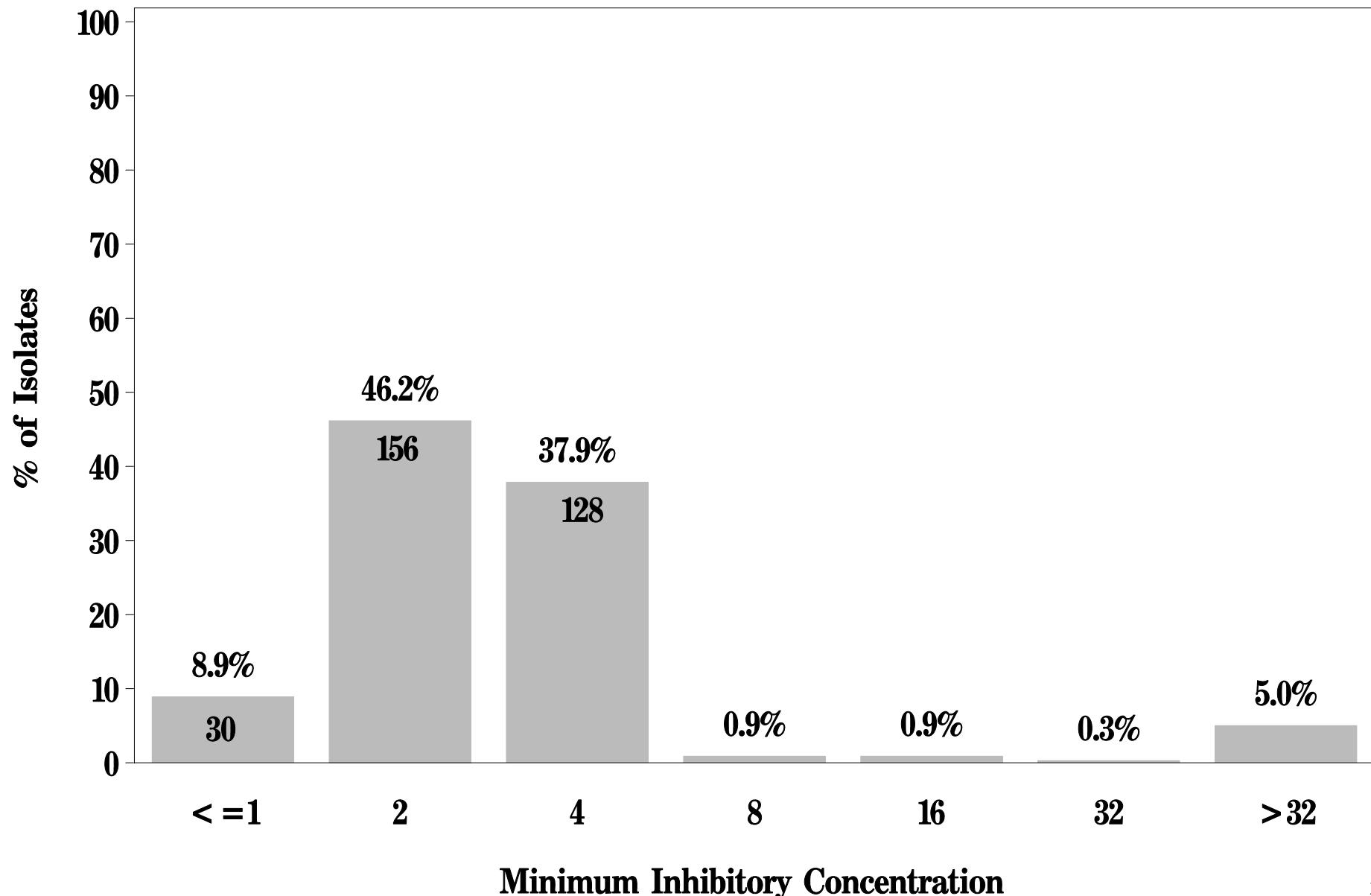
Breakpoints: Susceptible $\leq 8 \mu\text{g/mL}$ Resistant $\geq 32 \mu\text{g/mL}$



NARMS

**Figure 19c: Minimum Inhibitory Concentration of Ampicillin
for *Escherichia coli* in Ground Beef (N=338 Isolates)**

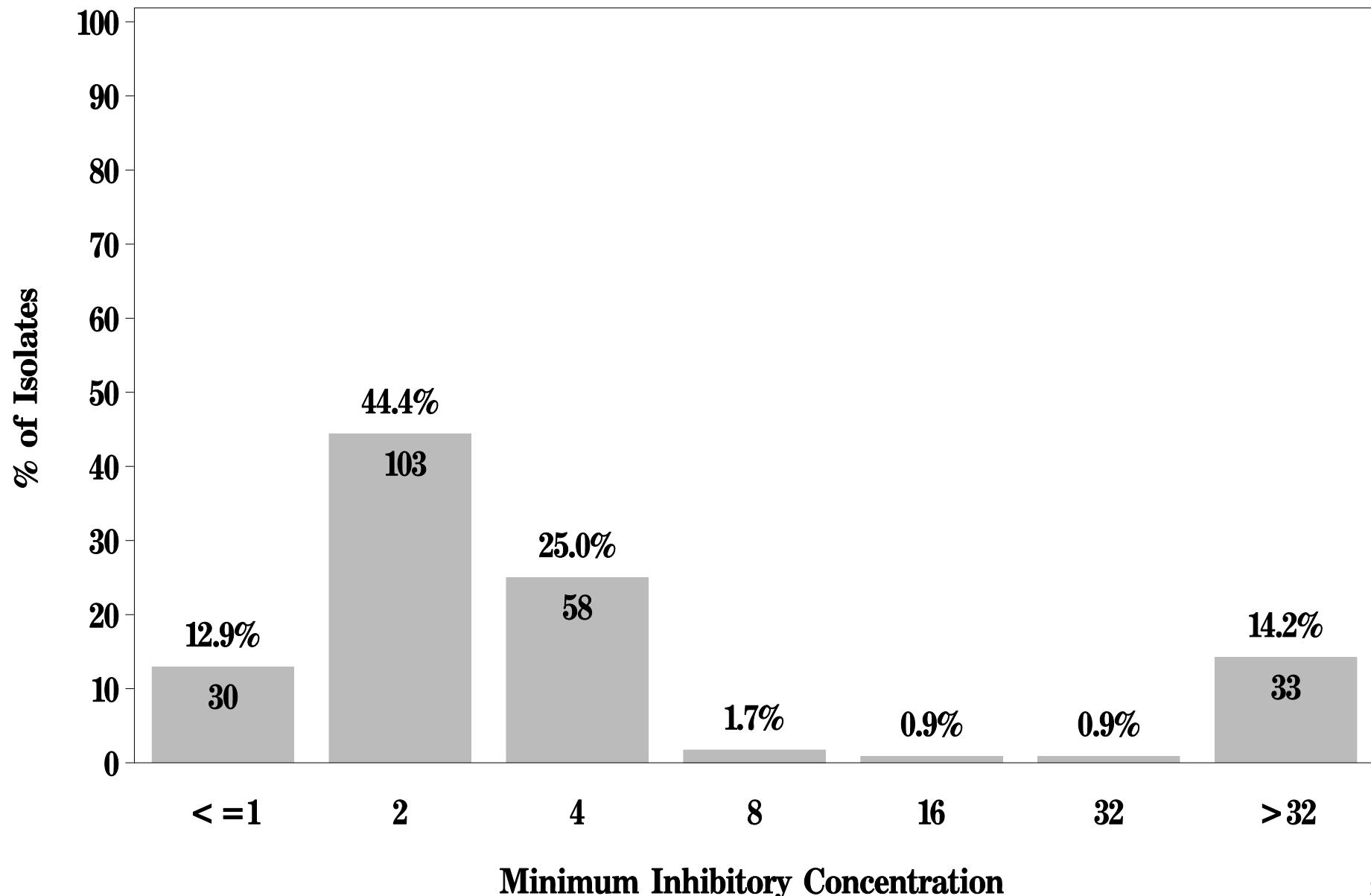
Breakpoints: Susceptible $\leq 8 \mu\text{g/mL}$ Resistant $\geq 32 \mu\text{g/mL}$



NARMS

**Figure 19c: Minimum Inhibitory Concentration of Ampicillin
for *Escherichia coli* in Pork Chop (N=232 Isolates)**

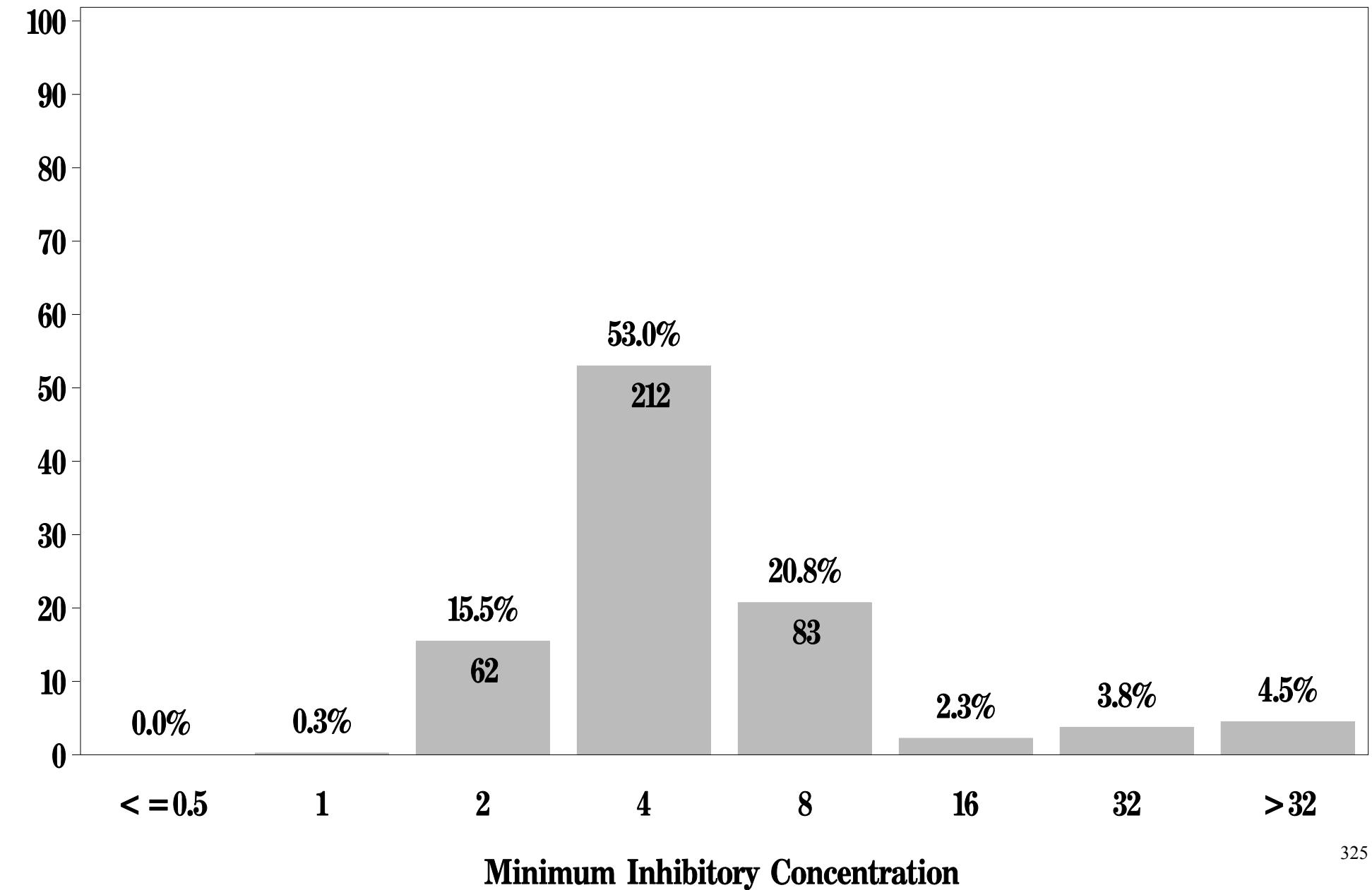
Breakpoints: Susceptible $\leq 8 \mu\text{g/mL}$ Resistant $\geq 32 \mu\text{g/mL}$



NARMS

Figure 19d: Minimum Inhibitory Concentration of Cefoxitin
for *Escherichia* in Chicken Breast (N=400 Isolates)

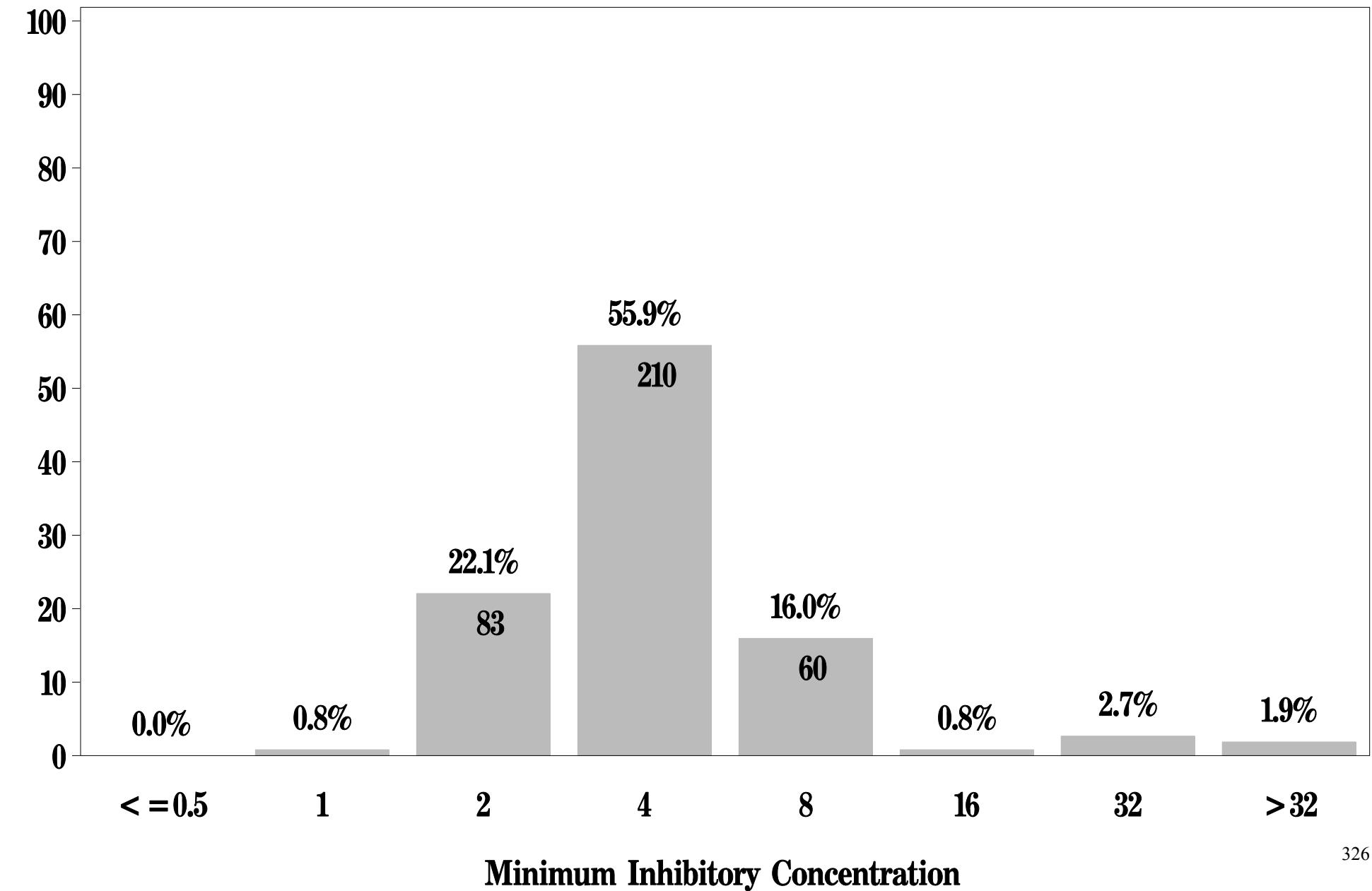
Breakpoints: Susceptible $\leq 8 \mu\text{g/mL}$ Resistant $\geq 32 \mu\text{g/mL}$



NARMS

Figure 19d: Minimum Inhibitory Concentration of Cefoxitin
for *Escherichia* in Ground Turkey (N=376 Isolates)

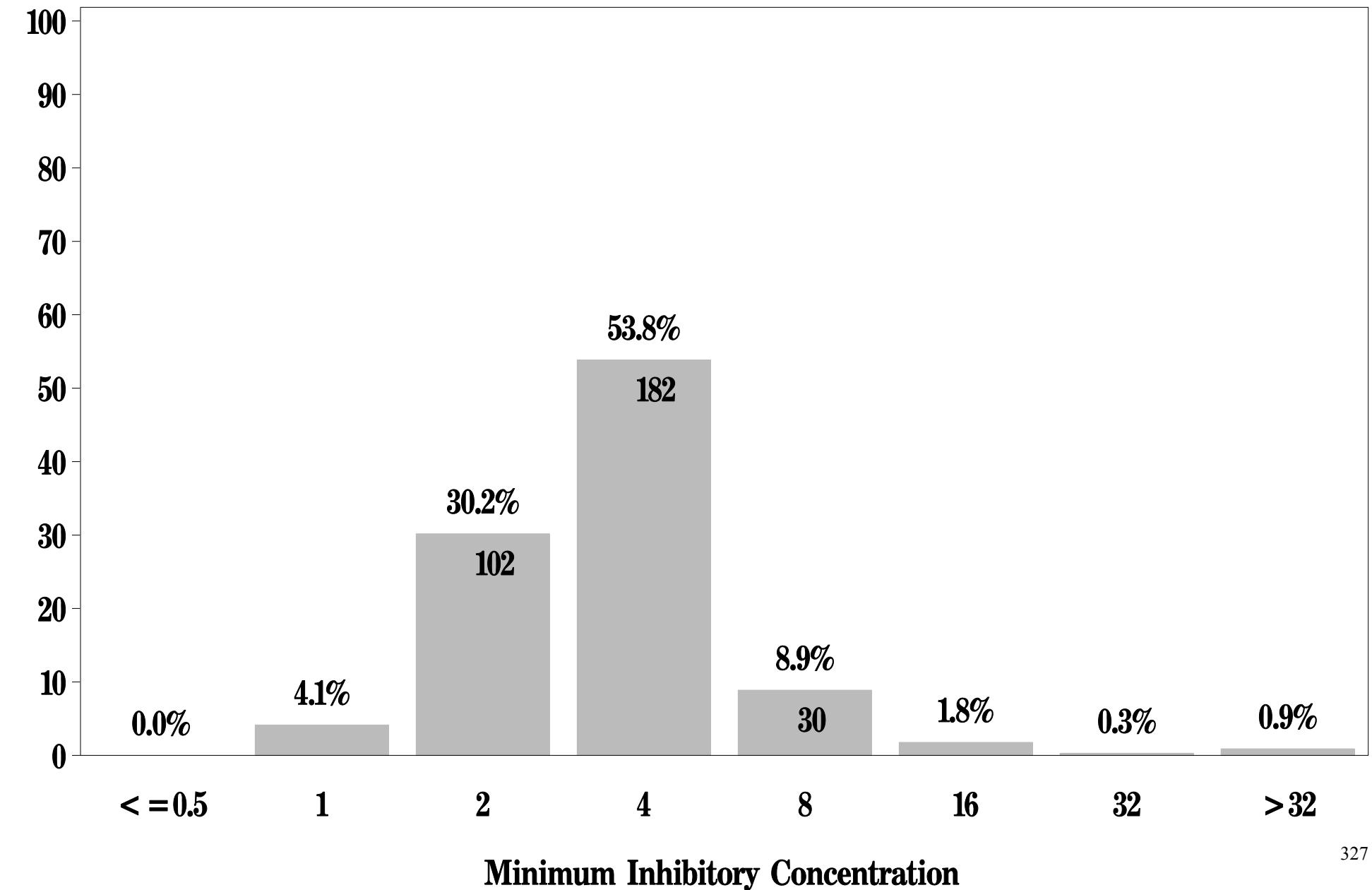
Breakpoints: Susceptible $\leq 8 \mu\text{g/mL}$ Resistant $\geq 32 \mu\text{g/mL}$



NARMS

Figure 19d: Minimum Inhibitory Concentration of Cefoxitin
for *Escherichia* in Ground Beef (N=338 Isolates)

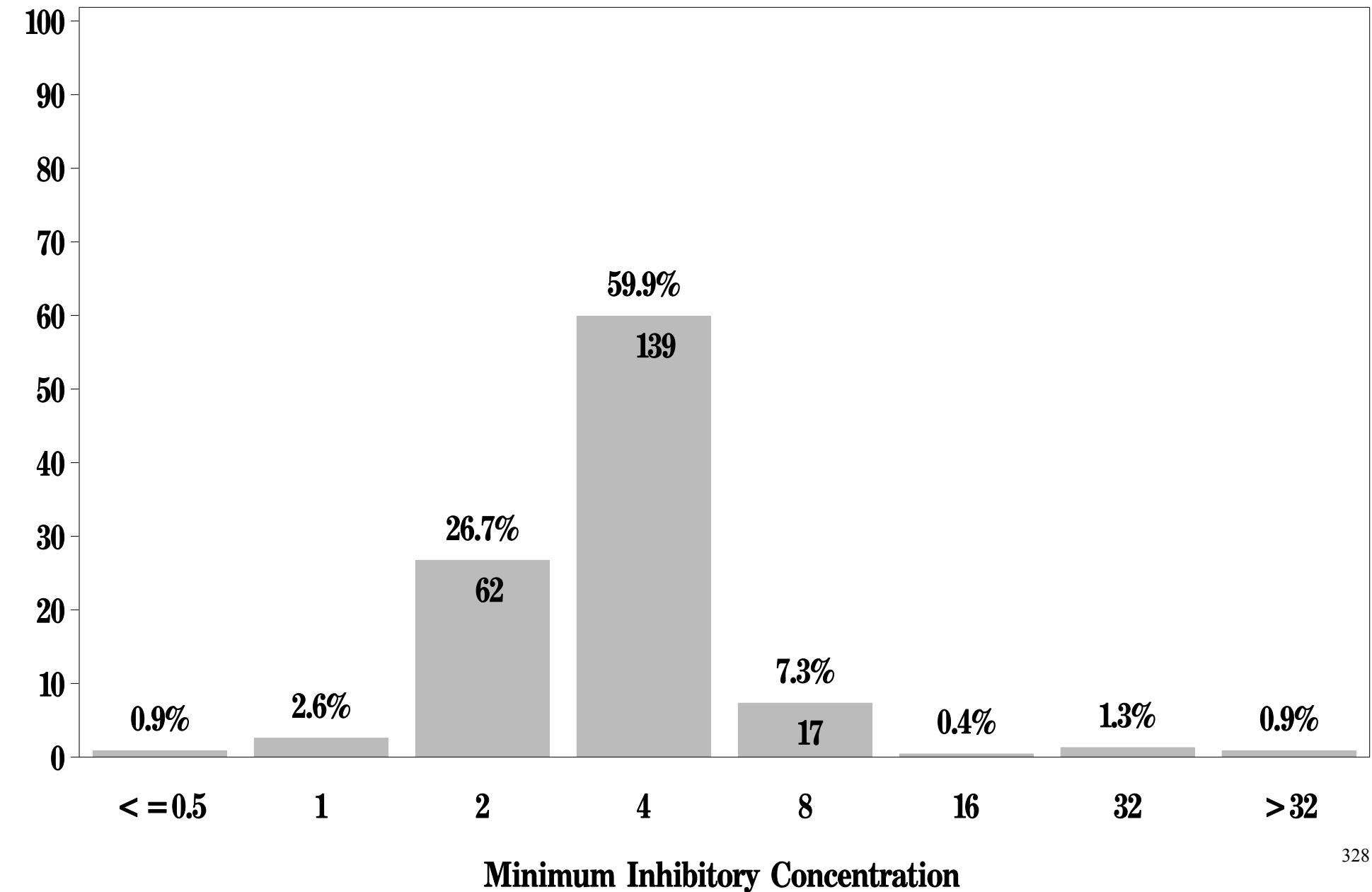
Breakpoints: Susceptible $\leq 8 \mu\text{g/mL}$ Resistant $\geq 32 \mu\text{g/mL}$



NARMS

Figure 19d: Minimum Inhibitory Concentration of Cefoxitin
for *Escherichia* in Pork Chop (N=232 Isolates)

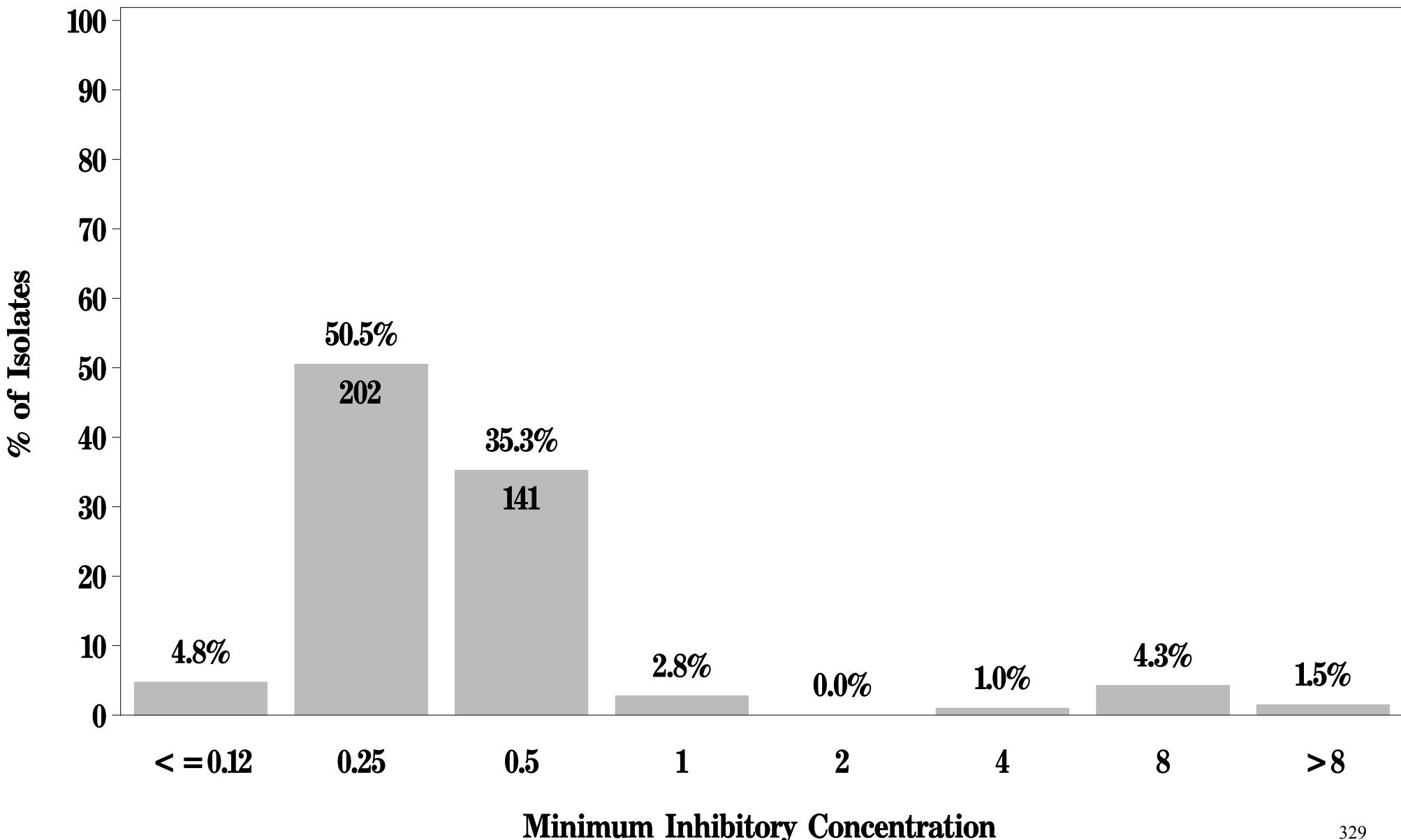
Breakpoints: Susceptible $\leq 8 \mu\text{g/mL}$ Resistant $\geq 32 \mu\text{g/mL}$



NARMS

Figure 19e: Minimum Inhibitory Concentration of Ceftiofur for *Escherichia coli* in Chicken Breast (N=400 Isolates)

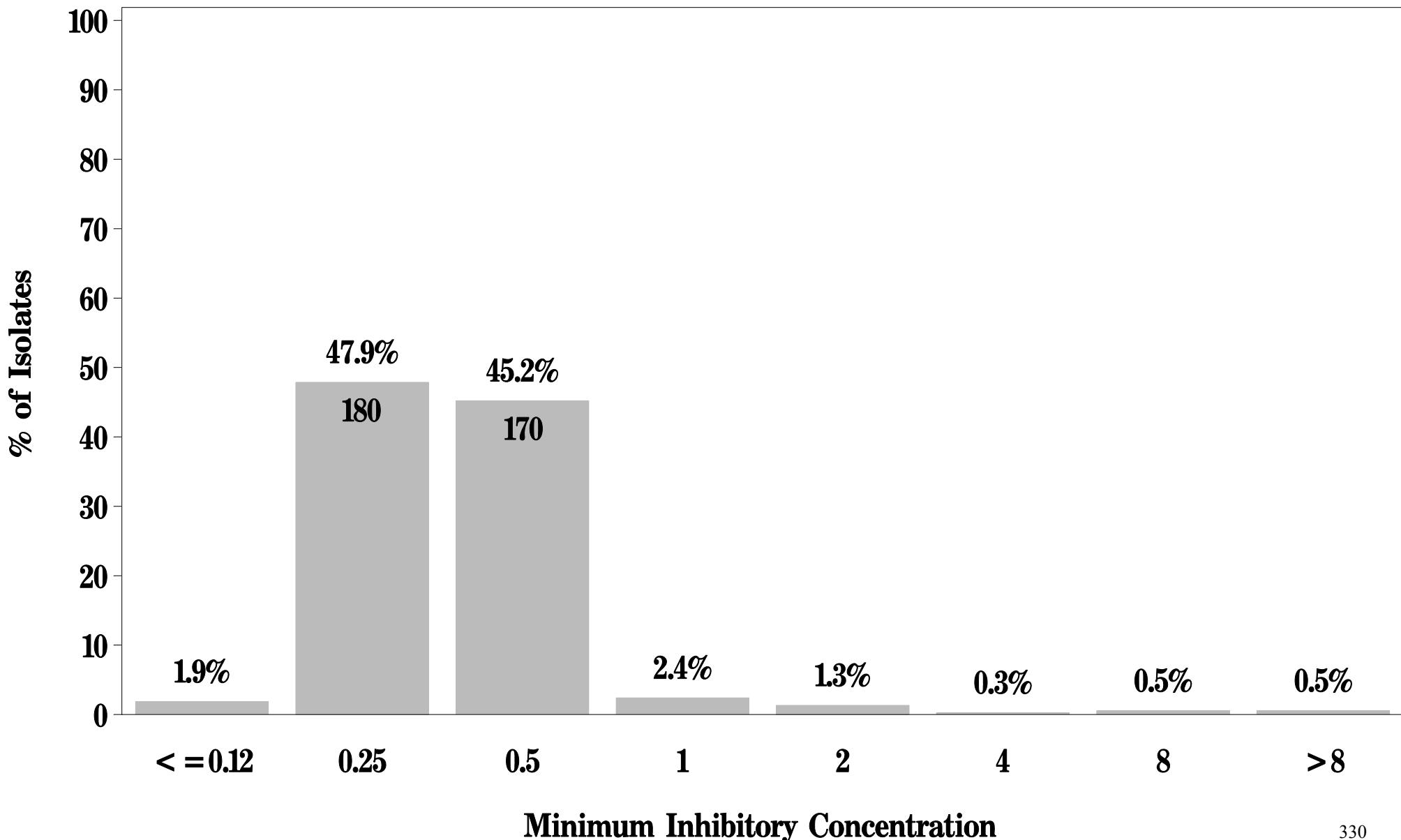
Breakpoints: Susceptible $\leq 2 \mu\text{g/mL}$ Resistant $\geq 8 \mu\text{g/mL}$



NARMS

Figure 19e: Minimum Inhibitory Concentration of Ceftiofur for *Escherichia coli* in Ground Turkey (N=376 Isolates)

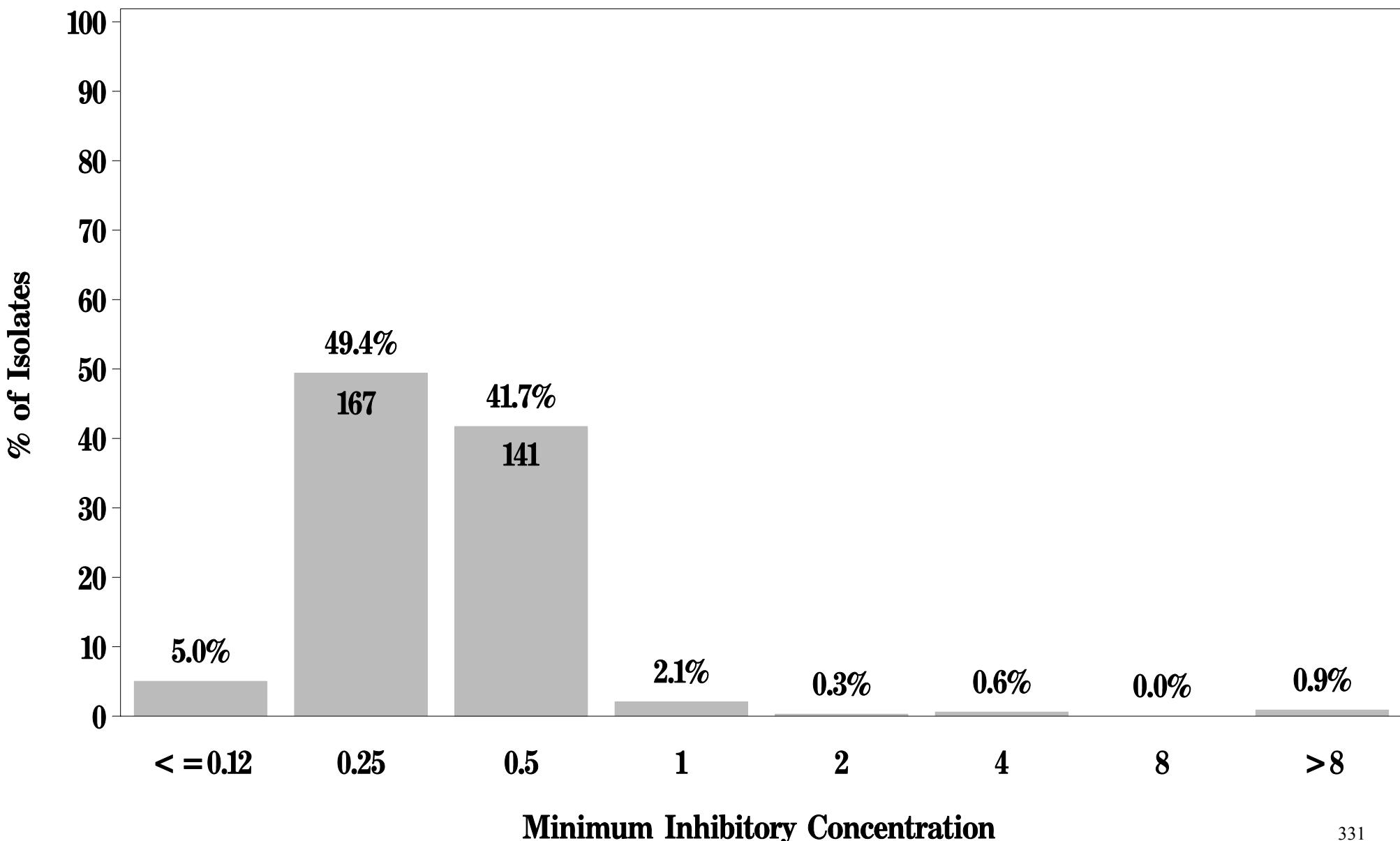
Breakpoints: Susceptible $\leq 2 \mu\text{g/mL}$ Resistant $\geq 8 \mu\text{g/mL}$



NARMS

**Figure 19e: Minimum Inhibitory Concentration of Ceftiofur
for *Escherichia coli* in Ground Beef (N=338 Isolates)**

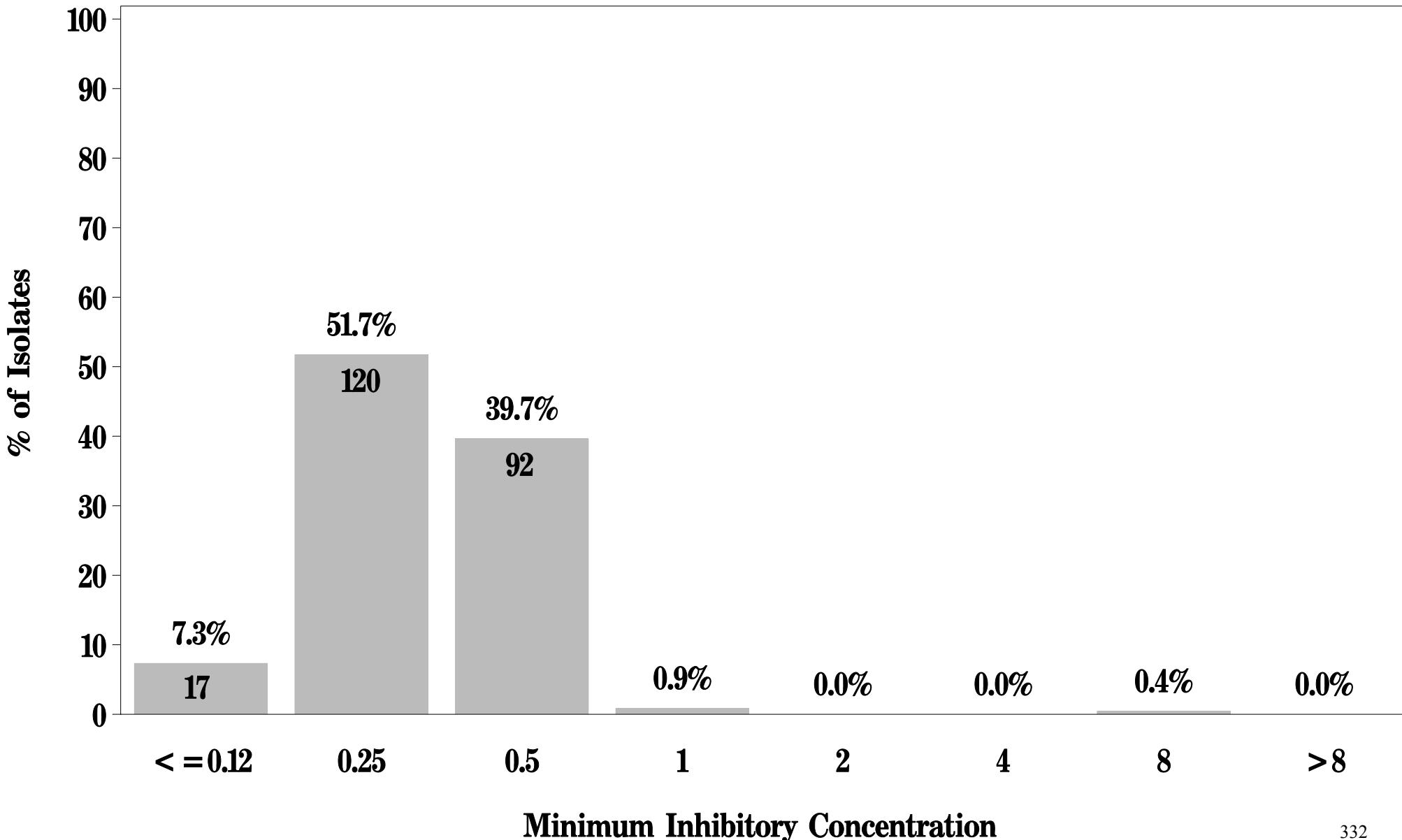
Breakpoints: Susceptible $\leq 2 \mu\text{g/mL}$ Resistant $\geq 8 \mu\text{g/mL}$



NARMS

**Figure 19e: Minimum Inhibitory Concentration of Ceftiofur
for *Escherichia coli* in Pork Chop (N=232 Isolates)**

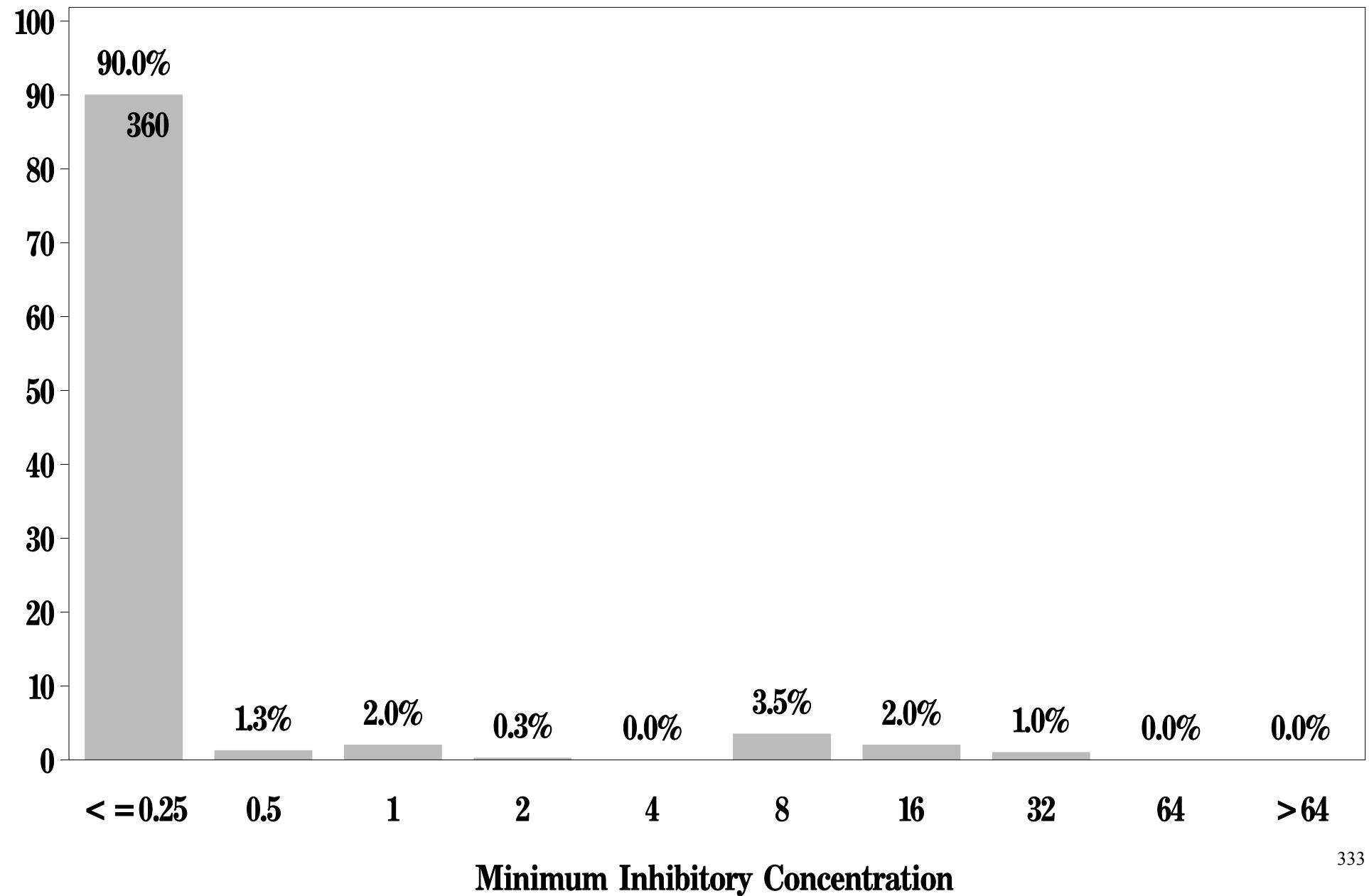
Breakpoints: Susceptible $\leq 2 \mu\text{g/mL}$ Resistant $\geq 8 \mu\text{g/mL}$



NARMS

**Figure 19f: Minimum Inhibitory Concentration of Ceftriaxone
for *Escherichia* in Chicken Breast (N=400 Isolates)**

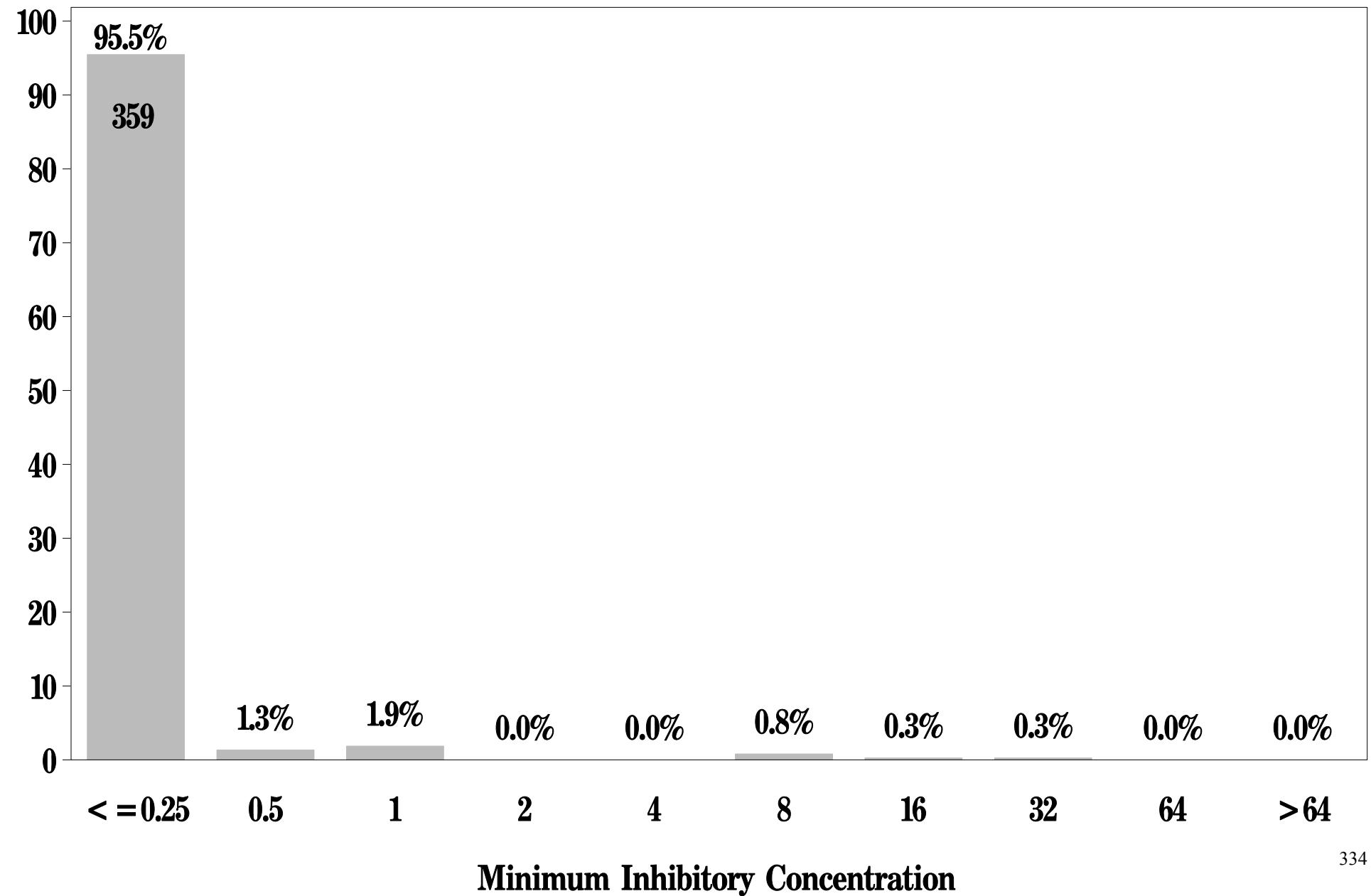
Breakpoints: Susceptible $\leq 8 \mu\text{g/mL}$ Resistant $\geq 16 \mu\text{g/mL}$



NARMS

**Figure 19f: Minimum Inhibitory Concentration of Ceftriaxone
for *Escherichia* in Ground Turkey (N=376 Isolates)**

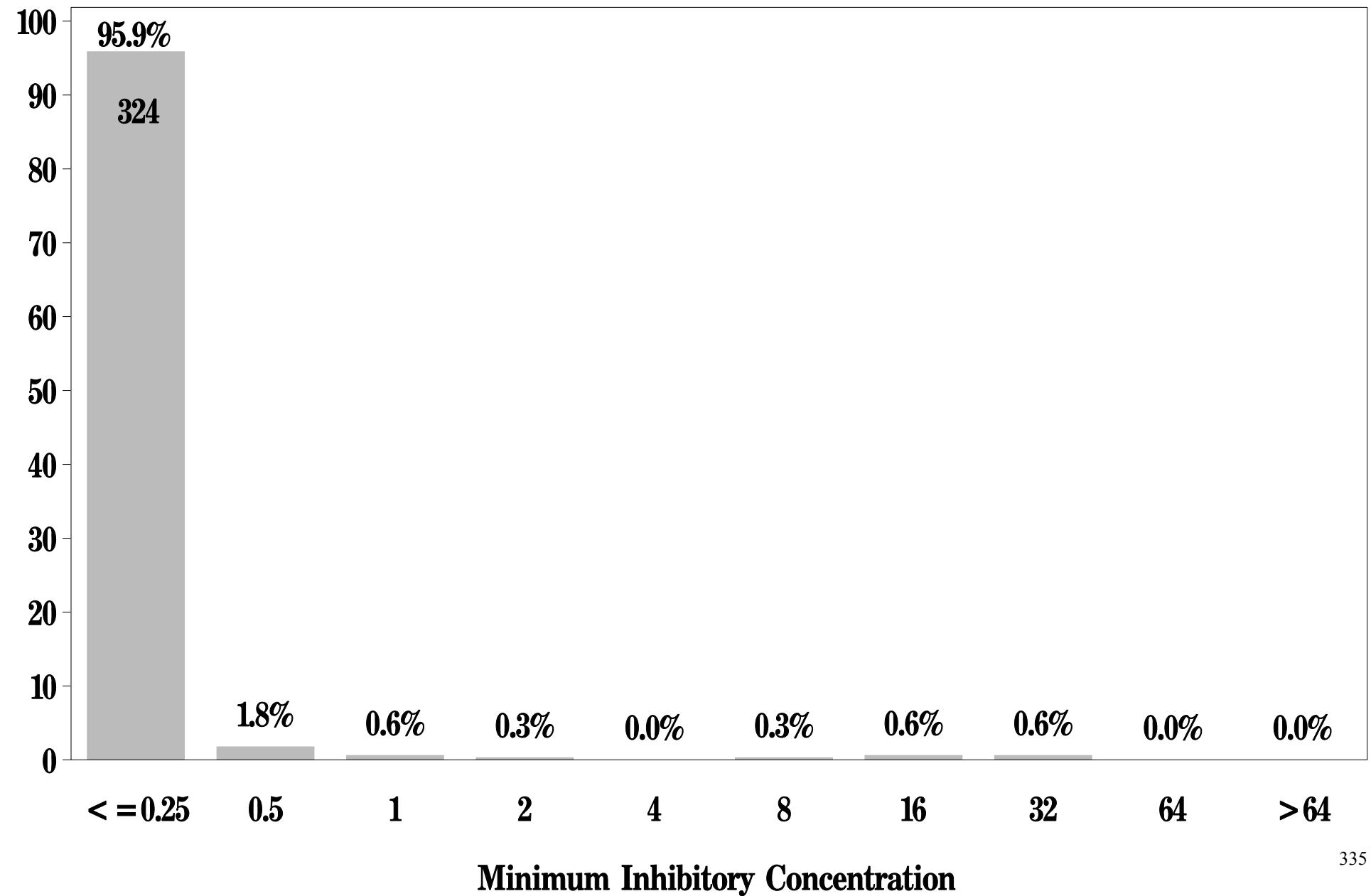
Breakpoints: Susceptible $\leq 8 \mu\text{g/mL}$ Resistant $\geq 64 \mu\text{g/mL}$



NARMS

**Figure 19f: Minimum Inhibitory Concentration of Ceftriaxone
for *Escherichia* in Ground Beef (N=338 Isolates)**

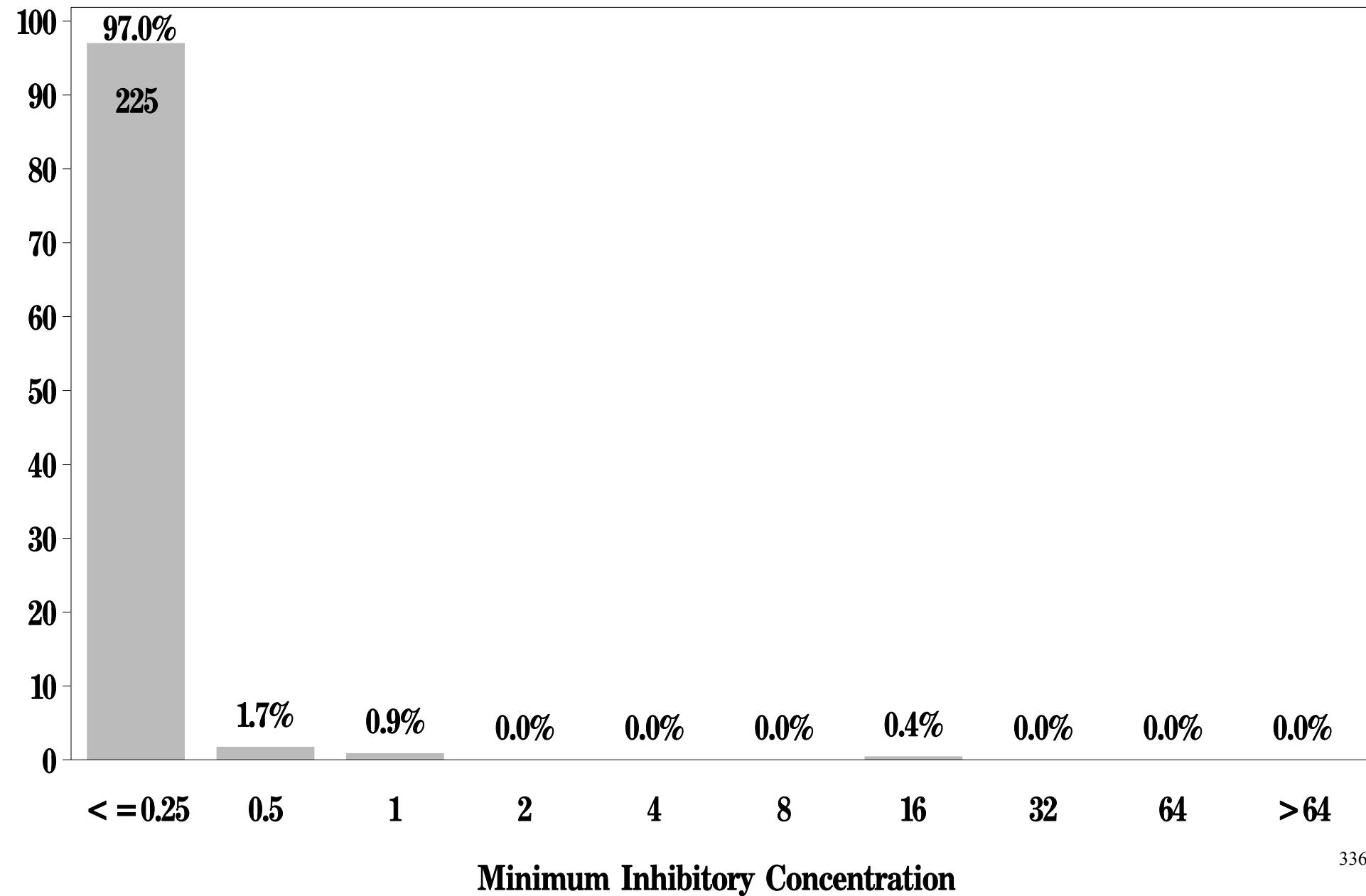
Breakpoints: Susceptible $\leq 8 \mu\text{g/mL}$ Resistant $\geq 64 \mu\text{g/mL}$



NARMS

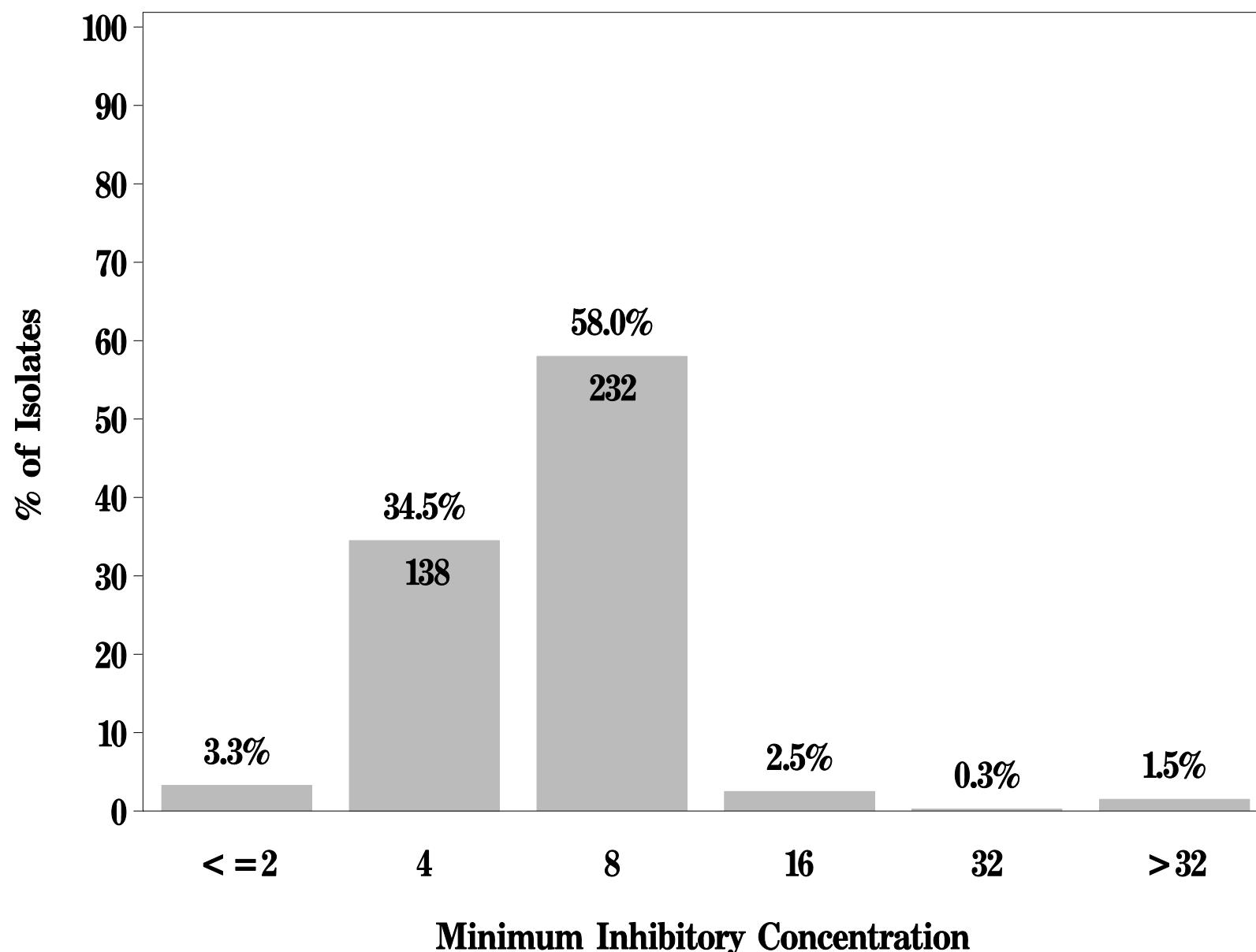
**Figure 19f: Minimum Inhibitory Concentration of Ceftriaxone
for *Escherichia* in Pork Chop (N=232 Isolates)**

Breakpoints: Susceptible $\leq 8 \mu\text{g/mL}$ Resistant $\geq 64 \mu\text{g/mL}$



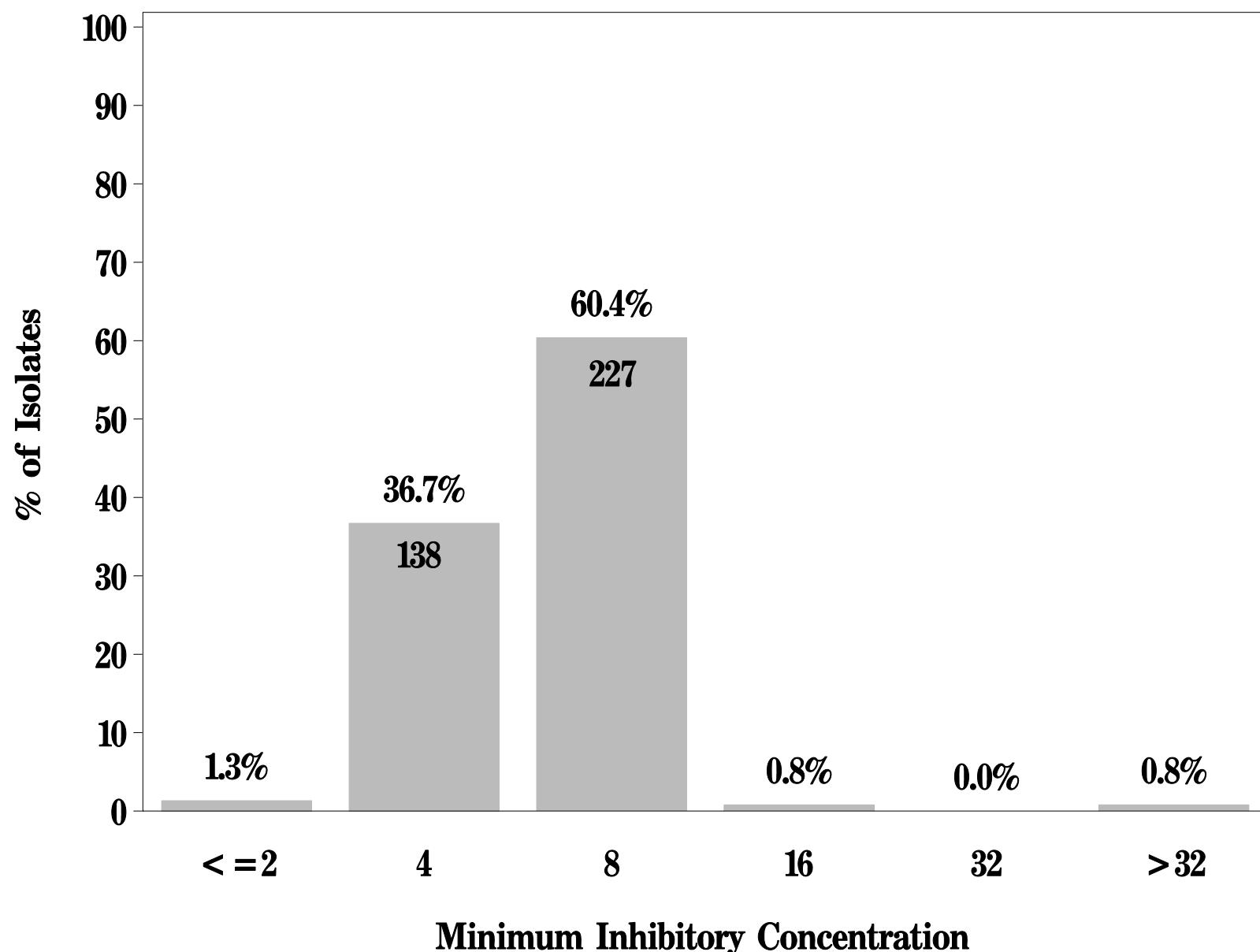
NARMS

**Figure 19g: Minimum Inhibitory Concentration of Chloramphenicol
for *Escherichia coli* in Chicken Breast (N=400 Isolates)**
Breakpoints: Susceptible $\leq 8 \text{ } \mu\text{g/mL}$ Resistant $\geq 32 \text{ } \mu\text{g/mL}$



NARMS

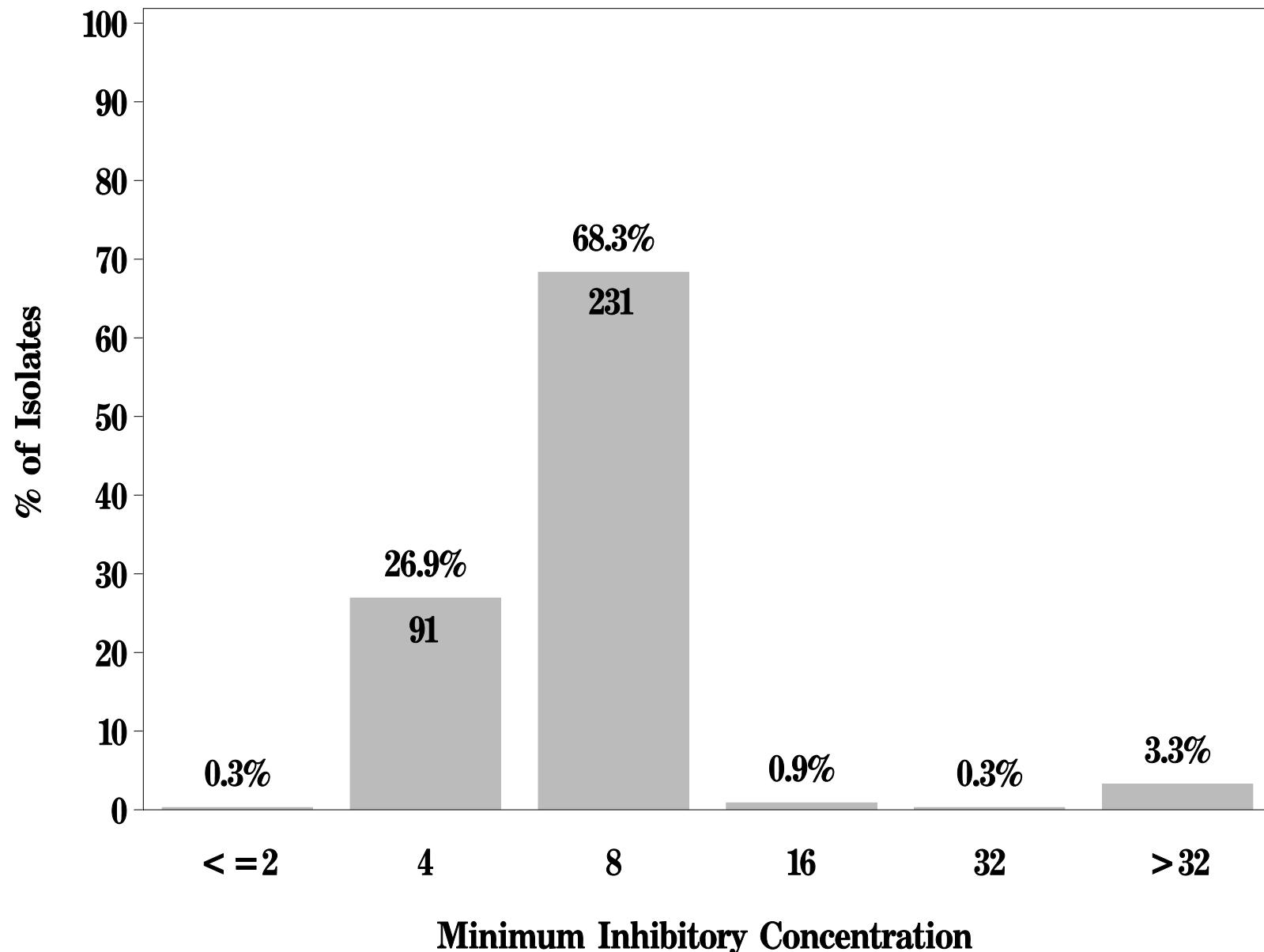
**Figure 19g: Minimum Inhibitory Concentration of Chloramphenicol
for *Escherichia coli* in Ground Turkey (N=376 Isolates)**
Breakpoints: Susceptible $\leq 8 \mu\text{g/mL}$ Resistant $\geq 32 \mu\text{g/mL}$



NARMS

**Figure 19g: Minimum Inhibitory Concentration of Chloramphenicol
for *Escherichia coli* in Ground Beef (N=338 Isolates)**

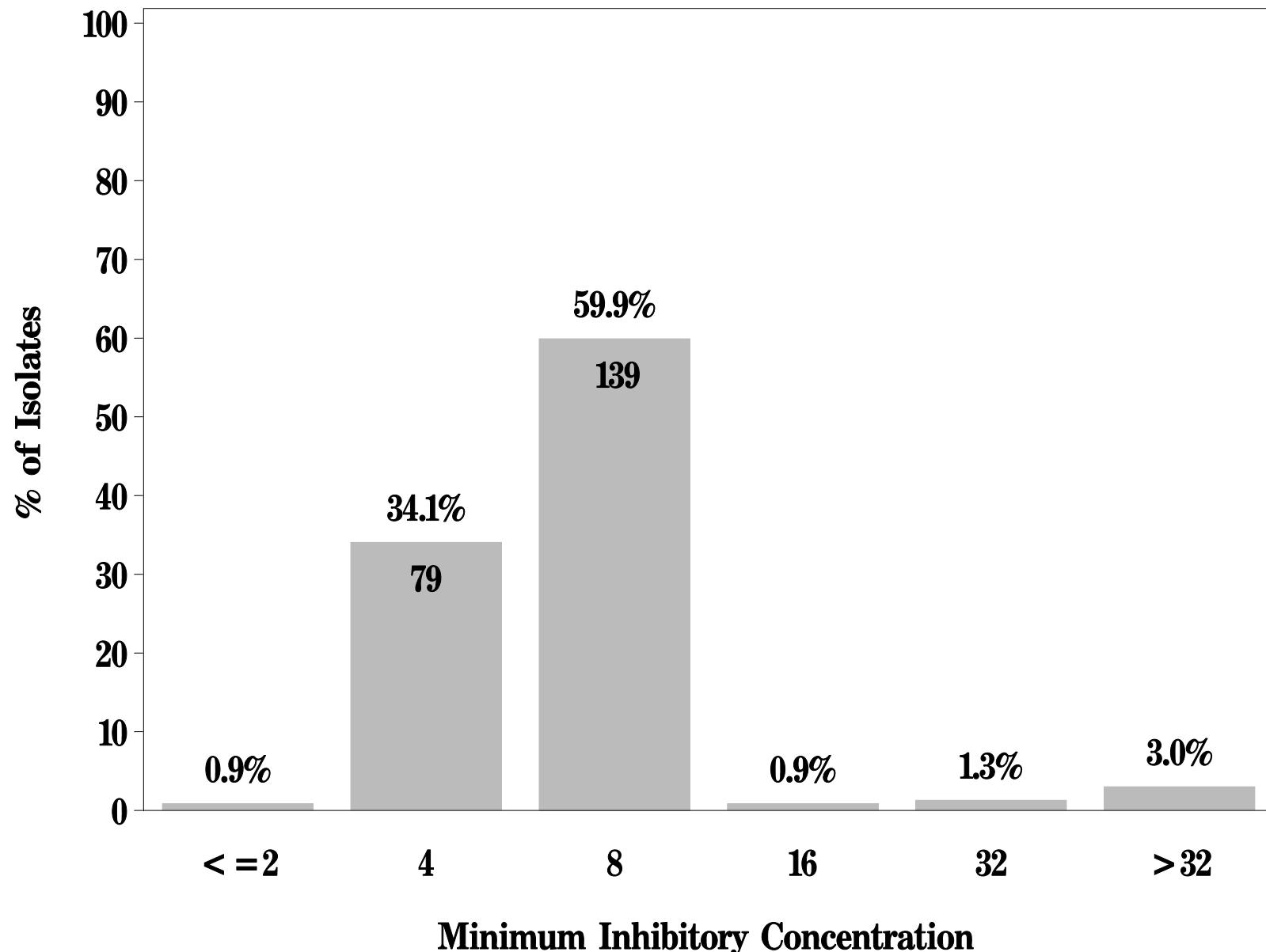
Breakpoints: Susceptible $\leq 8 \mu\text{g/mL}$ Resistant $\geq 32 \mu\text{g/mL}$



NARMS

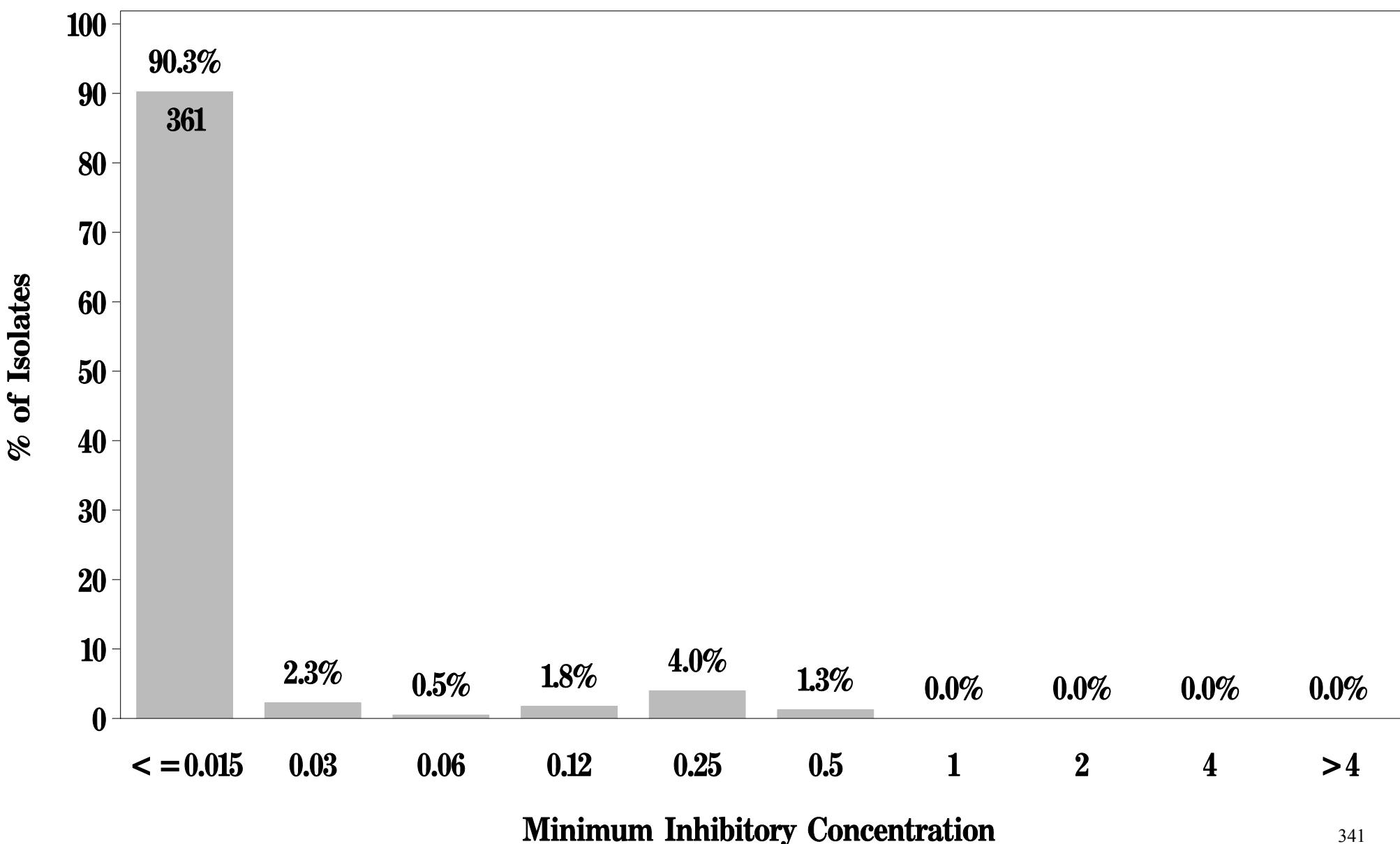
**Figure 19g: Minimum Inhibitory Concentration of Chloramphenicol
for *Escherichia coli* in Pork Chop (N=232 Isolates)**

Breakpoints: Susceptible $\leq 8 \mu\text{g/mL}$ Resistant $\geq 32 \mu\text{g/mL}$



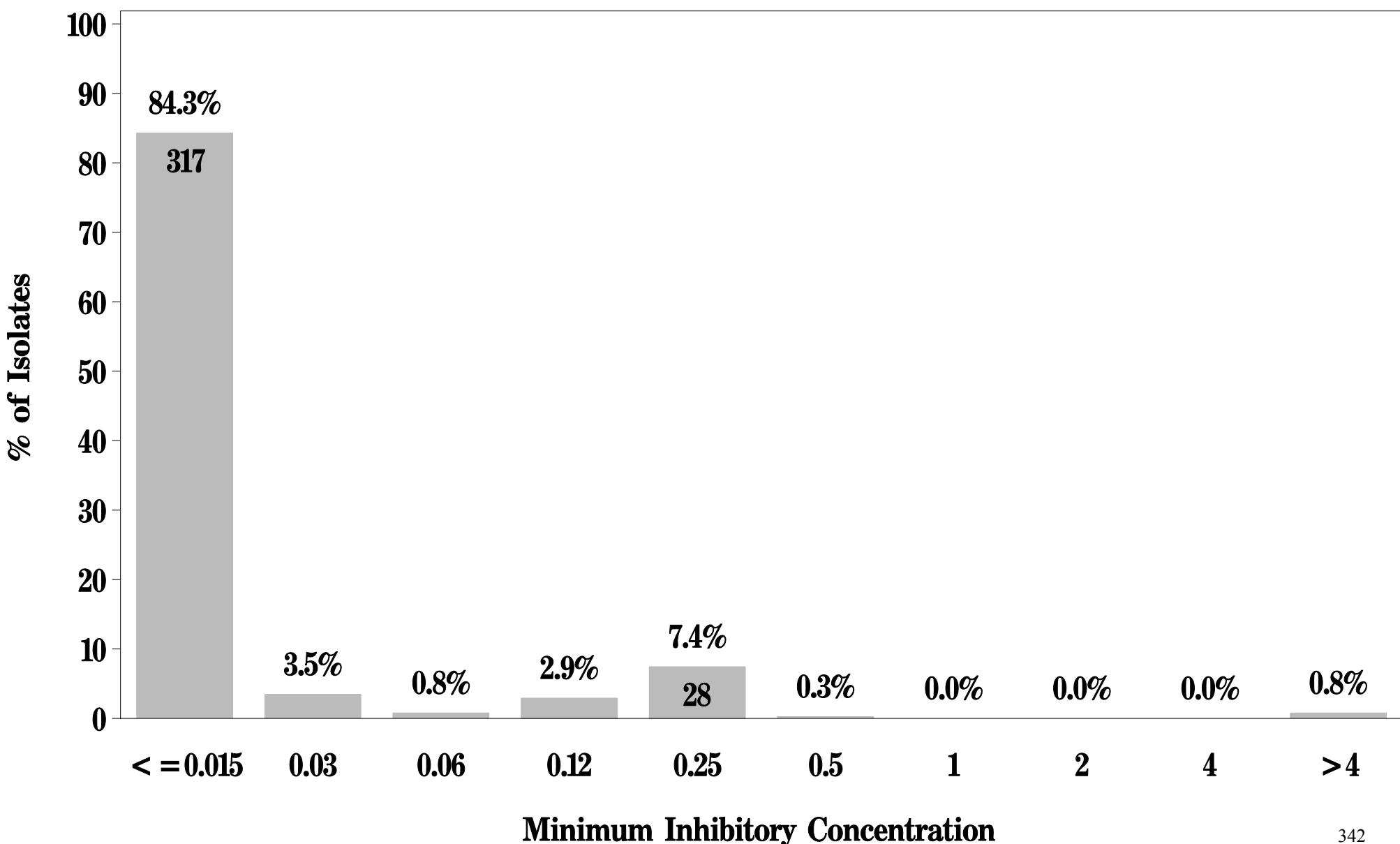
NARMS

**Figure 19h: Minimum Inhibitory Concentration of Ciprofloxacin
for *Escherichia coli* in Chicken Breast (N=400 Isolates)**
Breakpoints: Susceptible $\leq 1 \mu\text{g/mL}$ Resistant $\geq 4 \mu\text{g/mL}$



NARMS

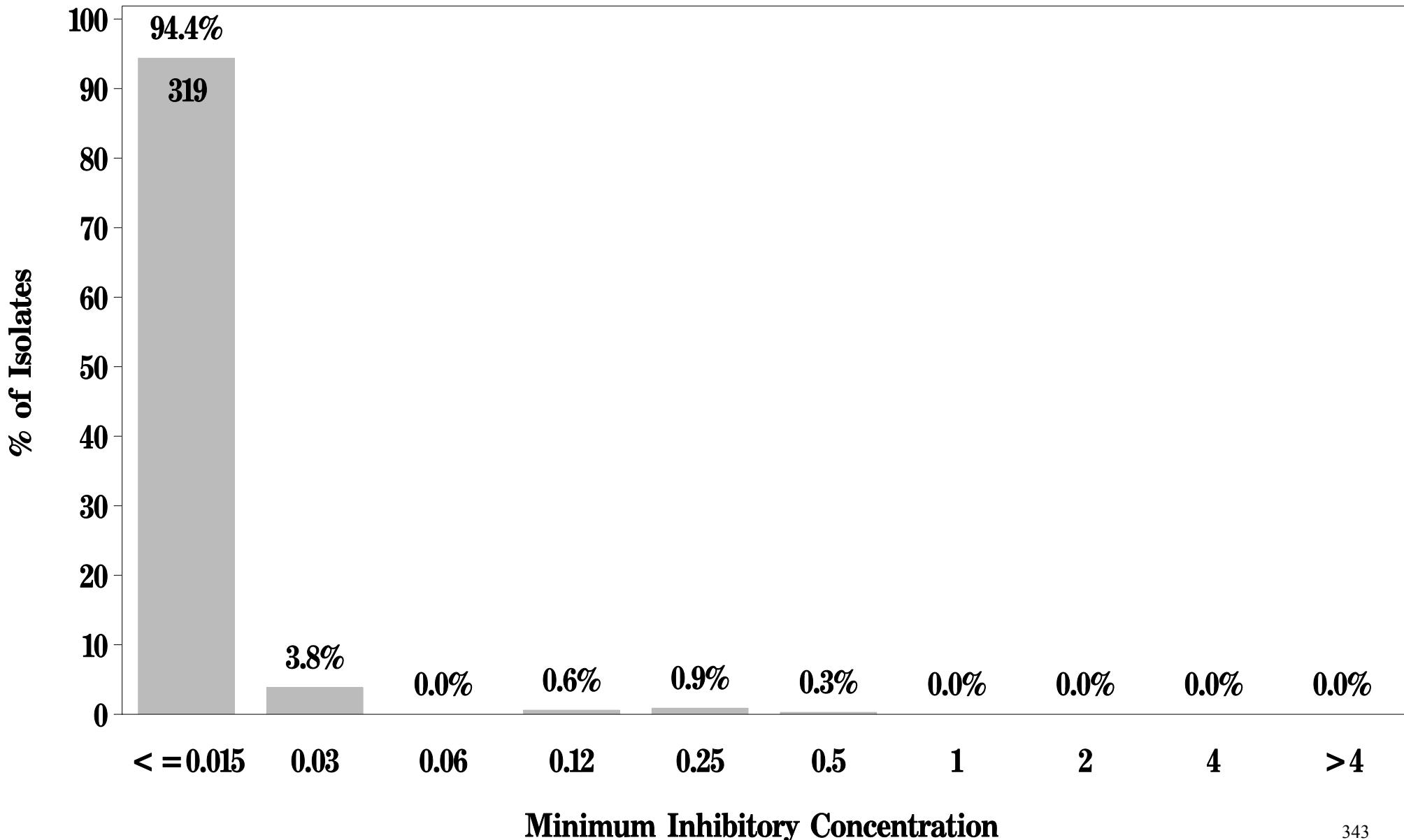
**Figure 19h: Minimum Inhibitory Concentration of Ciprofloxacin
for *Escherichia coli* in Ground Turkey (N=376 Isolates)**
Breakpoints: Susceptible $\leq 1 \mu\text{g/mL}$ Resistant $\geq 4 \mu\text{g/mL}$



NARMS

**Figure 19h: Minimum Inhibitory Concentration of Ciprofloxacin
for *Escherichia coli* in Ground Beef (N=338 Isolates)**

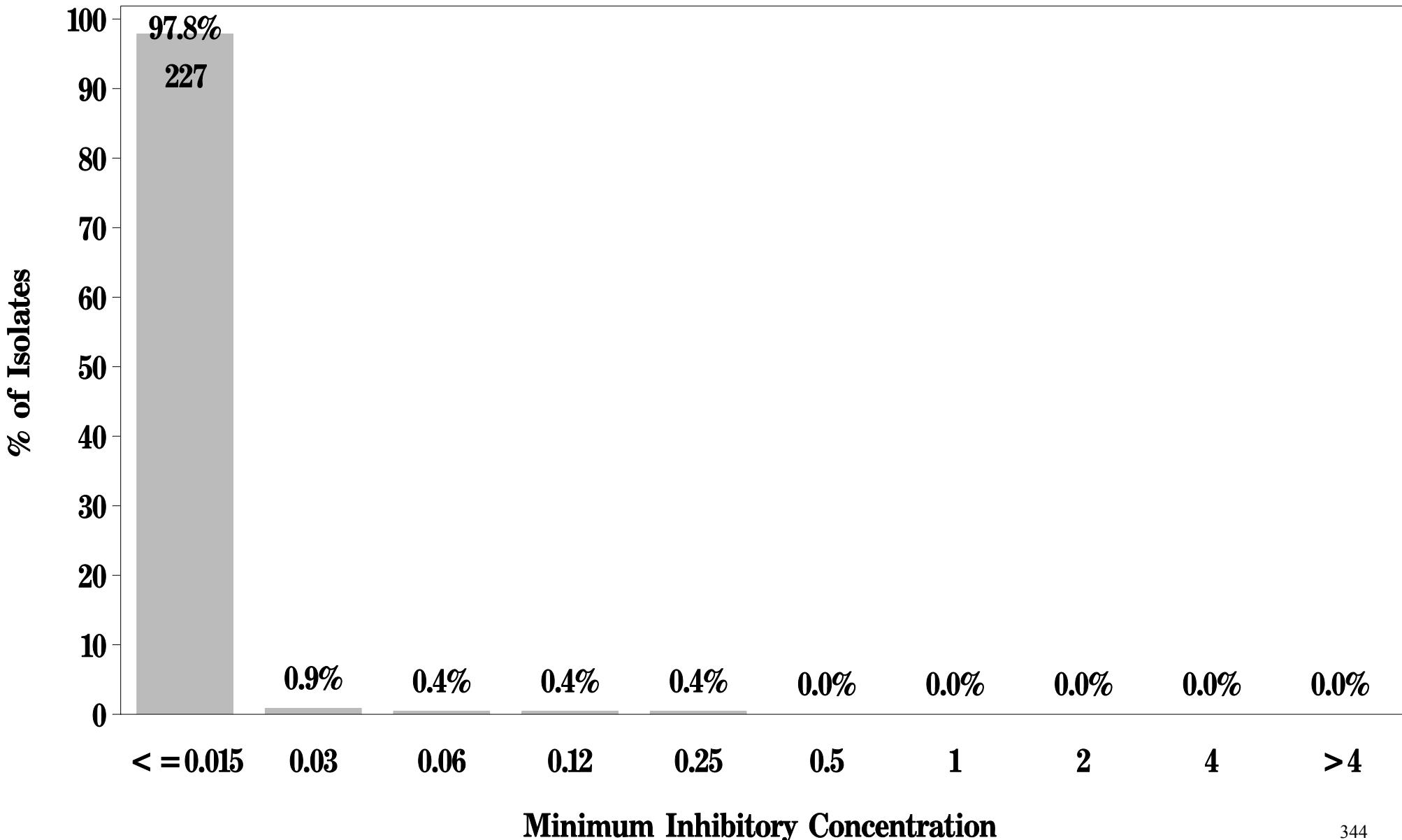
Breakpoints: Susceptible $\leq 1 \mu\text{g/mL}$ Resistant $\geq 4 \mu\text{g/mL}$



NARMS

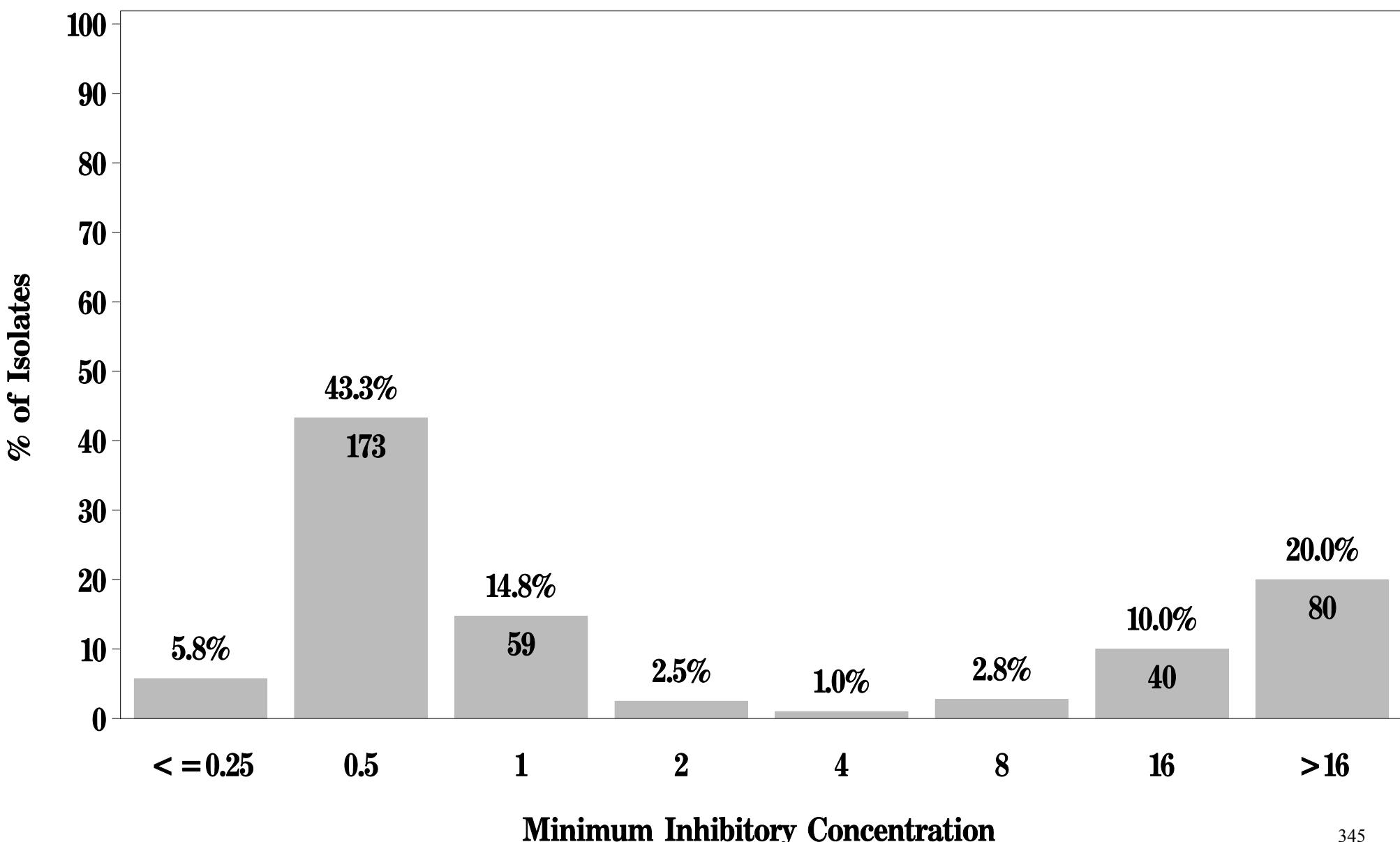
**Figure 19h: Minimum Inhibitory Concentration of Ciprofloxacin
for *Escherichia coli* in Pork Chop (N=232 Isolates)**

Breakpoints: Susceptible $\leq 1 \mu\text{g/mL}$ Resistant $\geq 4 \mu\text{g/mL}$



NARMS

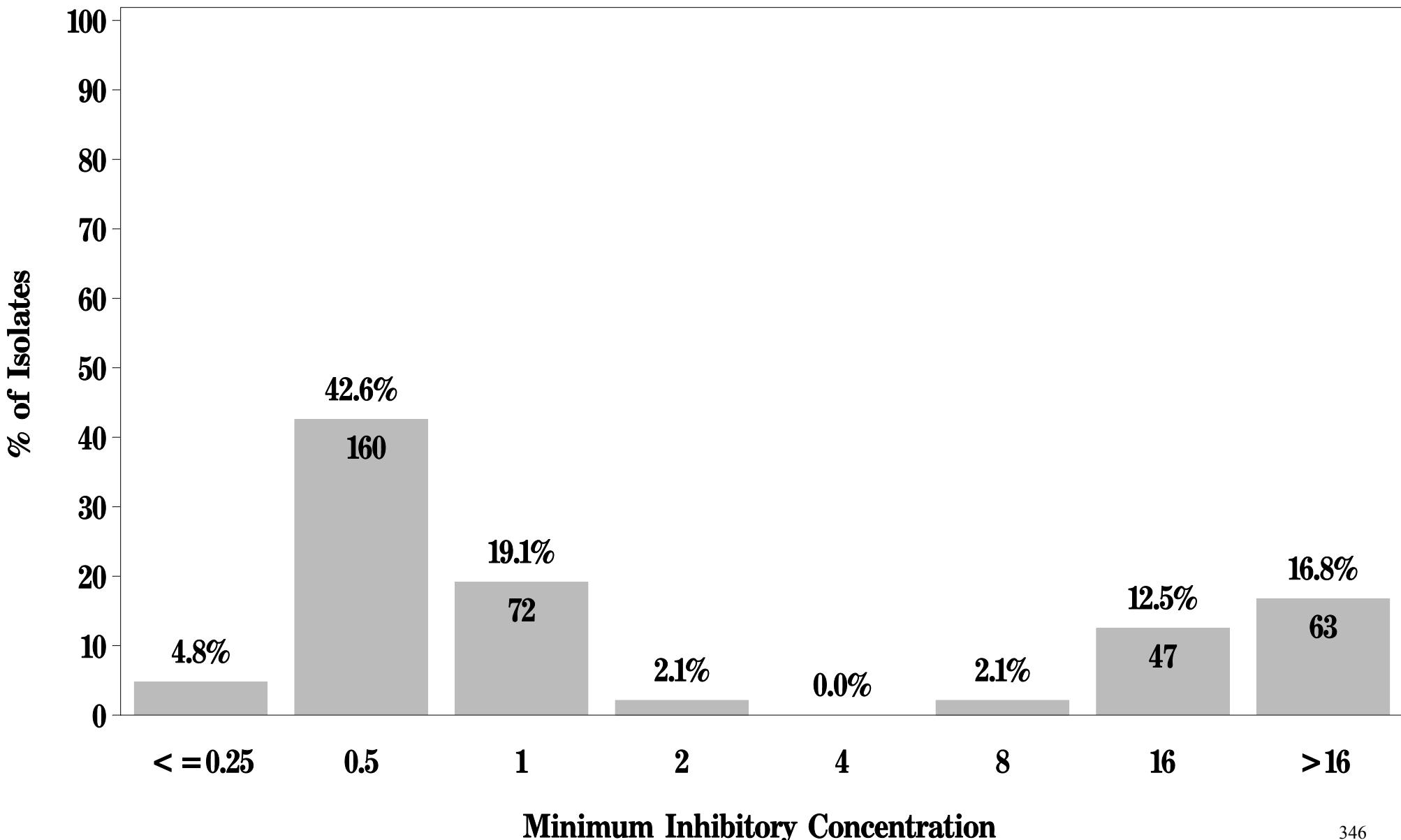
**Figure 19i: Minimum Inhibitory Concentration of Gentamicin
for *Escherichia coli* in Chicken Breast (N=400 Isolates)**
Breakpoints: Susceptible $\leq 4 \text{ } \mu\text{g/mL}$ Resistant $\geq 16 \text{ } \mu\text{g/mL}$



NARMS

**Figure 19i: Minimum Inhibitory Concentration of Gentamicin
for *Escherichia coli* in Ground Turkey (N=376 Isolates)**

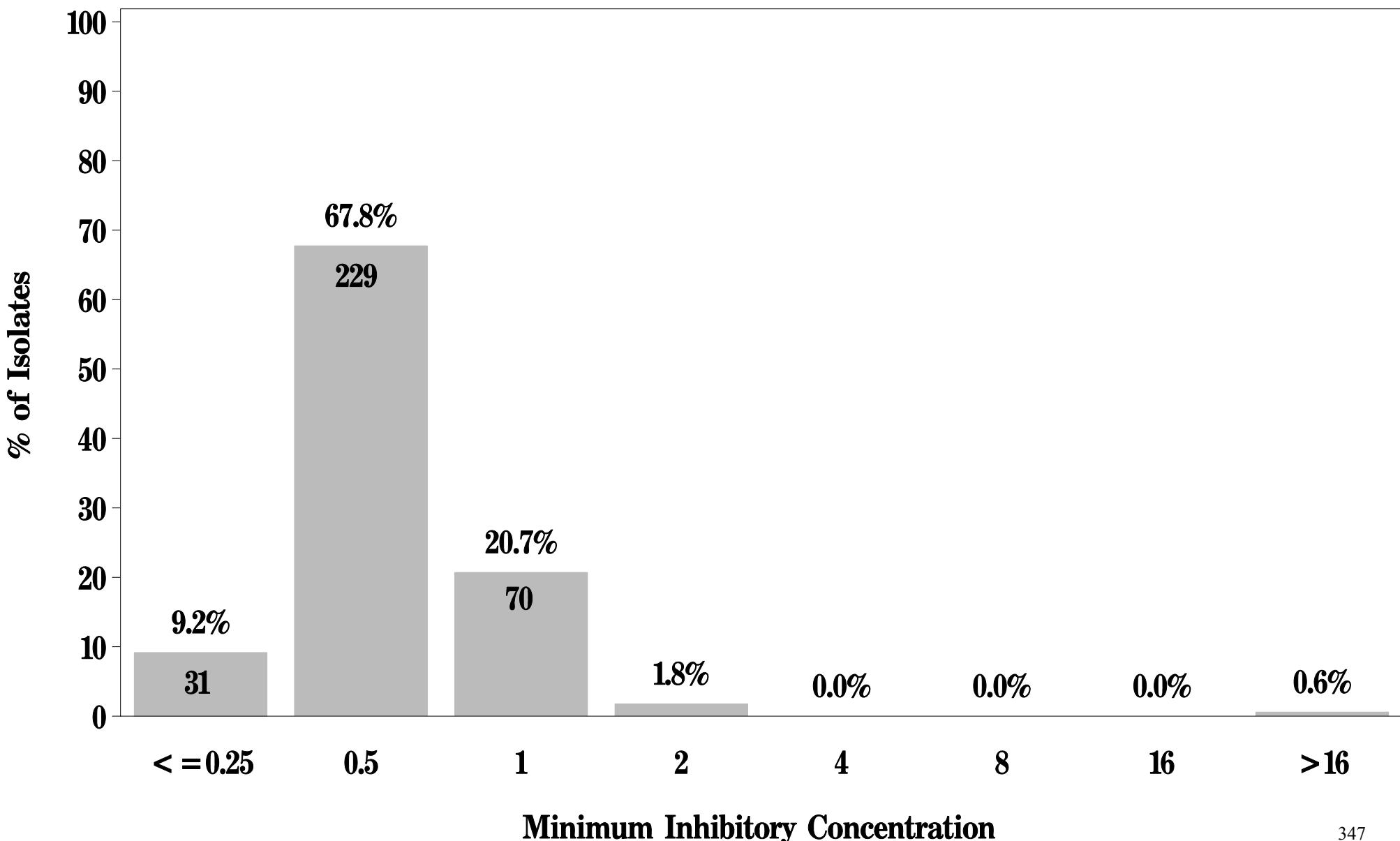
Breakpoints: Susceptible $\leq 4 \text{ } \mu\text{g/mL}$ Resistant $\geq 16 \text{ } \mu\text{g/mL}$



NARMS

**Figure 19i: Minimum Inhibitory Concentration of Gentamicin
for *Escherichia coli* in Ground Beef (N=338 Isolates)**

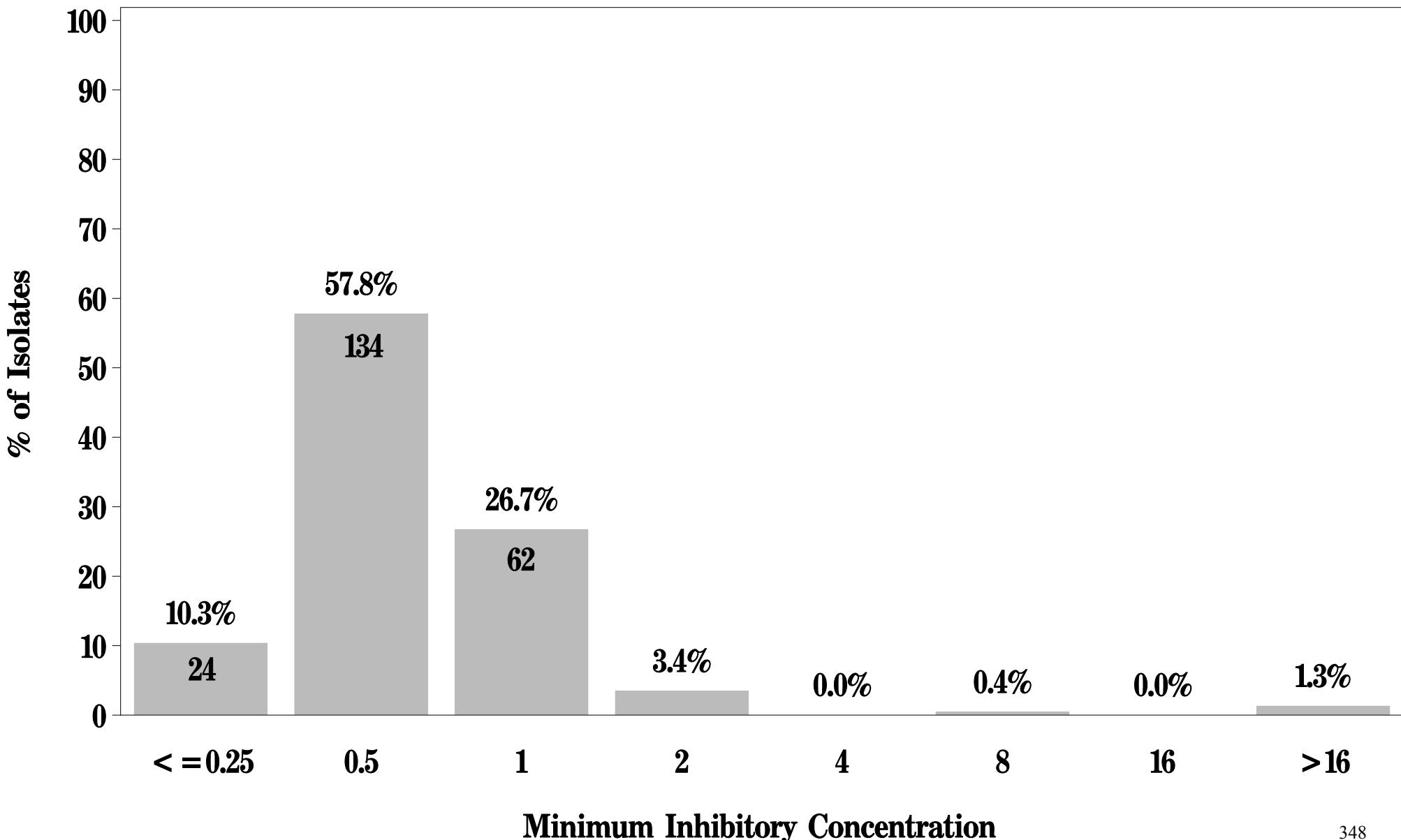
Breakpoints: Susceptible $\leq 4 \text{ } \mu\text{g/mL}$ Resistant $\geq 16 \text{ } \mu\text{g/mL}$



NARMS

**Figure 19i: Minimum Inhibitory Concentration of Gentamicin
for *Escherichia coli* in Pork Chop (N=232 Isolates)**

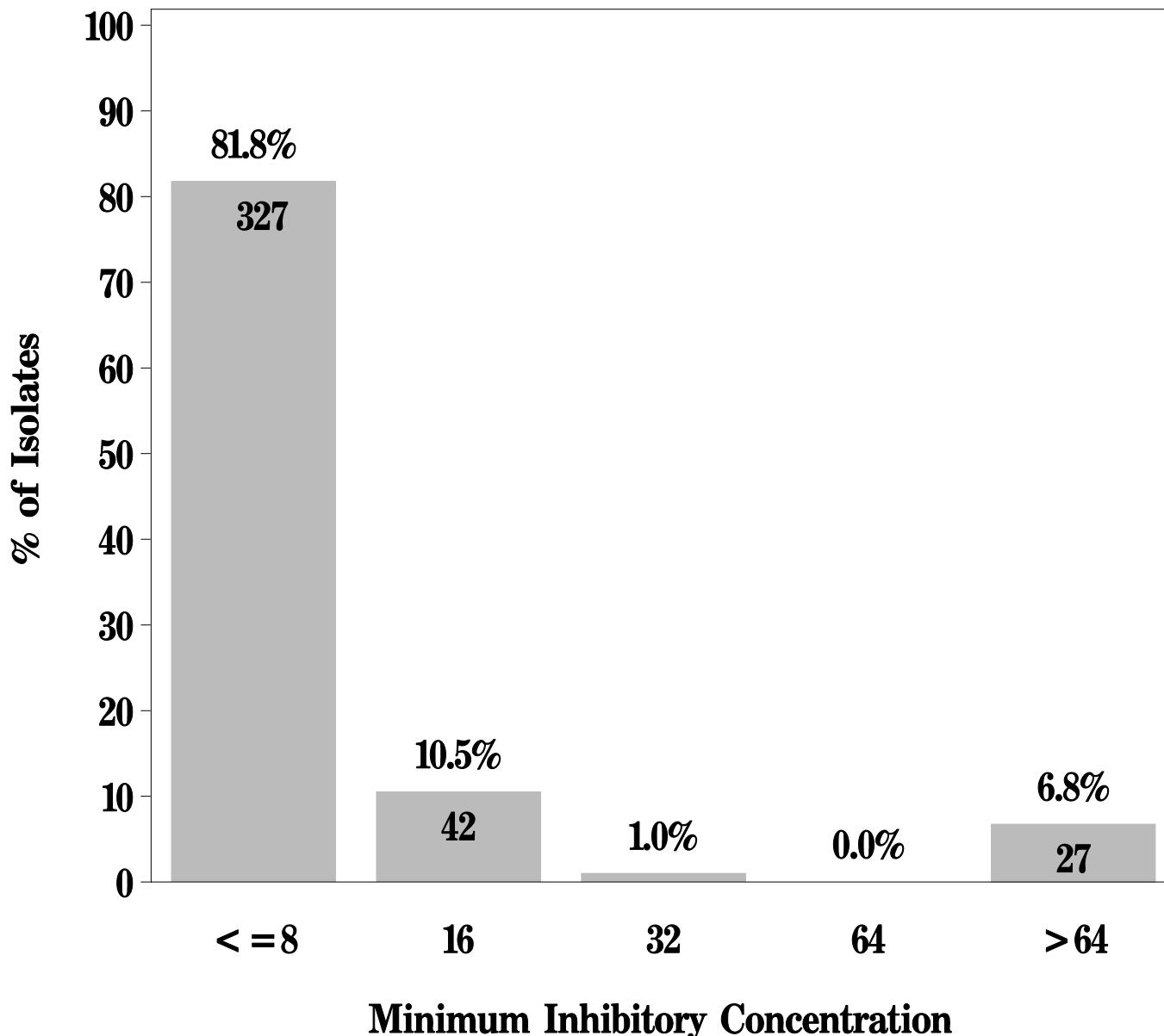
Breakpoints: Susceptible $\leq 4 \mu\text{g/mL}$ Resistant $\geq 16 \mu\text{g/mL}$



NARMS

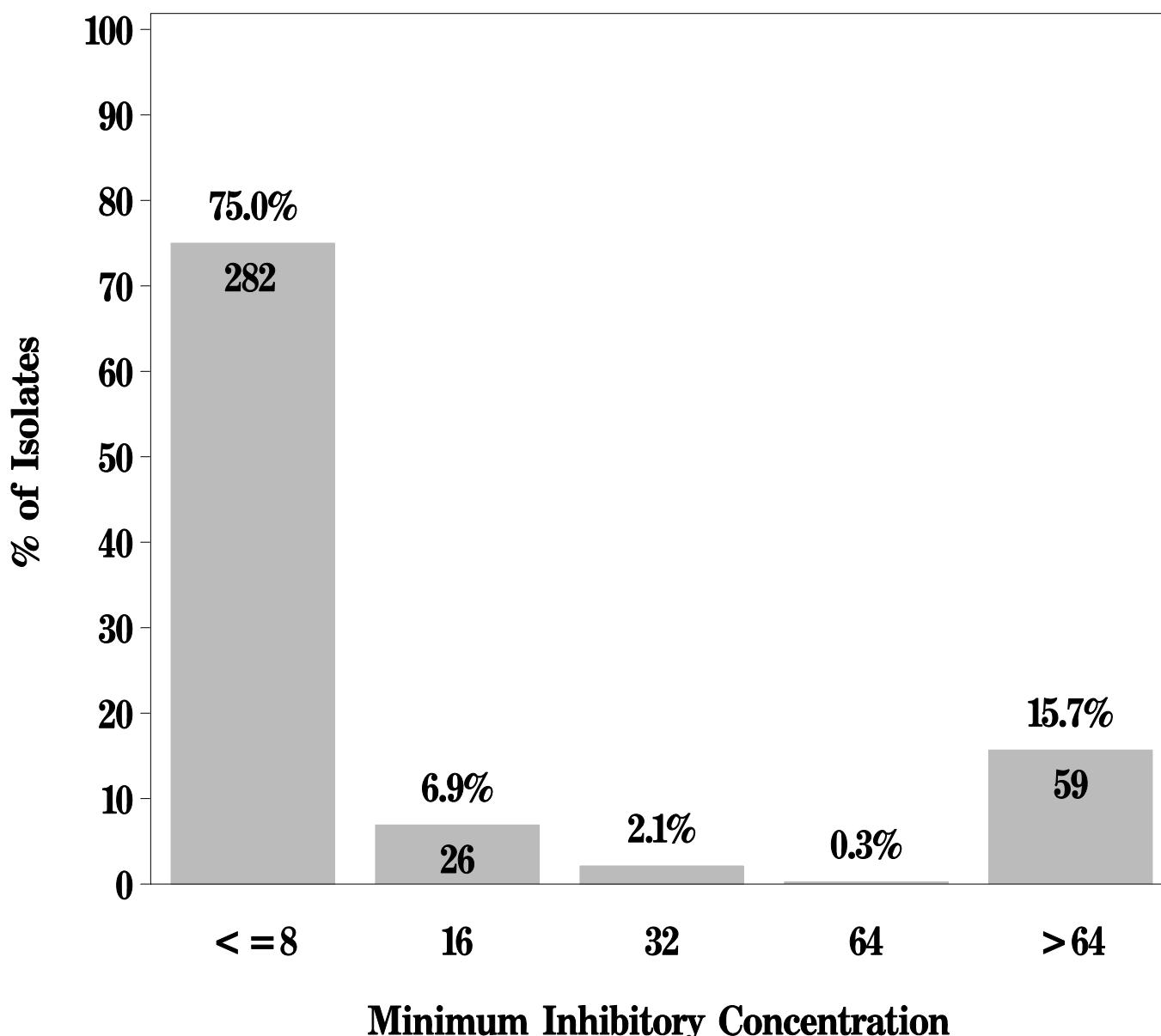
**Figure 19j: Minimum Inhibitory Concentration of Kanamycin
for *Escherichia coli* in Chicken Breast (N=400 Isolates)**

Breakpoints: Susceptible $\leq 16 \mu\text{g/mL}$ Resistant $\geq 64 \mu\text{g/mL}$



NARMS

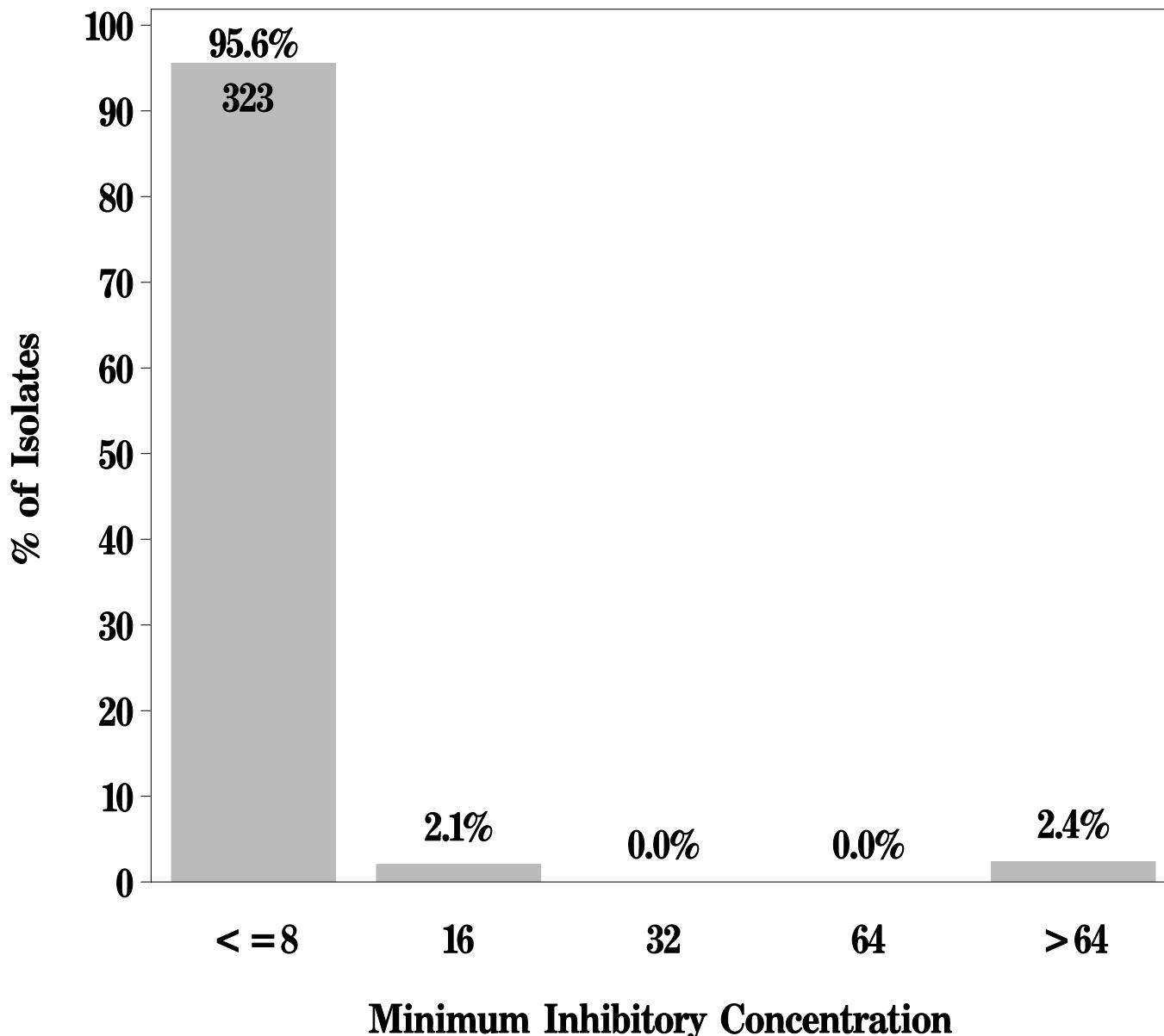
**Figure 19j: Minimum Inhibitory Concentration of Kanamycin
for *Escherichia coli* in Ground Turkey (N=376 Isolates)**
Breakpoints: Susceptible $\leq 16 \mu\text{g/mL}$ Resistant $\geq 64 \mu\text{g/mL}$



NARMS

**Figure 19j: Minimum Inhibitory Concentration of Kanamycin
for *Escherichia coli* in Ground Beef (N=338 Isolates)**

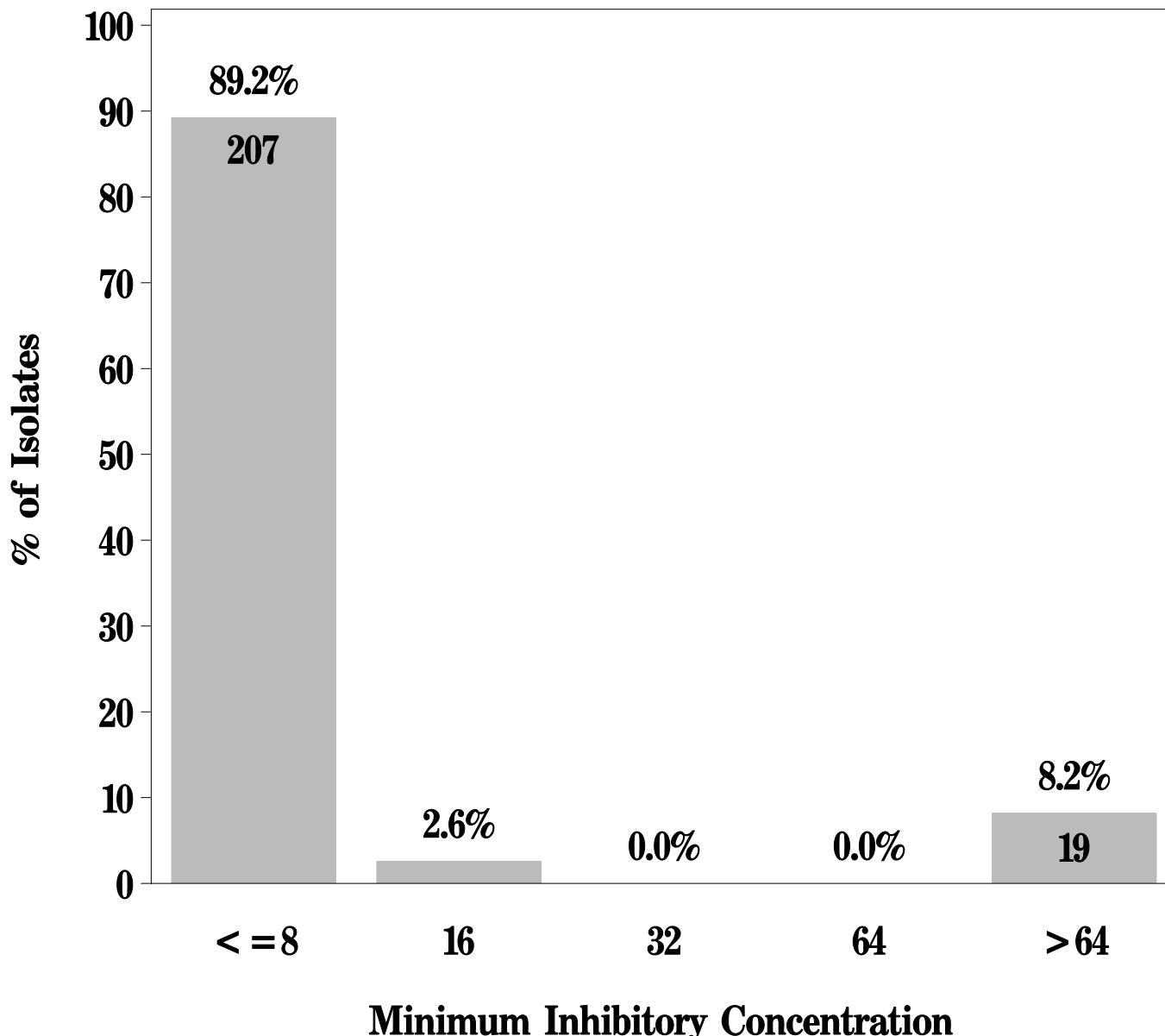
Breakpoints: Susceptible $\leq 16 \mu\text{g/mL}$ Resistant $\geq 64 \mu\text{g/mL}$



NARMS

**Figure 19j: Minimum Inhibitory Concentration of Kanamycin
for *Escherichia coli* in Pork Chop (N=232 Isolates)**

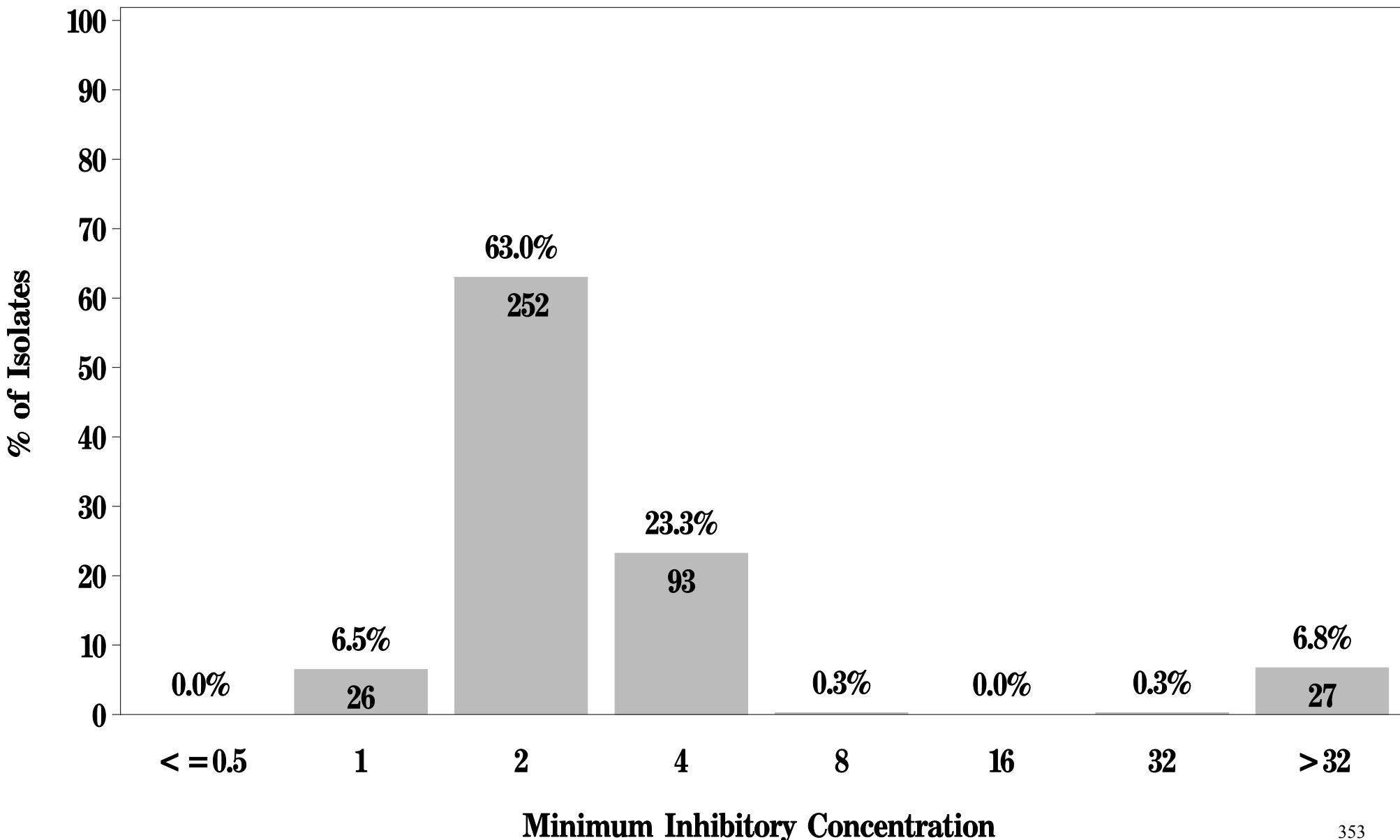
Breakpoints: Susceptible $\leq 16 \mu\text{g/mL}$ Resistant $\geq 64 \mu\text{g/mL}$



NARMS

**Figure 19k: Minimum Inhibitory Concentration of Nalidixic acid
for *Escherichia coli* in Chicken Breast (N=400 Isolates)**

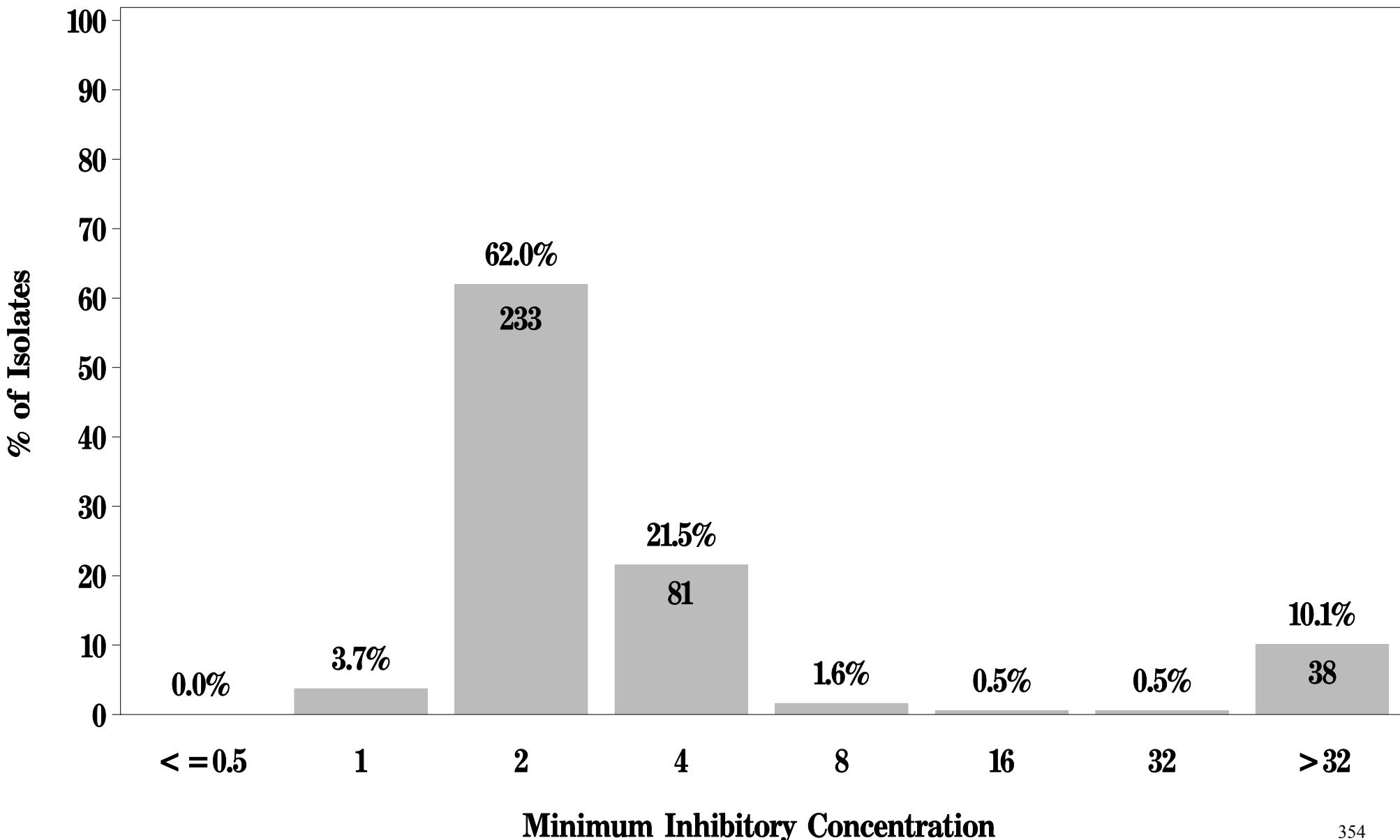
Breakpoints: Susceptible $\leq 16 \mu\text{g/mL}$ Resistant $\geq 32 \mu\text{g/mL}$



NARMS

**Figure 19k: Minimum Inhibitory Concentration of Nalidixic acid
for *Escherichia coli* in Ground Turkey (N=376 Isolates)**

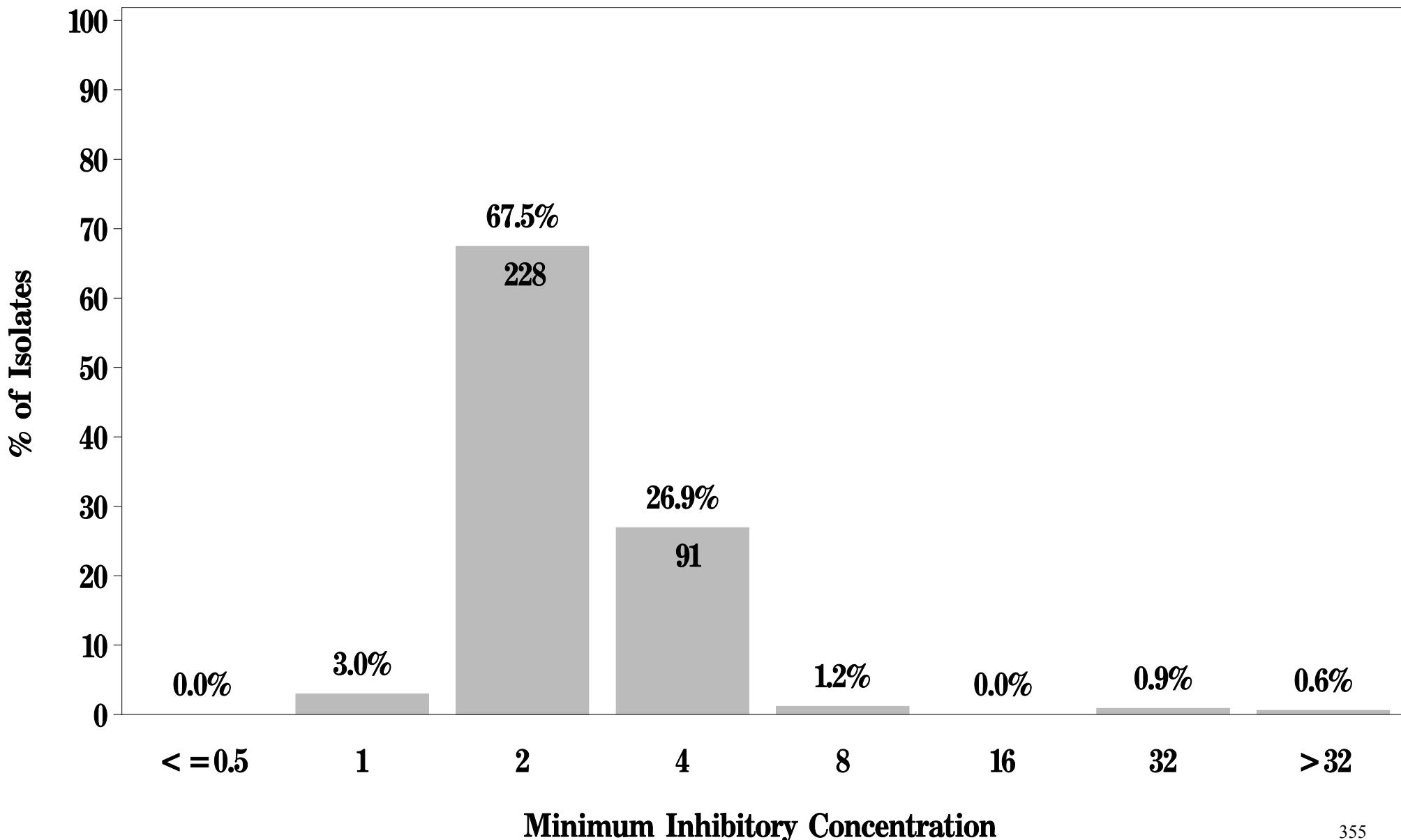
Breakpoints: Susceptible $\leq 16 \mu\text{g/mL}$ Resistant $\geq 32 \mu\text{g/mL}$



NARMS

**Figure 19k: Minimum Inhibitory Concentration of Nalidixic acid
for *Escherichia coli* in Ground Beef (N=338 Isolates)**

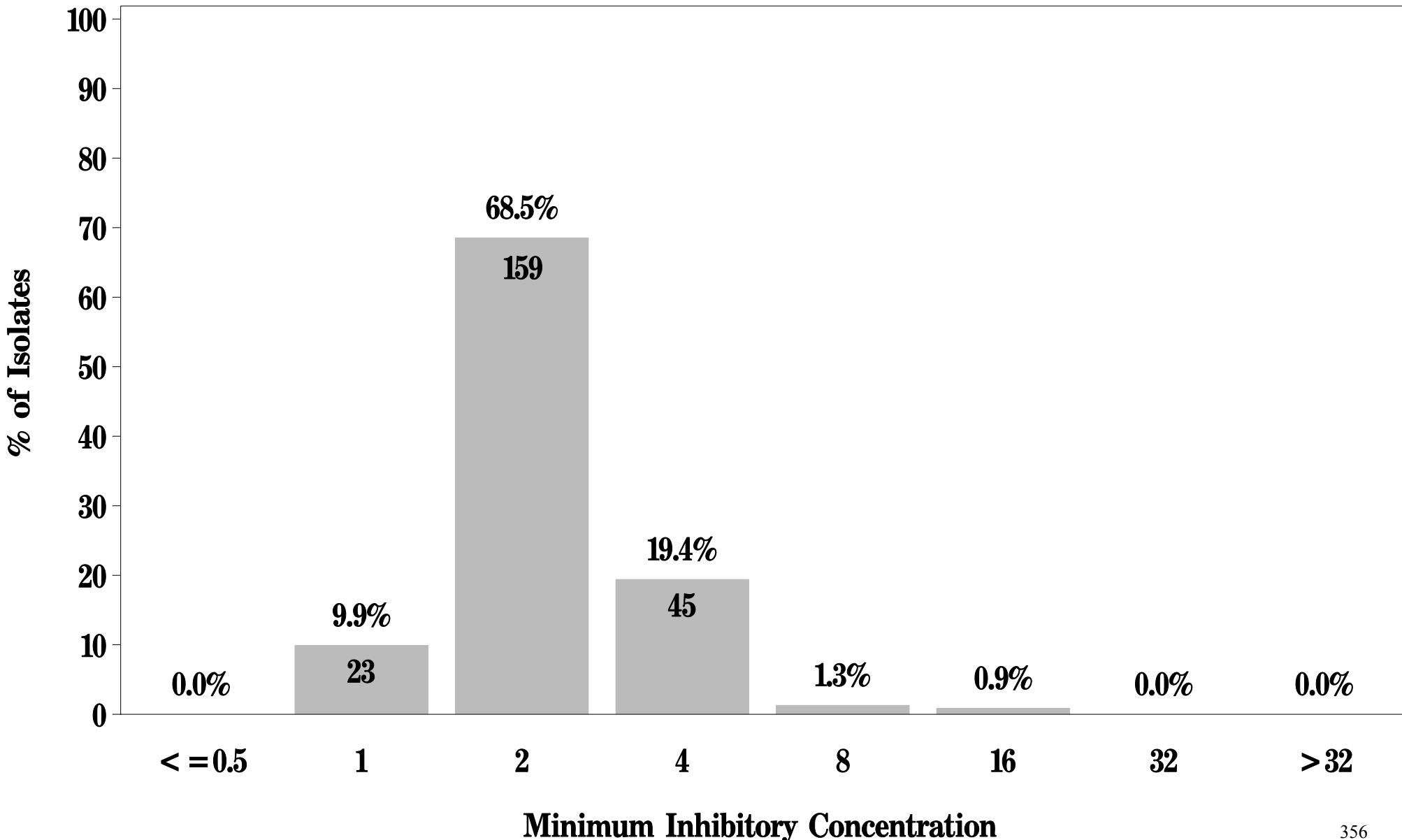
Breakpoints: Susceptible $\leq 16 \mu\text{g/mL}$ Resistant $\geq 32 \mu\text{g/mL}$



NARMS

**Figure 19k: Minimum Inhibitory Concentration of Nalidixic acid
for *Escherichia coli* in Pork Chop (N=232 Isolates)**

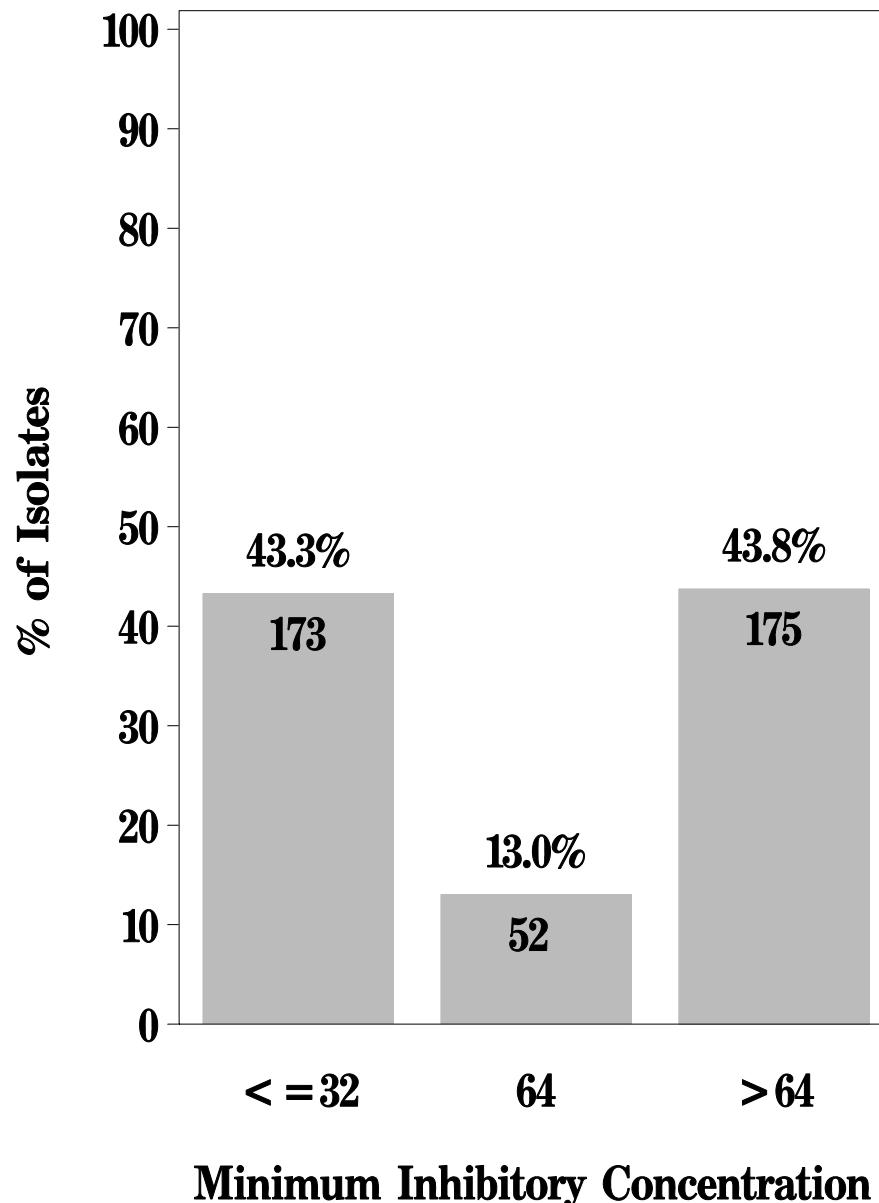
Breakpoints: Susceptible $\leq 16 \mu\text{g/mL}$ Resistant $\geq 32 \mu\text{g/mL}$



NARMS

Figure 19l: Minimum Inhibitory Concentration of Streptomycin for *Escherichia coli* in Chicken Breast (N=400 Isolates)

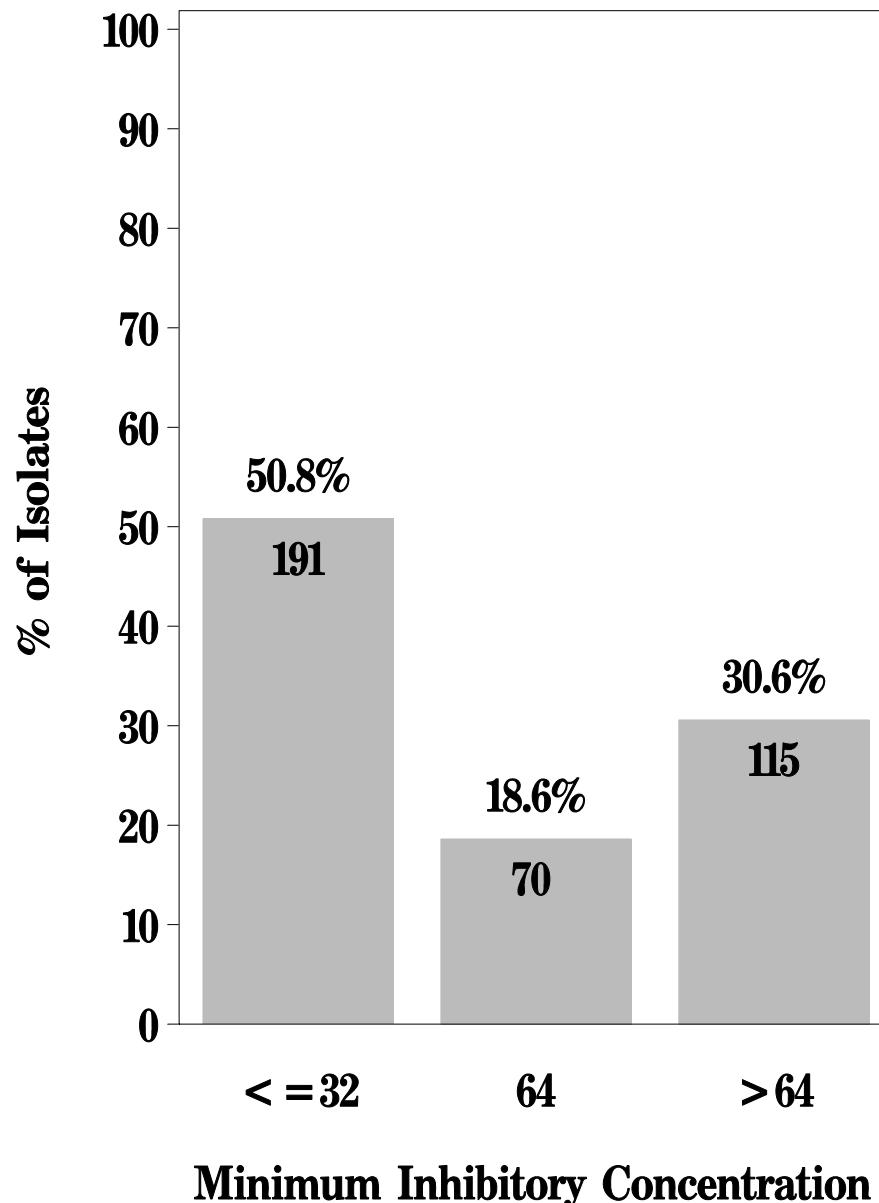
Breakpoints: Susceptible $\leq 32 \text{ } \mu\text{g/mL}$ Resistant $\geq 64 \text{ } \mu\text{g/mL}$



NARMS

**Figure 19l: Minimum Inhibitory Concentration of Streptomycin
for *Escherichia coli* in Ground Turkey (N=376 Isolates)**

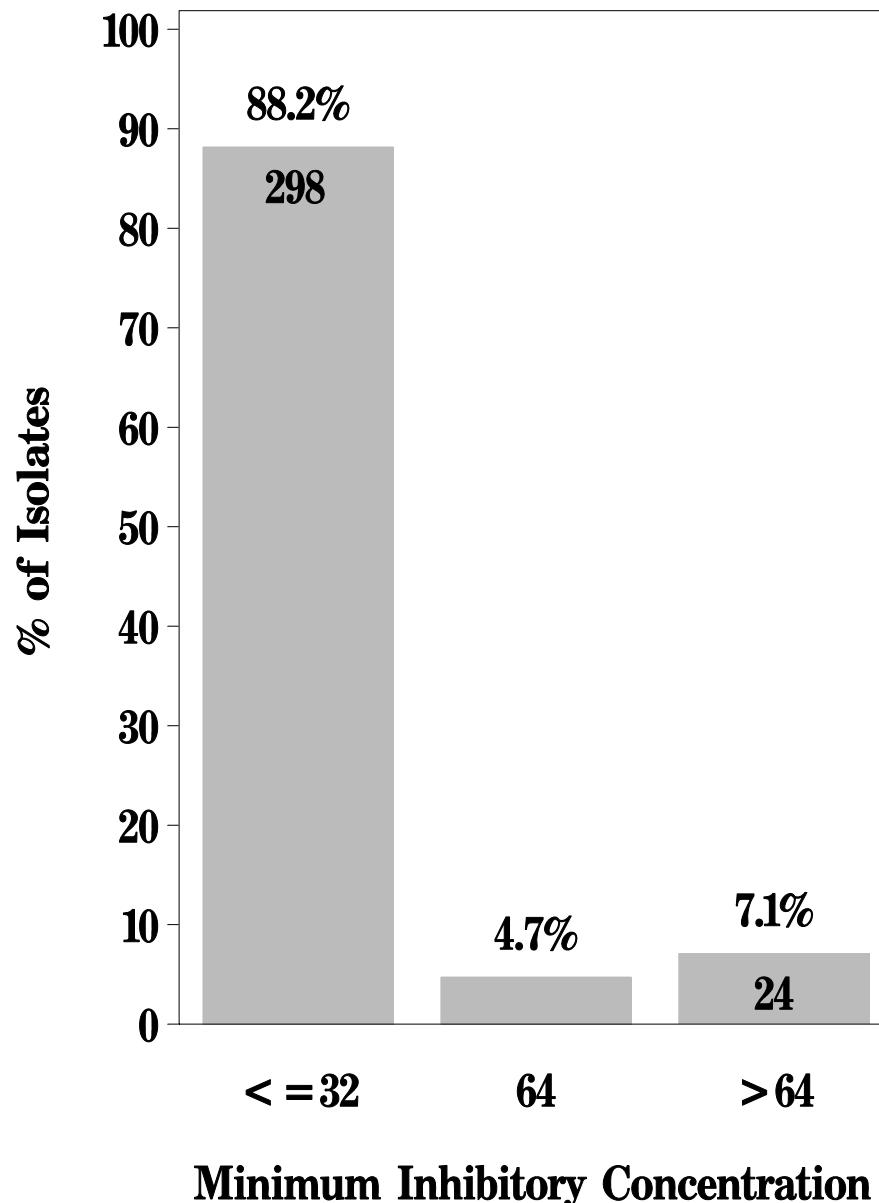
Breakpoints: Susceptible $\leq 32 \mu\text{g/mL}$ Resistant $\geq 64 \mu\text{g/mL}$



NARMS

**Figure 19l: Minimum Inhibitory Concentration of Streptomycin
for *Escherichia coli* in Ground Beef (N=338 Isolates)**

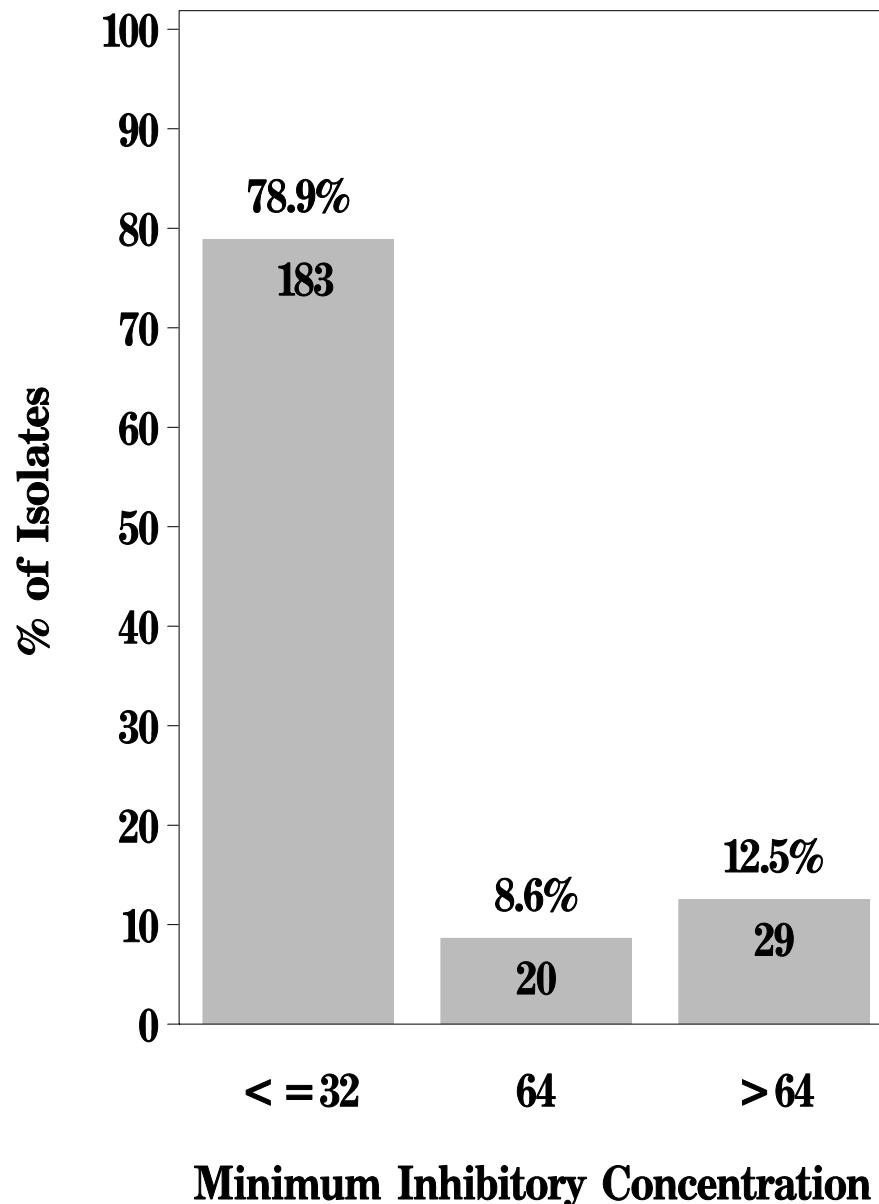
Breakpoints: Susceptible $\leq 32 \mu\text{g/mL}$ Resistant $\geq 64 \mu\text{g/mL}$



NARMS

**Figure 19l: Minimum Inhibitory Concentration of Streptomycin
for *Escherichia coli* in Pork Chop (N=232 Isolates)**

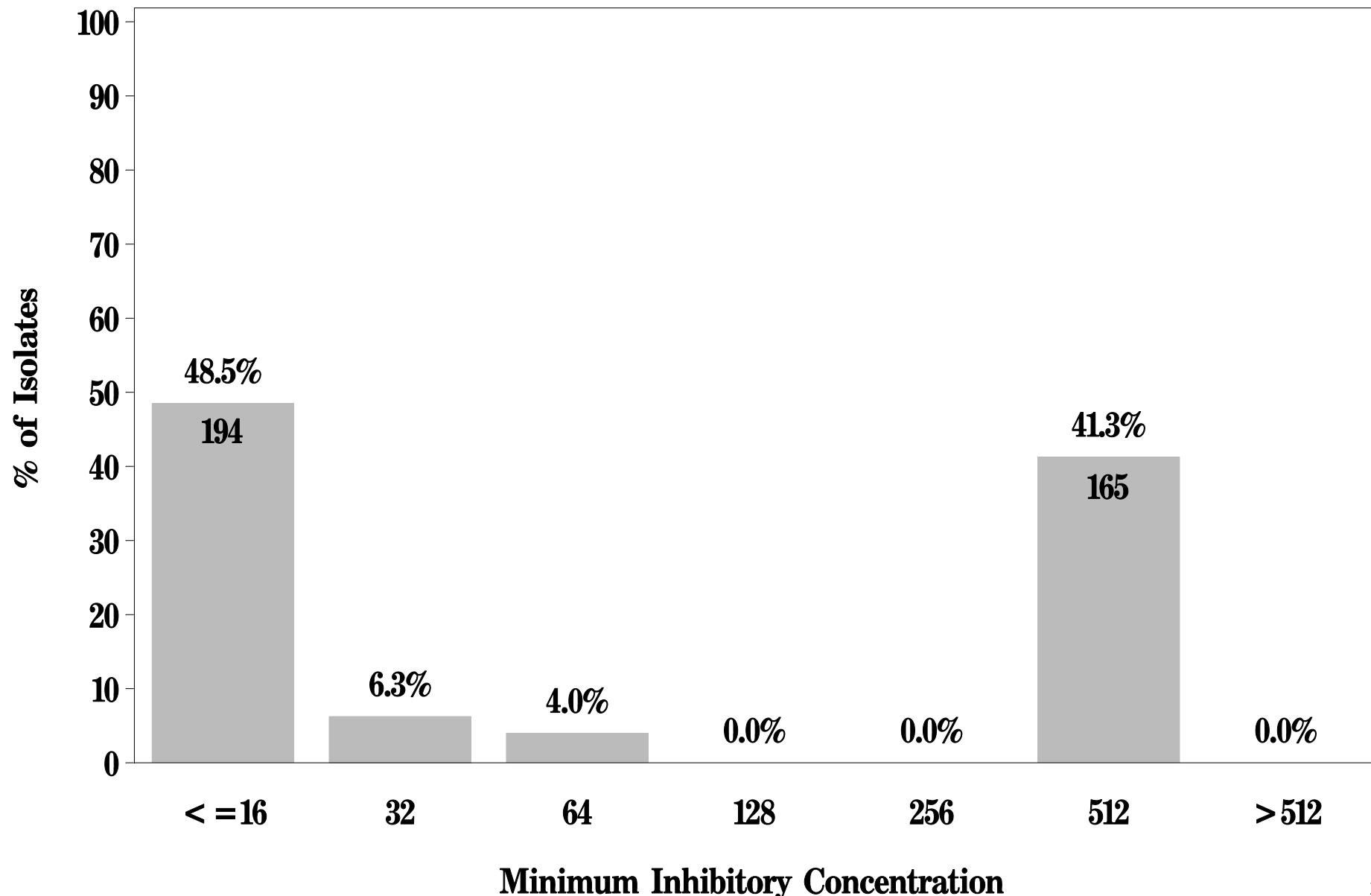
Breakpoints: Susceptible $\leq 32 \mu\text{g/mL}$ Resistant $\geq 64 \mu\text{g/mL}$



NARMS

**Figure 19m: Minimum Inhibitory Concentration of Sulfisoxazole
for *Escherichia coli* in Chicken Breast (N=400 Isolates)**

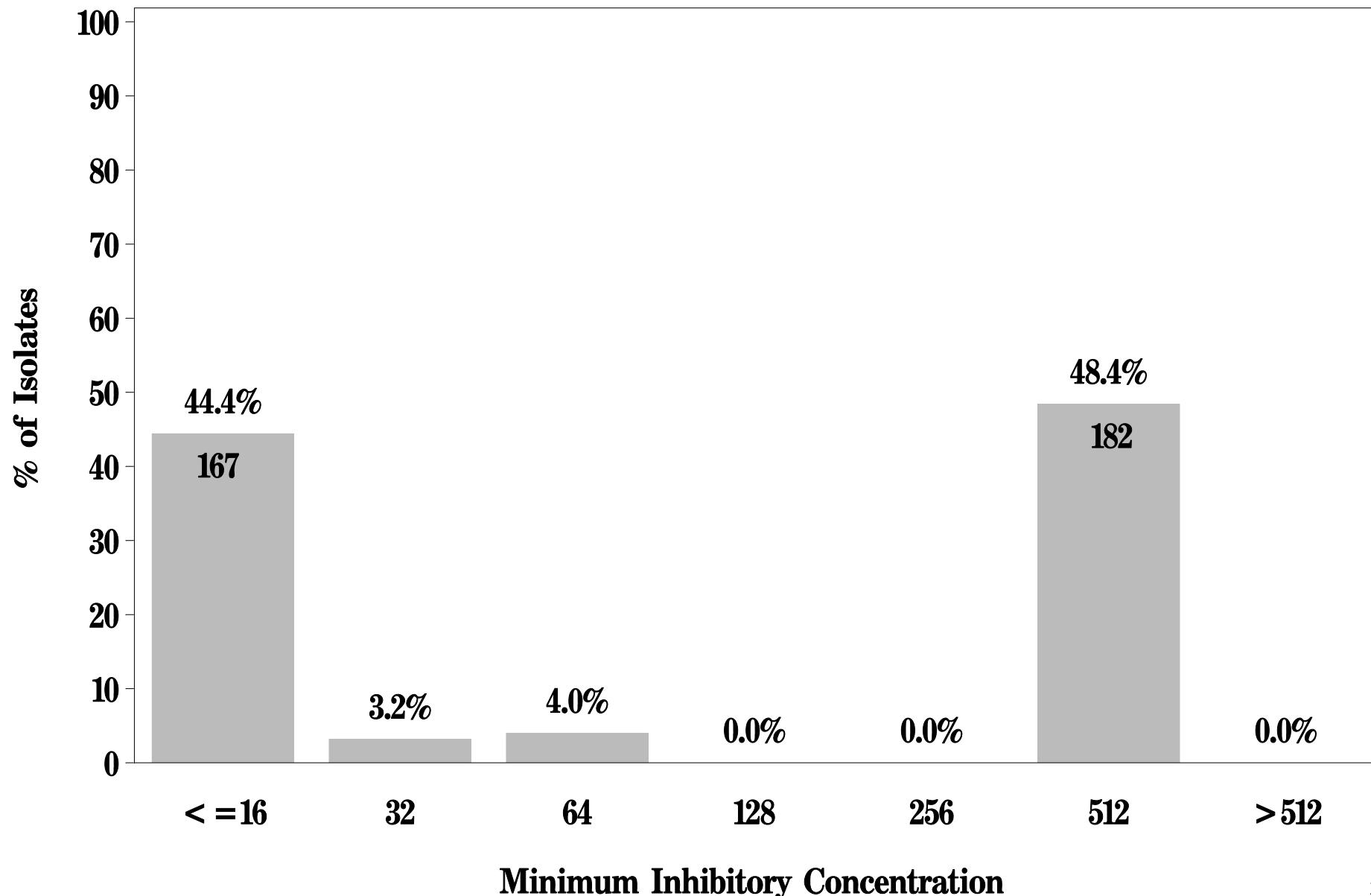
Breakpoints: Susceptible $\leq 256 \mu\text{g/mL}$ Resistant $\geq 512 \mu\text{g/mL}$



NARMS

**Figure 19m: Minimum Inhibitory Concentration of Sulfisoxazole
for *Escherichia coli* in Ground Turkey (N=376 Isolates)**

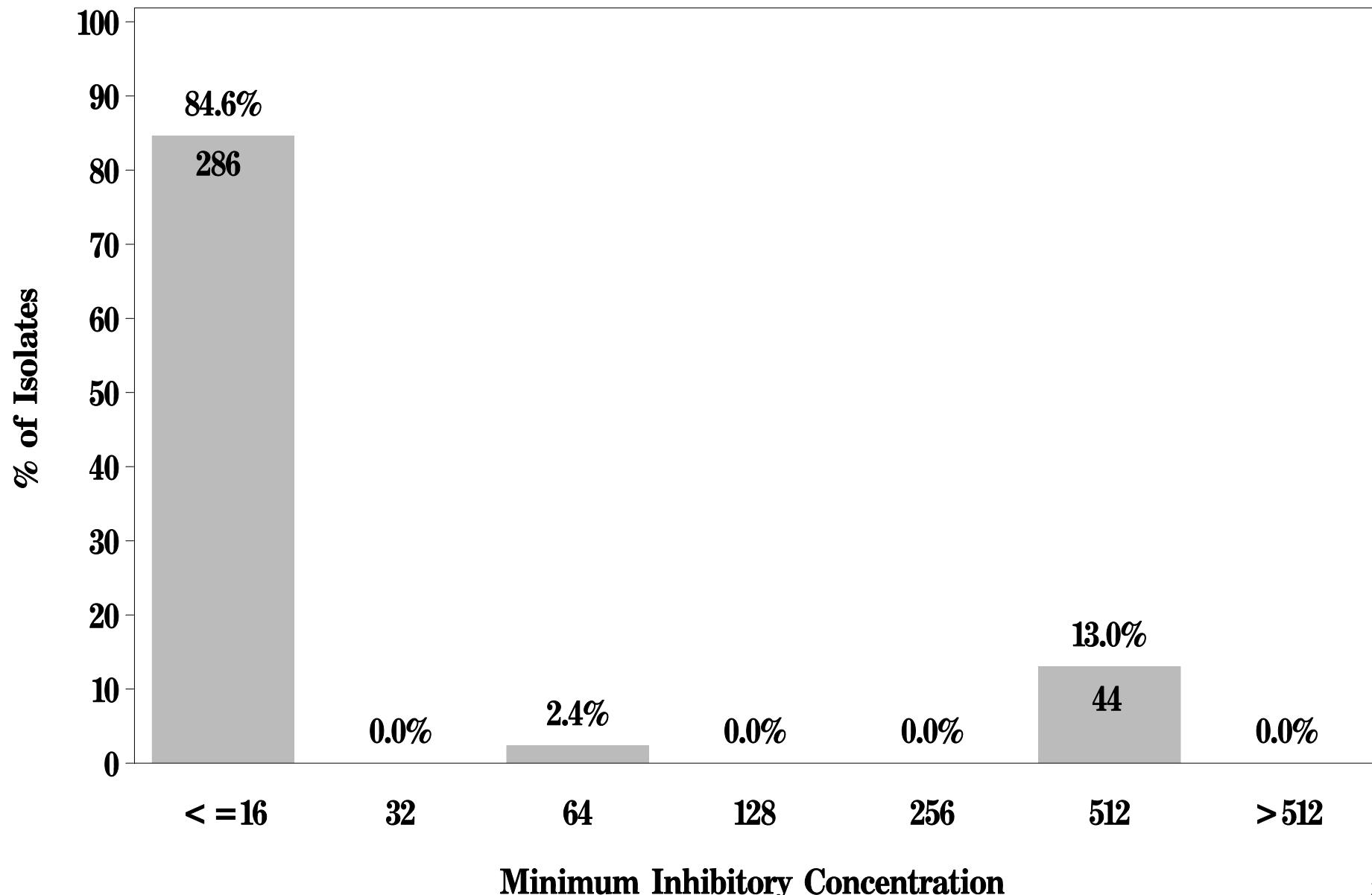
Breakpoints: Susceptible $\leq 256 \mu\text{g/mL}$ Resistant $\geq 512 \mu\text{g/mL}$



NARMS

**Figure 19m: Minimum Inhibitory Concentration of Sulfisoxazole
for *Escherichia coli* in Ground Beef (N=338 Isolates)**

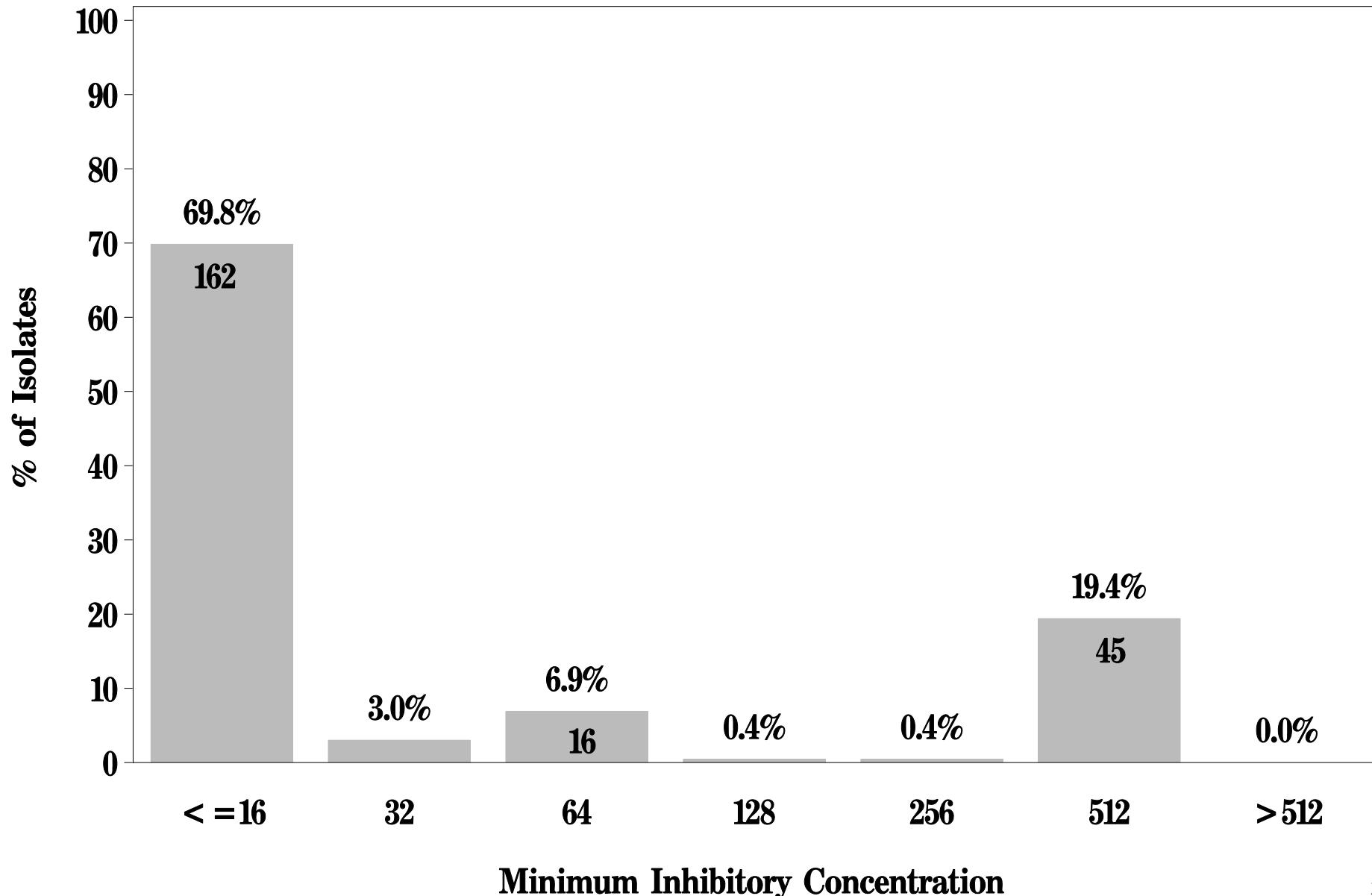
Breakpoints: Susceptible $\leq 256 \mu\text{g/mL}$ Resistant $\geq 512 \mu\text{g/mL}$



NARMS

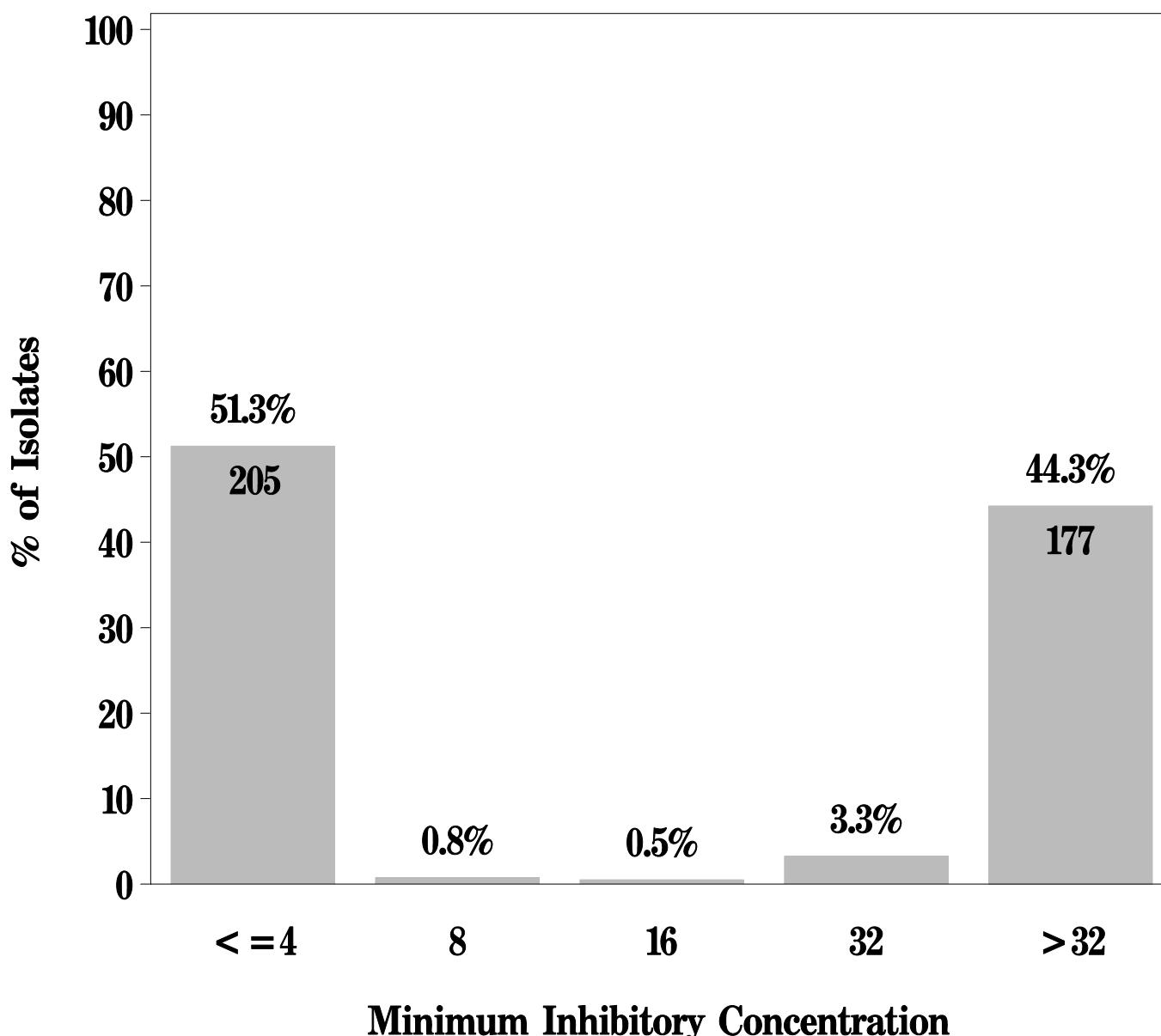
**Figure 19m: Minimum Inhibitory Concentration of Sulfisoxazole
for *Escherichia coli* in Pork Chop (N=232 Isolates)**

Breakpoints: Susceptible $\leq 256 \mu\text{g/mL}$ Resistant $\geq 512 \mu\text{g/mL}$



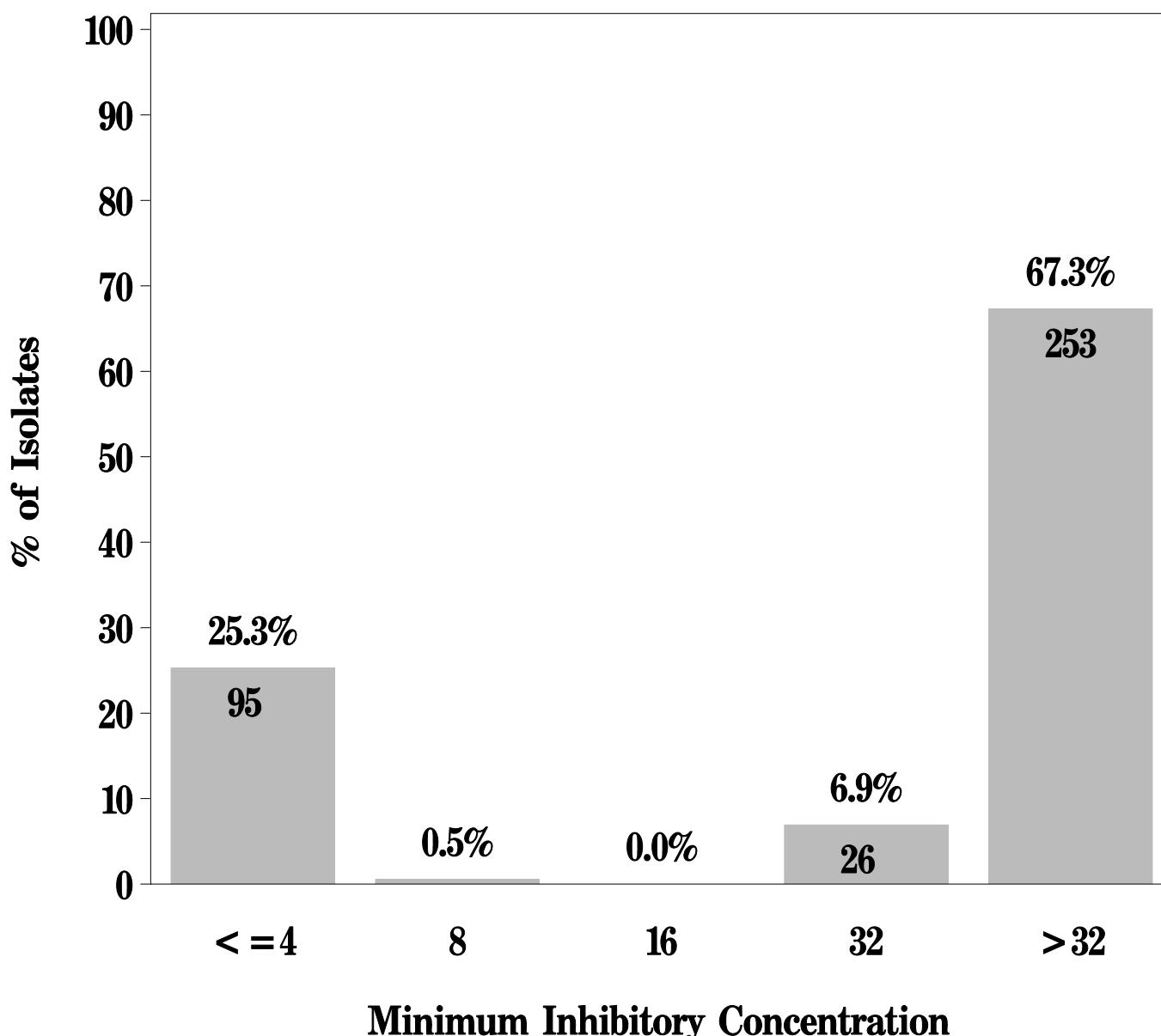
NARMS

**Figure 19n: Minimum Inhibitory Concentration of Tetracycline
for *Escherichia coli* in Chicken Breast (N=400 Isolates)**
Breakpoints: Susceptible $\leq 4 \text{ } \mu\text{g/mL}$ Resistant $\geq 16 \text{ } \mu\text{g/mL}$



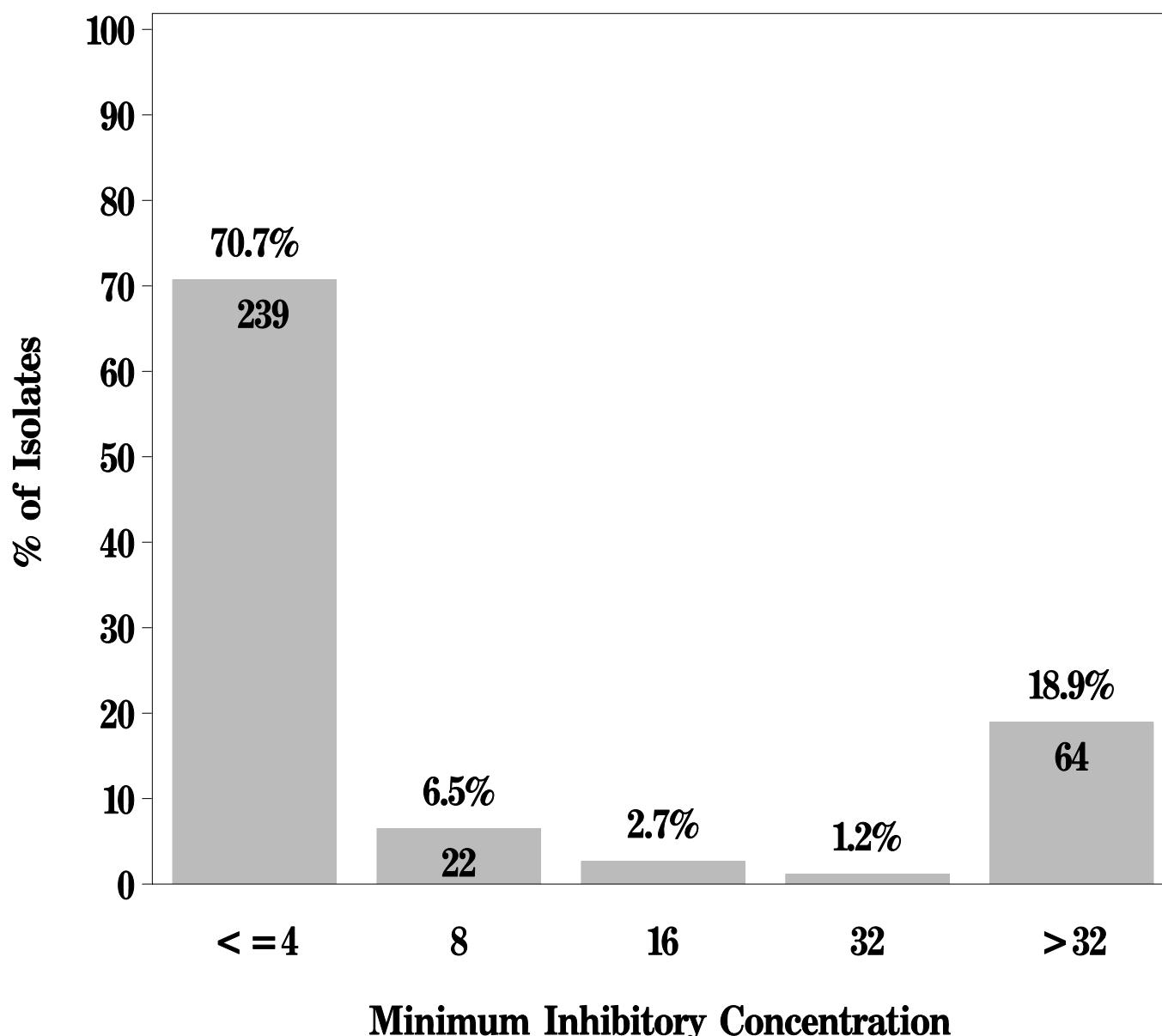
NARMS

**Figure 19n: Minimum Inhibitory Concentration of Tetracycline
for *Escherichia coli* in Ground Turkey (N=376 Isolates)**
Breakpoints: Susceptible $\leq 4 \text{ } \mu\text{g/mL}$ Resistant $\geq 16 \text{ } \mu\text{g/mL}$



NARMS

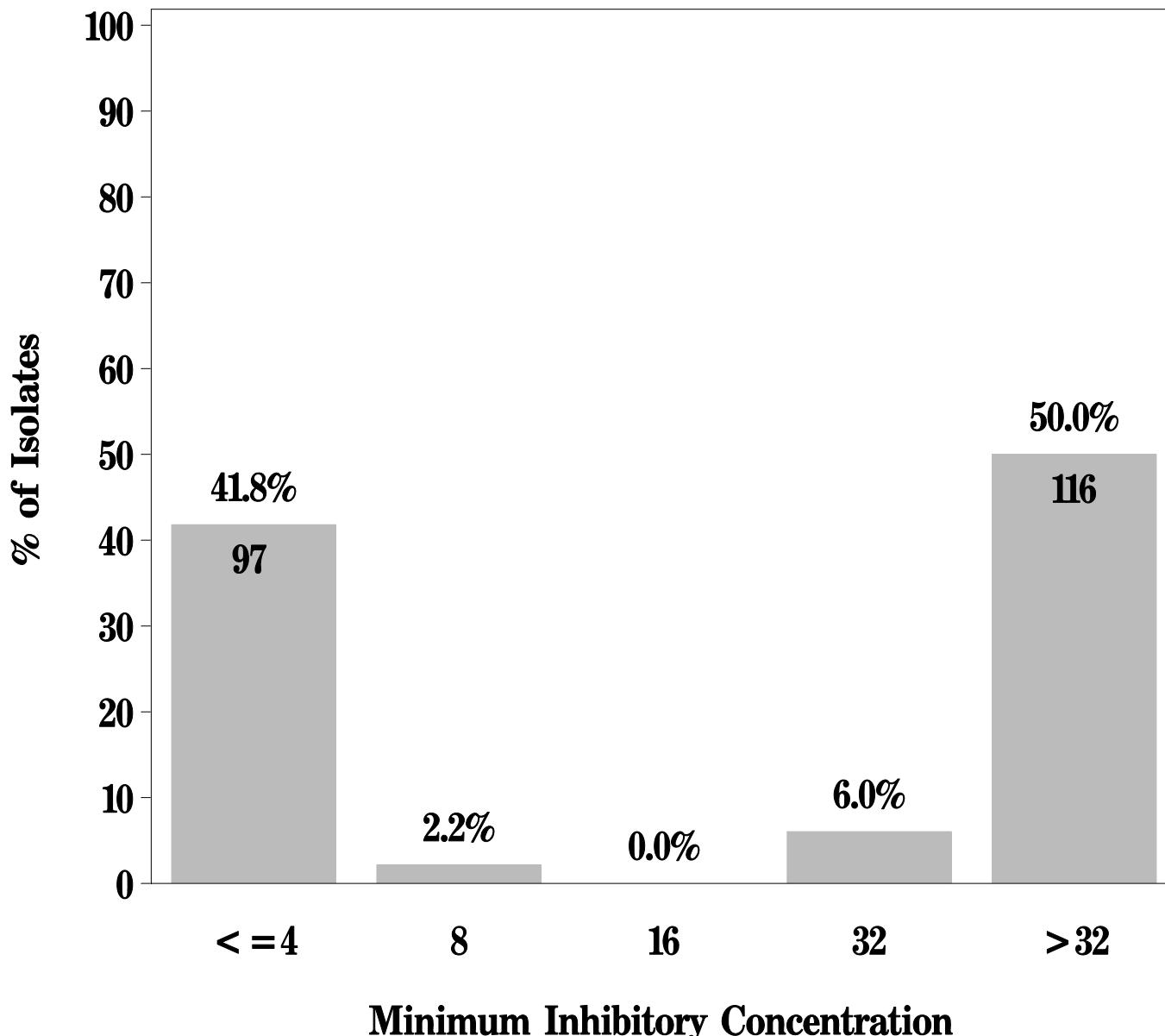
**Figure 19n: Minimum Inhibitory Concentration of Tetracycline
for *Escherichia coli* in Ground Beef (N=338 Isolates)**
Breakpoints: Susceptible $\leq 4 \text{ } \mu\text{g/mL}$ Resistant $\geq 16 \text{ } \mu\text{g/mL}$



NARMS

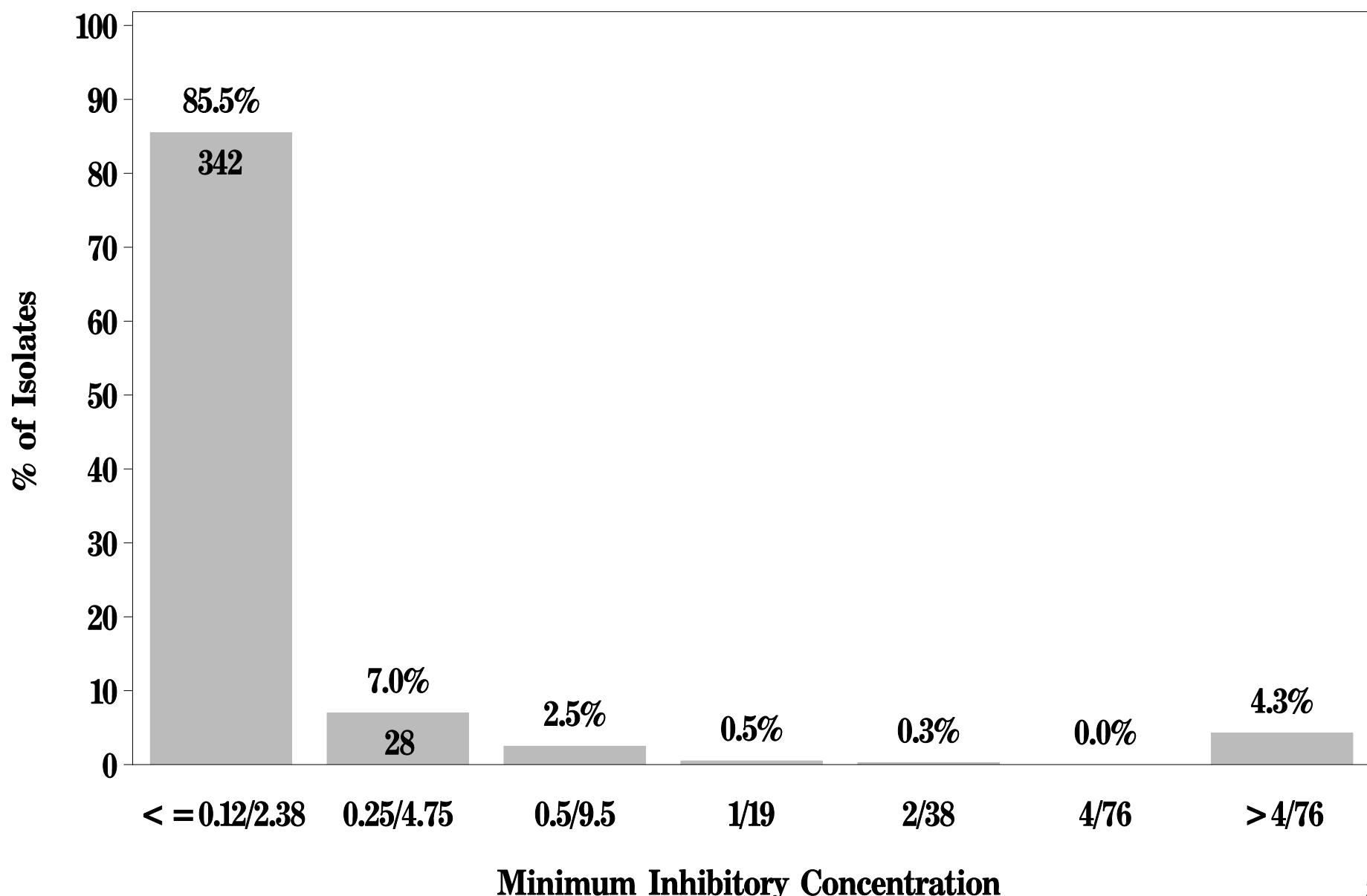
**Figure 19n: Minimum Inhibitory Concentration of Tetracycline
for *Escherichia coli* in Pork Chop (N=232 Isolates)**

Breakpoints: Susceptible $\leq 4 \text{ } \mu\text{g/mL}$ Resistant $\geq 16 \text{ } \mu\text{g/mL}$



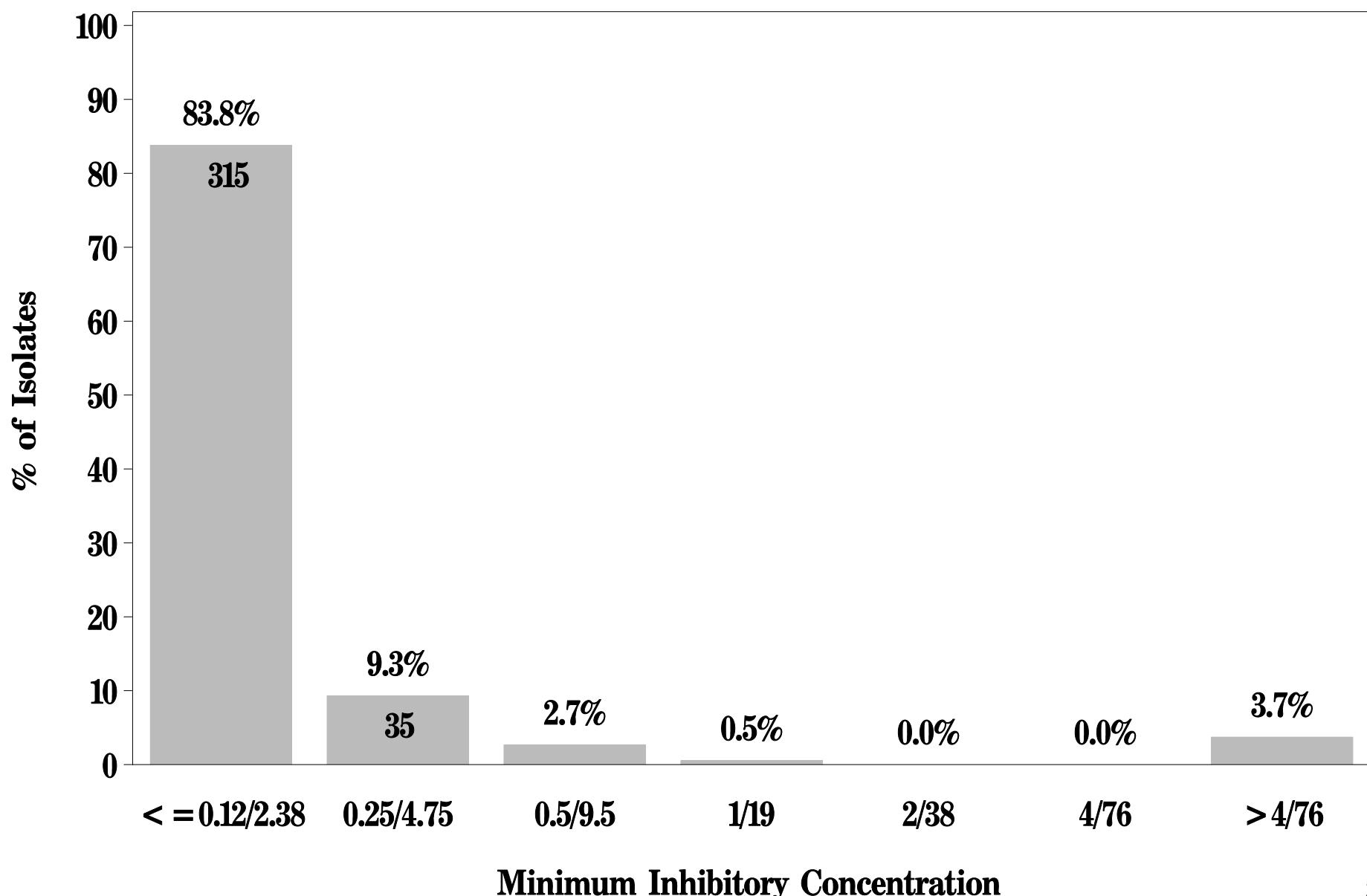
NARMS

Figure 19o: Minimum Inhibitory Concentration of Trimethoprim/sulfamethoxazole for *Escherichia coli* in Chicken Breast (N=400 Isolates)
Breakpoints: Susceptible $\leq 2 \mu\text{g/mL}$ Resistant $\geq 4 \mu\text{g/mL}$



NARMS

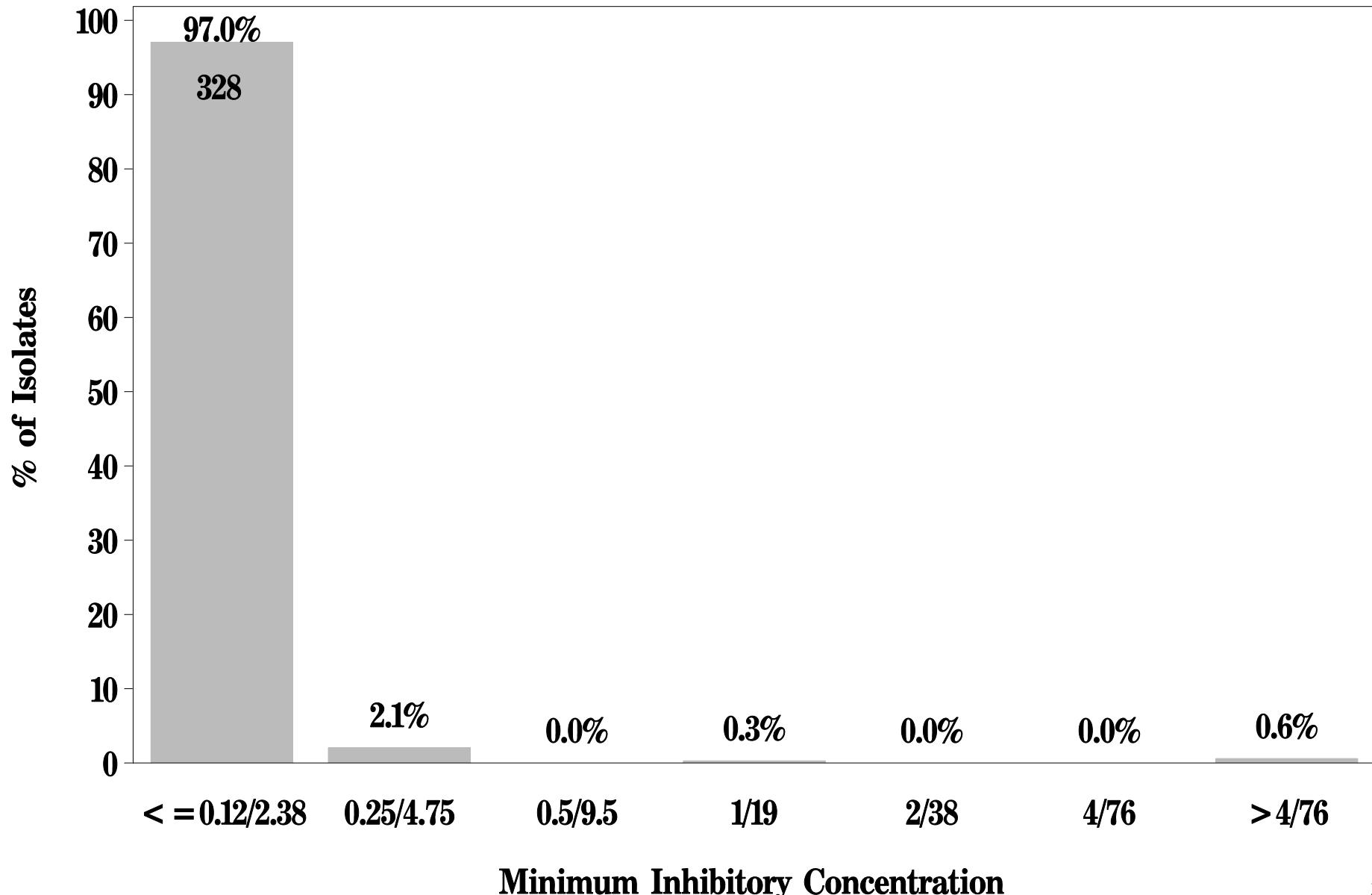
Figure 19o: Minimum Inhibitory Concentration of Trimethoprim/sulfamethoxazole for *Escherichia coli* in Ground Turkey (N=376 Isolates)
Breakpoints: Susceptible $\leq 2 \mu\text{g/mL}$ Resistant $\geq 4 \mu\text{g/mL}$



NARMS

Figure 19o: Minimum Inhibitory Concentration of Trimethoprim/sulfamethoxazole for *Escherichia coli* in Ground Beef (N=338 Isolates)

Breakpoints: Susceptible $\leq 2 \mu\text{g/mL}$ Resistant $\geq 4 \mu\text{g/mL}$



NARMS

Figure 19o: Minimum Inhibitory Concentration of Trimethoprim/sulfamethoxazole for *Escherichia coli* in Pork Chop (N=232 Isolates)

Breakpoints: Susceptible $\leq 2 \mu\text{g/mL}$ Resistant $\geq 4 \mu\text{g/mL}$

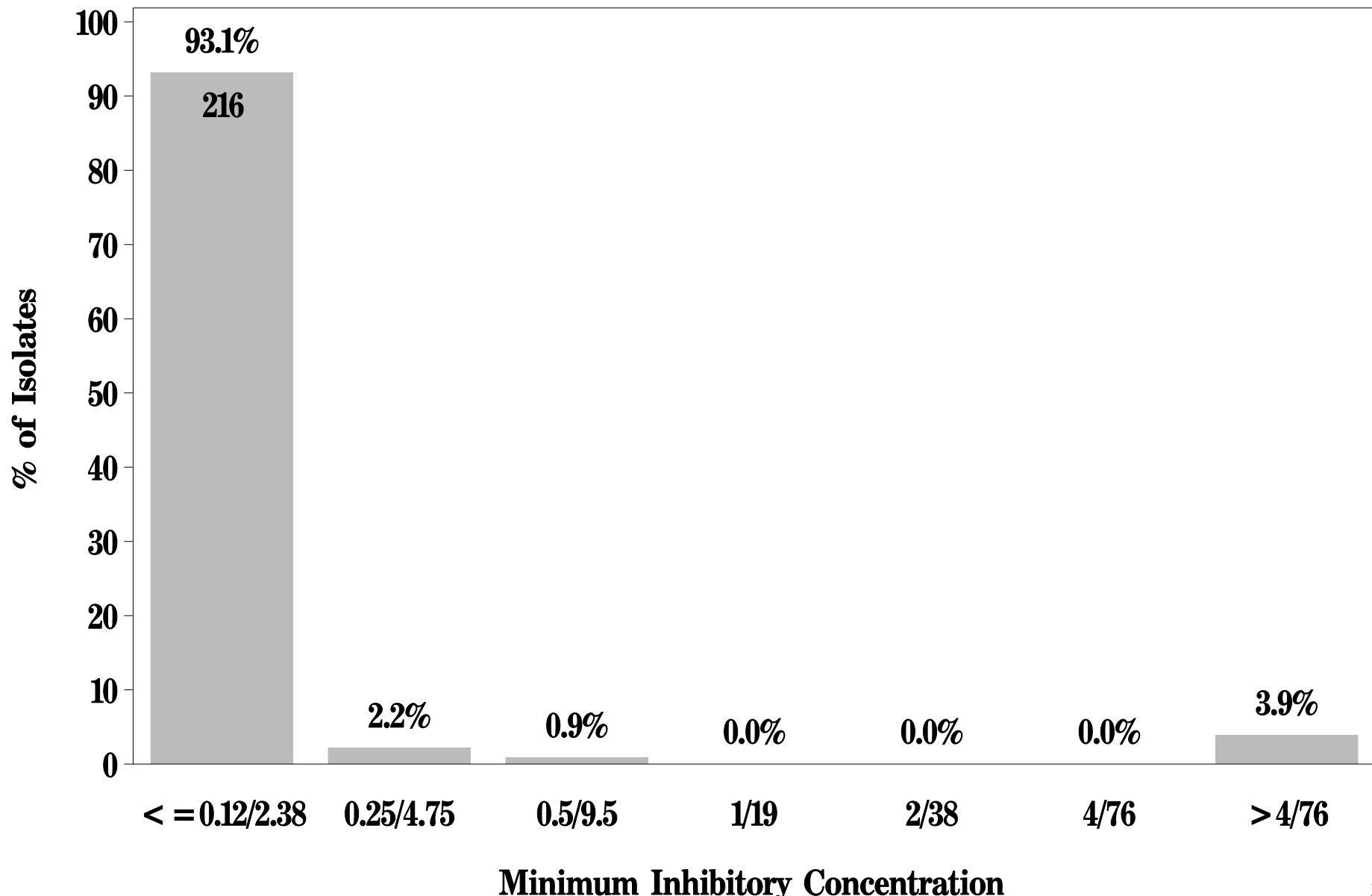


Table 41. Antimicrobial Resistance among *Escherichia coli* by Meat Type, 2004

Meat Type	Antimicrobial Agent														
	TET	STR	FIS	AMP	GEN	KAN	AMC	NAL	FOX	COT	CHL	TIO	CIP	AMI	AXO
Chicken Breast (n=400)	48.0%*	56.8%	41.3%	17.0%	30.0%	6.8%	10.0%	7.0%	8.3%	4.3%	1.8%	5.8%	-†	-	-
Ground Turkey (n=376)	74.2%	49.2%	48.4%	33.2%	29.3%	16.0%	5.3%	10.6%	4.5%	3.7%	0.8%	1.1%	0.8%	-	-
Ground Beef (n=338)	22.8%	11.8%	13.0%	5.3%	0.6%	2.4%	3.8%	1.5%	1.2%	0.6%	3.6%	0.9%	-	-	-
Pork Chop (n=232)	56.0%	21.1%	19.4%	15.1%	1.3%	8.2%	5.6%	-	2.2%	3.9%	4.3%	0.4%	-	-	-
Total (N=1346)	50.4%	37.2%	32.4%	18.3%	17.5%	8.5%	6.4%	5.4%	4.4%	3.1%	2.4%	2.3%	0.2%	-	-

* Where % Resistance = (# *E. coli* isolates resistant to antimicrobial) / (total # *E. coli* isolates).

† Dashes indicate 0.0% resistance to antimicrobial.

Table 42. Antimicrobial Resistance among *Escherichia coli* by Site, Meat Type, and Antimicrobial Agent, 2004

Site	Meat Type	Antimicrobial Agent														
		TET	STR	FIS	AMP	GEN	KAN	AMC	NAL	FOX	COT	CHL	TIO	CIP	AMI	AXO
GA	CB (n=115)	44.3%*	50.4%	54.8%	12.2%	46.1%	5.2%	7.8%	4.3%	6.1%	5.2%	0.9%	4.3%	-†	-	-
	GT (n=119)	74.8%	54.6%	46.2%	36.1%	29.4%	15.1%	3.4%	6.7%	3.4%	-	0.8%	0.8%	-	-	-
	GB (n=91)	18.7%	13.2%	14.3%	6.6%	1.1%	6.6%	4.4%	-	3.3%	-	2.2%	2.2%	-	-	-
	PC (n=64)	46.9%	12.5%	10.9%	10.9%	-	10.9%	-	-	1.6%	3.1%	1.6%	-	-	-	-
	Total (n=389)	48.1%	36.8%	35.5%	18.0%	22.9%	9.5%	4.4%	3.3%	3.9%	2.1%	1.3%	2.1%	-	-	-
MD	CB (n=110)	39.1%	58.2%	32.7%	20.0%	20.0%	3.6%	15.5%	10.9%	13.6%	3.6%	-	7.3%	-	-	-
	GT (n=109)	69.7%	40.4%	43.1%	36.7%	27.5%	14.7%	7.3%	16.5%	4.6%	5.5%	-	-	0.9%	-	-
	GB (n=83)	21.7%	10.8%	8.4%	2.4%	-	-	1.2%	1.2%	-	-	2.4%	-	-	-	-
	PC (n=62)	54.8%	14.5%	11.3%	17.7%	3.2%	4.8%	6.5%	-	1.6%	1.6%	4.8%	-	-	-	-
	Total (n=364)	47.0%	34.6%	26.6%	20.6%	14.8%	6.3%	8.2%	8.5%	5.8%	3.0%	1.4%	2.2%	0.3%	-	-
OR	CB (n=73)	57.5%	54.8%	30.1%	20.5%	15.1%	5.5%	9.6%	8.2%	6.8%	4.1%	4.1%	5.5%	-	-	-
	GT (n=53)	73.6%	49.1%	41.5%	17.0%	15.1%	18.9%	1.9%	11.3%	1.9%	3.8%	1.9%	-	-	-	-
	GB (n=99)	25.3%	11.1%	15.2%	7.1%	1.0%	1.0%	7.1%	-	1.0%	1.0%	8.1%	1.0%	-	-	-
	PC (n=51)	64.7%	35.3%	29.4%	25.5%	-	5.9%	5.9%	-	3.9%	7.8%	5.9%	2.0%	-	-	-
	Total (n=276)	50.4%	34.4%	26.8%	15.9%	7.2%	6.5%	6.5%	4.3%	3.3%	3.6%	5.4%	2.2%	-	-	-
TN	CB (n=102)	54.9%	63.7%	43.1%	16.7%	33.3%	12.7%	6.9%	4.9%	5.9%	3.9%	2.9%	5.9%	-	-	-
	GT (n=95)	78.9%	52.6%	61.1%	34.7%	38.9%	16.8%	7.4%	8.4%	7.4%	6.3%	1.1%	3.2%	2.1%	-	-
	GB (n=65)	26.2%	12.3%	13.8%	4.6%	-	1.5%	1.5%	6.2%	-	1.5%	-	-	-	-	-
	PC (n=55)	60.0%	25.5%	29.1%	7.3%	1.8%	10.9%	10.9%	-	1.8%	3.6%	5.5%	-	-	-	-
	Total (n=317)	57.1%	43.2%	40.1%	18.0%	22.7%	11.4%	6.6%	5.4%	4.4%	4.1%	2.2%	2.8%	0.6%	-	-
Total (N=1346)		50.4%	37.2%	32.4%	18.3%	17.5%	8.5%	6.4%	5.4%	4.4%	3.1%	2.4%	2.3%	0.2%	-	-

* Where % Resistance = (# isolates resistant to antimicrobial per meat type per site) / (total # isolates per meat type per site).

† Dashes indicate 0.0% resistance to antimicrobial.

Table 43. Number of *Escherichia coli* Resistant to Multiple Antimicrobial Agents, 2004

Meat Type	Number of Antimicrobials				
	0	1	2-4	5-7	<u>≥8</u>
Chicken Breast	86	97	190	23	4
Ground Turkey	74	61	212	24	5
Ground Beef	249	45	36	7	1
Pork Chop	90	72	64	6	0
Total	499	275	502	60	10

Appendix A-1. Number of Samples Tested by Site, Meat Type, and Month, 2004

Site: CA

Meat Type	Jan	Feb	Mar	Apr	May	Jun	Jul	Aug	Sep	Oct	Nov	Dec	Total
Chicken Breast	10	10	10	10	10	10	10	10	10	10	10	10	120
Ground Turkey	10	10	10	10	10	10	10	10	10	10	10	10	120
Ground Beef	10	10	10	10	10	10	10	10	10	10	10	10	120
Pork Chop	10	10	10	10	10	10	10	10	10	10	10	10	120
Total	40	480											

Site: CO

Meat Type	Jan	Feb	Mar	Apr	May	Jun	Jul	Aug	Sep	Oct	Nov	Dec	Total
Chicken Breast	1	6	7	6	7	10	10	10	10	10	10	10	97
Ground Turkey	5	4	9	6	7	10	10	10	10	10	10	10	101
Ground Beef	8	6	9	6	7	10	10	10	10	10	10	10	106
Pork Chop	2	6	8	6	7	10	10	10	10	10	10	10	99
Total	16	22	33	24	28	40	403						

Site: CT

Meat Type	Jan	Feb	Mar	Apr	May	Jun	Jul	Aug	Sep	Oct	Nov	Dec	Total
Chicken Breast	10	10	10	10	10	10	10	10	10	10	10	10	120
Ground Turkey	10	10	10	10	10	10	10	10	10	10	10	10	120
Ground Beef	10	10	10	10	10	10	10	10	10	10	10	10	120
Pork Chop	10	10	10	10	10	10	10	10	10	10	10	10	120
Total	40	480											

Site: GA

Meat Type	Jan	Feb	Mar	Apr	May	Jun	Jul	Aug	Sep	Oct	Nov	Dec	Total
Chicken Breast	10	10	10	10	10	10	10	10	10	10	10	10	120
Ground Turkey	10	10	10	10	10	10	10	10	10	10	10	10	120
Ground Beef	10	10	10	10	10	10	10	10	10	10	10	10	120
Pork Chop	10	10	10	10	10	10	10	10	10	10	10	10	120
Total	40	480											

Site: MD

Meat Type	Jan	Feb	Mar	Apr	May	Jun	Jul	Aug	Sep	Oct	Nov	Dec	Total
Chicken Breast	10	10	10	10	10	10	10	10	10	10	10	10	120
Ground Turkey	10	10	10	10	10	10	10	10	10	10	10	10	120
Ground Beef	10	10	10	10	10	10	10	10	10	10	10	10	120
Pork Chop	10	10	10	10	10	10	10	10	10	10	10	10	120
Total	40	480											

Site: MN

Meat Type	Jan	Feb	Mar	Apr	May	Jun	Jul	Aug	Sep	Oct	Nov	Dec	Total
Chicken Breast	10	10	10	10	10	10	10	10	10	10	10	10	120
Ground Turkey	10	10	10	10	10	10	10	10	10	10	10	10	120
Ground Beef	10	10	10	10	10	10	10	10	10	10	10	10	120
Pork Chop	10	10	10	10	10	10	10	10	10	10	10	10	120
Total:	20	40	480										

Site: NM

Meat Type	Jan	Feb	Mar	Apr	May	Jun	Jul	Aug	Sep	Oct	Nov	Dec	Total
Chicken Breast	9	10	10	10	10	10	10	10	10	10	10	10	119
Ground Turkey	9	10	9	10	10	10	10	10	10	10	10	10	120
Ground Beef	10	10	10	10	10	10	10	10	10	10	10	10	118
Pork Chop	10	10	9	10	10	10	10	10	10	10	10	10	119
Total:	38	40	38	40	476								

Site: NY

Meat Type	Jan	Feb	Mar	Apr	May	Jun	Jul	Aug	Sep	Oct	Nov	Dec	Total
Chicken Breast	10	10	10	10	10	10	10	10	10	10	10	10	120
Ground Turkey	10	10	10	10	10	10	10	10	10	10	10	10	120
Ground Beef	10	10	10	10	10	10	10	10	10	10	10	10	120
Pork Chop	10	10	10	10	10	10	10	10	10	10	10	10	120
Total	40	480											

Site: OR

Meat Type	Jan	Feb	Mar	Apr	May	Jun	Jul	Aug	Sep	Oct	Nov	Dec	Total
Chicken Breast	10	10	10	10	10	10	10	10	10	10	10	10	120
Ground Turkey	10	10	10	10	10	10	10	10	10	10	10	10	120
Ground Beef	10	10	10	10	10	10	10	10	10	10	10	10	120
Pork Chop	10	10	10	10	10	10	10	10	10	10	10	10	120
Total:	40	480											

Site: TN

Meat Type	Jan	Feb	Mar	Apr	May	Jun	Jul	Aug	Sep	Oct	Nov	Dec	Total
Chicken Breast	10	10	10	10	10	10	8	10	10	10	8	10	116
Ground Turkey	10	7	10	10	10	4	10	7	10	8	10	10	106
Ground Beef	10	10	10	10	10	10	10	10	10	10	10	10	120
Pork Chop	10	10	10	10	10	10	10	10	10	10	8	10	118
Total:	40	37	40	40	40	34	38	37	40	38	36	40	460

Total Year:**4699**

Appendix A-2. Percent Positive^{*} Samples by Month, Meat Type, and Bacterium, 2004

Month: January

Meat Type: Chicken Breast

Bacterium	# of Samples	# of Isolates	Positive (%)
<i>Campylobacter</i>	90	60	66.7%
<i>Salmonella</i>	90	16	17.8%
<i>Enterococcus</i>	40	38	95.0%
<i>Escherichia coli</i>	40	36	90.0%

Meat Type: Ground Turkey

Bacterium	# of Samples	# of Isolates	Positive (%)
<i>Campylobacter</i>	94	1	1.1%
<i>Salmonella</i>	94	22	23.4%
<i>Enterococcus</i>	40	39	97.5%
<i>Escherichia coli</i>	40	37	92.5%

Meat Type: Ground Beef

Bacterium	# of Samples	# of Isolates	Positive (%)
<i>Campylobacter</i>	98	0	0.0%
<i>Salmonella</i>	98	5	5.1%
<i>Enterococcus</i>	40	36	90.0%
<i>Escherichia coli</i>	40	24	60.0%

Meat Type: Pork Chop

Bacterium	# of Samples	# of Isolates	Positive (%)
<i>Campylobacter</i>	92	0	0.0%
<i>Salmonella</i>	92	6	6.5%
<i>Enterococcus</i>	40	35	87.5%
<i>Escherichia coli</i>	40	20	50.0%

* Where % Positive = (# isolates / # of samples).

Month: February

Meat Type: Chicken Breast

Bacterium	# of Samples	# of Isolates	Positive (%)
<i>Campylobacter</i>	96	59	61.5%
<i>Salmonella</i>	96	9	9.4%
<i>Enterococcus</i>	40	40	100.0%
<i>Escherichia coli</i>	40	37	92.5%

Meat Type: Ground Turkey

Bacterium	# of Samples	# of Isolates	Positive (%)
<i>Campylobacter</i>	91	0	0.0%
<i>Salmonella</i>	91	8	8.8%
<i>Enterococcus</i>	37	33	89.2%
<i>Escherichia coli</i>	37	25	67.6%

Meat Type: Ground Beef

Bacterium	# of Samples	# of Isolates	Positive (%)
<i>Campylobacter</i>	96	0	0.0%
<i>Salmonella</i>	96	0	0.0%
<i>Enterococcus</i>	40	39	97.5%
<i>Escherichia coli</i>	40	28	70.0%

Meat Type: Pork Chop

Bacterium	# of Samples	# of Isolates	Positive (%)
<i>Campylobacter</i>	96	0	0.0%
<i>Salmonella</i>	96	1	1.0%
<i>Enterococcus</i>	40	32	80.0%
<i>Escherichia coli</i>	40	16	40.0%

Month: March

Meat Type: Chicken Breast

Bacterium	# of Samples	# of Isolates	Positive (%)
<i>Campylobacter</i>	97	47	48.5%
<i>Salmonella</i>	97	18	18.6%
<i>Enterococcus</i>	40	39	97.5%
<i>Escherichia coli</i>	40	37	92.5%

Meat Type: Ground Turkey

Bacterium	# of Samples	# of Isolates	Positive (%)
<i>Campylobacter</i>	98	2	2.0%
<i>Salmonella</i>	98	2	2.0%
<i>Enterococcus</i>	40	34	85.0%
<i>Escherichia coli</i>	40	29	72.5%

Meat Type: Ground Beef

Bacterium	# of Samples	# of Isolates	Positive (%)
<i>Campylobacter</i>	99	0	0.0%
<i>Salmonella</i>	99	0	0.0%
<i>Enterococcus</i>	40	33	82.5%
<i>Escherichia coli</i>	40	26	65.0%

Meat Type: Pork Chop

Bacterium	# of Samples	# of Isolates	Positive (%)
<i>Campylobacter</i>	97	0	0.0%
<i>Salmonella</i>	97	1	1.0%
<i>Enterococcus</i>	40	34	85.0%
<i>Escherichia coli</i>	40	15	37.5%

Month: April

Meat Type: Chicken Breast

Bacterium	# of Samples	# of Isolates	Positive (%)
<i>Campylobacter</i>	96	35	36.5%
<i>Salmonella</i>	96	8	8.3%
<i>Enterococcus</i>	40	40	100.0%
<i>Escherichia coli</i>	40	34	85.0%

Meat Type: Ground Turkey

Bacterium	# of Samples	# of Isolates	Positive (%)
<i>Campylobacter</i>	96	0	0.0%
<i>Salmonella</i>	96	18	18.8%
<i>Enterococcus</i>	40	39	97.5%
<i>Escherichia coli</i>	40	31	77.5%

Meat Type: Ground Beef

Bacterium	# of Samples	# of Isolates	Positive (%)
<i>Campylobacter</i>	96	0	0.0%
<i>Salmonella</i>	96	1	1.0%
<i>Enterococcus</i>	40	39	97.5%
<i>Escherichia coli</i>	40	25	62.5%

Meat Type: Pork Chop

Bacterium	# of Samples	# of Isolates	Positive (%)
<i>Campylobacter</i>	96	0	0.0%
<i>Salmonella</i>	96	1	1.0%
<i>Enterococcus</i>	40	38	95.0%
<i>Escherichia coli</i>	40	25	62.5%

Month: May

Meat Type: Chicken Breast

Bacterium	# of Samples	# of Isolates	Positive (%)
<i>Campylobacter</i>	97	51	52.6%
<i>Salmonella</i>	97	7	7.2%
<i>Enterococcus</i>	40	39	97.5%
<i>Escherichia coli</i>	40	36	90.0%

Meat Type: Ground Turkey

Bacterium	# of Samples	# of Isolates	Positive (%)
<i>Campylobacter</i>	97	0	0.0%
<i>Salmonella</i>	97	17	17.5%
<i>Enterococcus</i>	40	39	97.5%
<i>Escherichia coli</i>	40	36	90.0%

Meat Type: Ground Beef

Bacterium	# of Samples	# of Isolates	Positive (%)
<i>Campylobacter</i>	97	0	0.0%
<i>Salmonella</i>	97	1	1.0%
<i>Enterococcus</i>	40	39	97.5%
<i>Escherichia coli</i>	40	33	82.5%

Meat Type: Pork Chop

Bacterium	# of Samples	# of Isolates	Positive (%)
<i>Campylobacter</i>	97	0	0.0%
<i>Salmonella</i>	97	0	0.0%
<i>Enterococcus</i>	40	36	90.0%
<i>Escherichia coli</i>	40	22	55.0%

Month: June

Meat Type: Chicken Breast

Bacterium	# of Samples	# of Isolates	Positive (%)
<i>Campylobacter</i>	100	59	59.0%
<i>Salmonella</i>	100	12	12.0%
<i>Enterococcus</i>	40	37	92.5%
<i>Escherichia coli</i>	40	34	85.0%

Meat Type: Ground Turkey

Bacterium	# of Samples	# of Isolates	Positive (%)
<i>Campylobacter</i>	94	2	2.1%
<i>Salmonella</i>	94	11	11.7%
<i>Enterococcus</i>	34	34	100.0%
<i>Escherichia coli</i>	34	24	70.6%

Meat Type: Ground Beef

Bacterium	# of Samples	# of Isolates	Positive (%)
<i>Campylobacter</i>	100	0	0.0%
<i>Salmonella</i>	100	0	0.0%
<i>Enterococcus</i>	40	35	87.5%
<i>Escherichia coli</i>	40	25	62.5%

Meat Type: Pork Chop

Bacterium	# of Samples	# of Isolates	Positive (%)
<i>Campylobacter</i>	100	1	1.0
<i>Salmonella</i>	100	0	0.0%
<i>Enterococcus</i>	40	30	75.0%
<i>Escherichia coli</i>	40	13	32.5%

Month: July

Meat Type: Chicken Breast

Bacterium	# of Samples	# of Isolates	Positive (%)
<i>Campylobacter</i>	98	67	68.4%
<i>Salmonella</i>	98	10	10.2%
<i>Enterococcus</i>	38	38	100.0%
<i>Escherichia coli</i>	38	29	76.3%

Meat Type: Ground Turkey

Bacterium	# of Samples	# of Isolates	Positive (%)
<i>Campylobacter</i>	100	1	1.0%
<i>Salmonella</i>	100	17	17.0%
<i>Enterococcus</i>	40	38	95.0%
<i>Escherichia coli</i>	40	33	82.5%

Meat Type: Ground Beef

Bacterium	# of Samples	# of Isolates	Positive (%)
<i>Campylobacter</i>	100	0	0.0%
<i>Salmonella</i>	100	5	5.0%
<i>Enterococcus</i>	40	36	90.0%
<i>Escherichia coli</i>	40	24	60.0%

Meat Type: Pork Chop

Bacterium	# of Samples	# of Isolates	Positive (%)
<i>Campylobacter</i>	100	0	0.0%
<i>Salmonella</i>	100	0	0.0%
<i>Enterococcus</i>	40	23	57.5%
<i>Escherichia coli</i>	40	21	52.5%

Month: August

Meat Type: Chicken Breast

Bacterium	# of Samples	# of Isolates	Positive (%)
<i>Campylobacter</i>	100	62	62.0%
<i>Salmonella</i>	100	16	16.0%
<i>Enterococcus</i>	40	40	100.0%
<i>Escherichia coli</i>	40	31	77.5%

Meat Type: Ground Turkey

Bacterium	# of Samples	# of Isolates	Positive (%)
<i>Campylobacter</i>	97	0	0.0%
<i>Salmonella</i>	97	17	17.5%
<i>Enterococcus</i>	37	33	89.2%
<i>Escherichia coli</i>	37	29	78.4%

Meat Type: Ground Beef

Bacterium	# of Samples	# of Isolates	Positive (%)
<i>Campylobacter</i>	100	0	0.0%
<i>Salmonella</i>	100	1	1.0%
<i>Enterococcus</i>	40	39	97.5%
<i>Escherichia coli</i>	40	35	87.5%

Meat Type: Pork Chop

Bacterium	# of Samples	# of Isolates	Positive (%)
<i>Campylobacter</i>	100	0	0.0%
<i>Salmonella</i>	100	0	0.0%
<i>Enterococcus</i>	40	36	90.0%
<i>Escherichia coli</i>	40	22	55.0%

Month: September

Meat Type: Chicken Breast

Bacterium	# of Samples	# of Isolates	Positive (%)
<i>Campylobacter</i>	100	72	72.0%
<i>Salmonella</i>	100	14	14.0%
<i>Enterococcus</i>	40	40	100.0%
<i>Escherichia coli</i>	40	31	77.5%

Meat Type: Ground Turkey

Bacterium	# of Samples	# of Isolates	Positive (%)
<i>Campylobacter</i>	100	1	1.0%
<i>Salmonella</i>	100	5	5.0%
<i>Enterococcus</i>	40	38	95.0%
<i>Escherichia coli</i>	40	31	77.5%

Meat Type: Ground Beef

Bacterium	# of Samples	# of Isolates	Positive (%)
<i>Campylobacter</i>	100	0	0.0%
<i>Salmonella</i>	100	0	0.0%
<i>Enterococcus</i>	40	39	97.5%
<i>Escherichia coli</i>	40	29	72.5%

Meat Type: Pork Chop

Bacterium	# of Samples	# of Isolates	Positive (%)
<i>Campylobacter</i>	100	0	0.0%
<i>Salmonella</i>	100	0	0.0%
<i>Enterococcus</i>	40	37	92.5%
<i>Escherichia coli</i>	40	20	50.0%

Month: October

Meat Type: Chicken Breast

Bacterium	# of Samples	# of Isolates	Positive (%)
<i>Campylobacter</i>	100	73	73.0%
<i>Salmonella</i>	100	20	20.0%
<i>Enterococcus</i>	40	39	97.5%
<i>Escherichia coli</i>	40	32	80.0%

Meat Type: Ground Turkey

Bacterium	# of Samples	# of Isolates	Positive (%)
<i>Campylobacter</i>	98	0	0.0%
<i>Salmonella</i>	98	4	4.1%
<i>Enterococcus</i>	38	34	89.5%
<i>Escherichia coli</i>	38	31	81.6%

Meat Type: Ground Beef

Bacterium	# of Samples	# of Isolates	Positive (%)
<i>Campylobacter</i>	100	0	0.0%
<i>Salmonella</i>	100	0	0.0%
<i>Enterococcus</i>	40	36	90.0%
<i>Escherichia coli</i>	40	33	82.5%

Meat Type: Pork Chop

Bacterium	# of Samples	# of Isolates	Positive (%)
<i>Campylobacter</i>	100	1	1.0%
<i>Salmonella</i>	100	0	0.0%
<i>Enterococcus</i>	40	32	80.0%
<i>Escherichia coli</i>	40	22	55.0%

Month: November

Meat Type: Chicken Breast

Bacterium	# of Samples	# of Isolates	Positive (%)
<i>Campylobacter</i>	98	58	59.2%
<i>Salmonella</i>	98	13	13.3%
<i>Enterococcus</i>	38	37	97.4%
<i>Escherichia coli</i>	38	29	76.3%

Meat Type: Ground Turkey

Bacterium	# of Samples	# of Isolates	Positive (%)
<i>Campylobacter</i>	100	5	5.0%
<i>Salmonella</i>	100	13	13.0%
<i>Enterococcus</i>	40	37	92.5%
<i>Escherichia coli</i>	40	34	85.0%

Meat Type: Ground Beef

Bacterium	# of Samples	# of Isolates	Positive (%)
<i>Campylobacter</i>	100	0	0.0%
<i>Salmonella</i>	100	0	0.0%
<i>Enterococcus</i>	40	40	100.0%
<i>Escherichia coli</i>	40	31	77.5%

Meat Type: Pork Chop

Bacterium	# of Samples	# of Isolates	Positive (%)
<i>Campylobacter</i>	98	1	1.0%
<i>Salmonella</i>	98	2	2.0%
<i>Enterococcus</i>	38	34	89.5%
<i>Escherichia coli</i>	38	19	50.0%

Month: December

Meat Type: Chicken Breast

Bacterium	# of Samples	# of Isolates	Positive (%)
<i>Campylobacter</i>	73	52	71.2%
<i>Salmonella</i>	73	4	5.5%
<i>Enterococcus</i>	38	38	100.0%
<i>Escherichia coli</i>	38	37	97.4%

Meat Type: Ground Turkey

Bacterium	# of Samples	# of Isolates	Positive (%)
<i>Campylobacter</i>	100	0	0.0%
<i>Salmonella</i>	100	8	8.0%
<i>Enterococcus</i>	40	39	97.5%
<i>Escherichia coli</i>	40	36	90.0%

Meat Type: Ground Beef

Bacterium	# of Samples	# of Isolates	Positive (%)
<i>Campylobacter</i>	100	0	0.0%
<i>Salmonella</i>	100	1	1.0%
<i>Enterococcus</i>	40	37	92.5%
<i>Escherichia coli</i>	40	25	62.5%

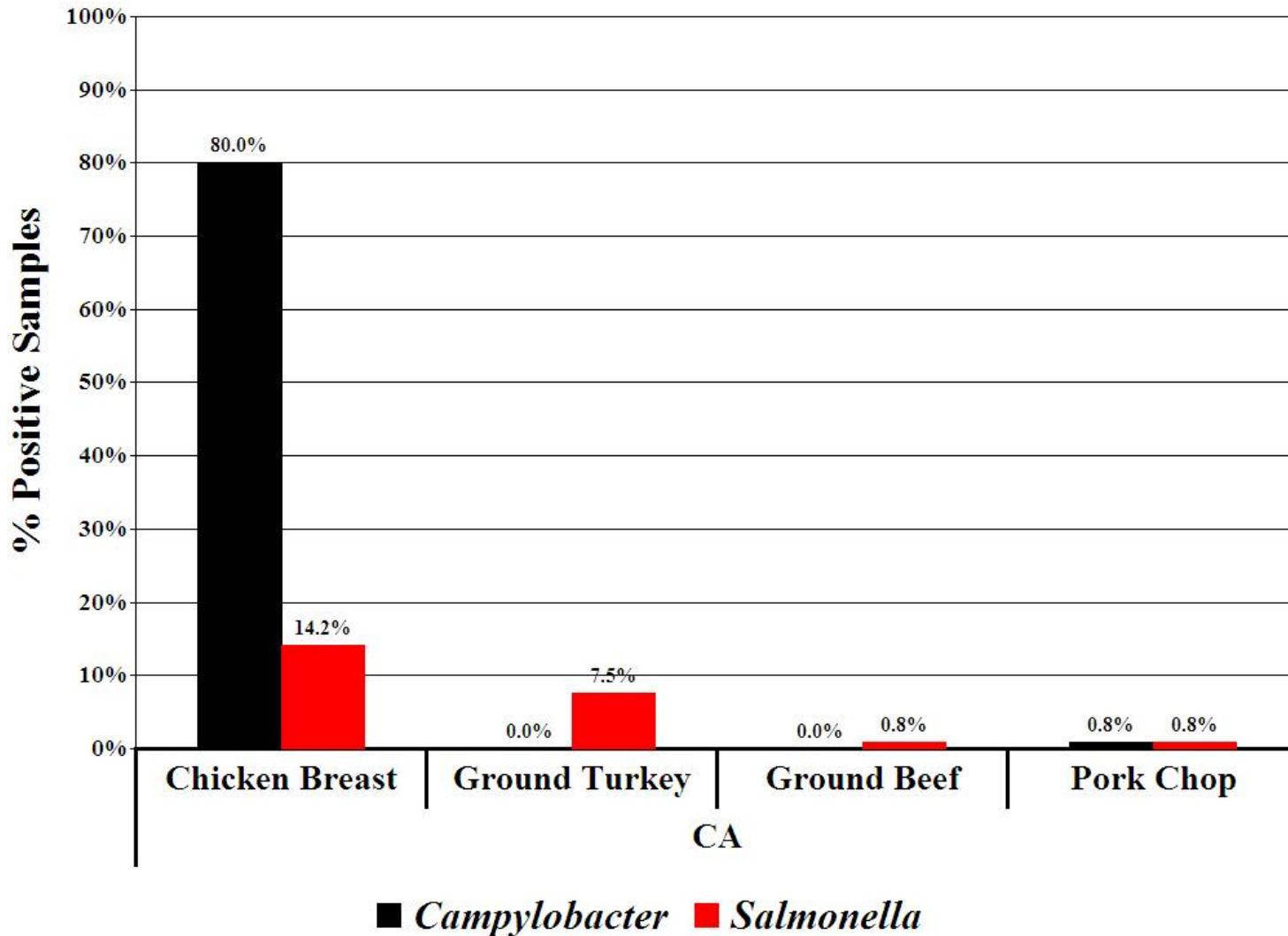
Meat Type: Pork Chop

Bacterium	# of Samples	# of Isolates	Positive (%)
<i>Campylobacter</i>	100	0	0.0%
<i>Salmonella</i>	100	0	0.0%
<i>Enterococcus</i>	40	37	92.5%
<i>Escherichia coli</i>	40	17	42.5%

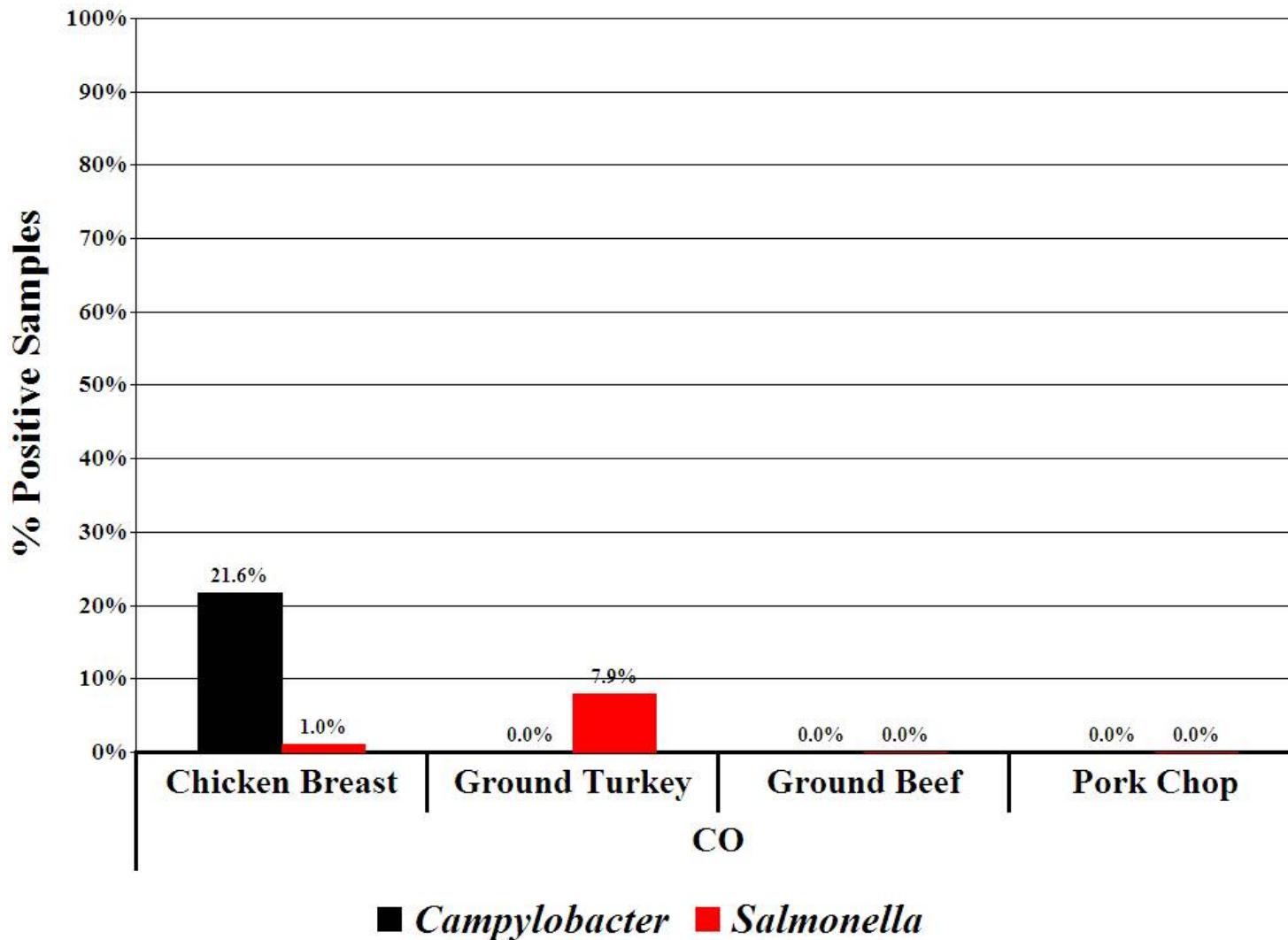
Appendix A-3. Percent Positive Samples by Meat Type, Bacterium, and Site, 2004

Meat Type	Site	<i>Campylobacter</i>			<i>Salmonella</i>			<i>Enterococcus</i>			<i>Escherichia coli</i>		
		N	Isolate	%Positive	N	Isolate	%Positive	N	Isolate	%Positive	N	Isolate	%Positive
Chicken Breast	CA	120	96	80.0%	120	17	14.2%						
	CO	97	21	21.6%	97	1	1.0%						
	CT	120	86	71.7%	120	30	25.0%						
	GA	120	61	50.8%	120	6	5.0%	120	120	100.0%	120	115	95.8%
	MD	120	76	63.3%	120	24	20.0%	120	114	95.0%	120	110	91.7%
	MN	120	73	60.8%	120	20	16.7%						
	NM	119	53	44.5%	119	3	2.5%						
	NY	120	96	80.0%	120	16	13.3%						
	OR	120	73	60.8%	120	25	20.8%	120	118	98.3%	120	73	60.8%
	TN	116	71	61.2%	116	15	12.9%	116	114	98.3%	116	102	87.9%
Total		1172	706	60.2%	1172	157	13.4%	476	466	97.9%	476	400	84.0%
Ground Turkey	CA	120	0	-	120	9	7.5%						
	CO	101	0	-	101	8	7.9%						
	CT	120	2	1.7%	120	26	21.7%						
	GA	120	1	0.8%	120	38	31.7%	120	120	100.0%	120	119	99.2%
	MD	120	2	1.7%	120	13	10.8%	120	106	88.3%	120	109	90.8%
	MN	120	6	5.0%	120	14	11.7%						
	NM	118	0	-	118	9	7.6%						
	NY	120	0	-	120	11	9.2%						
	OR	120	0	-	120	6	5.0%	120	105	87.5%	120	53	44.2%
	TN	106	1	0.9%	106	8	7.5%	106	106	100.0%	106	95	89.6%
Total		1165	12	1.0%	1165	142	12.2%	466	437	93.8%	466	376	80.7%
Ground Beef	CA	120	0	-	120	1	0.8%						
	CO	106	0	-	106	0	-						
	CT	120	0	-	120	5	4.2%						
	GA	120	0	-	120	1	0.8%	120	117	97.5%	120	91	75.8%
	MD	120	0	-	120	1	0.8%	120	100	83.3%	120	83	69.2%
	MN	120	0	-	120	0	-						
	NM	120	0	-	120	0	-						
	NY	120	0	-	120	0	-						
	OR	120	0	-	120	6	5.0%	120	115	95.8%	120	99	82.5%
	TN	120	0	-	120	0	-	120	116	96.7%	120	65	54.2%
Total		1186	0	-	1186	14	1.2%	480	448	93.3%	480	338	70.4%
Pork Chop	CA	120	1	0.8%	120	1	0.8%						
	CO	99	0	-	99	0	-						
	CT	120	1	0.8%	120	5	4.2%						
	GA	120	0	-	120	0	-	120	116	96.7%	120	64	53.3%
	MD	120	0	-	120	0	-	120	77	64.2%	120	62	51.7%
	MN	120	0	-	120	0	-						
	NM	119	1	0.8%	119	0	-						
	NY	120	0	-	120	3	2.5%						
	OR	120	0	-	120	2	1.7%	120	108	90.0%	120	51	42.5%
	TN	118	0	-	118	0	-	118	103	87.3%	118	55	46.6%
Total		1176	3	0.3%	1176	11	0.9%	478	404	84.5%	478	232	48.5%
Total		4699	721	15.3%	4699	324	6.9%	1900	1755	92.4%	1900	1346	70.8%

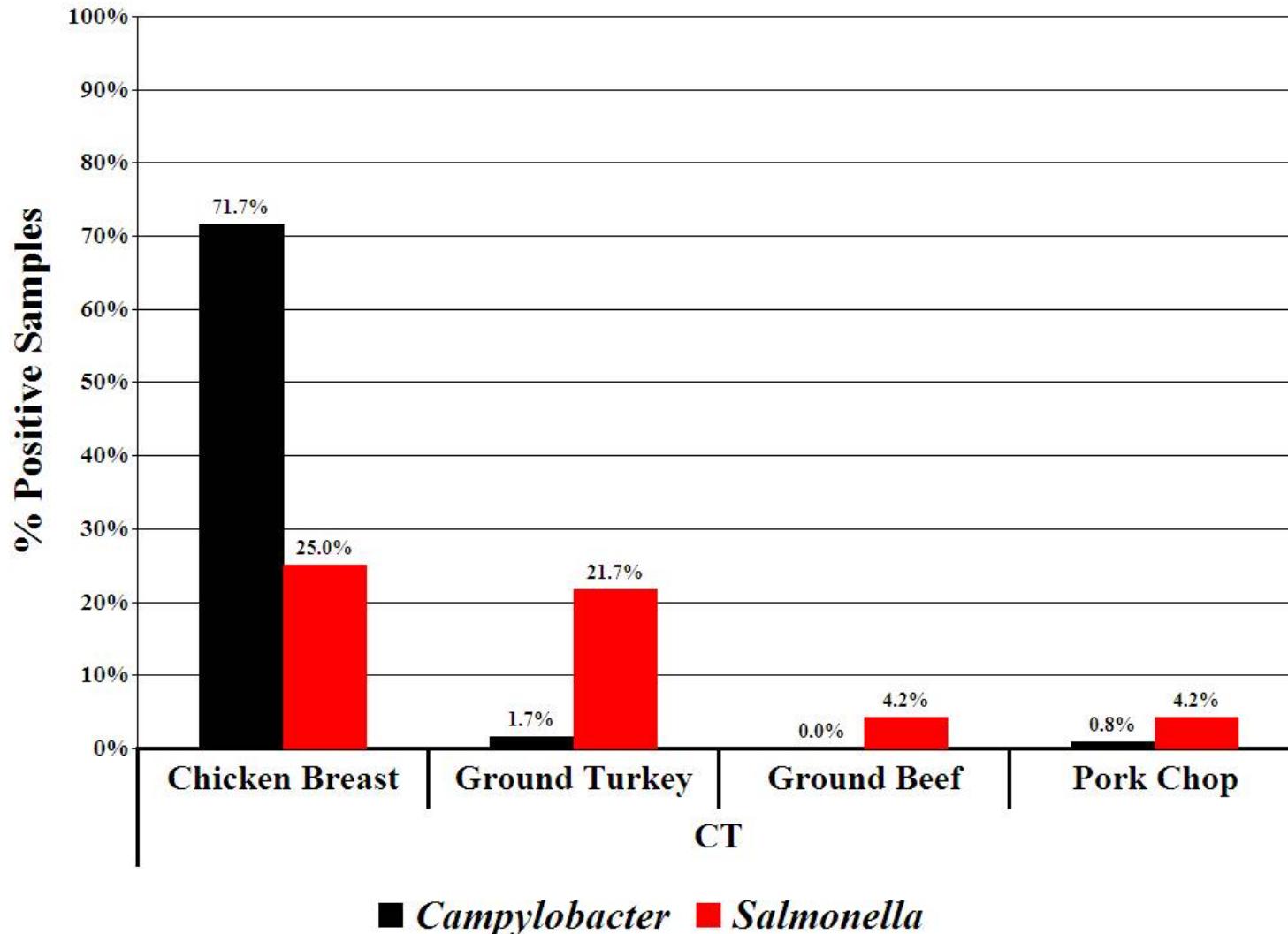
Appendix 3a. Percent Positive Samples by Meat Type, Bacterium in California, 2004



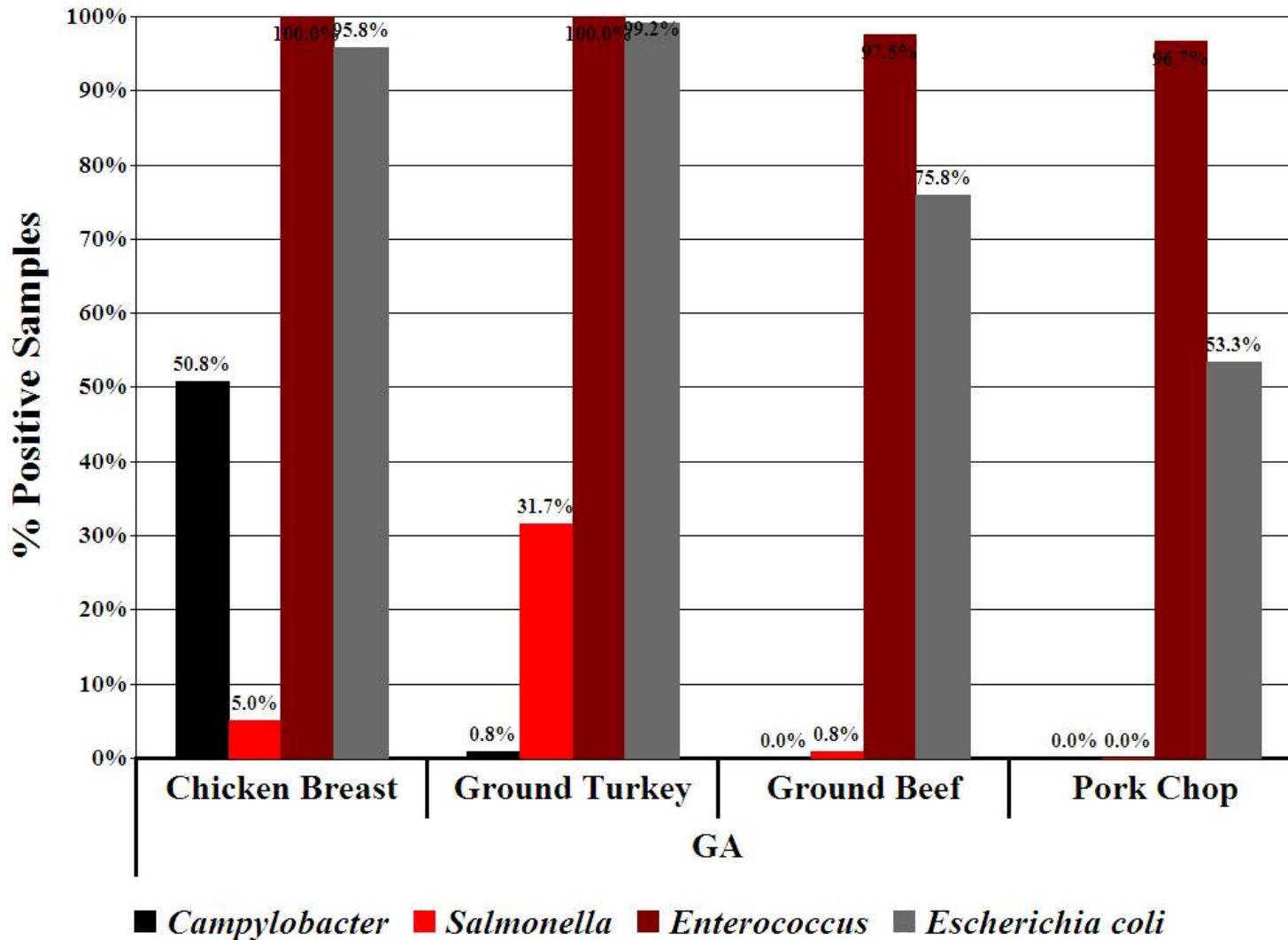
Appendix 3b. Percent Positive Samples by Meat Type, Bacterium in Colorado, 2004



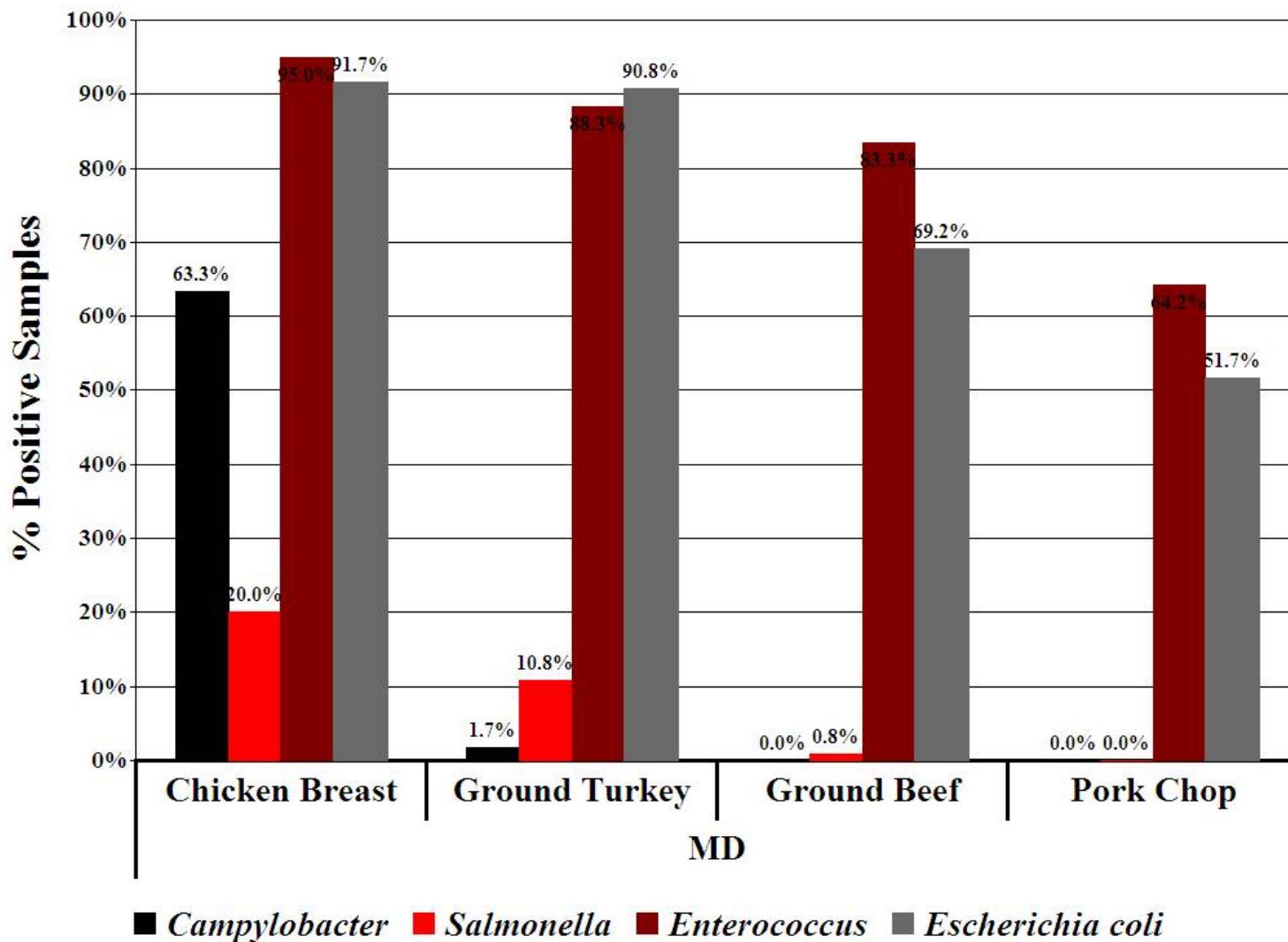
Appendix 3c. Percent Positive Samples by Meat Type, Bacterium in Connecticut, 2004



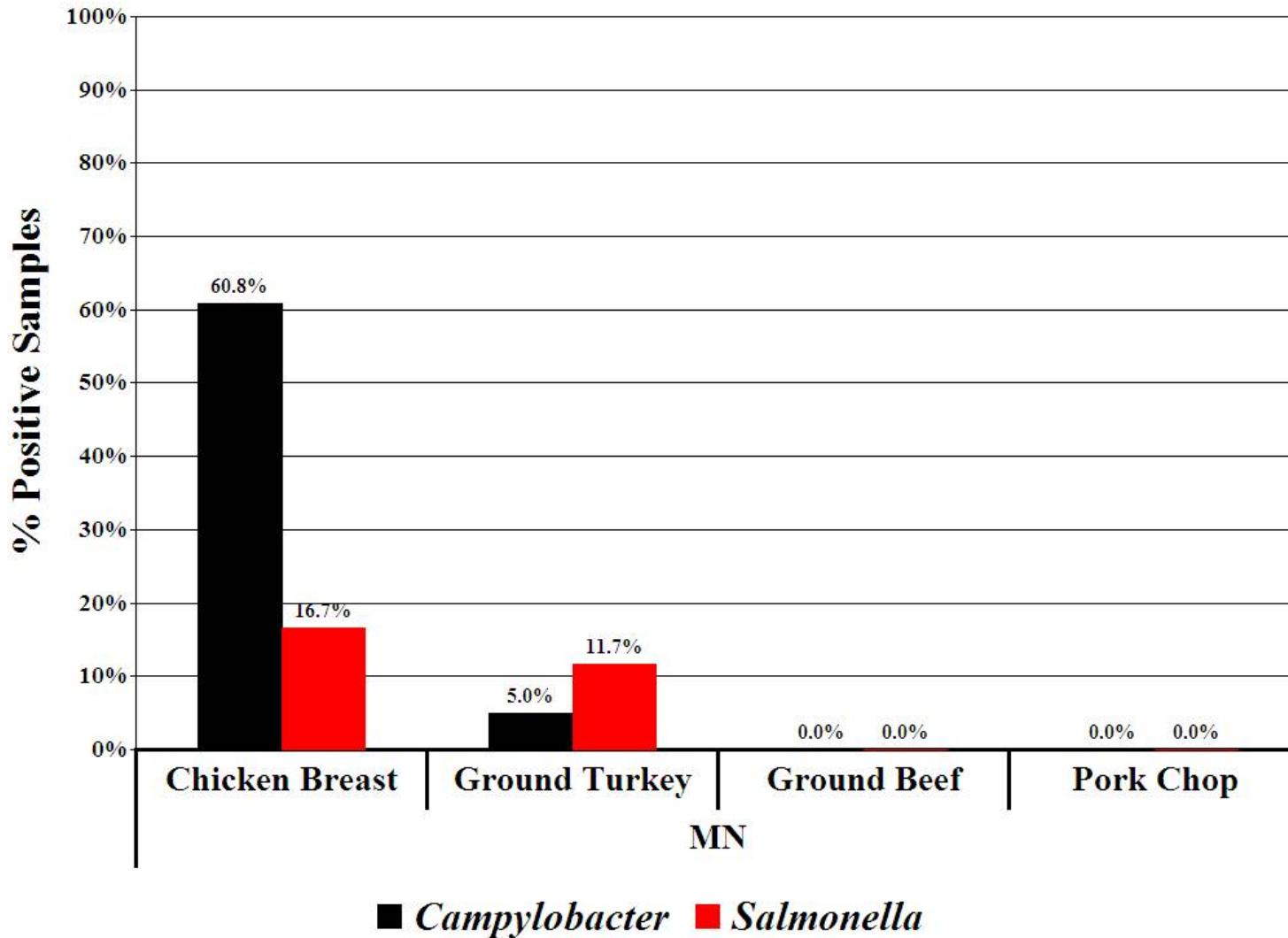
Appendix 3d. Percent Positive Samples by Meat Type, Bacterium in Georgia, 2004



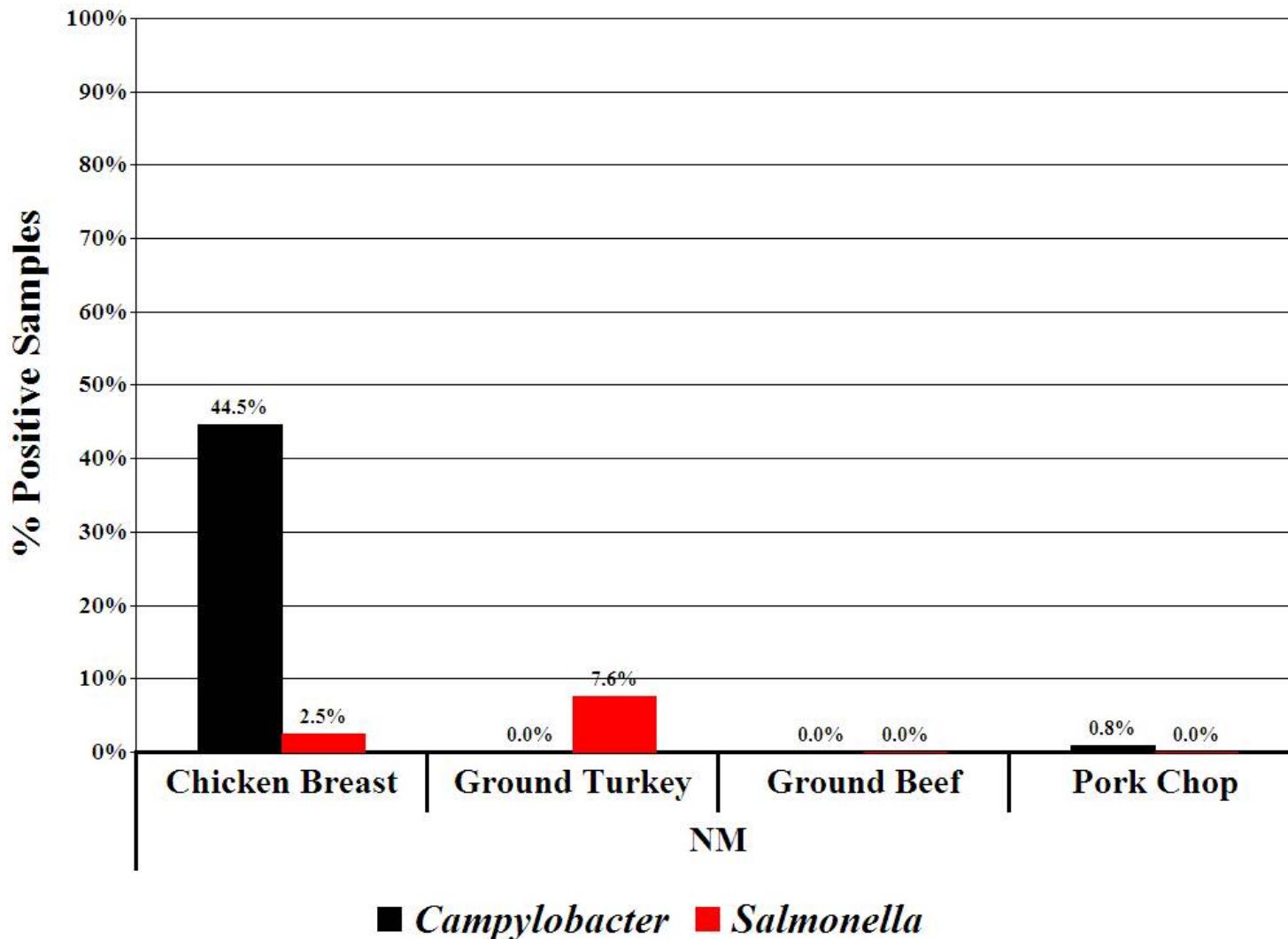
Appendix 3e. Percent Positive Samples by Meat Type, Bacterium in Maryland, 2004



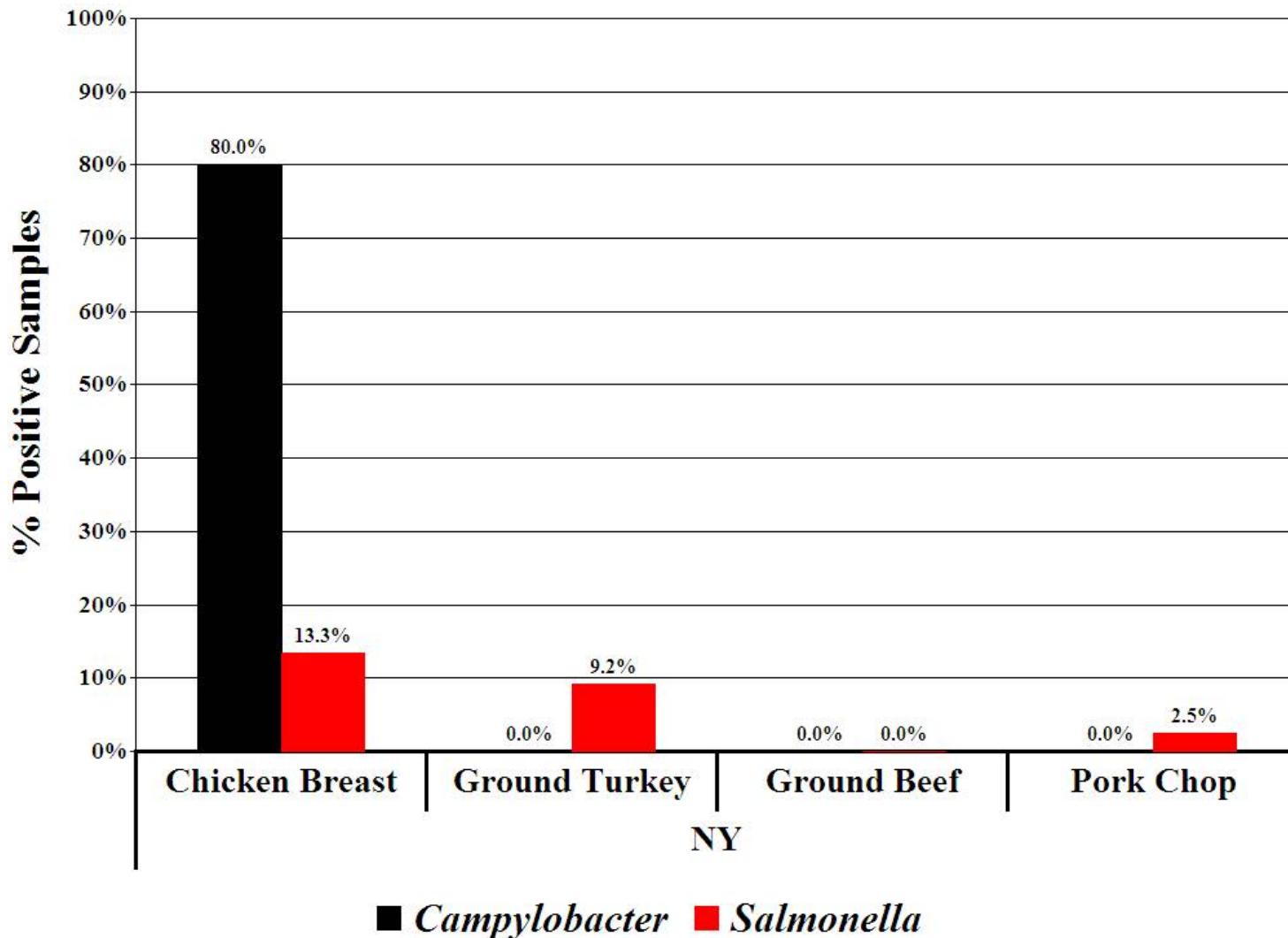
Appendix 3f. Percent Positive Samples by Meat Type, Bacterium in Minnesota, 2004



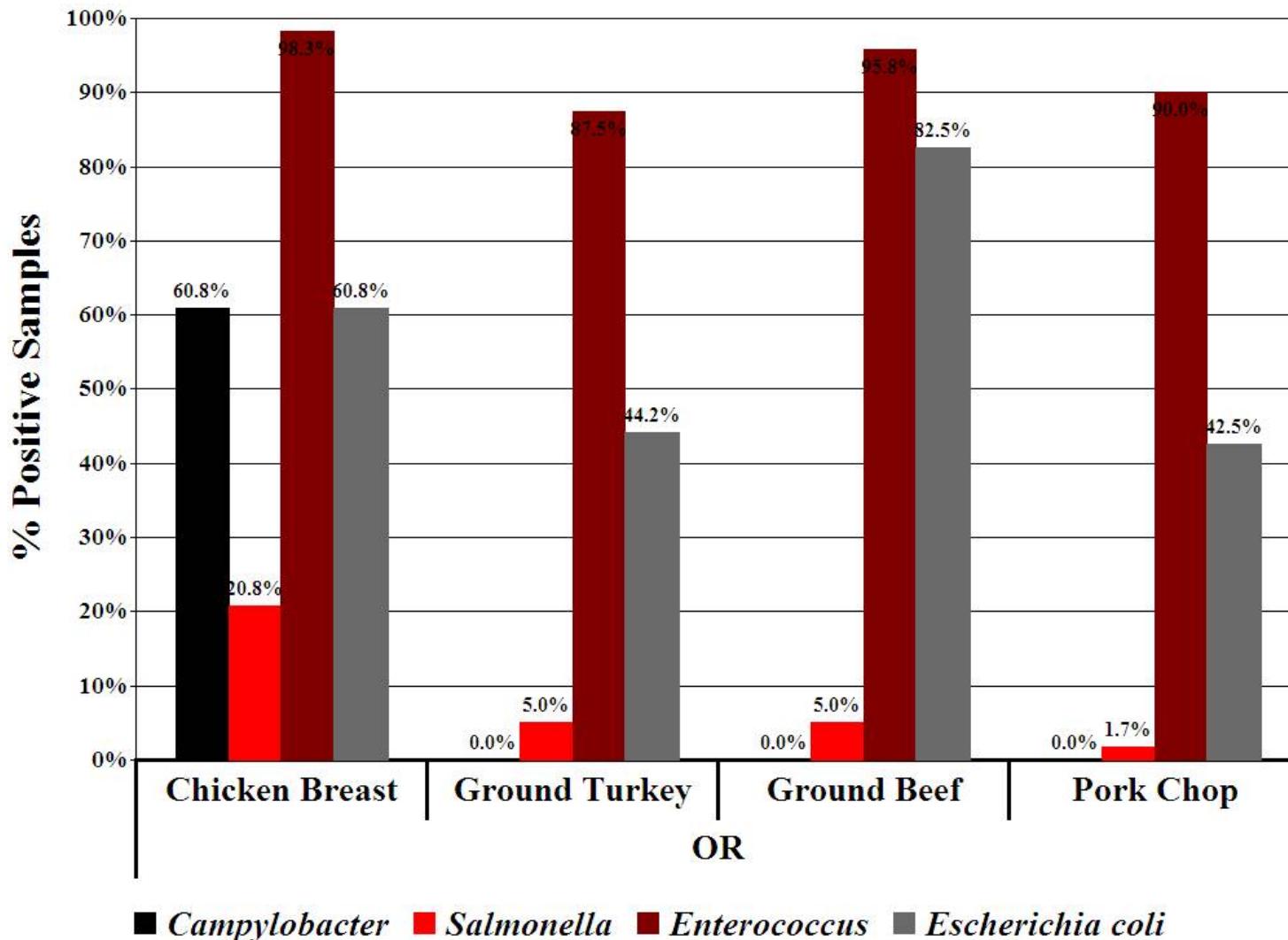
Appendix 3g. Percent Positive Samples by Meat Type, Bacterium in New Mexico, 2004



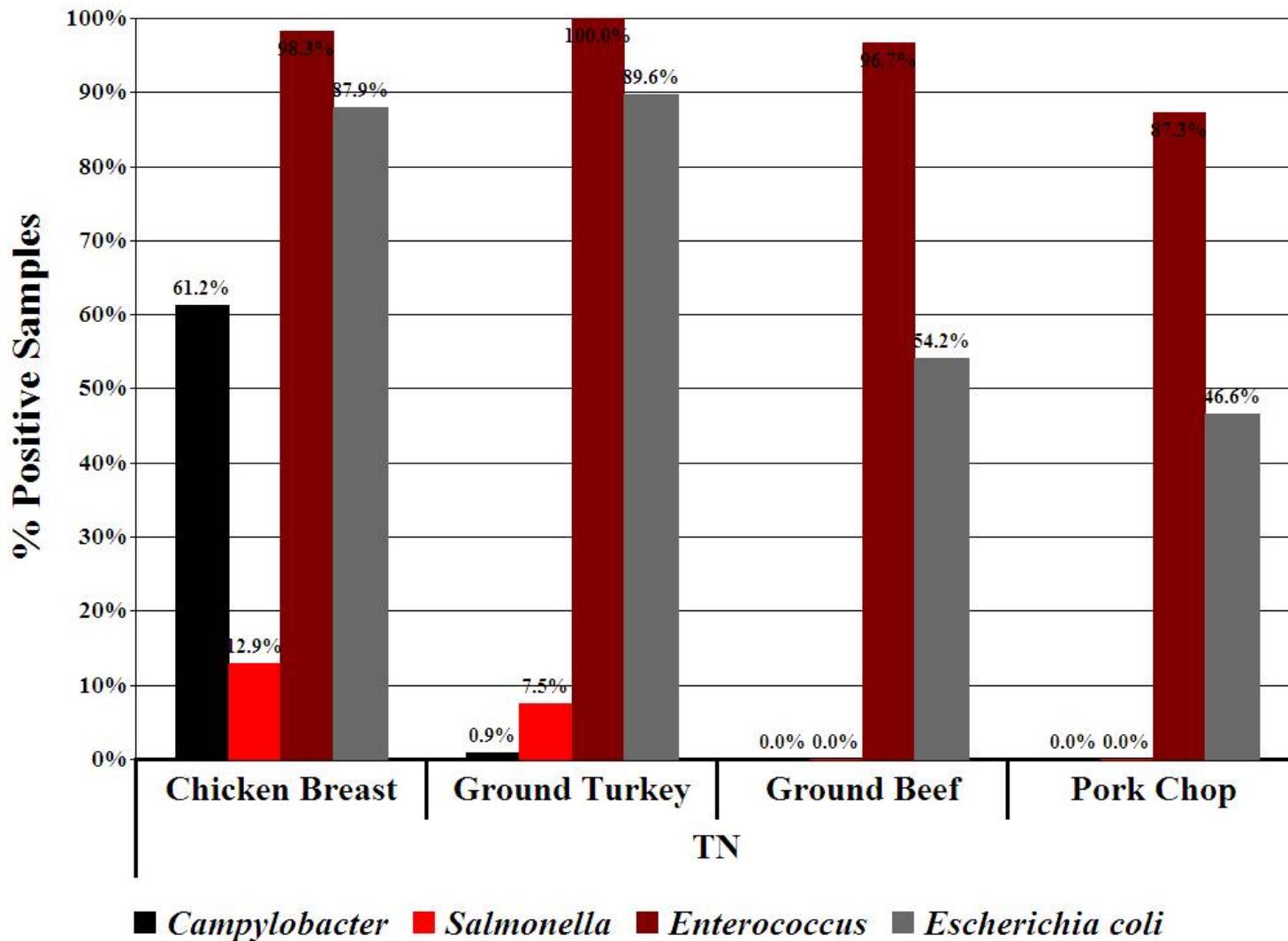
Appendix 3h. Percent Positive Samples by Meat Type, Bacterium in New York, 2004



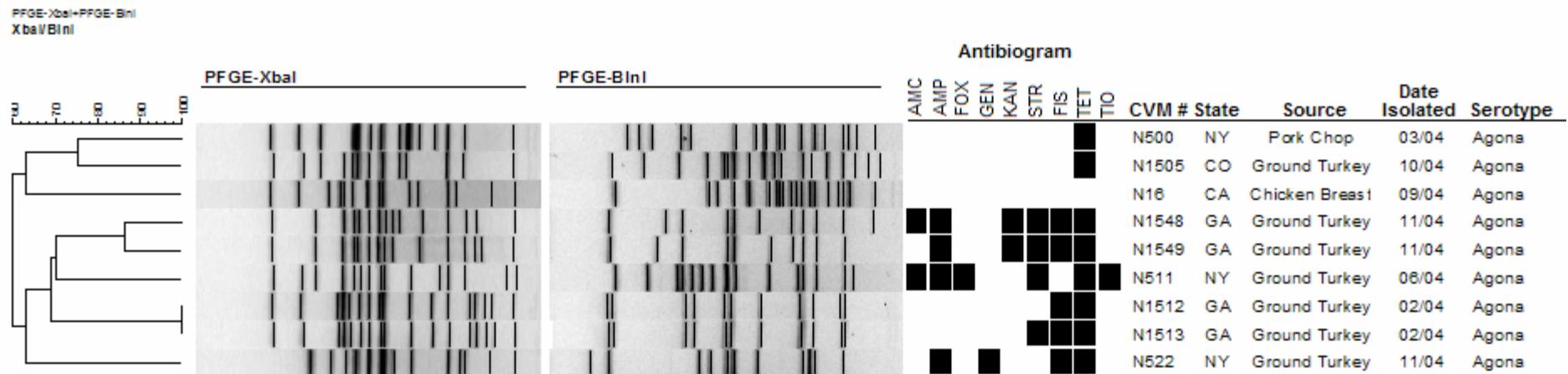
Appendix 3i. Percent Positive Samples by Meat Type, Bacterium in Oregon, 2004



Appendix 3j Percent Positive Samples by Meat Type, Bacterium in Tennessee, 2004

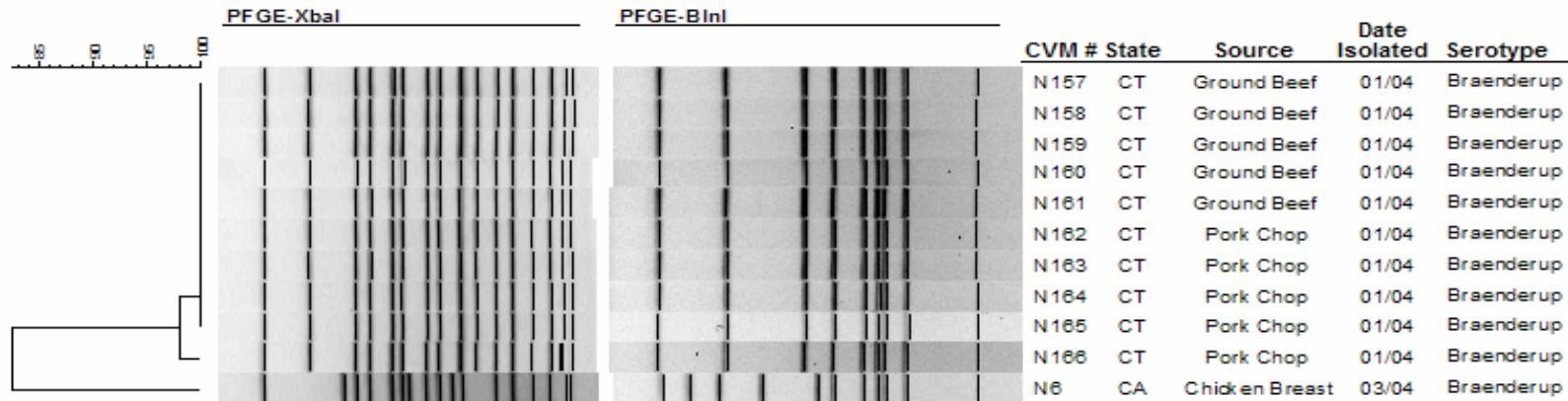


A-4a. PFGE Profiles for *Salmonella* Agona

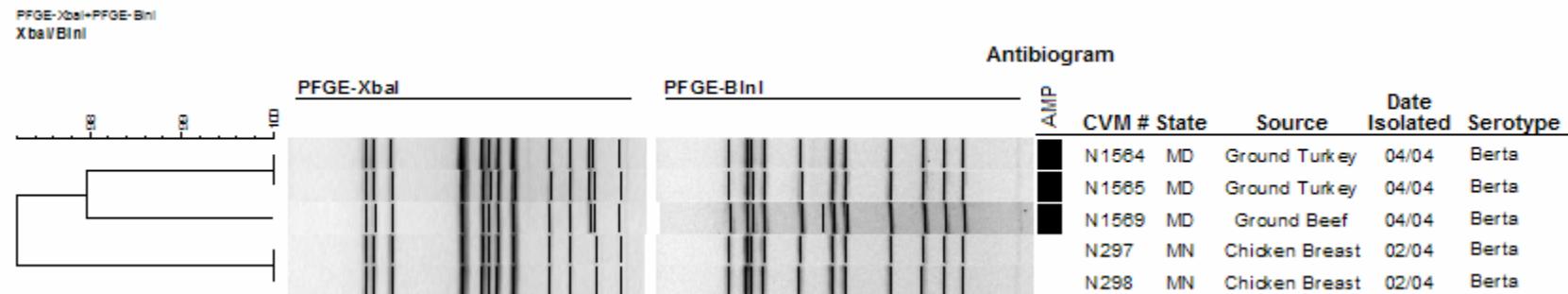


A-4b. PFGE Profiles for *Salmonella* Braenderup

PFGE-XbaI+PFGE-BlnI
XbaI/BlnI

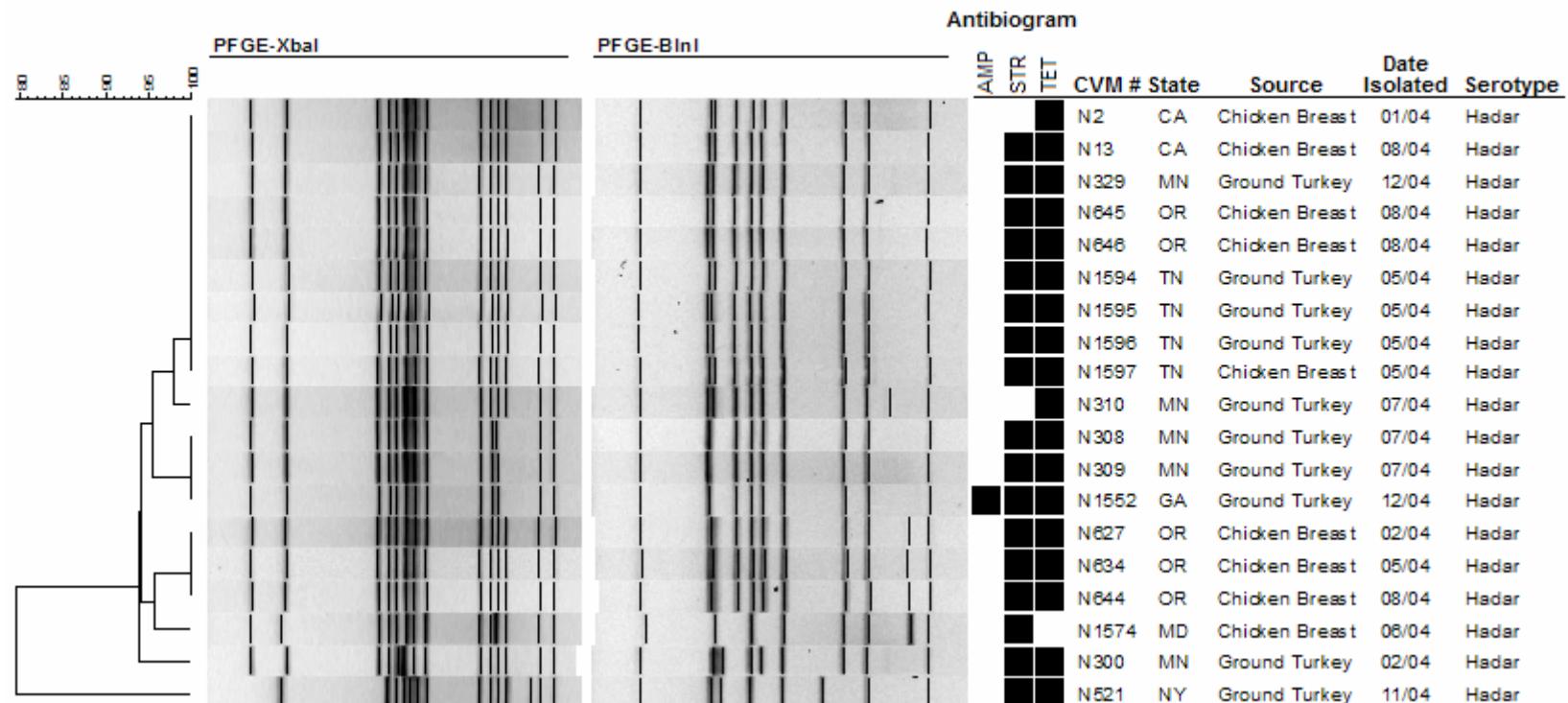


A-4c. PFGE Profiles for *Salmonella* Berta



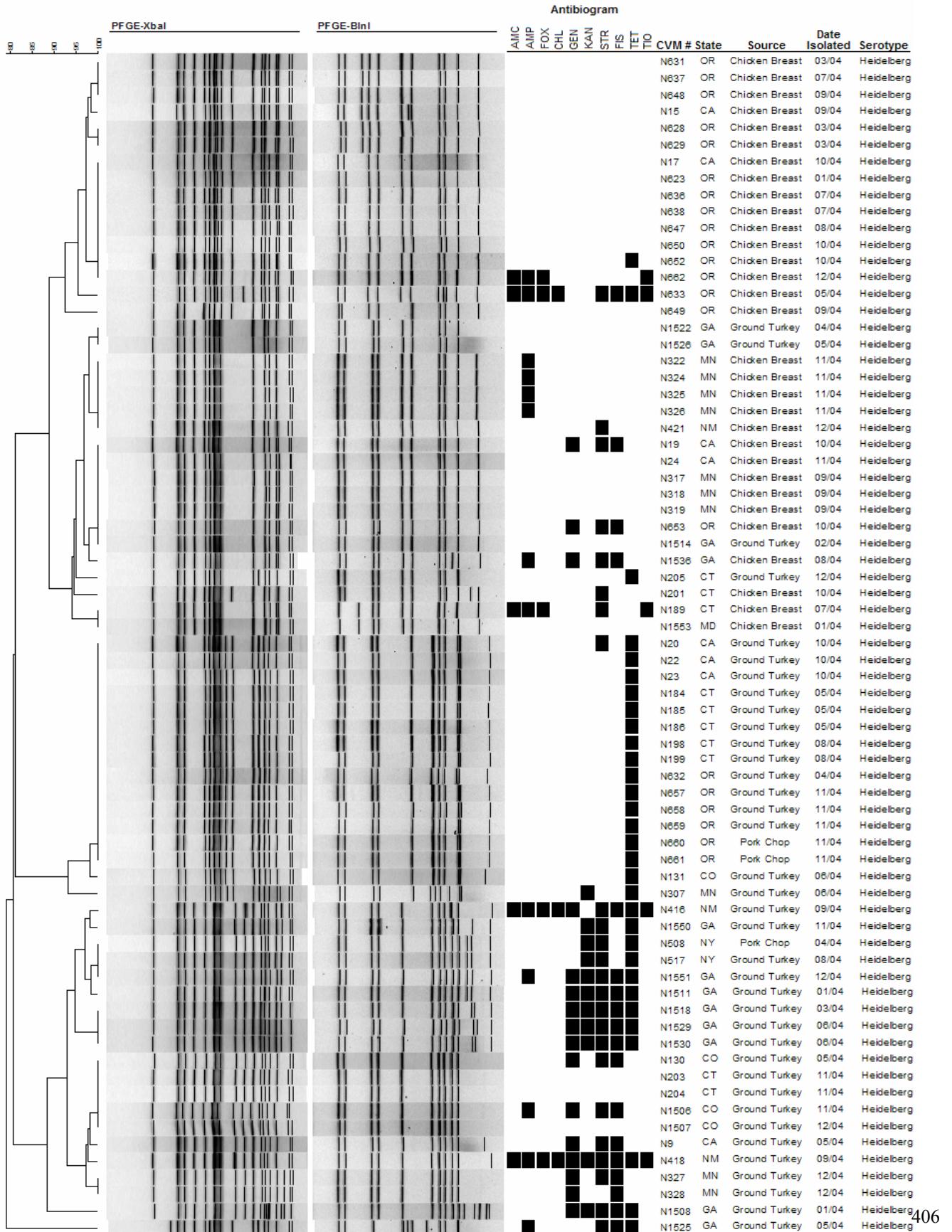
A-4d. PFGE Profiles for *Salmonella* Hadar

PFGE-XbaI+PFGE-BlnI
XbaI/BlnI

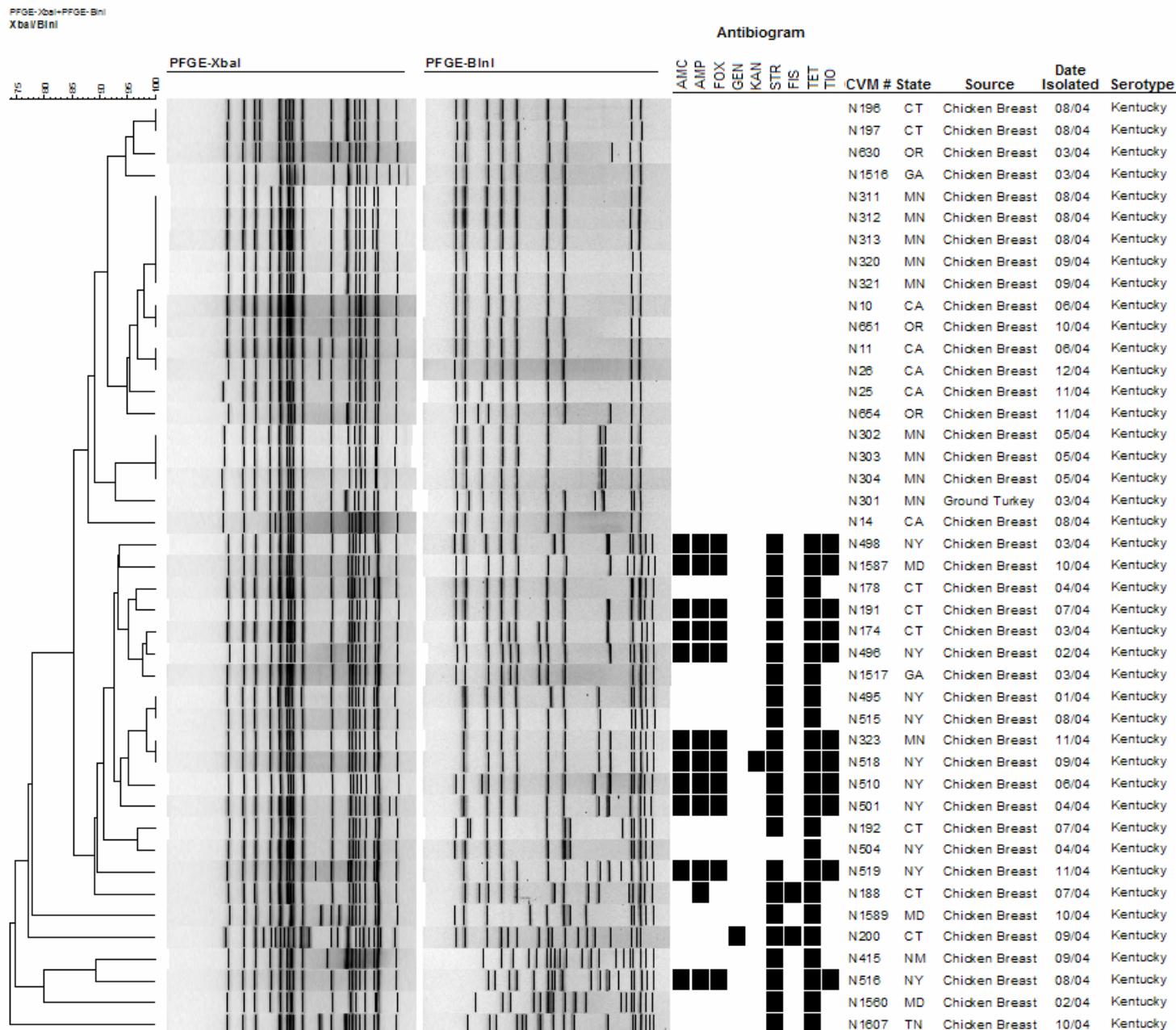


A-4e. PFGE Profiles for *Salmonella* Heidelberg

PFGE-XbaI+PFGE-BlnI
XbaI/BlnI

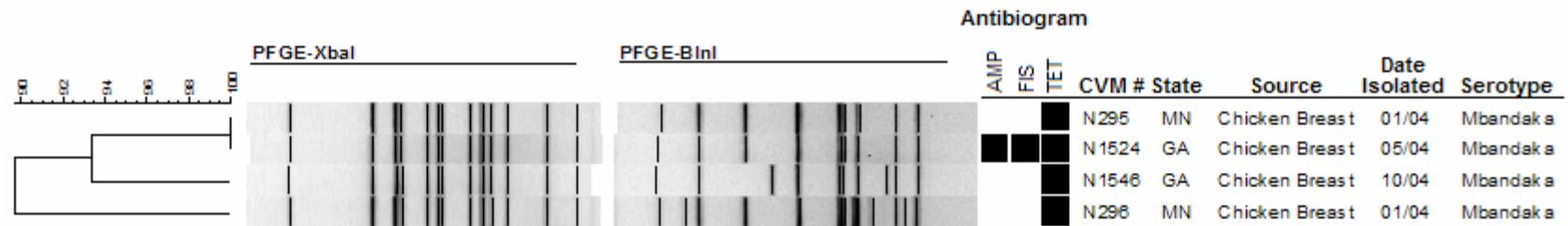


A-4f. PFGE Profiles for *Salmonella* Kentucky

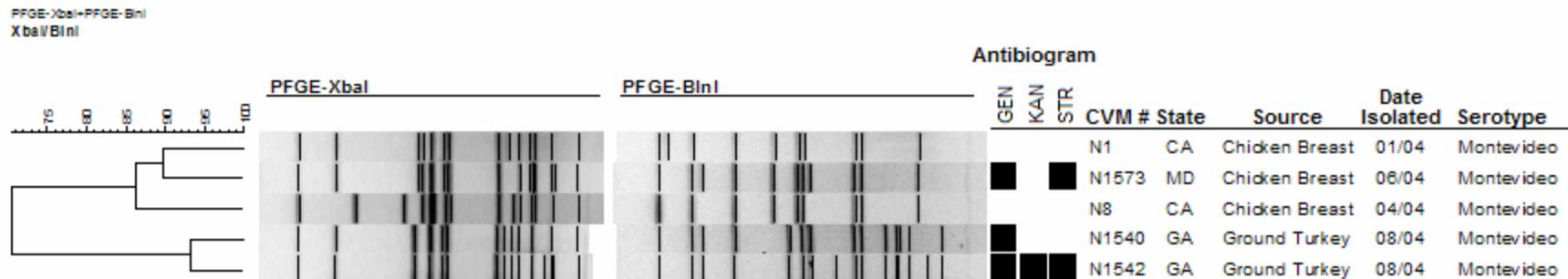


A-4g. PFGE Profiles for *Salmonella* Mbandaka

PFGE-XbaI+PFGE-BlnI
XbaI/BlnI

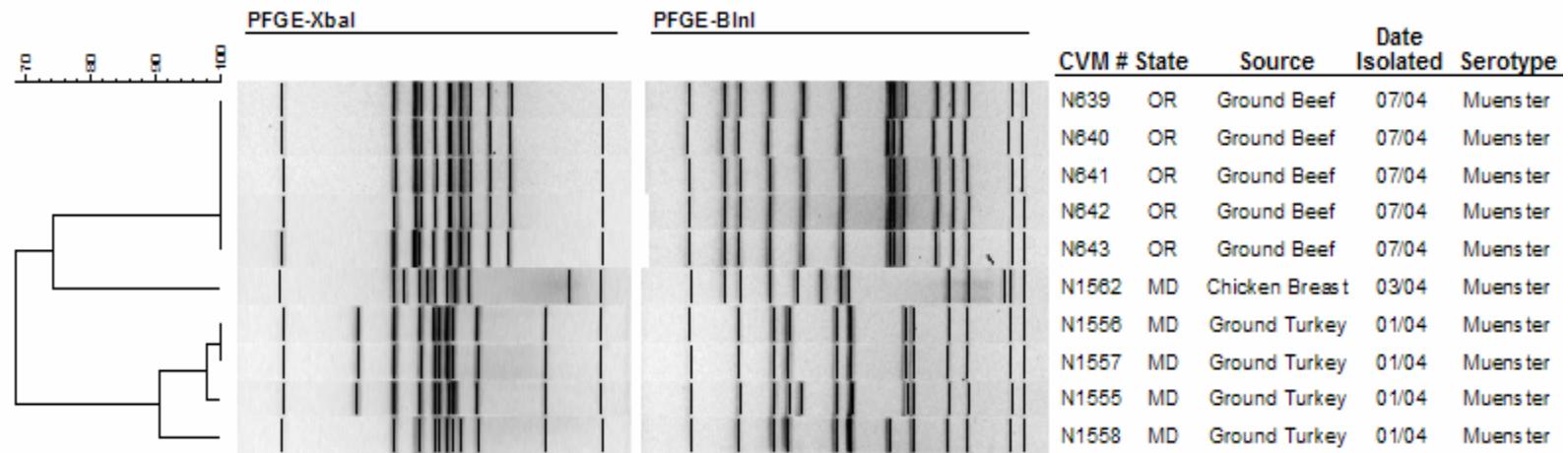


A-4h. PFGE Profiles for *Salmonella* Montevideo



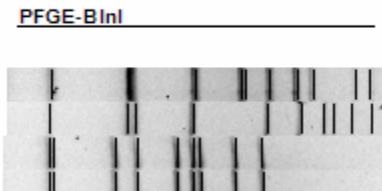
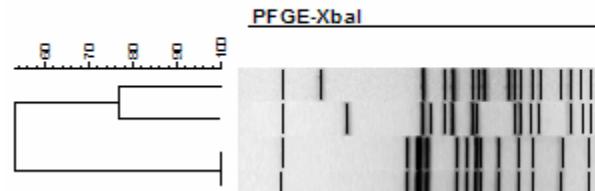
A-4i. PFGE Profiles for *Salmonella* Muenster

PFGE-XbaI+PFGE-BlnI
XbaI/BlnI



A-4j. PFGE Profiles for *Salmonella* Newport

PFGE-XbaI>PFGE-BlnI
XbaI/BlnI

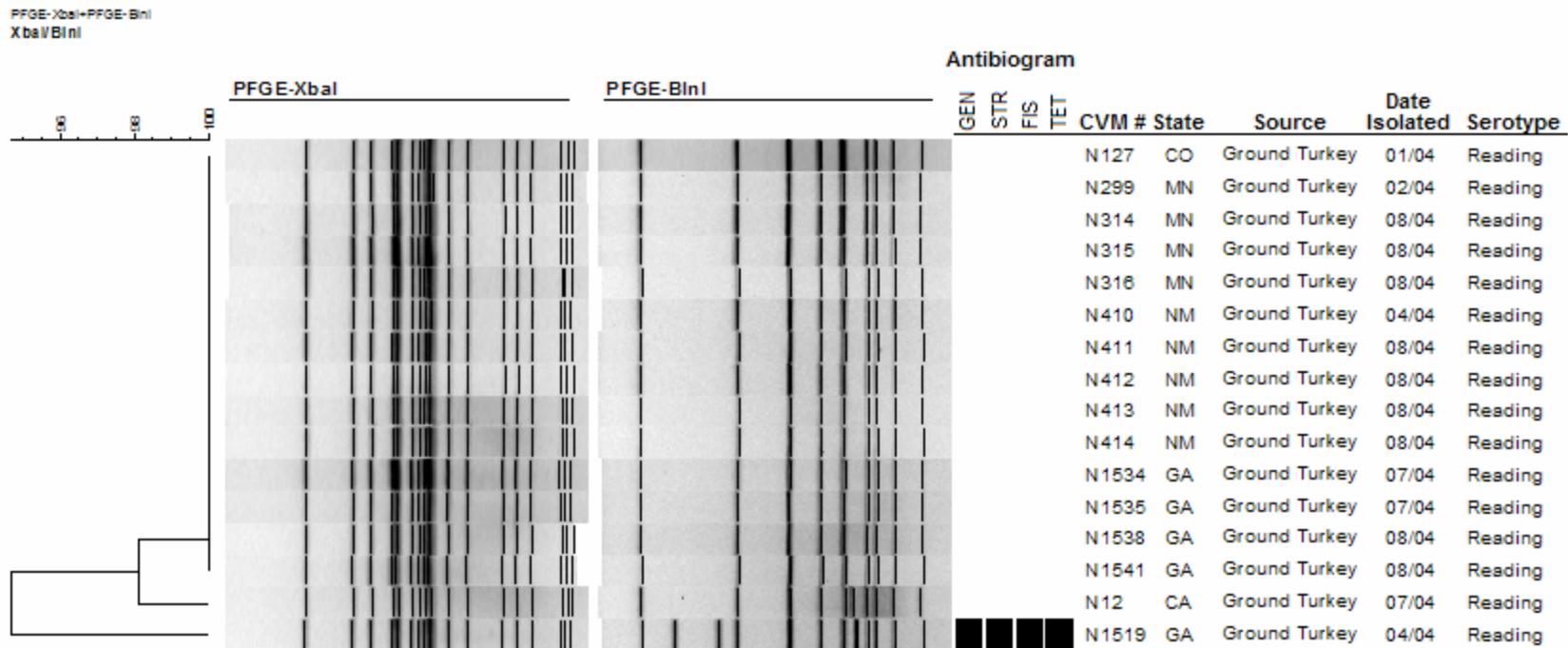


Antibiogram



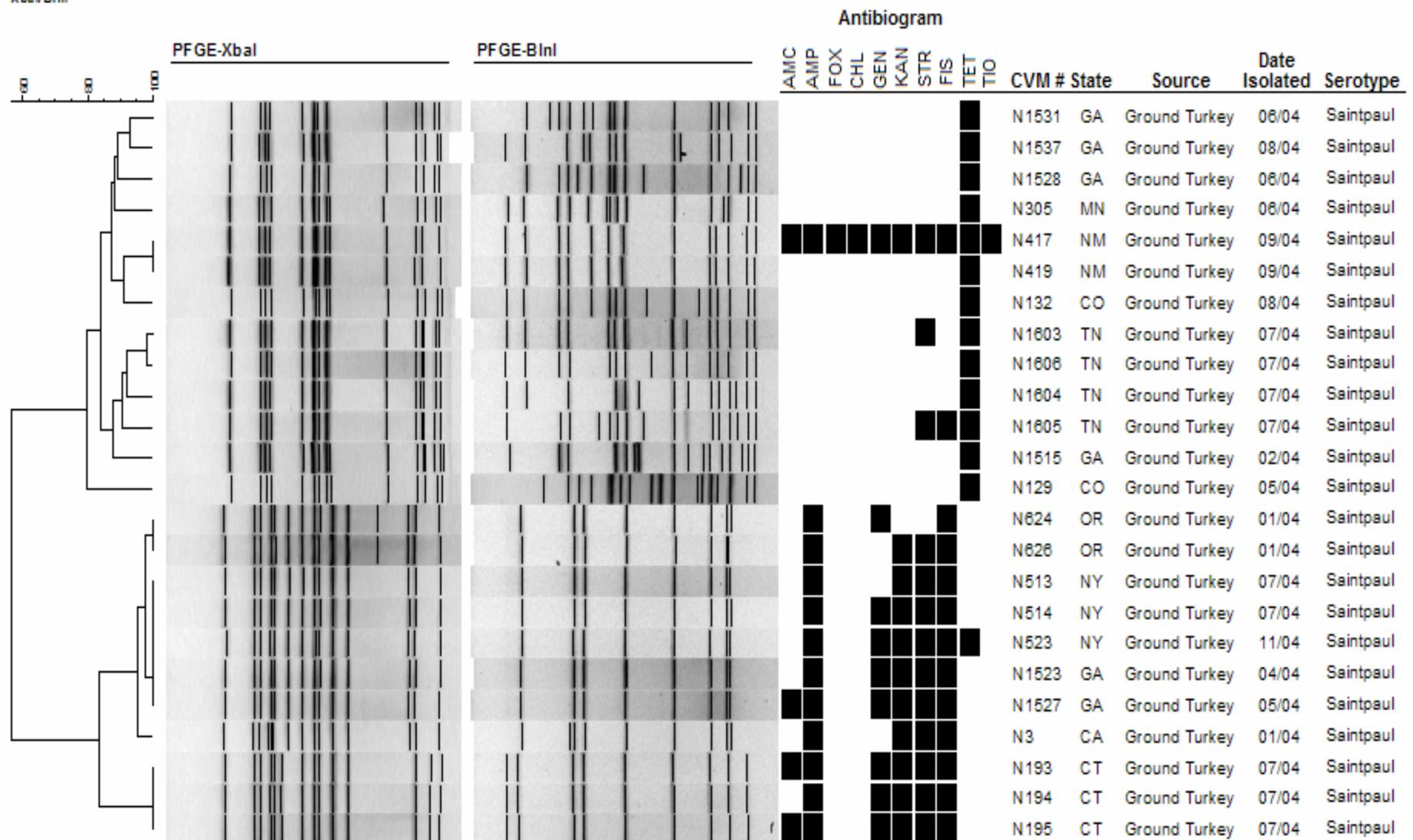
CVM #	State	Source	Date Isolated	Serotype
N635	OR	Ground Beef	05/04	Newport
N1543	GA	Ground Beef	08/04	Newport
N1509	GA	Ground Turkey	01/04	Newport
N1510	GA	Ground Turkey	01/04	Newport

A-4k. PFGE Profiles for *Salmonella* Reading

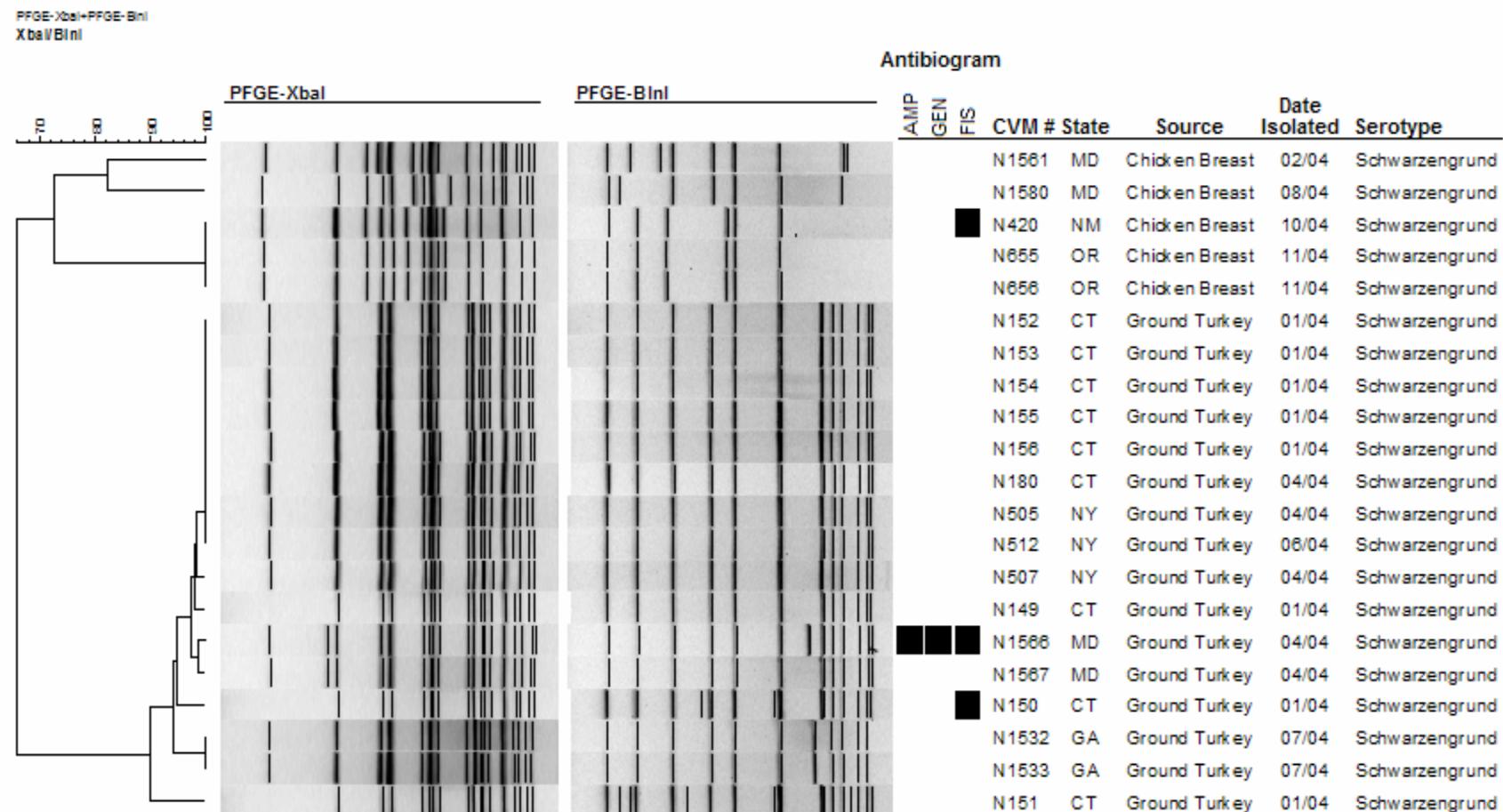


A-4l. PFGE Profiles for *Salmonella* Saintpaul

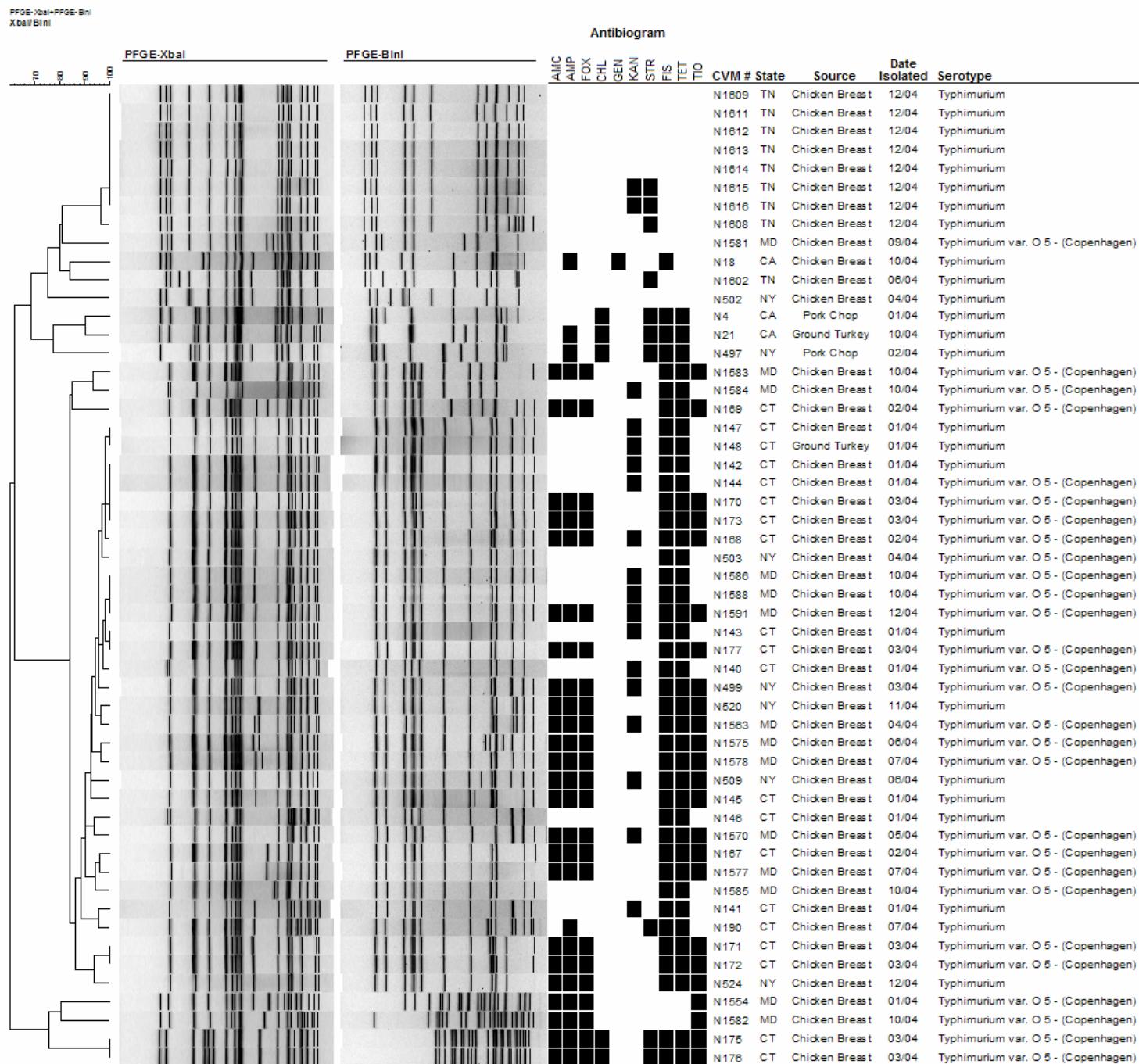
PFGE-XbaI+PFGE-BlnI
XbaI/BlnI



A-4m. PFGE Profiles for *Salmonella* Schwarzengrund

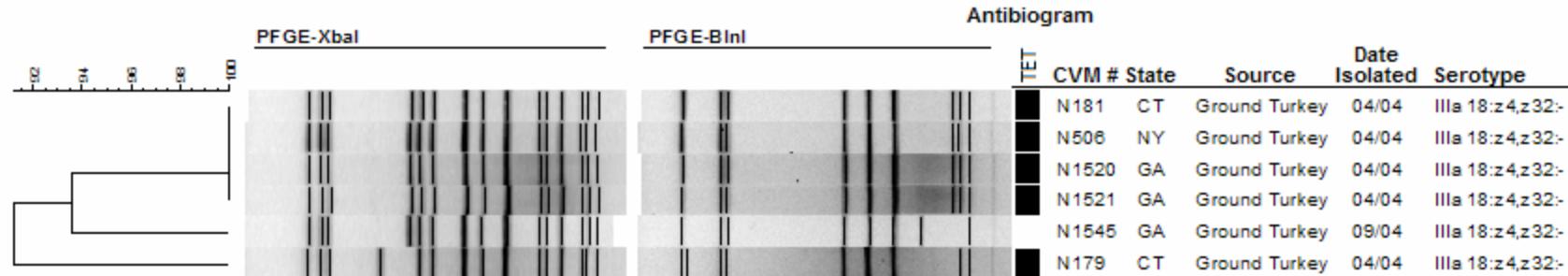


A-4n. PFGE Profiles for *Salmonella* Typhimurium



A-4o. PFGE Profiles for *Salmonella* IIIa 18:z4,z32:-

PFGE-XbaI+PFGE-BlnI
XbaI/BlnI

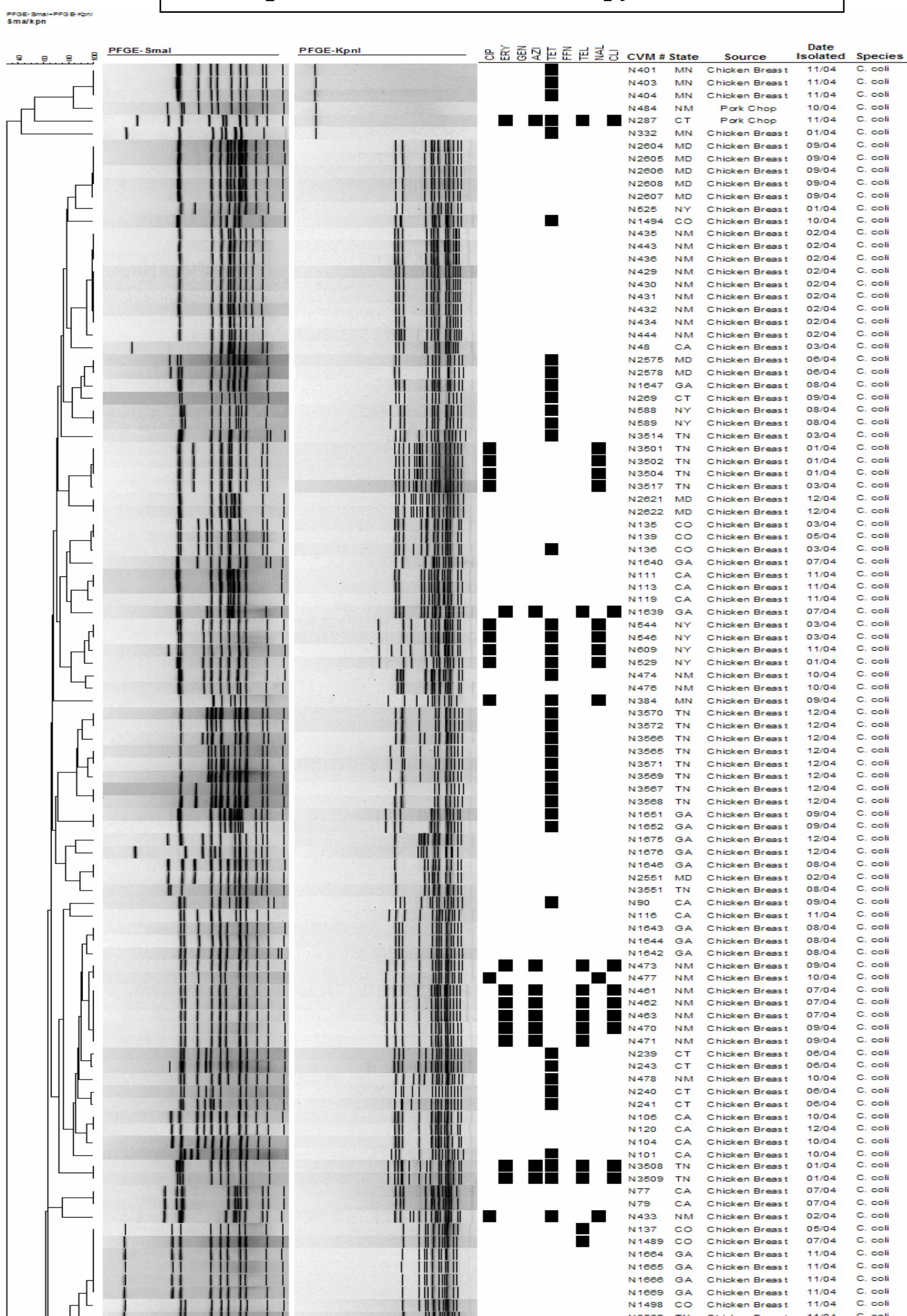


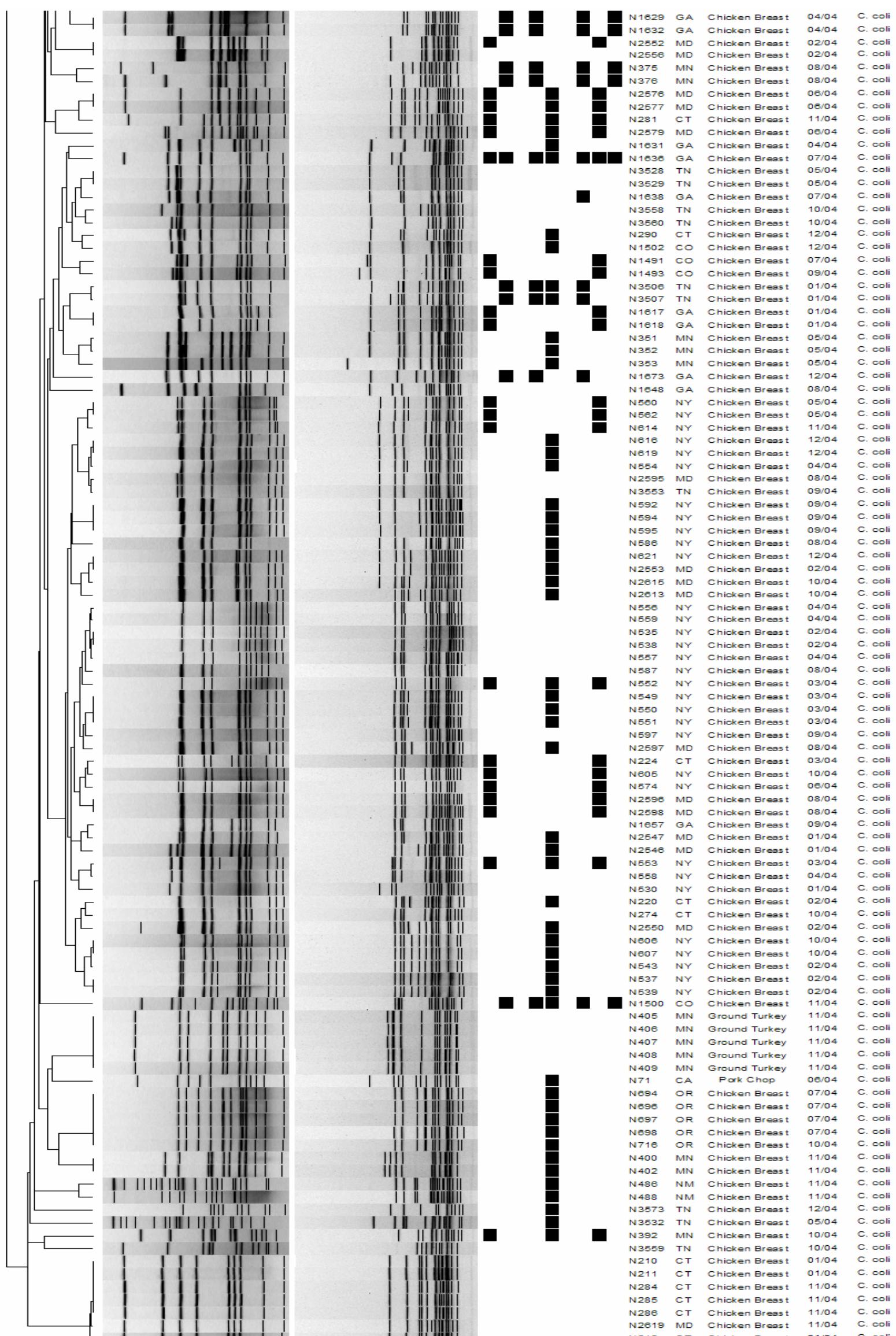
A-4p. PFGE Profiles for *Salmonella* 4,12:i:-

PFGE-XbaI+PFGE-BlnI
XbaI/BlnI

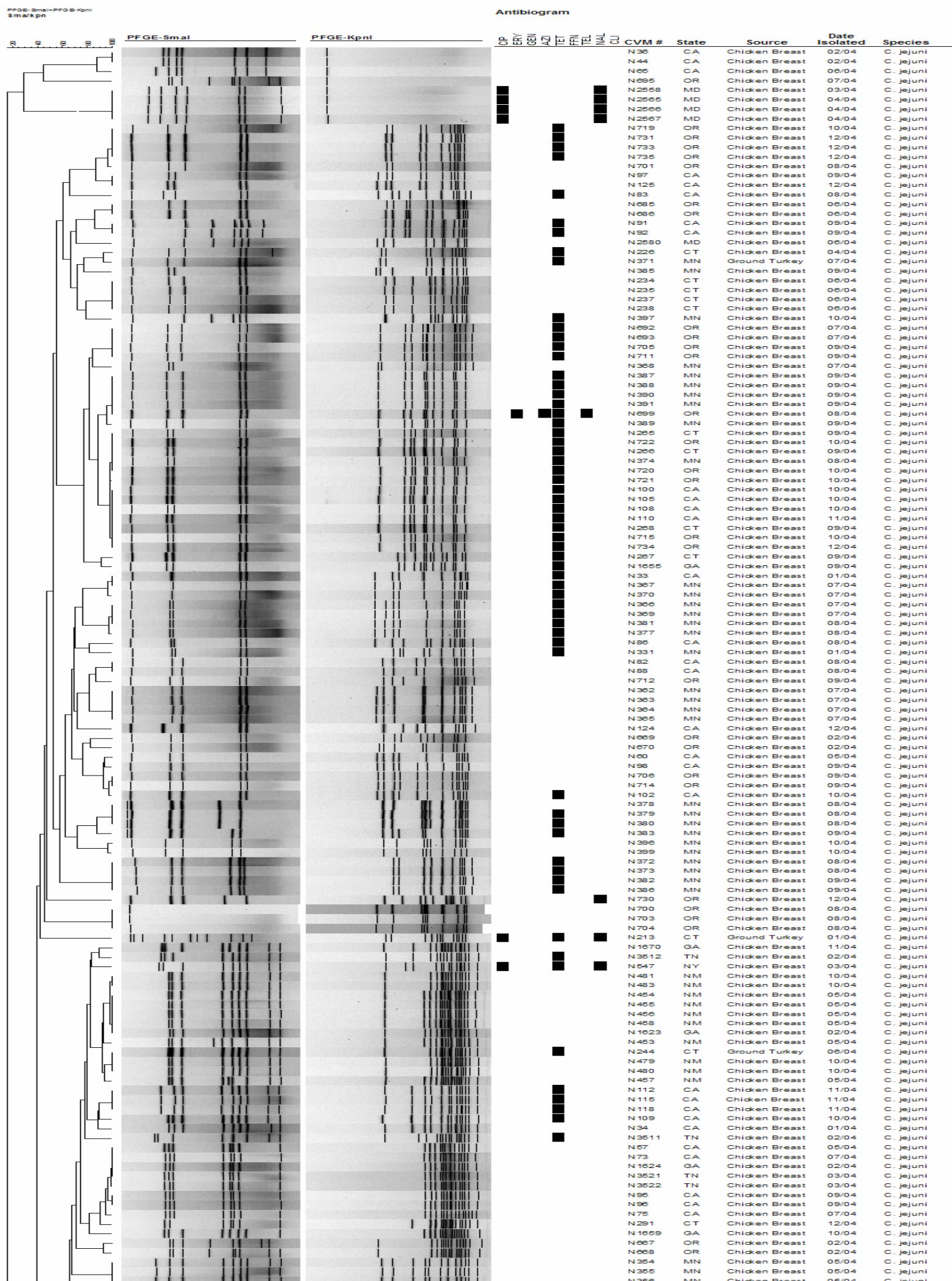


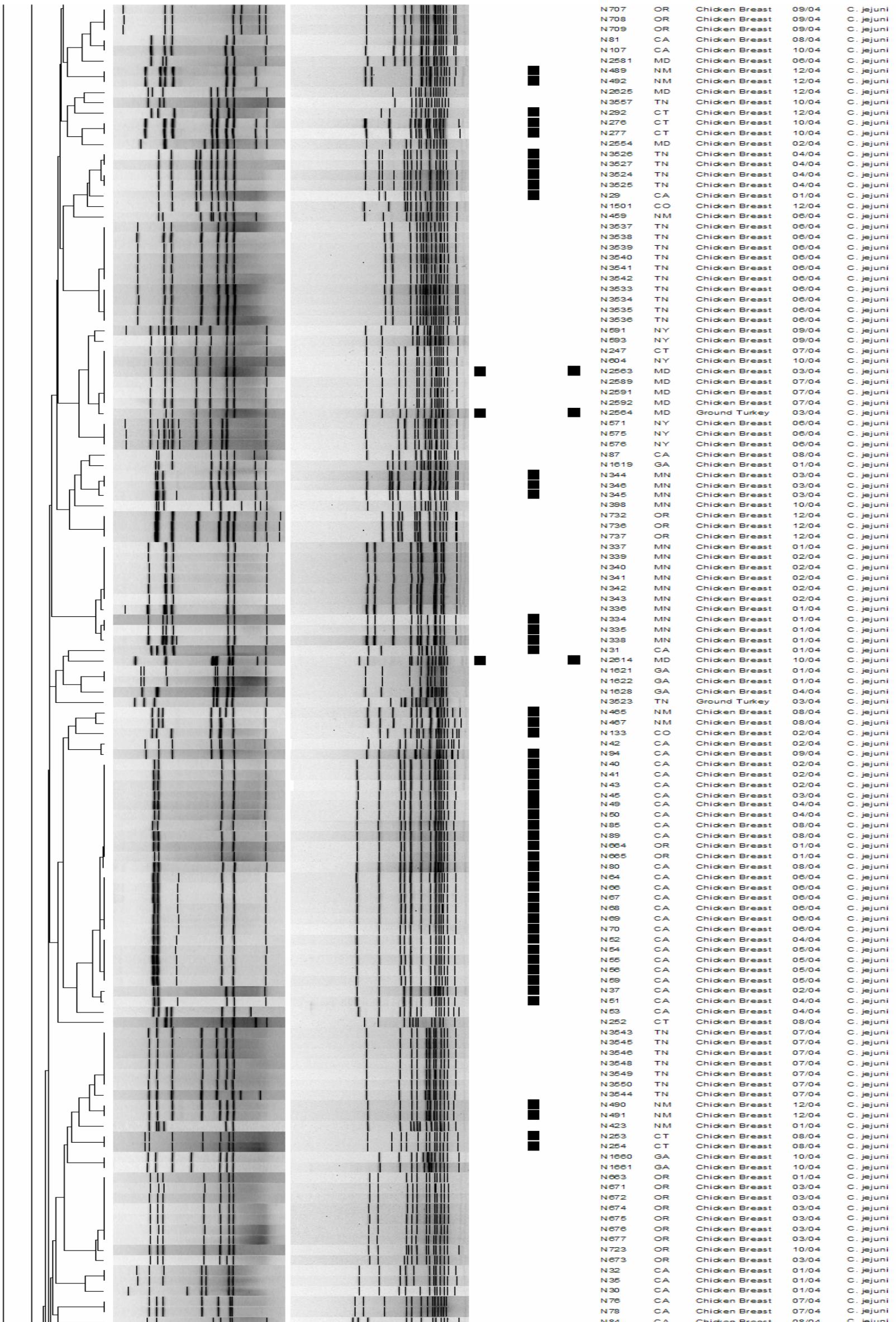
A-4q. PFGE Profiles for *Campylobacter coli*

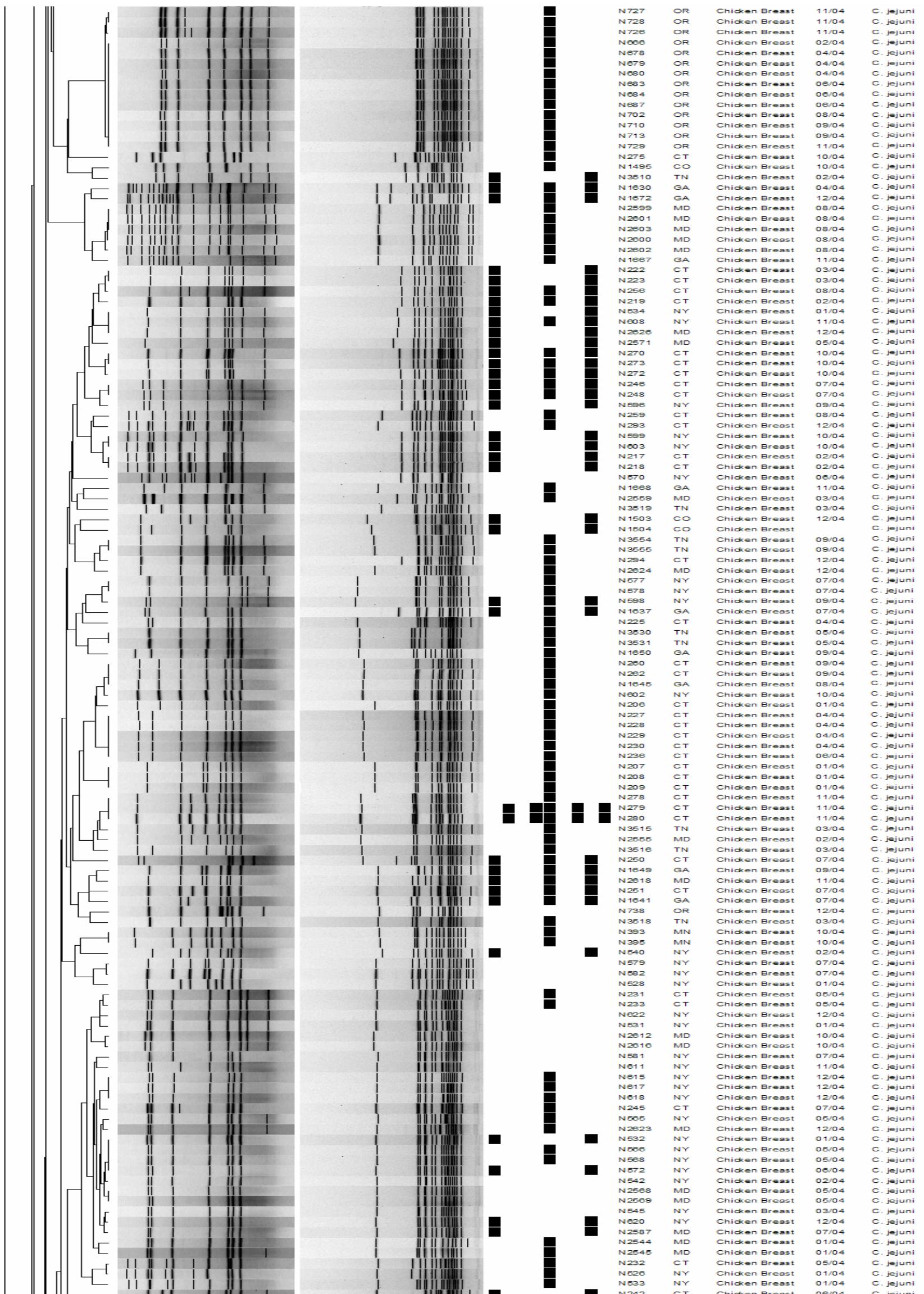




A-4r. PFGE Profiles for *Campylobacter jejuni*







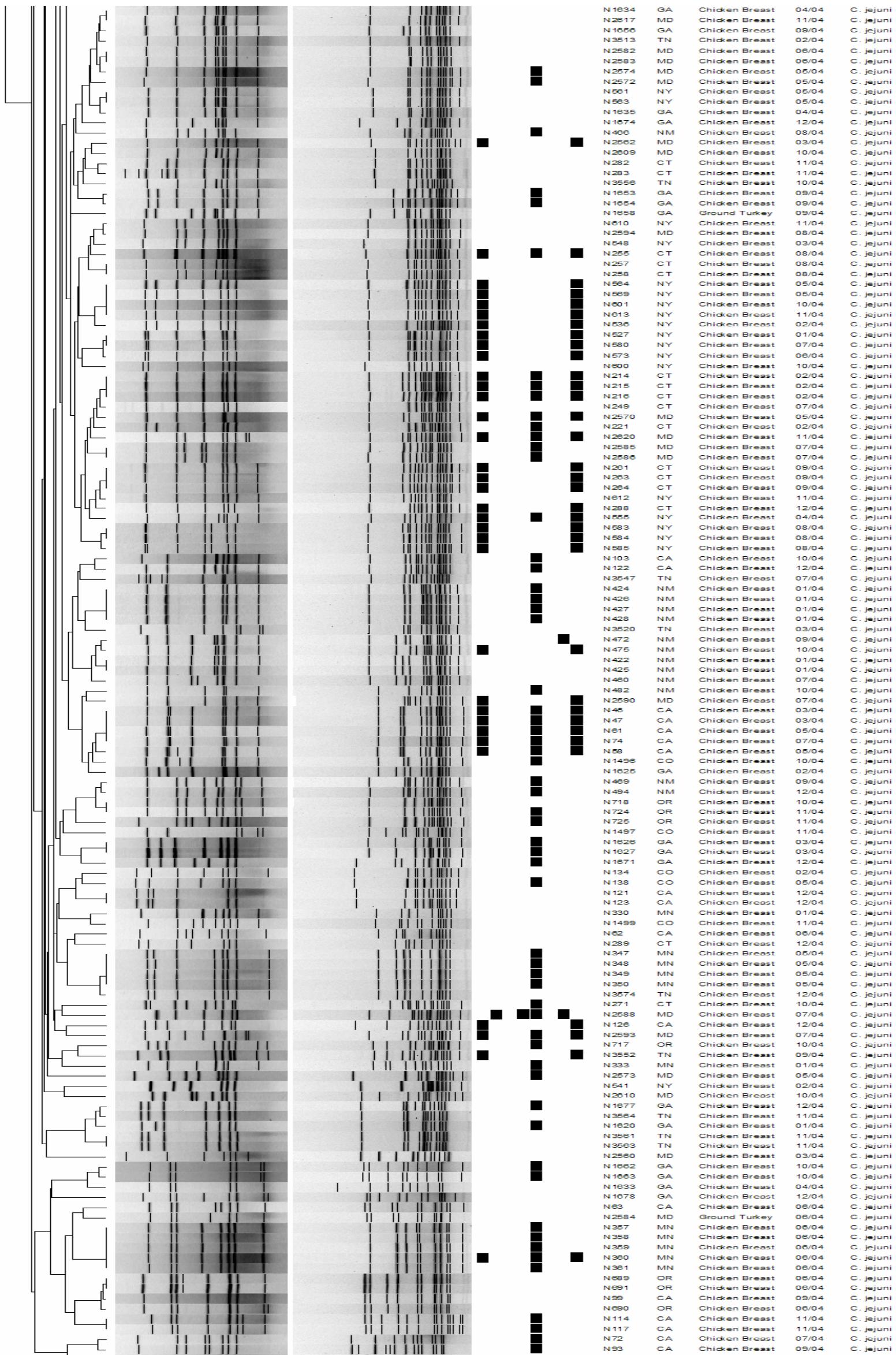
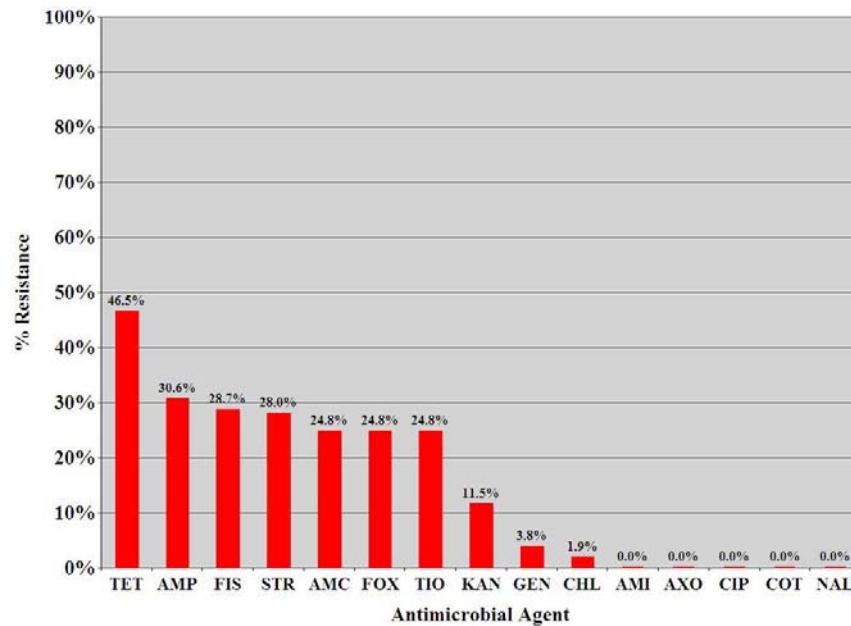
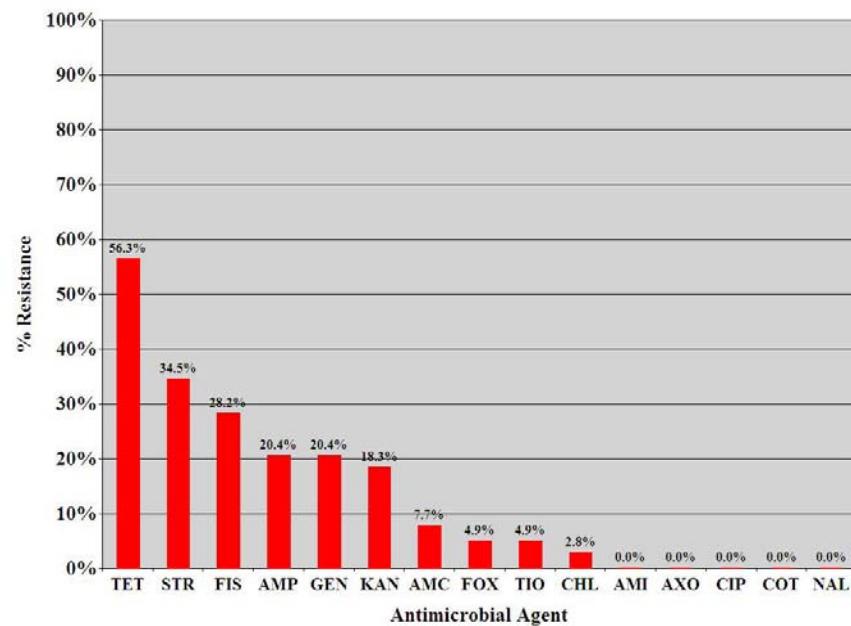


Figure A-5. Antimicrobial Resistance among *Salmonella* by Meat Type, 2004

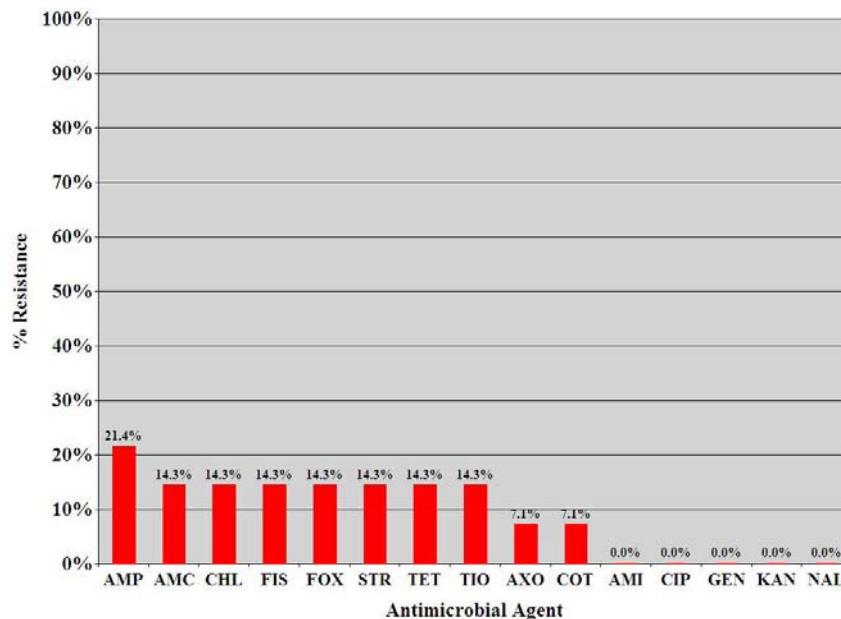
Chicken Breast (n=157)



Ground Turkey (n=142)



Ground Beef (n=14)



Pork Chop (n=11)

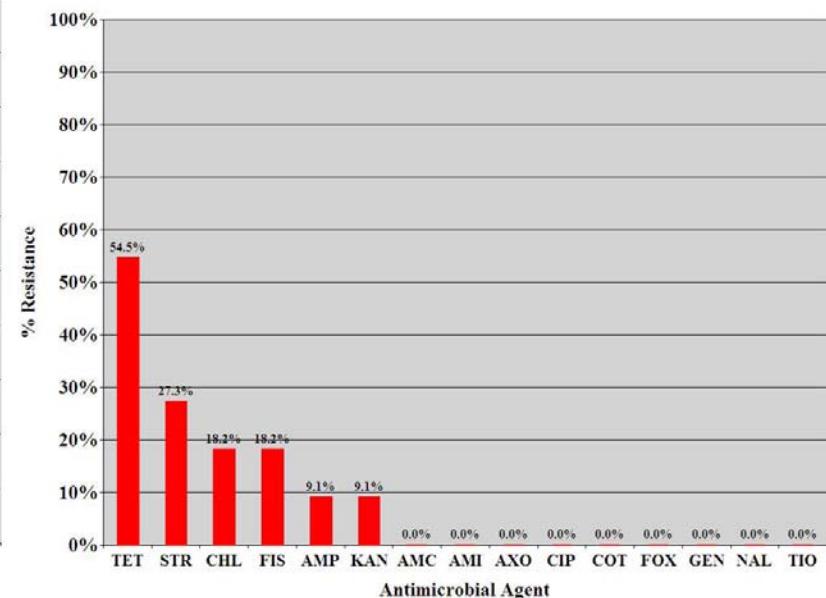


Figure A-6. Antimicrobial Resistance among *Campylobacter* by Meat Type, 2004

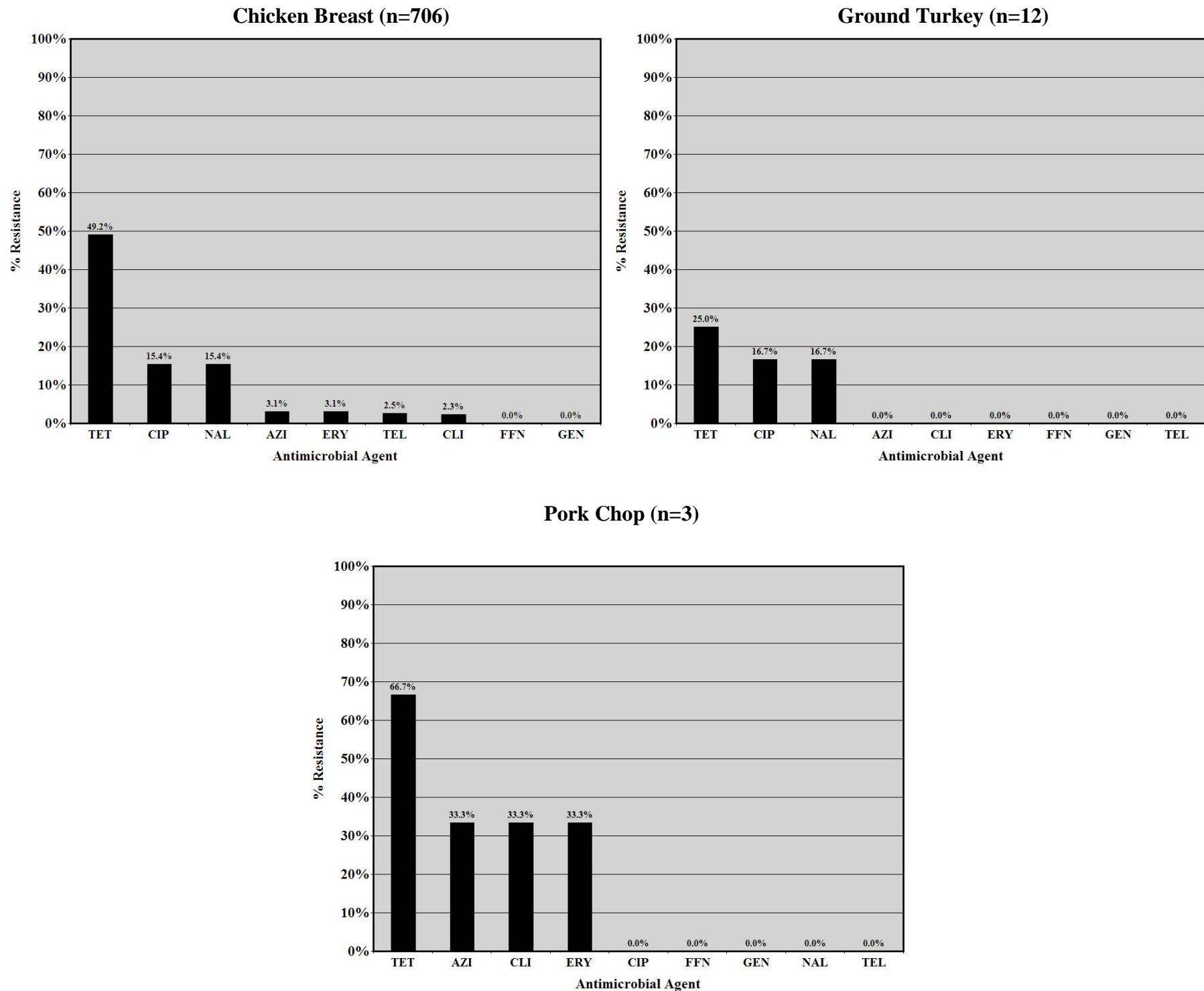


Figure A-6a. Antimicrobial Resistance among *Campylobacter jejuni* Meat Type, 2004

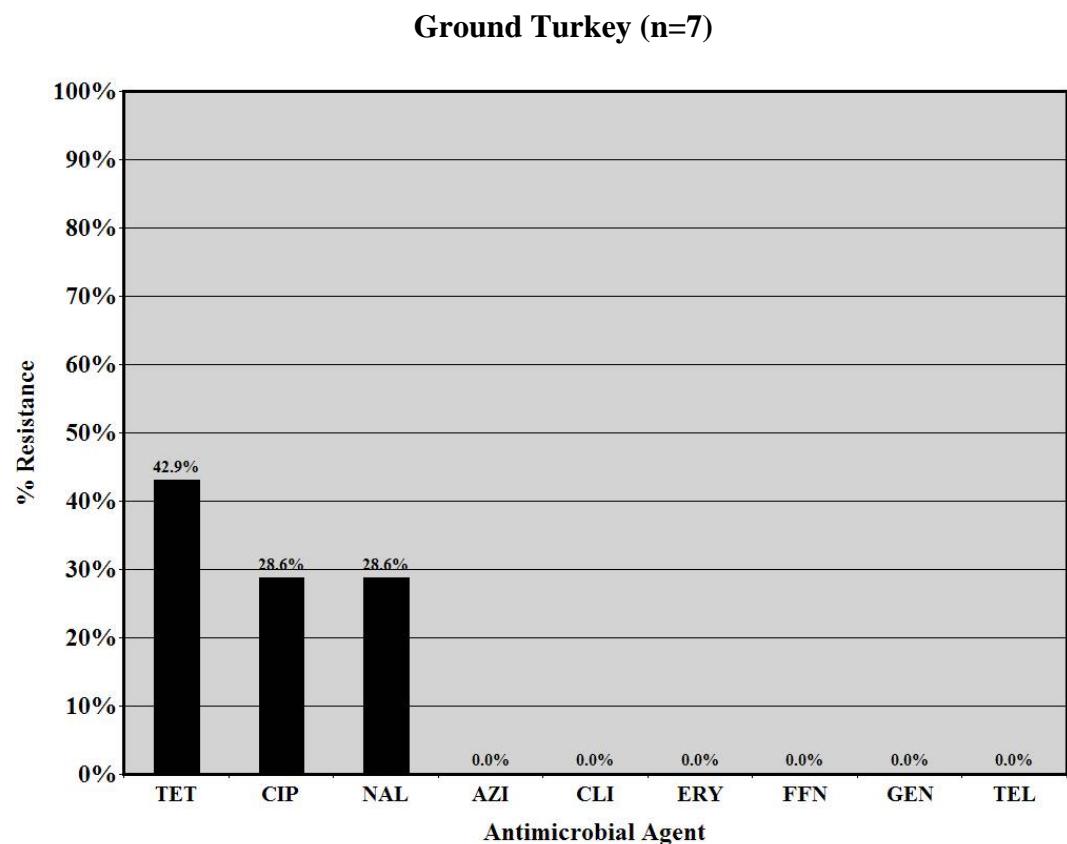
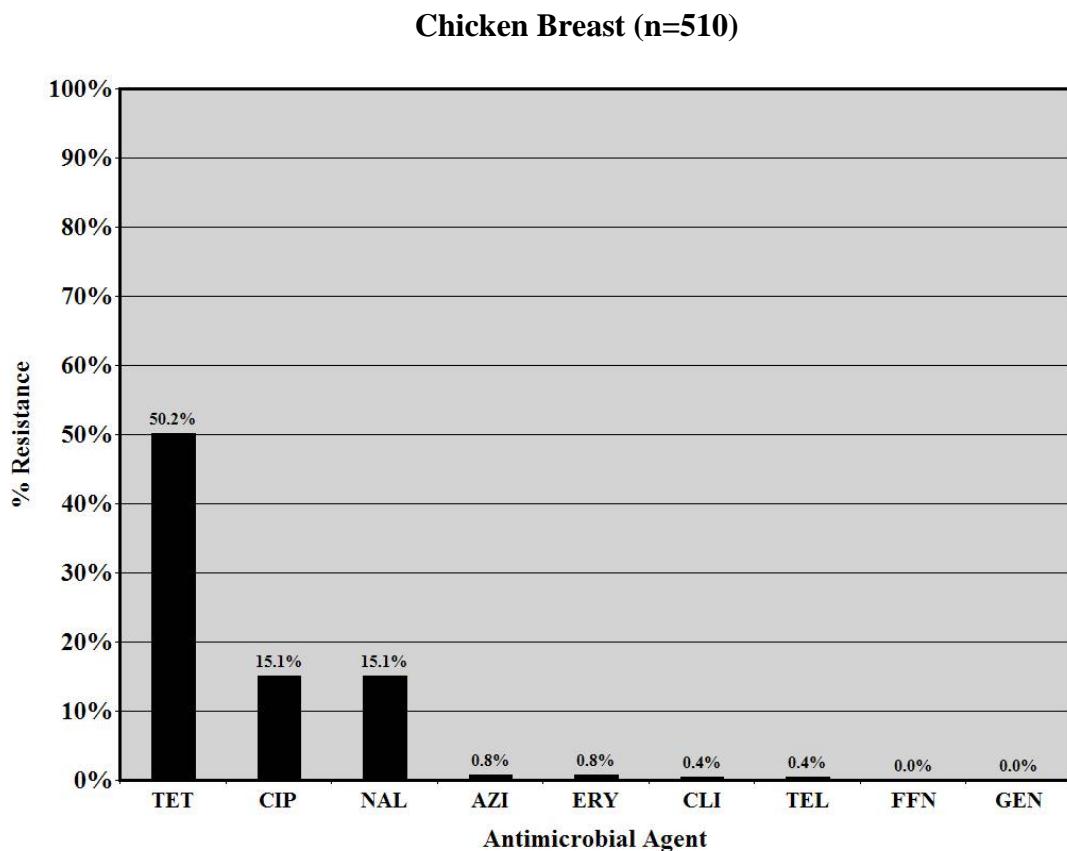


Figure A-6b. Antimicrobial Resistance among *Campylobacter coli* by Meat Type, 2004

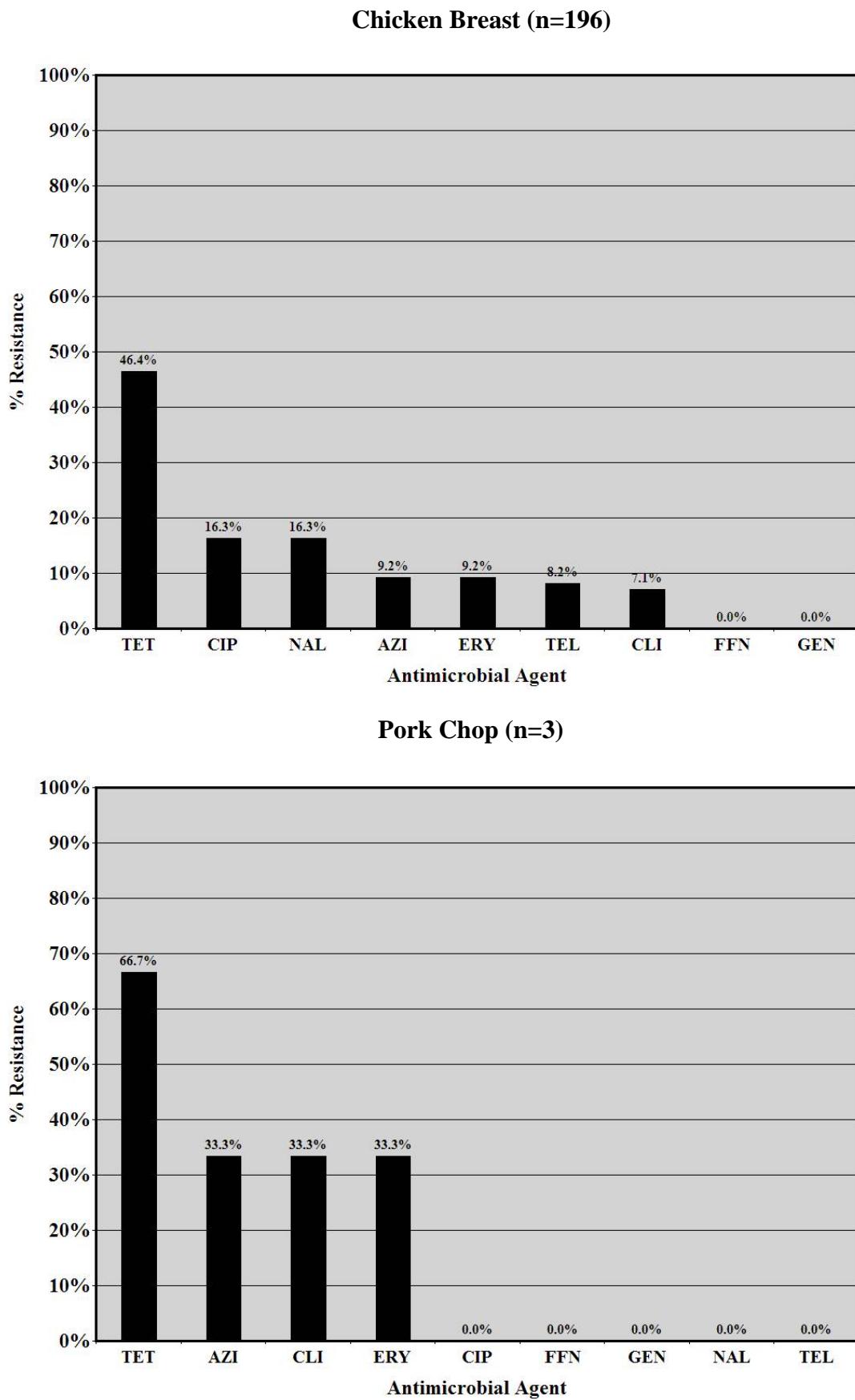
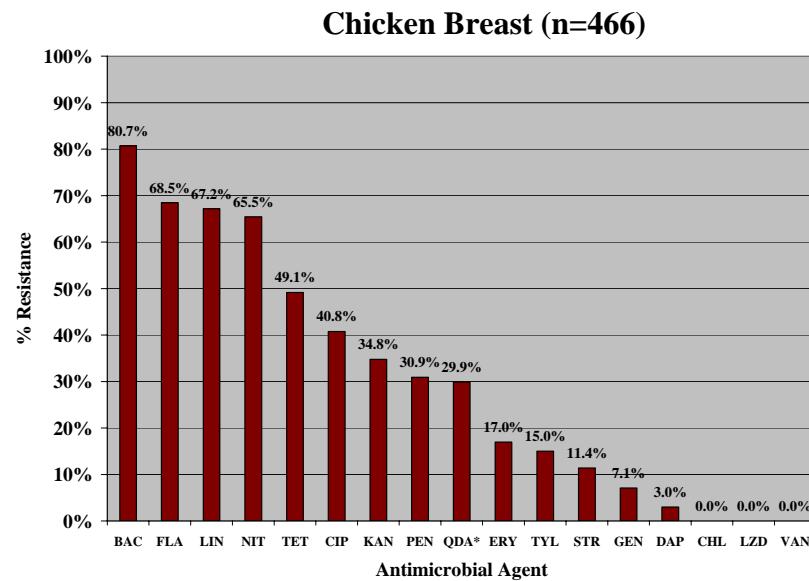
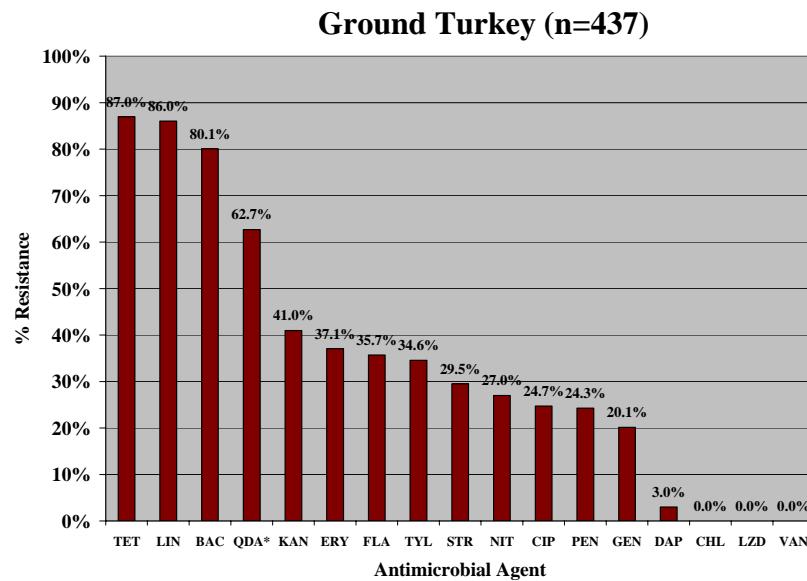


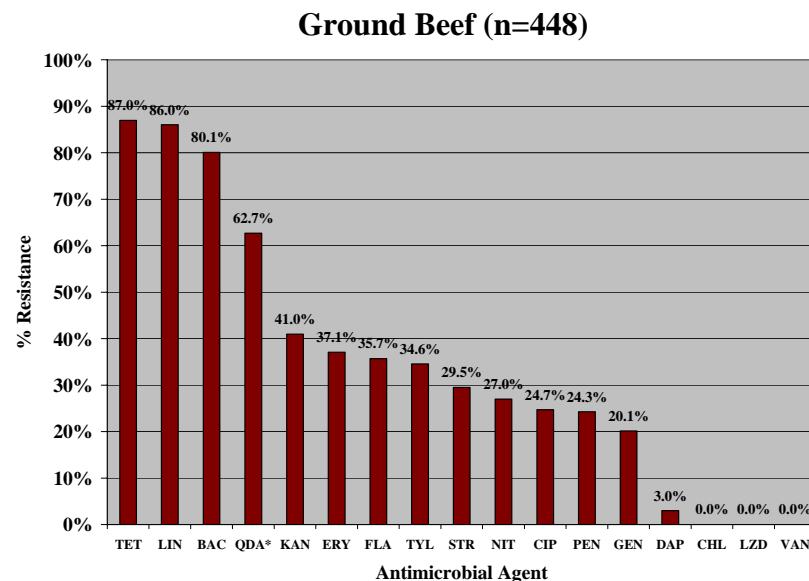
Figure A-7. Antimicrobial Resistance among *Enterococcus* by Meat Type, 2004



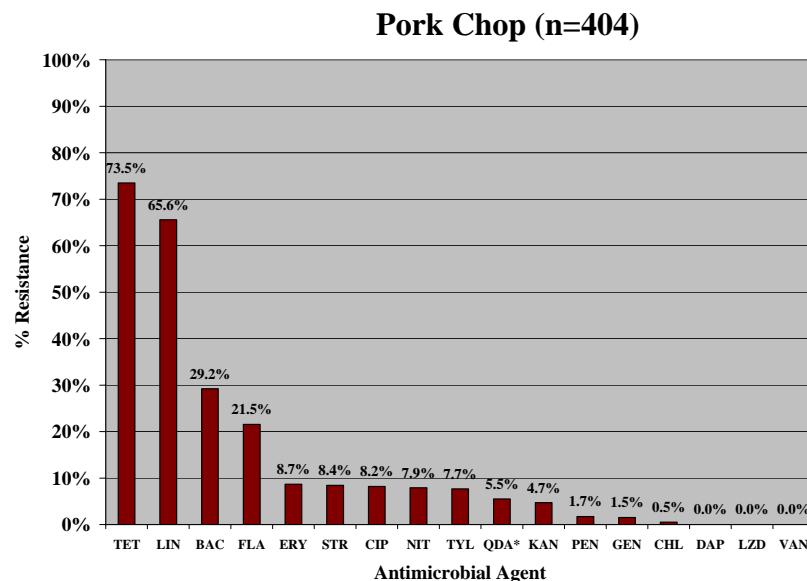
* Presented for all species except *E. faecalis* in QDA (n=466-88= 378 non *E. faecalis*)



* Presented for all species except *E. faecalis* in QDA (n=437-260= 177 non *E. faecalis*)



* Presented for all species except *E. faecalis* in QDA (n=448-194= 254 non *E. faecalis*)



*Presented for all species except *E. faecalis* in QDA (n=404-313= 91 non *E. faecalis*)

Figure A-7a. Antimicrobial Resistance among *Enterococcus faecium* by Meat Type, 2004

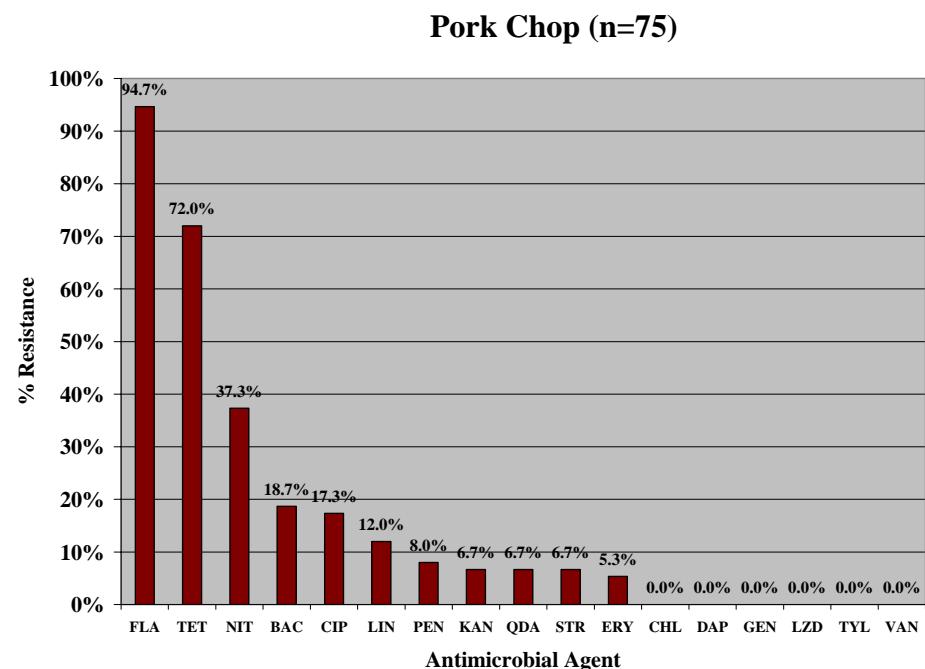
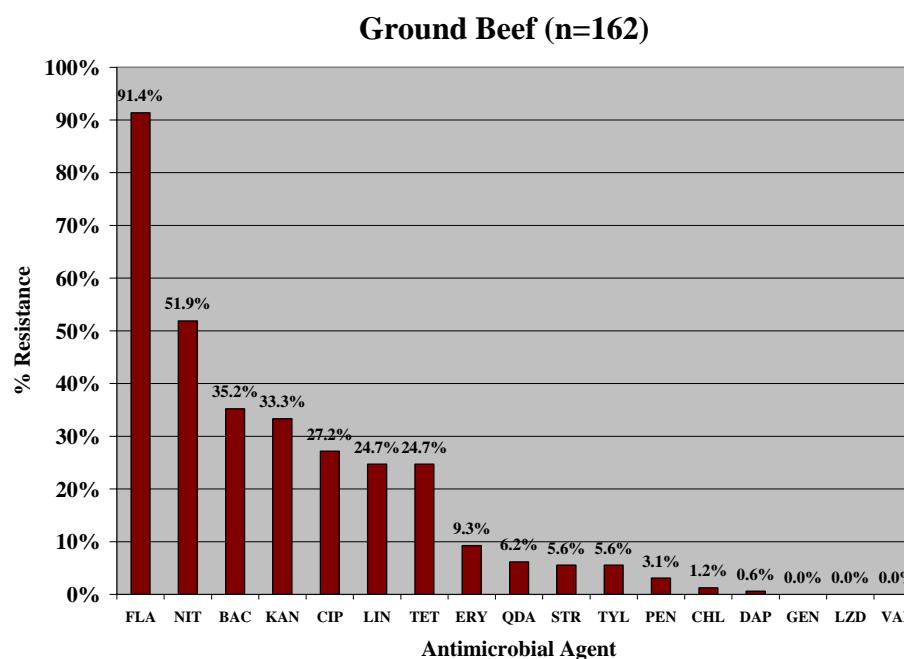
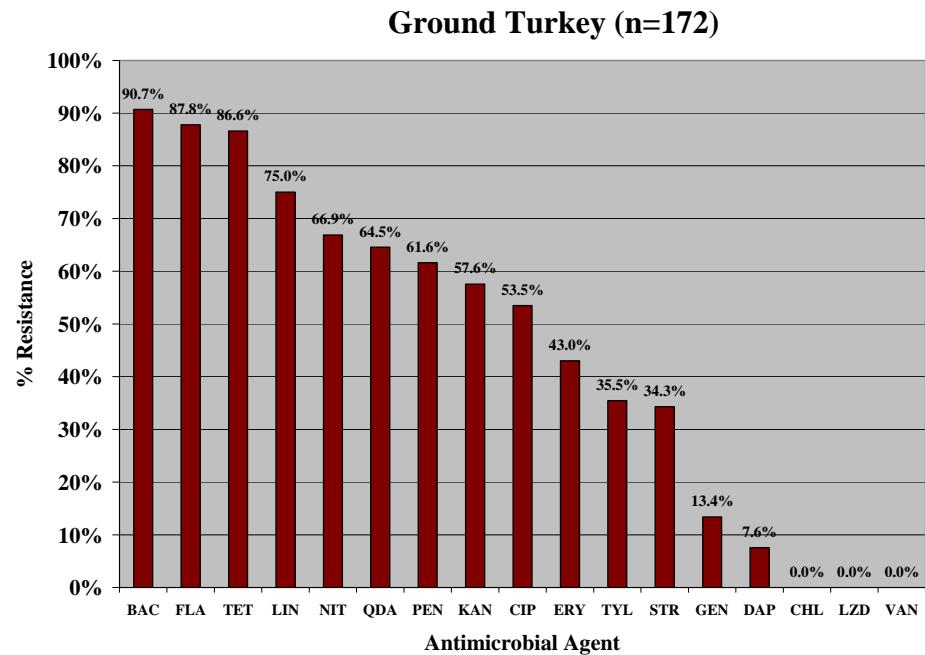
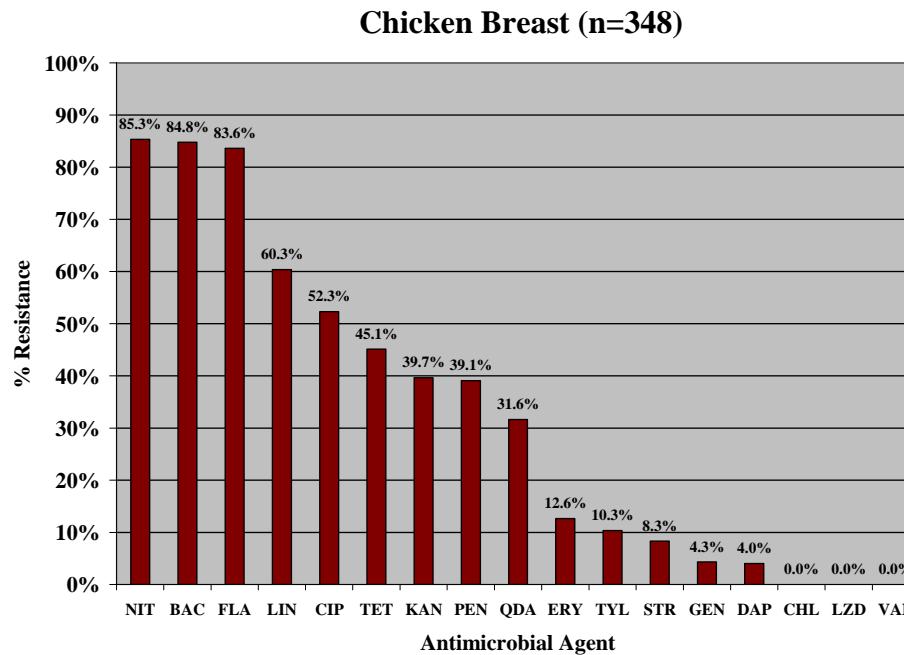
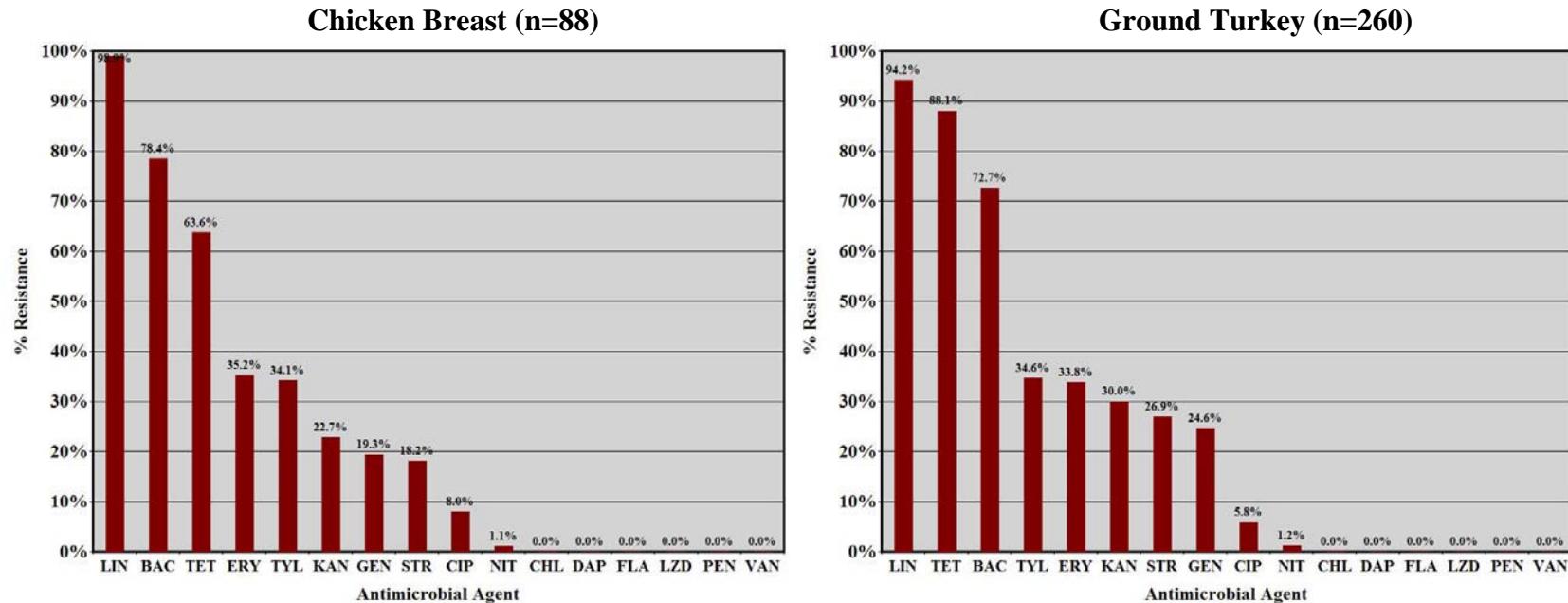
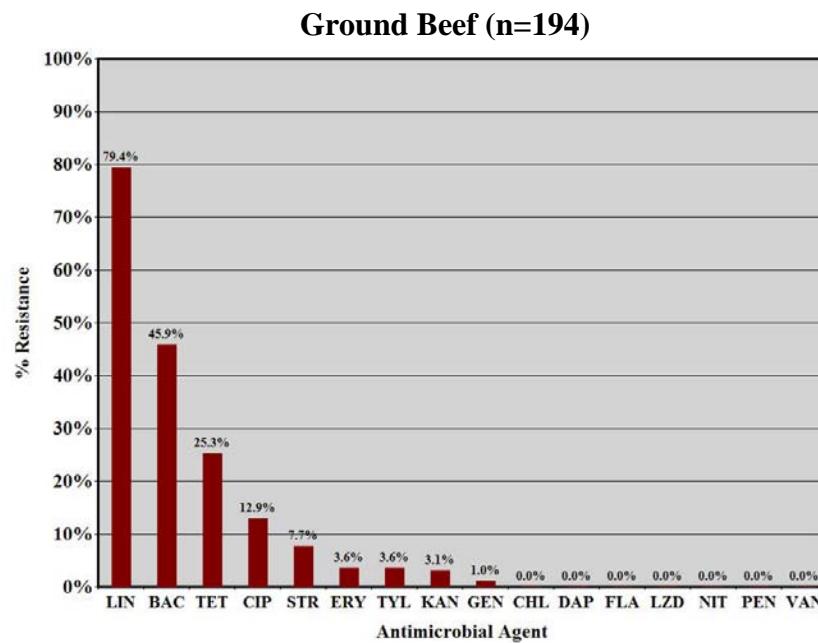


Figure A-7b. Antimicrobial Resistance among *Enterococcus faecalis by Meat Type, 2004**

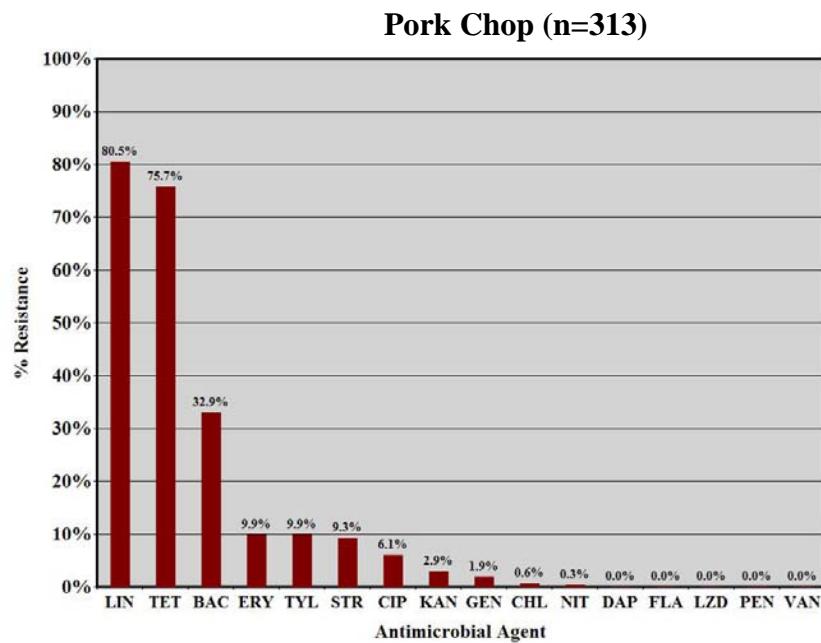


* Data does not include QDA, as *E. faecalis* is considered intrinsically resistant.

* Data does not include QDA, as *E. faecalis* is considered intrinsically resistant.

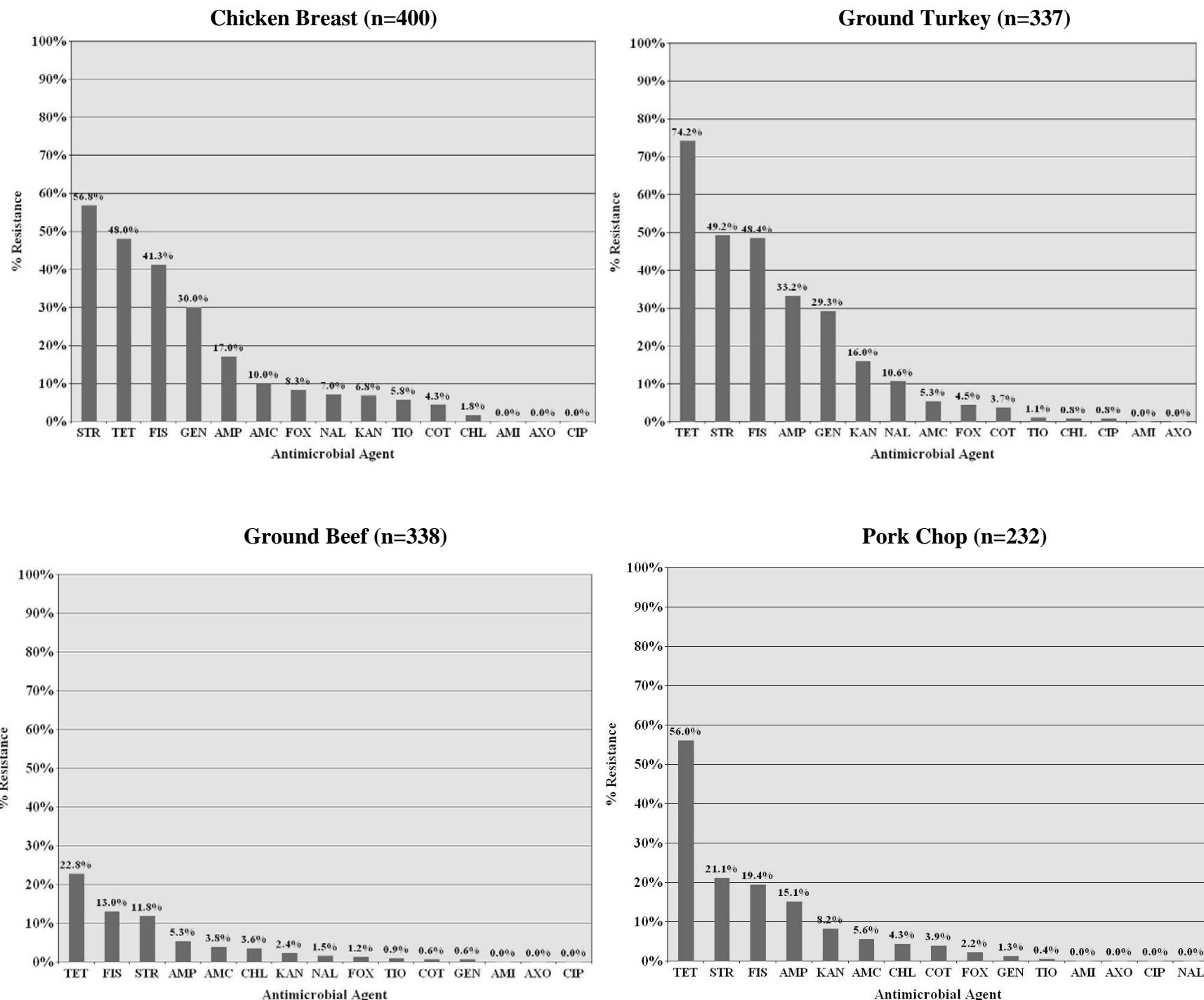


* Data does not include QDA, as *E. faecalis* is considered intrinsically resistant.



* Data does not include ODA, as *E. faecalis* is considered intrinsically resistant.

Figure A-8. Antimicrobial Resistance among *E. coli* by Meat Type, 2004



NATIONAL ANTIMICROBIAL RESISTANCE MONITORING SYSTEM – RETAIL FOOD STUDY ISOLATES MONTHLY LOG SHEET

STATE _____ MONTH _____ YEAR _____

Completed By (Initials): _____

Circle One → CHICKEN BREAST GROUND TURKEY GROUND BEEF PORK CHOP

PART I

	Sample ID Number	Store Name, City	Brand Name	Lot Number	Cut/Ground IN-STORE (\ One) Y N	Sell-by Date (M / D / Y)	Purchase Date (M / D / Y)	Lab Process Date (M / D / Y)
1								
2								
3								
4								
5								
6								
7								
8								
9								
10								

PART II

CONT. ↓	Growth (\ One) Y N	Salmonella		Species	Campylobacter		Growth (\ One) Y N	E. coli (GA, MD, TN, OR)		Growth (\ One) Y N	Enterococci (GA,MD,TN, OR)	
		Serotype	IF GROWTH		IF GROWTH	Isolate ID Number		IF GROWTH	Isolate ID Number		IF GROWTH	Isolate ID Number
1												
2												
3												
4												
5												
6												
7												
8												
9												
10												

Fax log sheet to CDC at 404-371-5444; send original log sheet with specimens to FDA-CVM and keep a copy for your records. Thank you.

NARMS Retail Meat, 2004

Experimental Design and Procedures:

Microbiological analysis:

In the laboratory, samples were refrigerated at 4°C and processed no later than 96 hours after purchase. After microbiological examination, recordings were made on the log sheets whether or not the meat and poultry samples were presumptively positive for *Salmonella*, *Campylobacter*, *E. coli*, and *Enterococcus*. Each laboratory used essentially the same procedure for sample collection. Retail meat and poultry packages were kept intact until they were aseptically opened in the laboratory at the start of examination. For chicken and pork samples, one piece of meat was examined, whereas, 25 g of ground product was examined for ground beef and ground turkey samples. The analytical portions from each sample were placed in separate sterile plastic bags, 250 mL of buffered peptone water was added to each bag, and the bags were vigorously shaken. Fifty mL of the rinsate from each sample was transferred to separate sterile flasks (or other suitable sterile containers) for isolation and identification of *Salmonella*, *Campylobacter*, *E. coli*, or *Enterococcus* using standard microbiological procedures. Once isolated and identified, bacterial isolates were sent to FDA's CVM Office of Research for further characterization including species confirmation, antimicrobial susceptibility testing and PFGE analysis (*Salmonella* and *Campylobacter* only).

Salmonella isolation:

Fifty mL of double strength lactose broth was added to each flask containing the 50 mL of rinsate to be used for *Salmonella* isolation. The contents were mixed thoroughly and incubated at 35°C for 24 hours. From each flask, 0.1 ml was then transferred to 9.9 mL tubes of RVR10 medium. The tubes of RVR10 medium were incubated in a water bath at 42°C for 16-20 hours before transferring one ml to pre-warmed (35-37°C) 10 mL tubes of M Broth. The inoculated M Broth tubes were incubated in a water bath at 35-37°C for 6-8 hours. From each M

Broth culture, one ml was heated at 100°C for 15 minutes, and the remaining portion was refrigerated. The heated portion from each culture was cooled to room temperature and tested using the TECRA *Salmonella* Visual Immunoassay kit (International BioProducts, Bothell, WA) or the VIDAS® *Salmonella* Immunoassay kit (bioMerieux, Hazelwood, MO) according to the manufacturers' instructions. If the TECRA or VIDAS assay was negative, the sample was considered negative for *Salmonella*. If the TECRA or VIDAS assay was positive, a loopful of the corresponding, unheated M Broth culture was streaked for isolation onto a XLD agar plate. The inoculated plate was incubated at 35°C for 24 hours. Each XLD agar plate was examined for typical *Salmonella* colonies (pink colonies with or without black centers). If no *Salmonella* like growth was observed on XLD agar, the sample was considered negative and the appropriate documentation was made on the log sheet accompanying the sample. When *Salmonella* like growth was observed, one well-isolated colony was streaked for isolation onto a trypticase soy agar plate supplemented with 5% defibrinated sheep blood (BAP). The BAP(s) were incubated at 35°C for 18-24 hours before sub-culturing an isolated colony for further biochemical identification and serotyping using the FoodNet laboratory's standard procedures. *Salmonella* isolates were subsequently frozen at -60 to -80°C in Brucella broth with 20% glycerol and shipped in cryo-vials on dry ice to FDA-CVM. Upon arrival at CVM, every isolate was streaked for purity on a BAP before being confirmed as *Salmonella* using the Vitek microbial identification system (bioMérieux, Hazelwood, MO). These isolates were further serotyped for O and H antigens using either commercially available (Difco-Becton Dickinson, Sparks, MD) or CDC antisera.

Campylobacter isolation:

Fifty mL of double strength Bolton broth was added to each flask containing the 50 mL of rinsate to be used for *Campylobacter* isolation. The broth and rinsate were mixed thoroughly, but gently to avoid aeration, and incubated at 42°C for 24 hours in a reduced oxygen atmosphere

that was obtained using a commercial gas generating envelope or a gas mixture containing 85% nitrogen, 10% carbon dioxide, and 5% oxygen. Using a swab, the first quadrant of a CCA Plate was inoculated with the incubated Bolton broth culture. The remainder of each plate was then streaked with a loop to obtain isolated colonies, and the CCA plates were incubated at 42°C in the above atmosphere for 24 to 48 hours. Each CCA plate was examined for typical *Campylobacter* colonies (round to irregular with smooth edges; thick translucent white growth to spreading, film-like transparent growth). If no *Campylobacter* like growth was observed on a CCA plate, the sample was considered negative and the appropriate documentation was made on the log sheet accompanying the sample. When *Campylobacter* like growth was observed, one typical well-isolated *Campylobacter* like colony from each positive CCA plate was sub-cultured to a BAP and incubated as described for the CCA plates. Following incubation, one typical well-isolated *Campylobacter* like colony was gram stained and tested using a smear catalase, oxidase, hippurate and/or motility test. If the Gram stain showed small, Gram- negative, curved rods, and the isolate was positive with the other test(s) that were conducted, a sample was considered presumptively positive for *Campylobacter*. If the CCA plates or BAPs had no typical colonies or isolate testing was inconsistent with *Campylobacter*, a sample was considered negative. All isolates presumptively identified as *Campylobacter* were frozen at -60 to -80°C in Brucella broth with 20% glycerol and shipped in cryo-vials on dry ice to FDA-CVM. Upon arrival at CVM, isolates were twice streaked for purity on a BAP before being identified to the species level using PCR assays previously described (2, 6).

E. coli isolation (Georgia, Maryland, Oregon and Tennessee)

Fifty mL of double strength MacConkey broth was added to each flask containing the 50 mL of rinsate to be used for *E. coli* isolation. The contents were mixed thoroughly and incubated at 35°C for 24 hours. One loopful from each flask was then transferred to an EMB agar plate and streaked for isolation. Agar plates were then incubated at 35°C for 24 hours in ambient air

and examined for typical *E. coli* colonies (colonies having a dark center and usually a green metallic sheen). If no typical growth was observed on an EMB agar plate, the sample was considered negative and the appropriate documentation was made on the log sheet accompanying the sample. When *E. coli*-like growth was present, one typical, well-isolated colony was streaked for isolation onto a BAP. The BAPs were incubated at 35°C for 24 hours in ambient air and examined for purity. One typical, well-isolated colony was subcultured for indole and oxidase tests. Indole positive and oxidase negative isolates were considered presumptively positive as *E. coli*. Presumptive *E. coli* isolates were subsequently frozen at -60 to -80°C in Brucella broth with 20% glycerol and shipped in cryo-vials on dry ice to FDA-CVM. Upon arrival at CVM, every isolate was streaked for purity on a BAP before being confirmed as *E. coli* using the Vitek microbial identification system (bioMérieux, Hazelwood, MO).

Enterococcus isolation (Georgia, Maryland, Oregon and Tennessee)

Fifty mL of double strength Enterococcosel broth was added to each flask containing the 50 ml of rinsate to be used for *Enterococcus* isolation. The contents were mixed thoroughly and incubated at 45°C for 24 hours in ambient air. If no typical growth or blackening was observed in the flask, the sample was considered negative and the appropriate documentation was made on the log sheet accompanying the sample. If blackening of the broth was observed, a loopful was streaked for isolation onto an EA plate. The plates were then incubated at 35°C for 24 hours in ambient air and examined for enterococcal-like colonies (small colonies surrounded by a blackening of the agar). If no typical growth was observed on the EA plate, the sample was considered negative and the appropriate documentation was made on the log sheet accompanying the sample. If enterococcal-like growth was present, one well-isolated colony was streaked for isolation onto a BAP, and incubated at 35°C for 24 hours in ambient air. Presumptive *Enterococcus* isolates were subsequently frozen at -60 to -80°C in Brucella broth with 20% glycerol and shipped in cryo-vials on dry ice to FDA-CVM. Upon arrival at CVM,

every isolate was streaked for purity on a BAP before being confirmed as *Enterococcus* using the Vitek microbial identification system (bioMérieux, Hazelwood, MO).

Antimicrobial Susceptibility Testing:

Antimicrobial MICs were determined using a 96 well broth microdilution method (Sensititre, Trek Diagnostic Systems, Westlake, OH) according to CLSI standards (3, 4, 5). *Salmonella* and *E. coli* isolates were tested using a custom plate developed for Gram negative bacteria, catalog # CMV1AGNF; *Enterococcus* isolates were tested using a custom plate developed for Gram positive bacteria, catalog # CMV1AGPF; and *Campylobacter* isolates were tested using a custom plate developed for *Campylobacter*, catalog # CAMPY ([Table 1](#)). CLSI recommended QC organisms were used each time that antimicrobial susceptibility testing was performed. The QC organisms included *Escherichia coli* ATCC 25922, *Enterococcus faecalis* ATCC 29212, *Staphylococcus aureus* ATCC 29213, *Pseudomonas aeruginosa* ATCC 27853, and *Campylobacter jejuni* ATCC 33560 (3, 4, 5).

CLSI approved interpretive criteria were used when available; otherwise tentative NARMS breakpoints were used ([Table 1](#)). All antimicrobial susceptibility testing was conducted in the laboratories of the Division of Animal and Food Microbiology, CVM-FDA, Laurel, MD.

Pulsed Field Gel Electrophoresis (PFGE):

Pulsed-field gel electrophoresis was used to assess genetic relatedness among *Salmonella* and *Campylobacter* isolates. The PFGE was performed according to protocols developed by CDC (1). Agarose-embedded DNA was digested with the enzyme *Xba*I for *Salmonella* isolates and *Sma*II for *Campylobacter* isolates. DNA restriction fragments were separated by electrophoresis using a Chef Mapper electrophoresis system (Bio-Rad, Hercules, CA). Genomic-DNA profiles or “fingerprints” were analyzed using BioNumerics software (Applied-Maths, Kortrijk, Belgium), and banding patterns were compared using Dice coefficients with a

1.5% band position tolerance. PFGE analysis was conducted in the laboratories of the Division of Animal and Food Microbiology, CVM-FDA, Laurel, MD.

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